

## **Cancer Type/Site Classification using Deep-Learning (Preliminary presentation slides)**

**S. Ravichandran**  
BIDS, FNLCR

(in preparation)

# Acknowledgements

- **NCI-DOE Pilot-1 Team**
  - Maulik Shukla
- **BIDS**
  - Drs. George Zaki, Andrew Weissman, Mark Jensen and Eric Stahlberg
  - Amar Khalsa, Dr. Deb Hope
  - Colleagues who reviewed the material

# Feel free to follow-along

## CBIIT

- <https://cbiit.github.io/sdsi/workshops> (landing site; creation in progress)

## Github

- <https://github.com/ravichas/ML-TC1> (in progress)

# Introduction

- **This is part of the NCI-DOE knowledge/capability transfer efforts**
- **Share tools/techniques/solutions for cancer related problems. We often take a test-case and show how it works**
- **You will be able to take the test-case (code/scripts) and tune it to your needs**

# Motivation: Cancer Prediction vs Cancer Detection

- **Cancer Prediction has been the major focus**
  - Prognosis, Recurrence, Susceptibility
- **Cancer Detection (classification of tumors/cancers) is lagging behind Prediction and we would like to share an application that might be useful**
  - Detect/Identify cancer type at an early stage

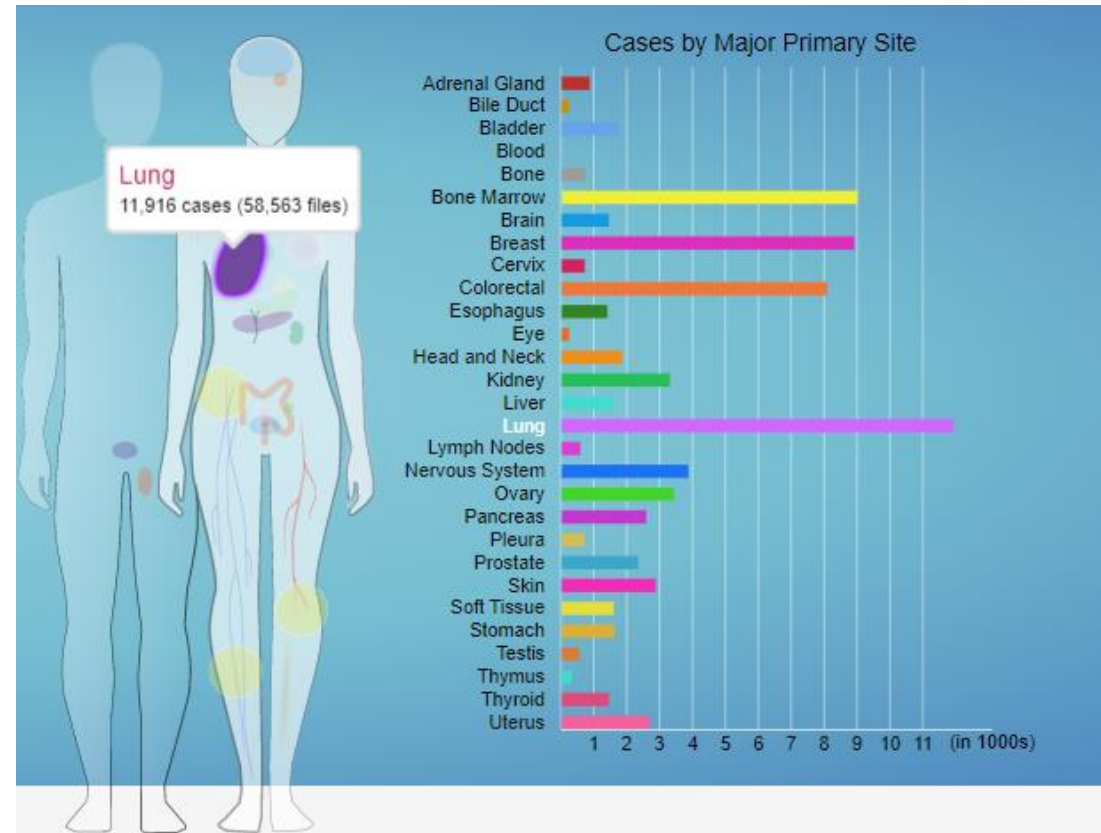
## Goal(s)/Questions

- **Take unstructured genomic expression data from tumor/cancer samples and apply Deep-Learning to create Cancer types/site(s) classifier models**
- **Are the expression profiles unique?**
- **Can we use the model as early cancer type detection**
  - Improving chance of early detection cure/survival?

# Cancer Burden

- **Cancer is a group of diseases and world-wide risk**
- **Acquired or somatic changes causes 90-95% of cancer (all types)**
  - *Source TCGA*
- **~ 200 forms of cancer**
  - *DOI: 10.5114/wo.2014.47136*
- **For 2020**
  - ~1.8M new cancer cases are expected
  - ~600K deaths will occur

Figure from Genomic Data Commons



# Expected New Cases/Deaths in 2020

## New Cancer Cases

Between 2010 and 2020, we expect the number of new cancer cases in the United States to go up about 24% in men to more than 1 million cases per year, and by about 21% in women to more than 900,000 cases per year.

