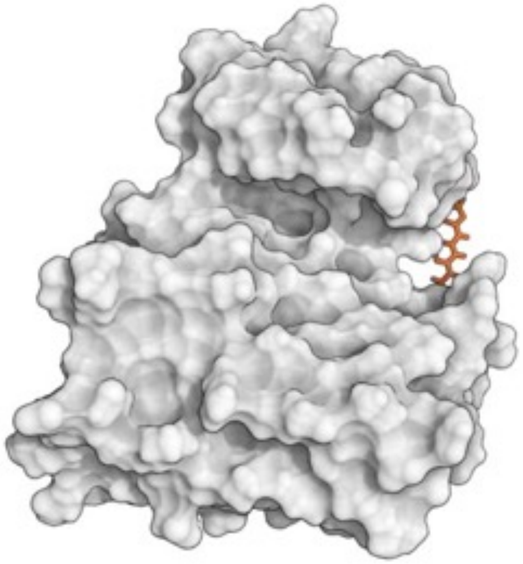
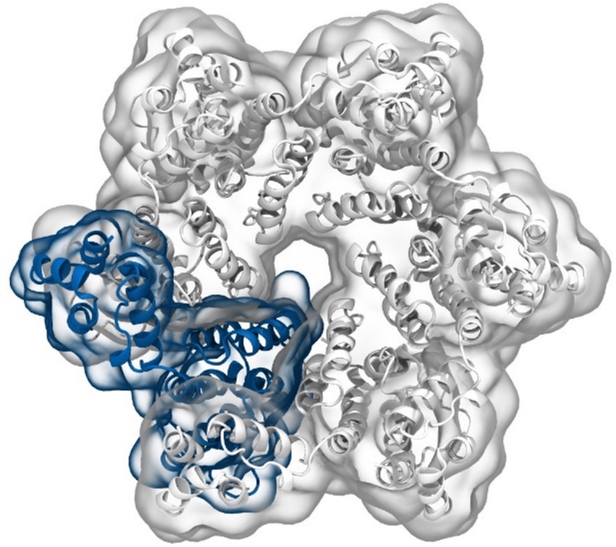
Simulation of Biomolecules



Lecture 2:
Protein preparation

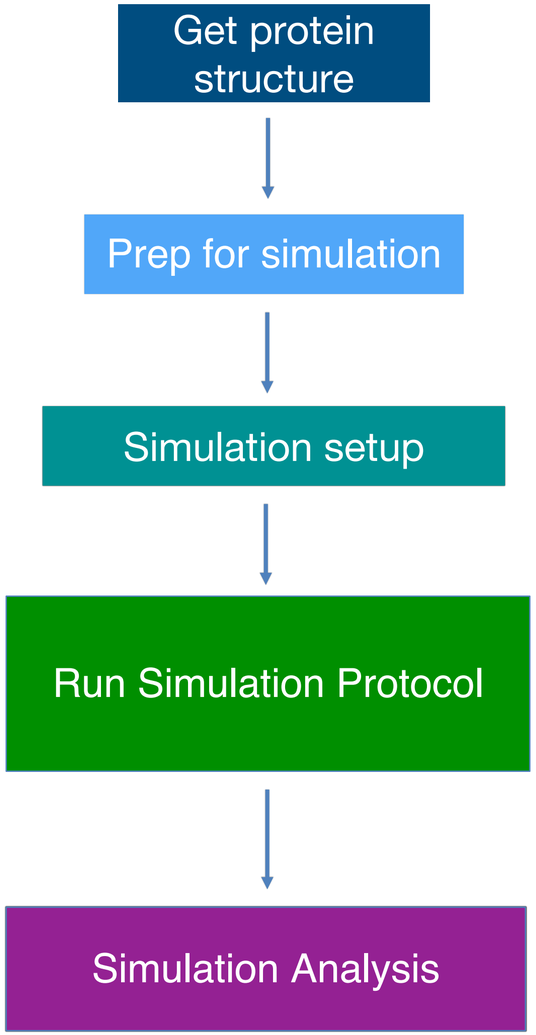


2024 CCP5 Summer School

Dr Matteo Degiacomi
Durham University
matteo.t.degiacomi@durham.ac.uk

Dr Antonia Mey
University of Edinburgh
antonia.mey@ed.ac.uk

A typical workflow for molecular dynamics



pdb api

pdb2gmx

SCHRÖDINGER Maestro

RDkit

OpenMM tools

SeeSAR The drug design dashboard

Amber tools

[...]

