# Survey of SARS-CoV, main protease assays for creating machine-learning models

Goal is to identify CoV related biochemical assay datasets. Other sources of data from NCATS, PubChem Bioassays, CheMBL had been identified and are being reviewed by ATOM team.

Identification of SARS-CoV-2 3CL Protease Inhibitors by a Quantitative High-throughput Screening

Wei Zhu, Miao Xu, Catherine Z. Chen, Hui Guo, Min Shen, Xin Hu, Paul Shinn, Carleen Klumpp-Thomas, Samuel G. Michael, Wei Zheng bioRxiv 2020.07.17.207019; doi: https://doi.org/10.1101/2020.07.17.207019 Remove from Selected Citations

A Fluorescence-based High Throughput-Screening assay for the SARS-CoV RNA synthesis complex

Cecilia Eydoux, Veronique Fattorini, Ashleigh Shannon, Thi-Tuyet-Nhung Le, Bruno Didier, Bruno Canard, Jean-Claude Guillemot bioRxiv 2020.07.07.192005; doi: https://doi.org/10.1101/2020.07.07.192005 Remove from Selected Citations

• Structure of papain-like protease from SARS-CoV-2 and its complexes with non-covalent inhibitors

Jerzy Osipiuk, Saara-Anne Azizi, Steve Dvorkin, Michael Endres, Robert Jedrzejczak, Krysten A. Jones, Soowon Kang, Rahul S. Kathayat, Youngchang Kim, Vladislav G. Lisnyak, Samantha L. Maki, Vlad Nicolaescu, Cooper A. Taylor, Christine Tesar, Yu-An Zhang, Zhiyao Zhou, Glenn Randall, Karolina Michalska, Scott A. Snyder, Bryan C. Dickinson, Andrzej Joachimiak bioRxiv 2020.08.06.240192; doi: https://doi.org/10.1101/2020.08.06.240192 Remove from Selected Citations

• Lead compounds for the development of SARS-CoV-2 3CL protease inhibitors

Sho Iketani, Farhad Forouhar, Hengrui Liu, Seo Jung Hong, Fang-Yu Lin, Manoj S. Nair, Arie Zask, Yaoxing Huang, Li Xing, Brent R. Stockwell, Alejandro Chavez, David D. Ho bioRxiv 2020.08.03.235291; doi: https://doi.org/10.1101/2020.08.03.235291 Remove from Selected Citations

Crystallographic and electrophilic fragment screening of the SARS-CoV-2 main protease

Alice Douangamath, Daren Fearon, Paul Gehrtz, Tobias Krojer, Petra Lukacik, C.
David Owen, Efrat Resnick, Claire Strain-Damerell, Anthony Aimon, Péter Ábrányi-Balogh, José Brandaő-Neto, Anna Carbery, Gemma Davison, Alexandre Dias, Thomas
D Downes, Louise Dunnett, Michael Fairhead, James D. Firth, S. Paul Jones, Aaron Keely, György
M. Keserü, Hanna F Klein, Mathew P. Martin, Martin E.
M. Noble, Peter O'Brien, Ailsa Powell, Rambabu Reddi, Rachael Skyner, Matthew Snee, Michael
J. Waring, Conor Wild, Nir London, Frank von Delft, Martin A. Walsh

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A drug repurposing screen identifies hepatitis C antivirals as inhibitors of the SARS-CoV-2 main protease

Jeremy D. Baker, Rikki L. Uhrich, Gerald C. Kraemer, Jason E. Love, Brian C. Kraemer bioRxiv 2020.07.10.197889; doi: https://doi.org/10.1101/2020.07.10.197889
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Development of a fluorescence based, high-throughput SARS-CoV-2 3CLpro reporter assay

Heather M. Froggatt, Brook E. Heaton, Nicholas S. Heaton bioRxiv 2020.06.24.169565; doi: https://doi.org/10.1101/2020.06.24.169565 Remove from Selected Citations