US population gender	Cancers that are expected to increase
Men	Prostrate, Kidney, Liver and Bladder
Women	Lung, Breast, Uterine and Thyroid



# Dynamic genomic changes result in Cancer

Cancer Genome (changes) → Transcript alterations

## Article

### Genomic basis for RNA alterations in cancer

<https://doi.org/10.1038/s41586-020-1970-0>

Received: 29 March 2018

Accepted: 11 December 2019

Published online: 5 February 2020

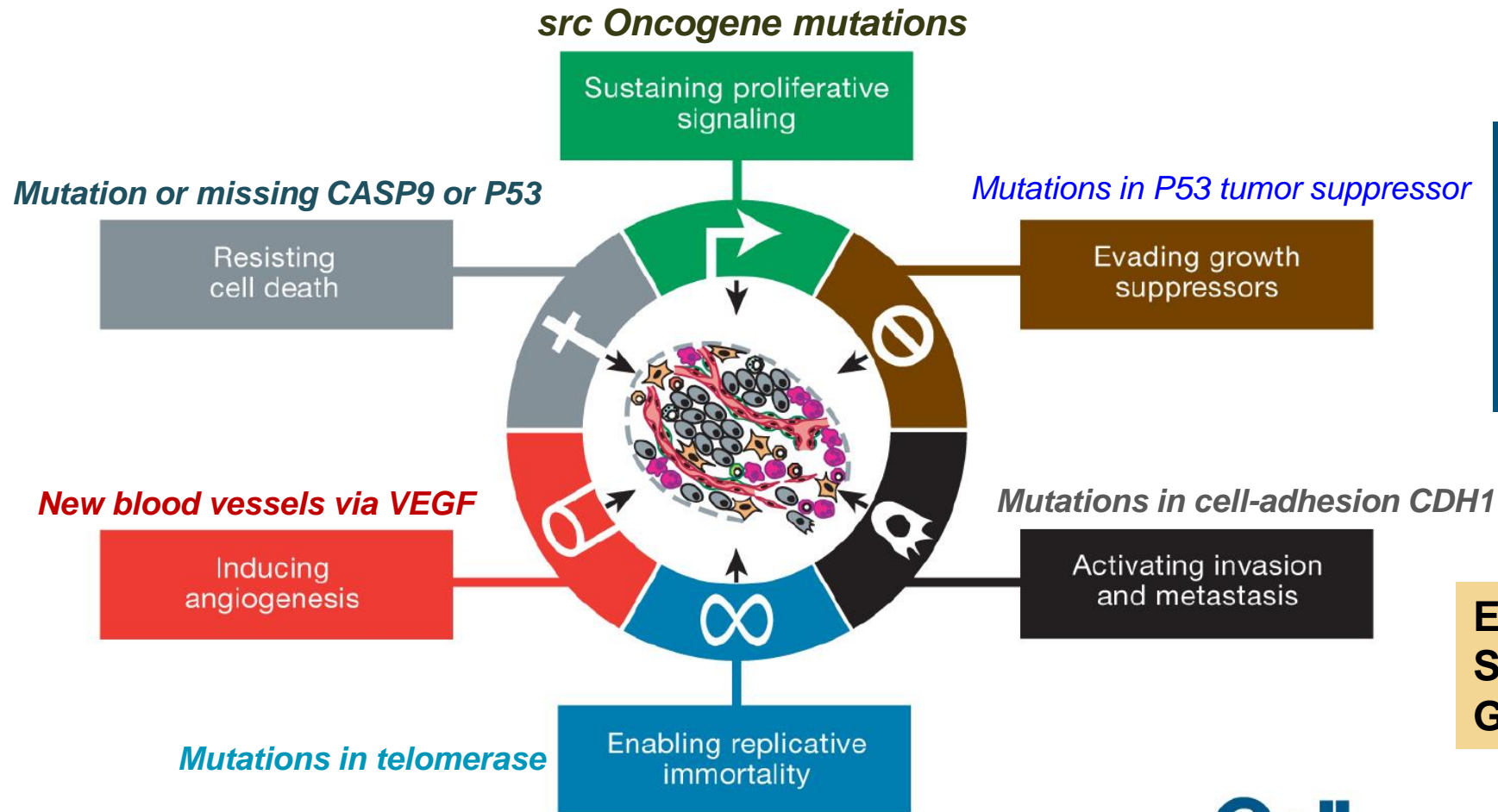
Transcript alterations often result from somatic changes in cancer genomes. Various forms of RNA alterations have been described in cancer, including overexpression, altered splicing and gene fusions; however, it is difficult to attribute these to underlying genomic changes owing to heterogeneity among patients and tumor types, and the relatively small cohorts of patients for whom samples have been analyzed by both transcriptome and whole-genome sequencing.

Expression changes in oncogenes; What type of changes?