GUI, TCL, bash, python, Perl … get creative

Gromacs

OpenMM

Python API

Amber

NAMD

DLPoly

SOMD

Charmm

mostly C++ commandline

BioSimSpace

[...]

PyEMMA

MDAnalysis

md-analysis-tools

MDTraj

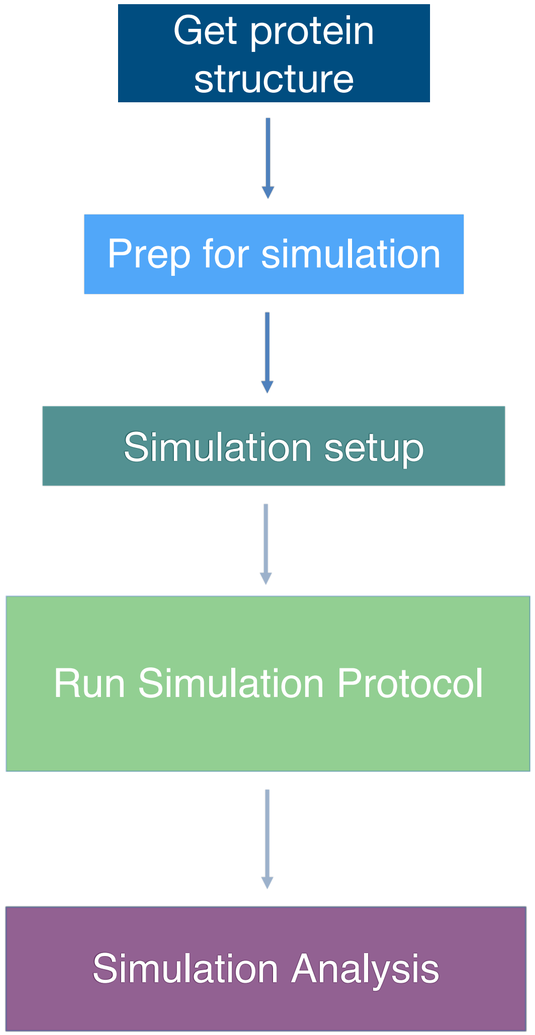
MDAnalysis

VMD

[...]

TCL, bash, python, Perl … get creative

Let’s get started with understanding protein structures



OpenEye SCIENTIFIC

pdb2gmx

pdb api

RDkit

SCHRÖDINGER Maestro

OpenMM tools

Amber tools

SeeSAR
The drug design dashboard

[…]

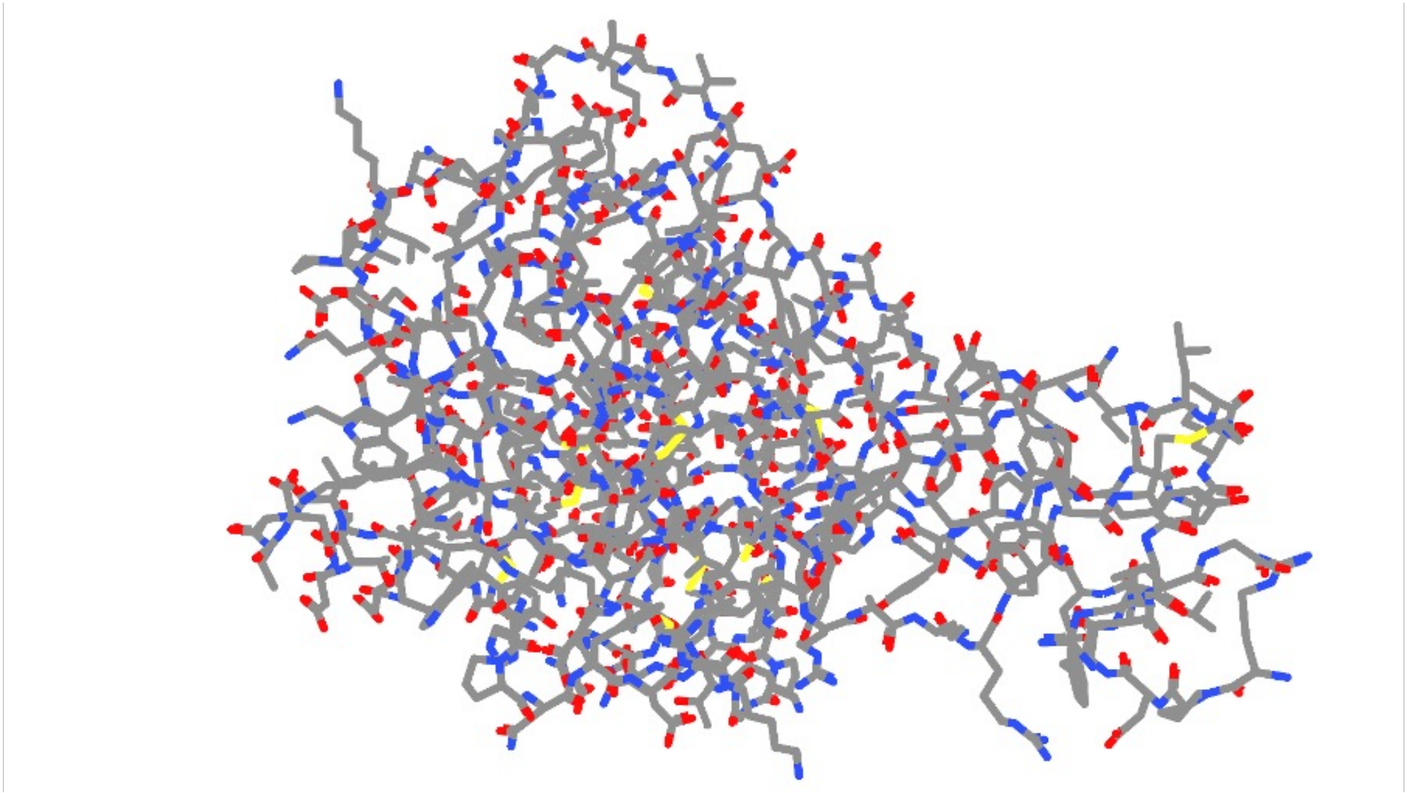
GUI, TCL, bash, python, Perl … get creative