• Feline coronavirus drug inhibits the main protease of SARS-CoV-2 and blocks virus replication

Wayne Vuong, Muhammad

Bashir Khan, Conrad Fischer, Elena Arutyunova, Tess Lamer, Justin Shields, Holly A. Saffran, Ryan T. McKay, Marco J. van Belkum, Michael Joyce, Howard S. Young, D. Lorne Tyrrell, John C. Vederas, M. Joanne Lemieux

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• Inhibition of Severe Acute Respiratory Syndrome Coronavirus 2 main protease by tafenoquine in vitro

Yeh Chen, Wen-Hao Yang, Li-Min Huang, Yu-Chuan Wang, Chia-Shin Yang, Yi-Liang Liu, Mei-Hui Hou, Chia-Ling Tsai, Yi-Zhen Chou, Bao-Yue Huang, Chian-Fang Hung, Yu-Lin Hung, Jin-Shing Chen, Yu-Ping Chiang, Der-Yang Cho, Long-Bin Jeng, Chang-Hai Tsai, Mien-Chie Hung bioRxiv 2020.08.14.250258; doi: https://doi.org/10.1101/2020.08.14.250258

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A simple protein-based SARS-CoV-2 surrogate neutralization assay

Kento T. Abe, Zhijie Li, Reuben Samson, Payman Samavarchi-Tehrani, Emelissa

J. Valcourt, Heidi Wood, Patrick Budylowski, Alan Dupuis, Roxie

C. Girardin, Bhavisha Rathod, Karen Colwill, Allison J McGeer, Samira Mubareka, Jennifer

L. Gommerman, Yves Durocher, Mario Ostrowski, Kathleen A. McDonough, Michael A. Drebot, Steven

J. Drews, James M. Rini, Anne-Claude Gingras

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Machine Learning Models Identify Inhibitors of SARS-CoV-2

Victor O. Gawriljuk, Phyo Phyo Kyaw Zin, Daniel

H. Foil, Jean Bernatchez, Sungjun Beck, Nathan Beutler, James Ricketts, Linlin Yang, Thomas Rogers, Ana

C. Puhl, Kimberley M. Zorn, Thomas R. Lane, Andre S. Godoy, Glaucius Oliva, Jair L. Siqueira-Neto, Peter B. Madrid, Sean Ekins

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 Development and validation of IMMUNO-COV™: a high-throughput clinical assay for detecting antibodies that neutralize SARS-CoV-2

Rianna Vandergaast, Timothy Carey, Samantha Reiter, Patrycja Lech, Clement Gnanadurai, Mulu Tesfay, J ason Buehler, Lukkana Suksanpaisan, Shruthi Naik, Bethany Brunton, Jordan Recker, Michelle Haselton, Christopher Ziegler, Anne Roesler, John R. Mills, Elitza Theel, Scott C. Weaver, Grace Rafael, Matthew M. Roforth, Calvin Jerde, Sheryl Tran, Rosa Maria Diaz, Alice Bexon, Alina Baum, Christos A. Kyratsous, Kah Whye Peng, Stephen J. Russell bioRxiv 2020.05.26.117549; doi: https://doi.org/10.1101/2020.05.26.117549

 An Enzymatic TMPRSS2 Assay for Assessment of Clinical Candidates and Discovery of Inhibitors as Potential Treatment of COVID-19

Jonathan H. Shrimp, Stephen C. Kales, Philip E. Sanderson, Anton Simeonov, Min Shen, Matthew D. Hall bioRxiv 2020.06.23.167544; doi: https://doi.org/10.1101/2020.06.23.167544

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 Identifying SARS-CoV-2 entry inhibitors through drug repurposing screens of SARS- S and MERS-S pseudotyped particles