# Hallmarks of cancer: Acquired capabilities (mutations) that drive cancer

Hallmarks of Cancer: The Next Generation



Hanahan and Weinberg, 2011



REVIEW | VOLUME 100, ISSUE 1, P57-70, JANUARY 07, 2000

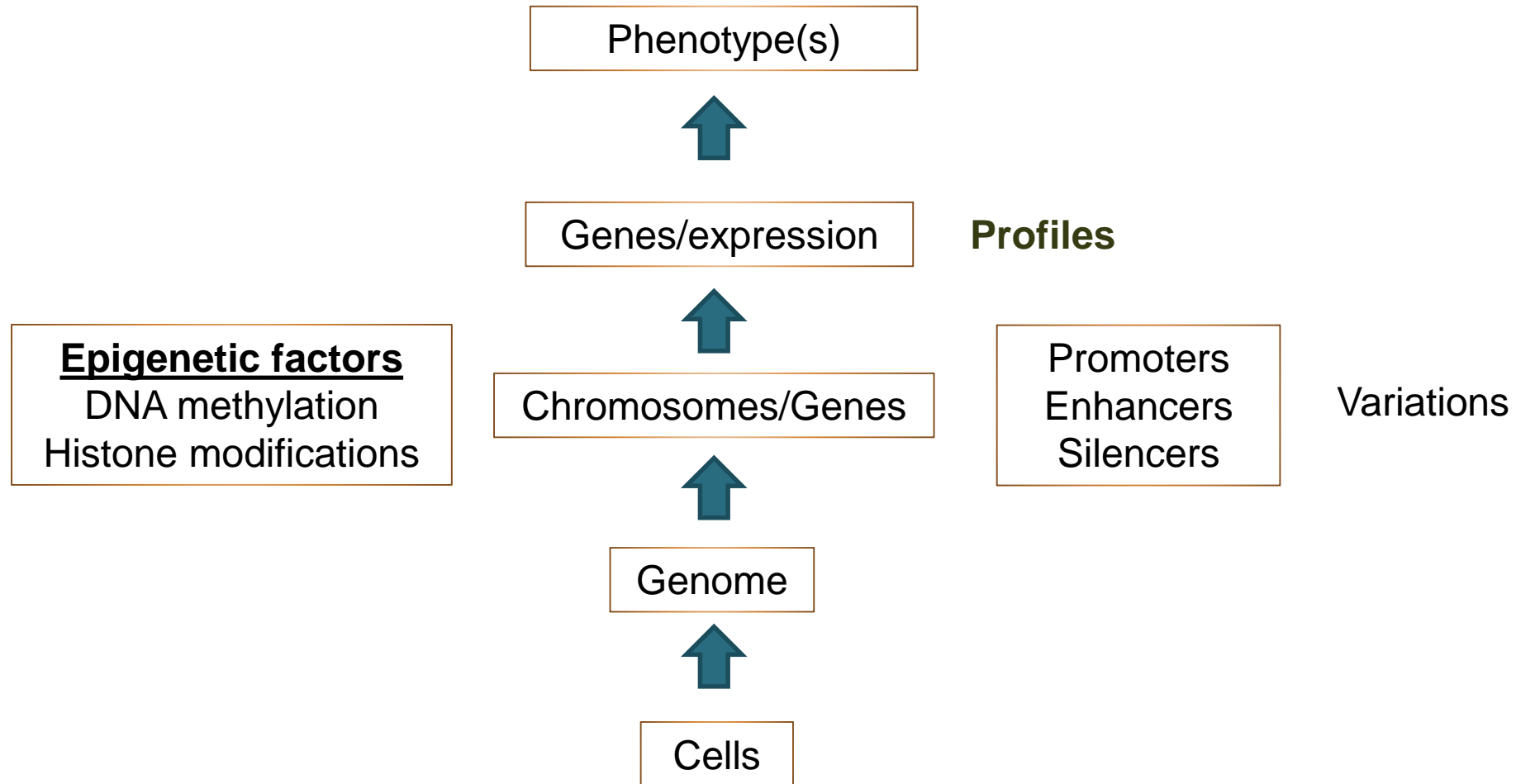
## The Hallmarks of Cancer

Douglas Hanahan • Robert A Weinberg

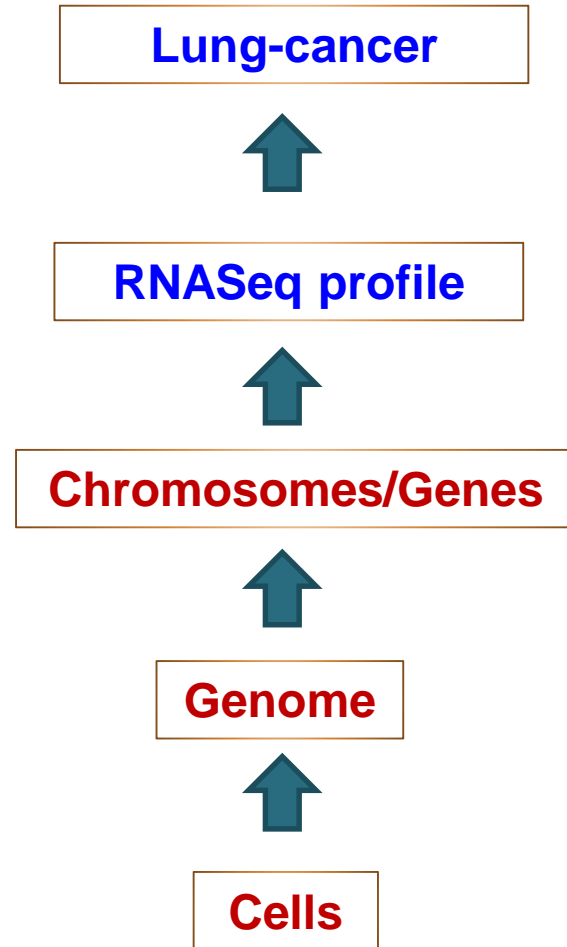
Open Archive • DOI: [https://doi.org/10.1016/S0092-8674\(00\)81683-9](https://doi.org/10.1016/S0092-8674(00)81683-9)

Expression changes in oncogenes;  
Six capabilities; Overview of  
Genotype/phenotypes?