Part 1: I have a protein I want to model, now what?

What things are important to look out for in protein prep?

https://www.menti.com/alc5aozap924 58950948

Crystal structures provide electron densities

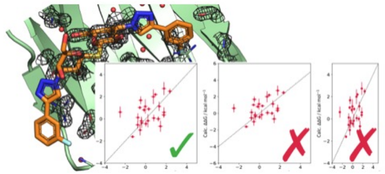


Crystal structures are models with potential errors

HOME / ARCHIVES / VOL. 4 NO. 1 (2022) / Articles

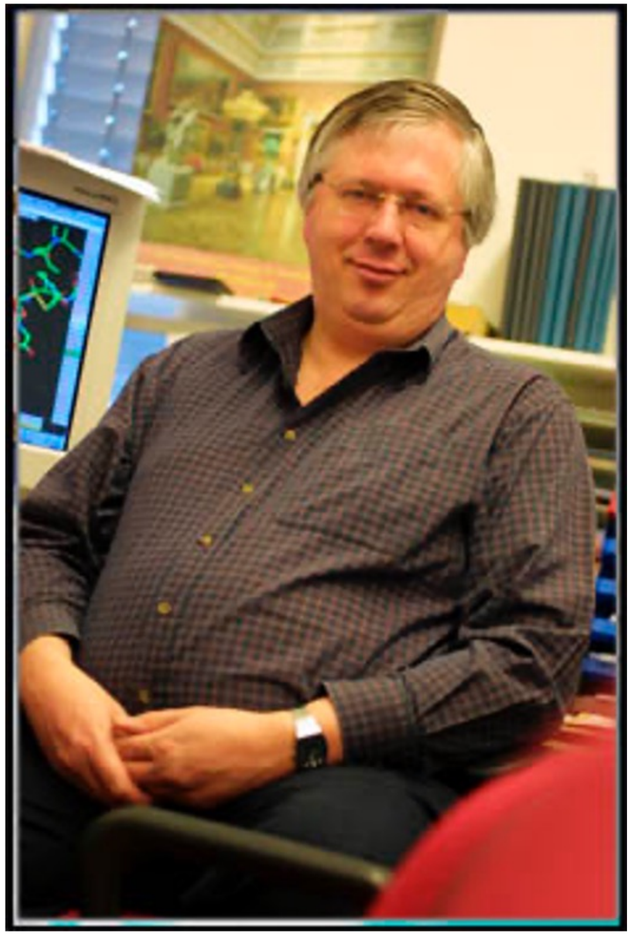
Best Practices for Constructing, Preparing, and Evaluating Protein-Ligand Binding Affinity Benchmarks [Article v1.0]

David F. Hahn
Computational Chemistry, Janssen Research & Development, Turnhoutseweg 30, Beerse B-2340, Belgium
https://orcid.org/0000-0003-2830-6880