Catherine Z. Chen, Miao Xu, Manisha Pradhan, Kirill Gorshkov, Jennifer Petersen, Marco R. Straus, Wei Zhu, Paul Shinn, Hui Guo, Min Shen, Carleen Klumpp-Thomas, Samuel G. Michael, Joshua Zimmerberg, Wei Zheng, Gary R Whittaker bioRxiv 2020.07.10.197988; doi: https://doi.org/10.1101/2020.07.10.197988 Remove from Selected Citations

• In vitro screening of a FDA approved chemical library reveals potential inhibitors of SARS-CoV-2 replication

Franck Touret, Magali Gilles, Karine Barral, Antoine Nougairède, Etienne Decroly, Xavier de Lamballerie, Bruno Coutard bioRxiv 2020.04.03.023846; doi: https://doi.org/10.1101/2020.04.03.023846 Remove from Selected Citations

Discovery of COVID-19 Inhibitors Targeting the SARS-CoV2 Nsp13 Helicase

Mark Andrew White, Wei Lin, Xiaodong Cheng bioRxiv 2020.08.09.243246; doi: https://doi.org/10.1101/2020.08.09.243246 Remove from Selected Citations • Rapid in vitro assays for screening neutralizing antibodies and antivirals against SARS-CoV-2

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Swab-Seq: A high-throughput platform for massively scaled up SARS-CoV-2 testing

Joshua S. Bloom, Eric M. Jones, Molly Gasperini, Nathan B. Lubock, Laila Sathe, Chetan Munugala, A. Sina Booeshaghi, Oliver F. Brandenberg, Longhua Guo, Scott

W. Simpkins, Isabella Lin, Nathan LaPierre, Duke Hong, Yi Zhang, Gabriel Oland, Bianca Judy Choe, Sukantha Chandrasekaran, Evann E. Hilt, Manish J. Butte, Robert Damoiseaux, Aaron R. Cooper, Yi Yin, Lior Pachter, Omai

B. Garner, Jonathan Flint, Eleazar Eskin, Chongyuan Luo, Sriram Kosuri, Leonid Kruglyak, Valerie A. Arboleda

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 Neutralizing antibody against SARS-CoV-2 spike in COVID-19 patients, health care workers and convalescent plasma donors: a cohort study using a rapid and sensitive high-throughput neutralization assay

Cong Zeng, John P Evans, Rebecca Pearson, Panke Qu, Yi-Min Zheng, Richard T Robinson, Luanne Hall-Stoodley, Jacob Yount, Sonal Pannu, Rama K Mallampalli, Linda Saif, Eugene Oltz, Gerard Lozanski, Shan-Lu Liu

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A facile Q-RT-PCR assay for monitoring SARS-CoV-2 growth in cell culture

Christian Shema Mugisha, Hung R. Vuong, Maritza Puray-Chavez, Sebla B. Kutluay bioRxiv 2020.06.26.174698; doi: https://doi.org/10.1101/2020.06.26.174698 Remove from Selected Citations

An enzyme-based immunodetection assay to quantify SARS-CoV-2 infection

Carina Conzelmann, Andrea Gilg, Rüdiger Groß, Desirée Schütz, Nico Preising, Ludger Ständker, Bernd Jahrsdörfer, Hubert Schrezenmeier, Konstantin M. J. Sparrer, Thomas Stamminger, Steffen Stenger, Jan Münch, Janis A. Müller bioRxiv 2020.06.14.150862; doi: https://doi.org/10.1101/2020.06.14.150862

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• The discovery of potential natural products for targeting SARS-CoV-2 spike protein by virtual screening

Guan-Yu Chen, Tsung-You Yao, Azaj Ahmed, Yi-Cheng Pan, Juan-Cheng Yang, Yang-Chang Wu bioRxiv 2020.06.25.170639; doi: https://doi.org/10.1101/2020.06.25.170639

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• Virus-free and live-cell visualizing SARS-CoV-2 cell entry for studies of neutralizing antibodies and compound inhibitors