# Influence of genomic features on phenotypes: An overview



# Influence of genomic features on phenotypes: An overview



Diagnosis/treatment vs Prediction



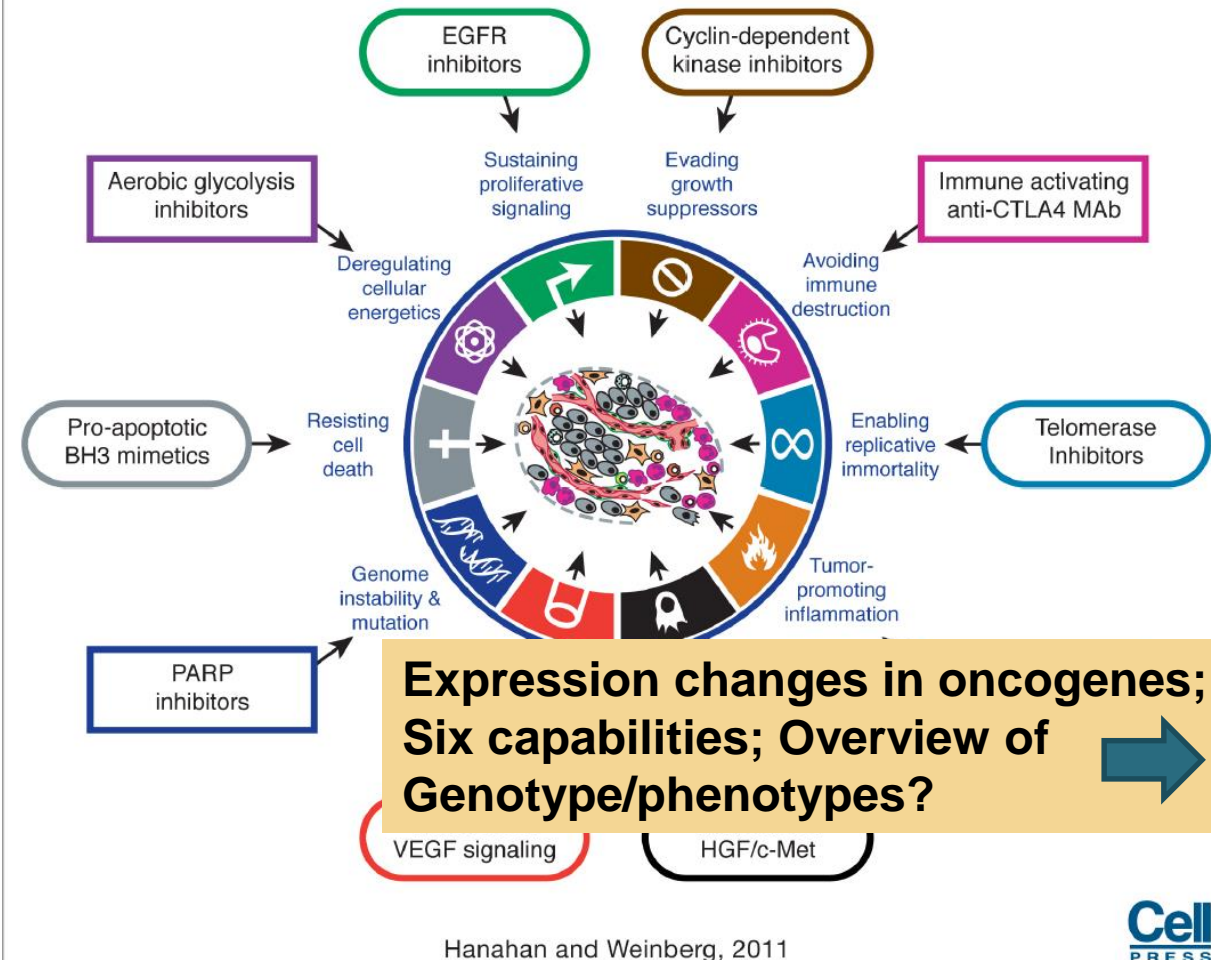
# Treatment vs Type-Prediction

- **Treatment**

- Gene-centric (or a slice of pathway)
- Imatinib targeting BCR/KIT

- **Detecting Type**

- “The architecture of occurring genetic aberrations such as somatic mutations, CNVs, changed gene expression profiles, and different epigenetic alterations, is unique for each type of cancer.”, DOI: 10.5114/wo.2014.47136
- Complex
- Multi-gene centric



*The architecture of occurring genetic aberrations such as somatic mutations, CNV, changed gene expression profiles, and different epigenetic alterations, is unique for each type of cancer*

**DOI: [10.5114/wo.2014.47136](https://doi.org/10.5114/wo.2014.47136)**

## PERSPECTIVE

# Understanding Genotype-Phenotype Effects in Cancer via Network Approaches

Yoo-Ah Kim, Dong-Yeon Cho, Teresa M. Przytycka\*

National Center for Biotechnology Information, National Library of Medicine, National Institutes of Health, Bethesda, Maryland, United States of America