Examples of errors/oddities in PDB structures

https://swift.cmbi.umcn.nl/teach/pdbad/

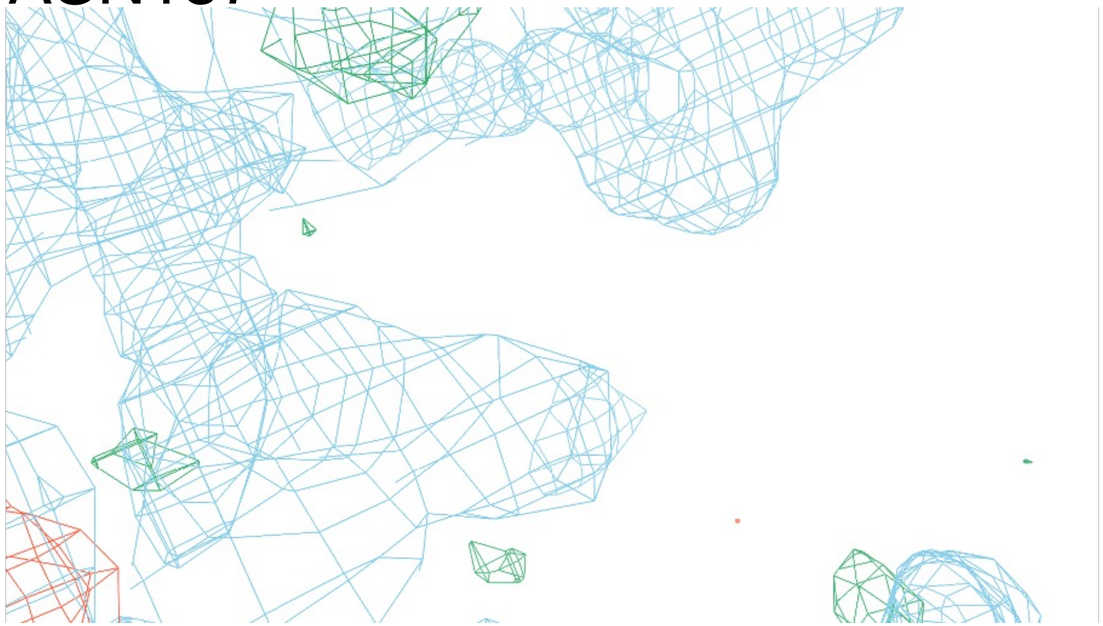


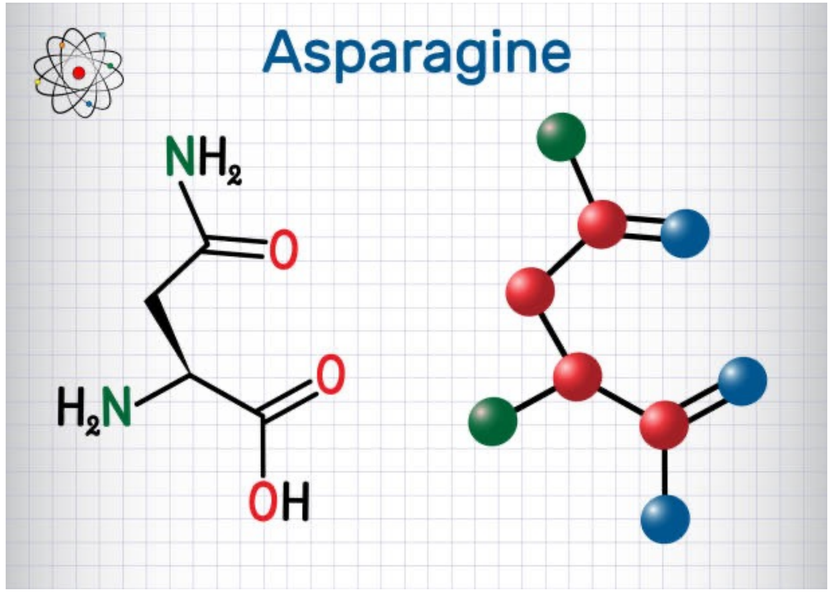
Gert Vriend (author of WHAT_CHECK)