Yali Zhang, Shaojuan Wang, Yangtao Wu, Wangheng Hou, Lunzhi Yuan, Chenguang Sheng, Juan Wang, Jia nghui Ye, Qingbing Zheng, Jian Ma, Jingjing Xu, Min Wei, Zonglin Li, Sheng Nian, Hualong Xiong, Liang Zhang, Yang Shi, Baorong Fu, Jiali Cao, Chuanlai Yang, Zhiyong Li, Ting Yang, Lei Liu, Hai Yu, Jianda Hu, Shen gxiang Ge, Yixin Chen, Tianying Zhang, Jun Zhang, Tong Cheng, Quan Yuan, Ningshao Xia bioRxiv 2020.07.22.215236; doi: https://doi.org/10.1101/2020.07.22.215236

 Discovery of Clioquinol and Analogues as Novel Inhibitors of Severe Acute Respiratory Syndrome Coronavirus 2 Infection, ACE2 and ACE2 - Spike Protein Interaction In Vitro

Omonike A. Olaleye, Manvir Kaur, Collins Onyenaka, Tolu Adebusuyi bioRxiv 2020.08.14.250480; doi: https://doi.org/10.1101/2020.08.14.250480 Remove from Selected Citations

 Rapid assessment of ligand binding to the SARS-CoV-2 main protease by saturation transfer difference NMR spectroscopy

Anastassia L. Kantsadi, Ioannis Vakonakis bioRxiv 2020.06.17.156679; doi: https://doi.org/10.1101/2020.06.17.156679 Remove from Selected Citations

• Single-cell screening of SARS-CoV-2 target cells in pets, livestock, poultry and wildlife

Dongsheng Chen, Jian Sun, Jiacheng Zhu, Xiangning Ding, Tianming Lan, Linnan Zhu, Rong Xiang, Peiwen Ding, Haoyu Wang, Xiaoling Wang, Weiying Wu, Jiaying Qiu, Shiyou Wang, Haimeng Li, Fuyu An, Heng Ba o, Le Zhang, Lei Han, Yixin Zhu, Xiran Wang, Feiyue Wang, Yuting Yuan, Wendi Wu, Chengcheng Sun, Hao rong Lu, Jihong Wu, Xinghuai Sun, Shenghai Zhang, Sunil

Kumar Sahu, Haixia Chen, Dongming Fang, Lihua Luo, Yuying Zeng, Yiquan Wu, ZeHua Cui, Qian He, Sanji e Jiang, Xiaoyan Ma, Weimin Feng, Yan Xu, Fang Li, Zhongmin Liu, Lei Chen, Fang Chen, Xin Jin, Wei Qiu, Huanming Yang, Jian Wang, Yan Hua, Yahong Liu, Huan Liu, Xun Xu bioRxiv 2020.06.13.149690; doi: https://doi.org/10.1101/2020.06.13.149690

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 Antiviral effects of miRNAs in extracellular vesicles against severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and mutations in SARS-CoV-2 RNA virus

Jae Hyun Park, Yuri Choi, Chul-Woo Lim, Ji-Min Park, Shin-Hye Yu, Yujin Kim, Hae Jung Han, Chun-Hyung Kim, Young-Sook Song, Chul Kim, Jisook Moon bioRxiv 2020.07.27.190561; doi: https://doi.org/10.1101/2020.07.27.190561

 A nanoluciferase SARS-CoV-2 for rapid neutralization testing and screening of anti-infective drugs for COVID-19

Xuping Xie, Antonio E. Muruato, Xianwen Zhang, Kumari G. Lokugamage, Camila R. Fontes-Garfias, Jing Zou, Jianying Liu, Ping Ren, Mini Balakrishnan, Tomas Cihlar, Chien-Te K. Tseng, Shinji Makino, Vineet D. Menachery, John P. Bilello, Pei-Yong Shi bioRxiv 2020.06.22.165712; doi: https://doi.org/10.1101/2020.06.22.165712

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 High-Throughput Human Primary Cell-Based Airway Model for Evaluating Influenza, Coronavirus, or other Respiratory Viruses in vitro