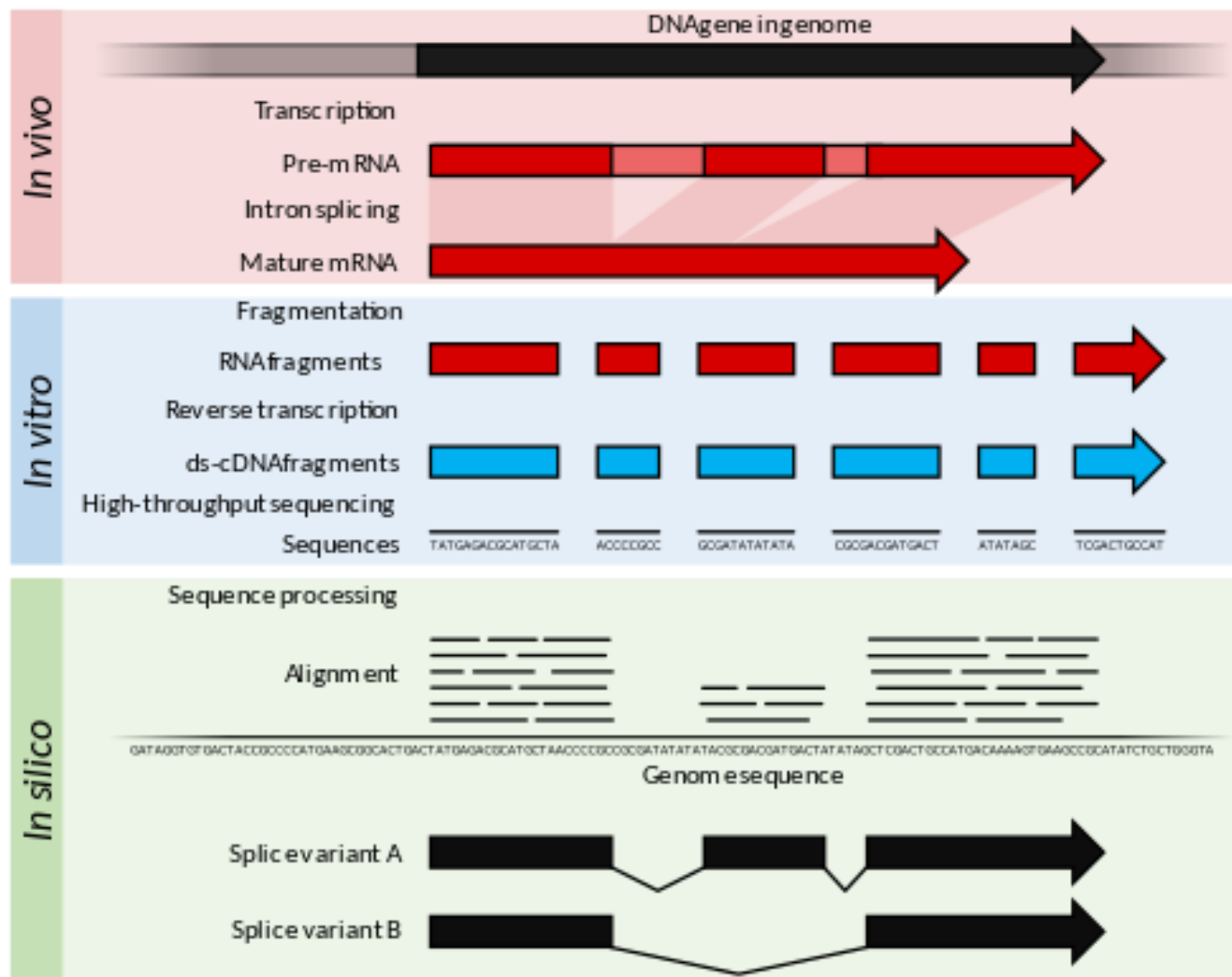
\* [przytyck@ncbi.nlm.nih.gov](mailto:przytyck@ncbi.nlm.nih.gov)

## Author Summary

Cancer is now increasingly studied from the perspective of dysregulated pathways, rather than as a disease resulting from mutations of individual genes. A pathway-centric view acknowledges the heterogeneity between genomic profiles from different cancer patients while assuming that the mutated genes are likely to belong to the same pathway and cause similar disease phenotypes. Indeed, network-centric approaches have proven to be helpful for finding genotypic causes of diseases, classifying disease subtypes, and identifying drug targets. In this review, we discuss how networks can be used to help understand patient-to-patient variations and how one can leverage this variability to elucidate interactions between cancer drivers.

# What kind of data do we need?

NGS



READS

NGS



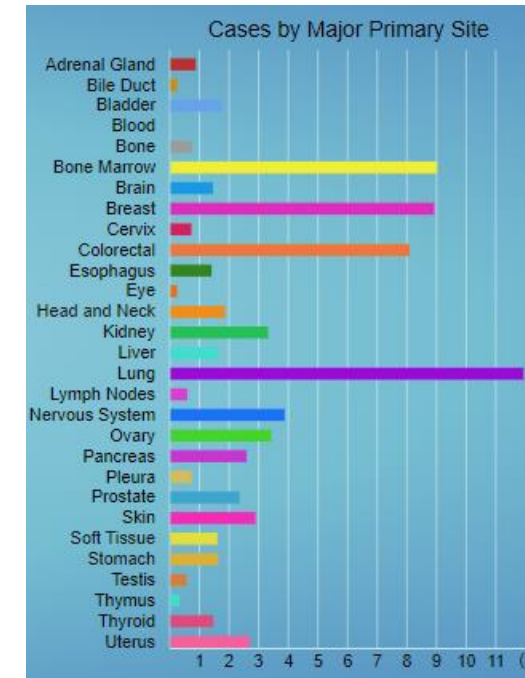
## Data source: The Cancer Genome Atlas (TCGA)

- NIH launched TCGA Pilot Project – a public funded project
- Goal of creating a comprehensive “atlas” of cancer genomic profiles.
- Large cohorts of over 30 human tumors through large-scale genome sequencing and integrated multi-dimensional analyses.
- Contains Microarray and NGS data
  - RNASeq
  - miRNA seq
  - SNP based platforms
  - .....
- TCGA data is available via GDC



# Data Harmonization: GDC

- Data and metadata is submitted to the GDC in standard data types and file formats. Other data sources (Ex. TCGA) are also included
- Data are harmonized against a common reference genome (GRCh38)
- For this workshop, we will focus on TCGA Genomic expression data from GDC



# Expression Data Quantification

- $RC_g$ : Number of reads mapped to the gene
- $RC_{g75}$ : The 75th percentile read count value for genes in the sample
- $L$ : Length of the gene in base pairs; Calculated as the sum of all exons in a gene

$$FPKM-UQ = \frac{RC_g \times 10^9}{RC_{g75} \times L}$$

FASTQ

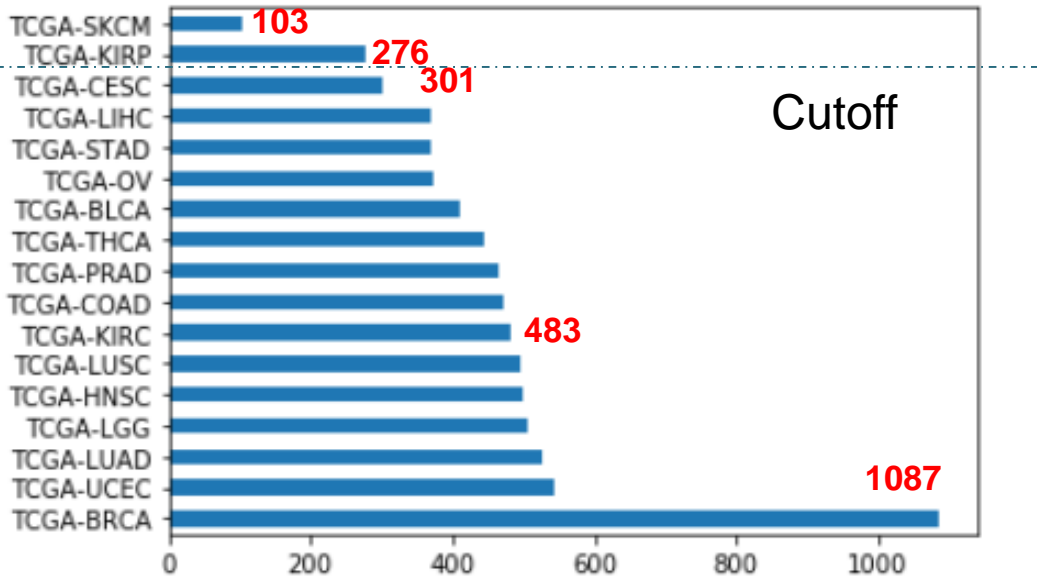
Alignment to Ref  
Genome (SAM/BAM)

Quantification HTSeq

Gene Expression  
(FPKM-UQ)

Fragments **P**er **K**ilobase of transcript per **M**illion mapped reads

# How much data for modeling?



CODE	Cancer Site/Type
BRCA	Breast invasive carcinoma
UCEC	Uterine Corpus Endometrial Carcinoma
LUAD	Lung adenocarcinoma
LGG	Brain Lower Grade Glioma
HNSC	Head and Neck squamous cell carcinoma
LUHSC	Lung squamous cell carcinoma
KIRC	Kidney renal clear cell carcinoma
PRAD	Prostate adenocarcinoma
COAD	Colon adenocarcinoma
THCA	Thyroid carcinoma
BLCA	Bladder Urothelial Carcinoma
OV	Ovarian serous cystadenocarcinoma
STAD	Stomach adenocarcinoma
LIHC	Liver hepatocellular carcinoma
CEC	Cervical squamous cell carcinoma and endocervical adenocarcinoma

300  
samples  
each

# Expression data from a sample

## TCGA-BRCA

Genes	Expression
ENSG00000242268.2	1658.464179
ENSG00000270112.3	460.2343433
ENSG00000167578.15	52440.10096
ENSG00000273842.1	0
ENSG00000078237.5	68165.45626
ENSG00000146083.10	255959.2351
ENSG00000225275.4	0
ENSG00000158486.12	104.9473768
ENSG00000198242.12	4968556.658
ENSG00000259883.1	6108.999052
ENSG00000231981.3	0
ENSG00000269475.2	0
ENSG00000201788.1	0
ENSG00000134108.11	957330.2056
ENSG00000263089.1	3484.027373
ENSG00000172137.17	41485.9507
ENSG00000167700.7	226717.4208
ENSG00000234943.2	2082.245035
ENSG00000240423.1	310.5246749
ENSG00000060642.9	155863.9216
ENSG00000271616.1	0
ENSG00000234881.1	0
ENSG00000236040.1	394.4755669
ENSG00000231105.1	1583.312582
ENSG00000243044.1	0
ENSG00000182141.8	45538.60648
ENSG00000269416.4	119.0847054
ENSG00000264981.1	0

60,483  
transcripts

Gene: AC090241.2 ENSG00000270112

Description novel transcript, antisense to ST8SIA5

Location [Chromosome 18: 46,756,487-46,802,449](#) forward strand.  
GRCh38:CM000680.2

About this gene This gene has 8 transcripts ([splice variants](#))

Transcripts [Hide transcript table](#)

Gene: DNAH3 ENSG00000158486

Description dynein axonemal heavy chain 3 [Source:HGNC Symbol;Acc:[HGNC:2949](#)]

Gene Synonyms DKFZp434N074, DLP3, Dnahc3b, Hsadhc3

Location [Chromosome 16: 20,933,111-21,159,441](#) reverse strand.  
GRCh38:CM000678.2

About this gene This gene has 6 transcripts ([splice variants](#)), [371 orthologues](#), [14 paralogues](#) and is a member of [1 Ensembl protein family](#).