A.L. Gard, R. Maloney, B.P. Cain, C.R. Miller, R.J. Luu, J.R. Coppeta, P. Liu, J.P. Wang, H. Azizgolshani, R.F. Fe zzie, J.L. Balestrini, B.C. Isenberg, R.W. Finberg, J.T. Borenstein bioRxiv 2020.05.23.112797; doi: https://doi.org/10.1101/2020.05.23.112797 Remove from Selected Citations

A CRISPR-based SARS-CoV-2 diagnostic assay that is robust against viral evolution and RNA editing

Kean Hean Ooi, Jie Wen Douglas Tay, Seok Yee Teo, Mengying Mandy Liu, Pornchai Kaewsapsak, Shengyang Jin, Yong-Gui Gao, Meng How Tan bioRxiv 2020.07.03.185850; doi: https://doi.org/10.1101/2020.07.03.185850 Remove from Selected Citations

 Quantifying absolute neutralization titers against SARS-CoV-2 by a standardized virus neutralization assay allows for cross-cohort comparisons of COVID-19 sera

Kasopefoluwa Y. Oguntuyo, Christian S Stevens, Chuan-Tien Hung, Satoshi Ikegame, Joshua A. Acklin, Shreyas S Kowdle, Jillian C. Carmichael, Hsin-ping Chiu, Kristopher D. Azarm, Griffin D. Haas, Fatima Amanat, Jeromine Klingler, Ian Baine, Suzanne Arinsburg, Juan C. Bandres, Mohammed NA Siddiquey, Robert M. Schilke, Matthew D. Woolard, Hongbo Zhang, COVIDAR Argentina Consortium, Andrew J. Duty, Thomas A. Kraus, Thomas M. Moran, Domenico Tortorella, Jean K. Lim, Andrea V. Gamarnik, Catarina E. Hioe, Susan Zolla-Pazner, Stanimir S. Ivanov, Jeremy P. Kamil, Florian Krammer, Benhur Lee medRxiv 2020.08.13.20157222; doi: https://doi.org/10.1101/2020.08.13.20157222 Remove from Selected Citations

 Catalytic cleavage of HEAT and subsequent covalent binding of the tetralone moiety by the SARS-CoV-2 main protease

Sebastian Günther, Patrick Y.

A. Reinke, Dominik Oberthuer, Oleksandr Yefanov, Helen Ginn, Susanne Meier, Thomas J. Lane, Kristina Lorenzen, Luca Gelisio, Wolfgang Brehm, Illona Dunkel, Martin Domaracky, Sofiane Saou ane, Julia Lieske, Christiane Ehrt, Faisal Koua, Alexandra Tolstikova, Thomas

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Rahmani Mashhour, Filip Guicking, Vincent Hennicke, Pontus Fischer, Cromarte Rogers, Diana C.

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Christian Schulz, Pedram Mehrabi, Christina Schmidt, Robin Schubert, Huijong Han, Boris Krichel, Yaiza Fernández-García, Beatriz Escudero-

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• SARS-coronavirus-2 replication in Vero E6 cells: replication kinetics, rapid adaptation and cytopathology

Natacha S. Ogando, Tim J. Dalebout, Jessika C. Zevenhoven-Dobbe, Ronald W. Limpens, Yvonne van der Meer, Leon Caly, Julian Druce, Jutte J. C. de

Vries, Marjolein Kikkert, Montserrat Bárcena, Igor Sidorov, Eric J. Snijder bioRxiv 2020.04.20.049924; doi: https://doi.org/10.1101/2020.04.20.049924 Remove from Selected Citations

• Scutellaria baicalensis extract and baicalein inhibit replication of SARS-CoV-2 and its 3C-like protease in vitro

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 A SARS-CoV-2 serological assay to determine the presence of blocking antibodies that compete for human ACE2 binding

James R. Byrnes, Xin X. Zhou, Irene Lui, Susanna K. Elledge, Jeff E. Glasgow, Shion A. Lim, Rita Loudermilk, Charles Y. Chiu, Michael R. Wilson, Kevin K. Leung, James A. Wells medRxiv 2020.05.27.20114652; doi: https://doi.org/10.1101/2020.05.27.20114652 Remove from Selected Citations

 Development of a simple in vitro assay to identify and evaluate nucleotide analogs against SARS-CoV-2 RNAdependent RNA polymerase

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 Robust neutralization assay based on SARS-CoV-2 S-bearing vesicular stomatitis virus (VSV) pseudovirus and ACE2-overexpressed BHK21 cells

Hua-Long Xiong, Yang-Tao Wu, Jia-Li Cao, Ren Yang, Jian Ma, Xiao-Yang Qiao, Xiang-Yang Yao, Bao-Hui Zhang, Ya-Li Zhang, Wang-Heng Hou, Yang-Shi, Jing-Jing Xu, Liang-Zhang, Shao-Juan Wang, Bao-Rong Fu, Ting Yang, Sheng-Xiang Ge, Jun Zhang, Quan Yuan, Bao-Ying Huang, Zhi-Yong Li, Tian-Ying Zhang, Ning-Shao Xia

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Shotgun Transcriptome and Isothermal Profiling of SARS-CoV-2 Infection Reveals Unique Host Responses,
 Viral Diversification, and Drug Interactions

#### Daniel

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A. Ivanov, Maria Sierra, Diana Pohle, Michael Zietz, Undina Gisladottir, Vijendra Ramlall, Craig

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J. Shemesh, Jenny Xiang, Jean Thierry-Mieg, Danielle Thierry-Mieg, Robert

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 Probing the dynamic structure-function and structure-free energy relationships of the corona virus main protease with Biodynamics theory

Hongbin Wan, Vibhas Aravamuthan, Robert A. Pearlstein bioRxiv 2020.07.19.211185; doi: https://doi.org/10.1101/2020.07.19.211185 Remove from Selected Citations

Physiologic RNA Targets and Refined Sequence Specificity of Coronavirus EndoU

Rachel Ancar, Yize Li, Eveline Kindler, Daphne A. Cooper, Monica Ransom, Volker Thiel, Susan R. Weiss, Jay R. Hesselberth, David J. Barton bioRxiv 2020.05.20.064436; doi: https://doi.org/10.1101/2020.05.20.064436 Remove from Selected Citations

Network-based Drug Repurposing for Human Coronavirus

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Roberto Benoni, Petra Krafcikova, Marek R. Baranowski, Joanna Kowalska, Evzen Boura, Hana Cahová bioRxiv 2020.07.30.228478; doi: https://doi.org/10.1101/2020.07.30.228478 Remove from Selected Citations

Multi-level proteomics reveals host-perturbation strategies of SARS-CoV-2 and SARS-CoV

Alexey Stukalov, Virginie Girault, Vincent Grass, Valter Bergant, Ozge Karayel, Christian Urban, Darya A. Haas, Yiqi Huang, Lila Oubraham, Anqi Wang, Sabri M. Hamad, Antonio Piras, Maria Tanzer, Fynn M. Hansen, Thomas Enghleitner, Maria Reinecke, Teresa M. Lavacca, Rosina Ehmann, Roman Wölfel, Jörg Jores, Bernhard Kuster, Ulrike Protzer, Roland Rad, John Ziebuhr, Volker Thiel, Pietro Scaturro, Matthias Mann, Andreas Pichlmair bioRxiv 2020.06.17.156455; doi: https://doi.org/10.1101/2020.06.17.156455