Transcripts [Hide transcript table](#)

Breast Cancer

60,484  
transcripts

ENSG00000070113	460	23443.43
ENSG00000065788.15	52440	10000
ENSG00000278421.0		
ENSG00000092875.5	68185	45626
ENSG00000248210.10	123	898.8
ENSG00000252574.0		
ENSG00000214846.12	104	9473.68
ENSG00000280422.12	418	6858.58
ENSG00000288811.10	118	108.99052
ENSG00000219813.3		
ENSG0000020475.2		
ENSG00000201881.0		
ENSG00000214308.11	95730	2056
ENSG00000205868.10	348	402.6
ENSG00000072317.17	1458	9507
ENSG00000076070.7	22617	4208
ENSG00000234463.2	2082	26025
ENSG00000205868.10	348	402.6
ENSG00000060420.9	153	3262.66
ENSG00000278161.0		
ENSG00000214881.1		
ENSG00000040041.2	394	4755669
ENSG00000211051.1	158	3112582
ENSG00000283044.1		
ENSG00000212414.8	455	438.6
ENSG00000209416.4	129	1870.54

# Merged Sample Expression Data

Genes

SAMPLES

	0	1	2	3	4	5	6	7	8	9	...	60474	60475	60476	60477	60478	60479	60480	60481	60482	submitter_id
0	574548	2263.14	983212	69718	54834.9	19718.1	175853	735123	38662.4	233190	...	0	0	0	0	0	0	0	0	0	TCGA-04-1331-01A-01R-1569-13
1	352295	4592.37	663107	39745.4	36553.5	41147.1	241313	396423	37567	128693	...	0	0	0	0	0	0	0	0	0	TCGA-04-1332-01A-01R-1564-13
2	295162	649.026	1.21115e+06	57385.5	33097.4	58051.8	228615	346066	105567	408267	...	0	0	0	0	0	0	0	0	0	TCGA-04-1338-01A-01R-1564-13
3	329580	1835.59	1.08437e+06	33812.3	24516.1	22330.6	42134.4	895558	56178	83847.3	...	0	0	0	0	0	0	0	0	0	TCGA-04-1341-01A-01R-1564-13
4	289269	40061.7	2.44837e+06	26399.5	18248	49610	74761.1	571992	71951.9	98726.4	...	0	0	0	0	0	0	0	0	0	TCGA-04-1343-01A-01R-1564-13
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
4495	1.18093e+06	0	1.01139e+06	67877.2	15005.7	50527.3	6.21536e+06	1.47373e+06	459656	167488	...	0	0	0	0	0	0	0	0	0	TCGA-ZS-A9CD-01A-11R-A37K-07
4496	929228	0	869800	95607.5	17188.6	9352.12	7.61121e+06	196838	354465	138074	...	0	0	0	0	0	0	0	0	0	TCGA-ZS-A9CE-01A-11R-A37K-07
4497	469276	476.683	516938	110051	34469.4	37334.7	5.95811e+06	427832	323833	154861	...	0	0	0	0	0	0	0	0	0	TCGA-ZS-A9CF-01A-11R-A38B-07
4498	2.44119e+06	18282.7	853547	79288.7	106926	42593.9	4.80111e+06	955338	331924	177020	...	0	0	0	0	0	0	0	0	0	TCGA-ZS-A9CG-01A-11R-A37K-07
4499	259853	505.488	591328	74253.7	42553.5	118772	148978	508465	153862	170412	...	0	0	0	0	0	0	0	0	0	TCGA-ZX-AA5X-01A-11R-A42T-07

4500 rows × 60484 columns

Transpose and  
add as a row

Genes	Expression
ENSG0000024298.2	3038.404179
ENSG00000276112.3	403.734143
ENSG0000026978.15	52440.1006
ENSG0000027840.1	0
ENSG0000028121.1	68285.4526
ENSG0000024293.10	25099.2351
ENSG0000025277.4	0
ENSG0000025486.12	154.947378
ENSG00000219842.12	406856.458
ENSG0000025881.1	6218.19052
ENSG0000021038.3	0
ENSG0000028071.2	0
ENSG0000026178.1	0
ENSG0000023428.11	90730.2056
ENSG0000026208.1	2484.0373
ENSG00000272137.17	41485.9507
ENSG00000257780.7	22672.4208
ENSG0000025484.2	2982.24035
ENSG00000240423.1	330.5246749
ENSG00000260342.9	125863.5216
ENSG00000271816.1	0
ENSG00000214881.1	0
ENSG00000230461.1	394.475669
ENSG00000231101.1	1583.112582
ENSG00000240464.1	0
ENSG00000252141.8	45338.40648
ENSG00000289416.4	119.0847054
ENSG00000254911.1	0

# Quantifying mRNA abundance and Scaling

- GDC harmonization data is provided in FPKM-UQ
- In our code, FPKM-UQ is rescaled to TPM using the following formula.

$$\text{TPM}_i = \left( \frac{\text{FPKM}_i}{\sum_j \text{FPKM}_j} \right) \cdot 10^6$$

- TPM has nice mathematical properties and a stable entity

<https://docs.gdc.cancer.gov/Encyclopedia/pages/HTSeq-FPKM-UQ/>

Mapping and quantifying mammalian transcriptomes  
by RNA-Seq

Ali Mortazavi<sup>1,2</sup>, Brian A Williams<sup>1,2</sup>, Kenneth McCue<sup>1</sup>, Lorian Schaeffer<sup>1</sup> & Barbara Wold<sup>1</sup>