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Zhenming Jin, Xiaoyu Du, Yechun Xu, Yongqiang Deng, Meiqin Liu, Yao Zhao, Bing Zhang, Xiaofeng Li, Lei ke Zhang, Chao Peng, Yinkai Duan, Jing Yu, Lin Wang, Kailin Yang, Fengjiang Liu, Rendi Jiang, Xinglou Yan g, Tian You, Xiaoce Liu, Xiuna Yang, Fang Bai, Hong Liu, Xiang Liu, Luke W. Guddat, Wenqing Xu, Gengfu Xiao, Chengfeng Qin, Zhengli Shi, Hualiang Jiang, Zihe Rao, Haitao Yang bioRxiv 2020.02.26.964882; doi: https://doi.org/10.1101/2020.02.26.964882 Remove from Selected Citations

Green tea and Spirulina extracts inhibit SARS, MERS, and SARS-2 spike pseudotyped virus entry in vitro

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Mechanism and inhibition of SARS-CoV-2 PLpro

Theresa Klemm, Gregor Ebert, Dale J. Calleja, Cody C. Allison, Lachlan W. Richardson, Jonathan P. Bernardini, Bernadine G. C. Lu, Nathan W. Kuchel, Christoph Grohmann, Yuri Shibata, Zhong Yan Gan, James P. Cooney, Marcel Doerflinger, Amanda E. Au, Timothy R. Blackmore, Paul P. Geurink, Huib Ovaa, Janet Newman, Alan Riboldi-Tunnicliffe, Peter E. Czabotar, Jeffrey P. Mitchell, Rebecca Feltham, Bernhard C. Lechtenberg, Kym N. Lowes, Grant Dewson, Marc Pellegrini, Guillaume Lessene, David Komander bioRxiv 2020.06.18.160614; doi: https://doi.org/10.1101/2020.06.18.160614 Remove from Selected Citations

Quantitative PCR for cannabis flower containing SARs-CoV-2

Kevin J. McKernan, Liam T. Kane, Yvonne Helbert bioRxiv 2020.06.06.112474; doi: https://doi.org/10.1101/2020.06.06.112474 Remove from Selected Citations

 Targeting ACE2-RBD interaction as a platform for COVID19 therapeutics: Development and drug repurposing screen of an AlphaLISA proximity assay

Quinlin M. Hanson, Kelli M. Wilson, Min Shen, Zina Itkin, Richard T. Eastman, Paul Shinn, Matthew D. Hall bioRxiv 2020.06.16.154708; doi: https://doi.org/10.1101/2020.06.16.154708 Remove from Selected Citations

• Potential antiviral options against SARS-CoV-2 infection

Aleksandr Ianevski, Rouan Yao, Mona

Høysæter Fenstad, Svetlana Biza, Eva Zusinaite, Tuuli Reisberg, Hilde Lysvand, Kirsti Løseth, Veslemøy Malm Landsem, Janne Fossum Malmring, Valentyn Oksenych, Sten Even Erlandsen, Per Arne Aas, Lars Hagen, Caroline H. Pettersen, Tanel Tenson, Jan Egil Afset, Svein Arne Nordbø, Magnar Bjørås, Denis E. Kainov bioRxiv 2020.05.12.091165; doi: https://doi.org/10.1101/2020.05.12.091165
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Human IgG neutralizing monoclonal antibodies block SARS-CoV-2 infection

Jinkai Wan, Shenghui Xing, Longfei Ding, Yongheng Wang, Dandan Zhu, Bowen Rong, Siqing Wang, Kun Chen, Chenxi He, Songhua Yuan, Chengli Qiu, Chen Zhao, Xiaoyan Zhang, Xiangxi Wang, Yanan Lu, Jianqing Xu, Fei Lan

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Drug Repurposing Screen for Compounds Inhibiting the Cytopathic Effect of SARS-CoV-2

### Catherine

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Specific viral RNA drives the SARS CoV-2 nucleocapsid to phase separate

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 Neuropilin-I facilitates SARS-CoV-2 cell entry and provides a possible pathway into the central nervous system

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Antibody responses to SARS-CoV2 are distinct in children with MIS-C compared to adults with COVID-19

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