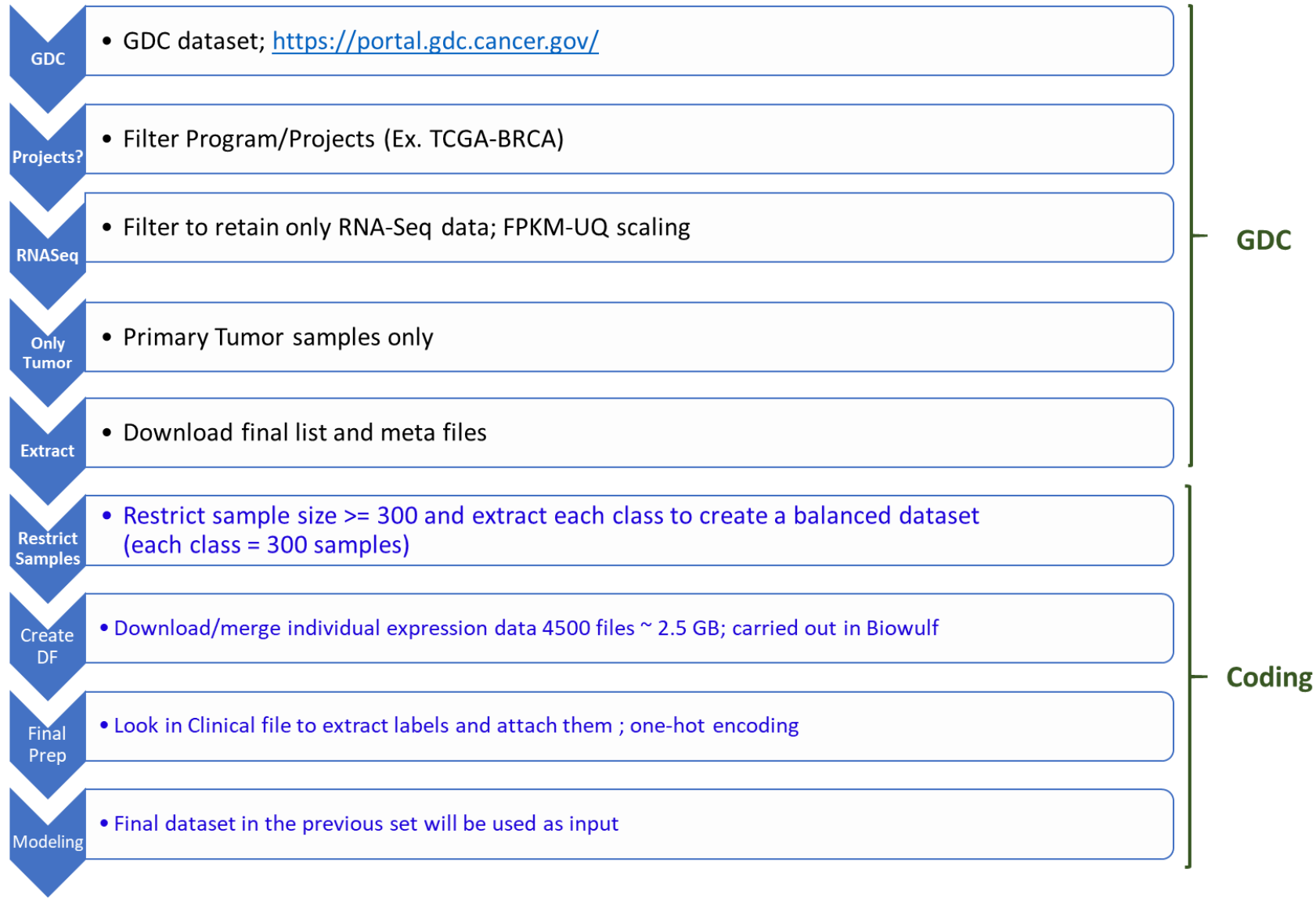


# One-hot encoding to convert Cancer types to numbers

- **Convert each class to a numerical quantity**
  - BRCA to 0 ; LUAD to 1 etc.
  - 0, 1, 2, 3, ..., 13, 14, 15

```
>>> encoded
array([[1., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0.],
       [0., 1., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0.],
       [0., 0., 1., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0.],
       [0., 0., 0., 1., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0.],
       [0., 0., 0., 0., 1., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0.],
       [0., 0., 0., 0., 0., 1., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0.],
       [0., 0., 0., 0., 0., 0., 1., 0., 0., 0., 0., 0., 0., 0., 0., 0.],
       [0., 0., 0., 0., 0., 0., 0., 1., 0., 0., 0., 0., 0., 0., 0., 0.],
       [0., 0., 0., 0., 0., 0., 0., 0., 1., 0., 0., 0., 0., 0., 0., 0.],
       [0., 0., 0., 0., 0., 0., 0., 0., 0., 1., 0., 0., 0., 0., 0., 0.],
       [0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 1., 0., 0., 0., 0., 0.],
       [0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 1., 0., 0., 0., 0.],
       [0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 1., 0., 0., 0.],
       [0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 1., 0., 0.],
       [0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 1., 0.],
       [0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 0., 1.]],
      dtype=float32)
```

# Data preparation steps summary



## Before we break for hands-on

- **Python as the programming language for this workshop, but similar libraries are available in R or other languages**



- **Will use Jupyter Notebook for sharing the code**
  - With little effort one can convert the Python code into R and still use Jupyter Notebook

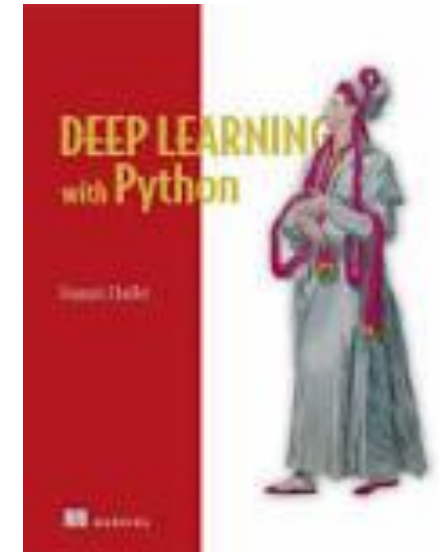
## To be continued after Code-Review

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<https://github.com/ravichas/ML-TC1>

## Before we break for hands-on

- Due to lack of time, I wont be covering the basics of Neural Network



- Following two are good books for beginners and up

# Convolutional Neural Networks

- Preparation in progress

# Thanks

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S. Ravichandran