Crystal structures are models derived from electron densities

1T38: HUMAN O6-ALKYLGUANINE-DNA ALKYLTRANSFERASE

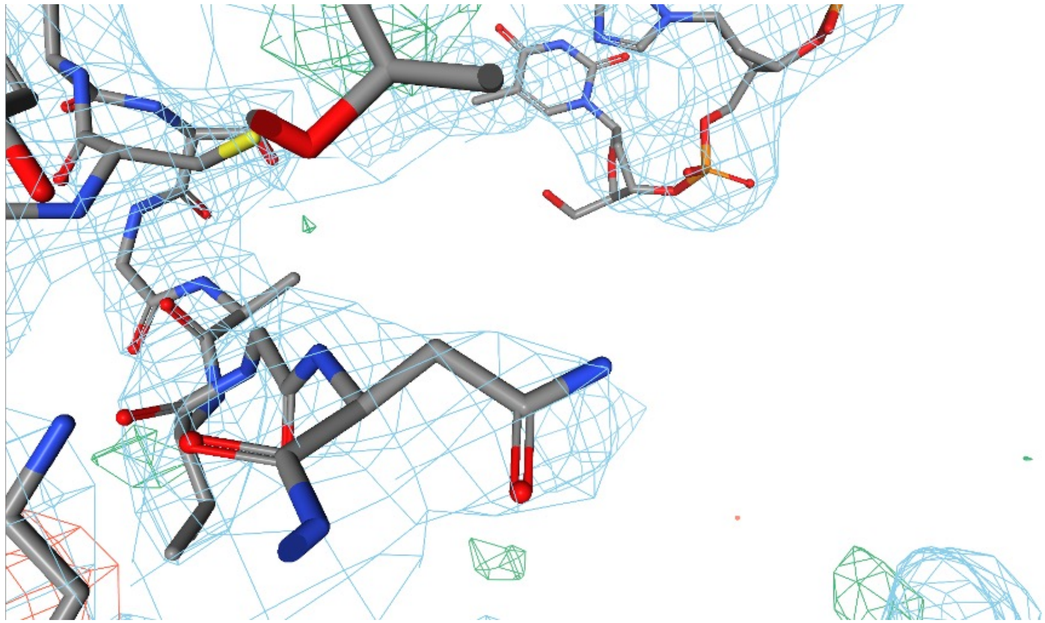
ASN157

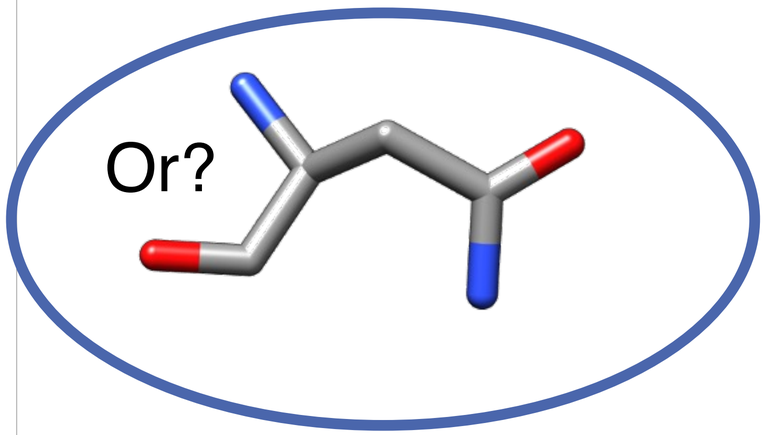




Crystal structures are models derived from electron densities

1T38: HUMAN O6-ALKYLGUANINE-DNA ALKYLTRANSFERASE

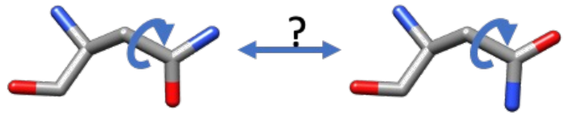




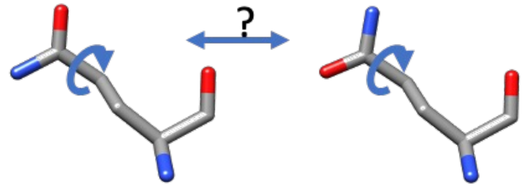
Alternative conformations of side chains - NGH flips

Typically the crystallographers will have assigned the orientation based on potential H-bonding interactions, etc.

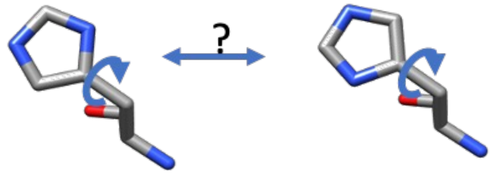
No standard protocol for this: always worth double-checking.



Asparagine (N)



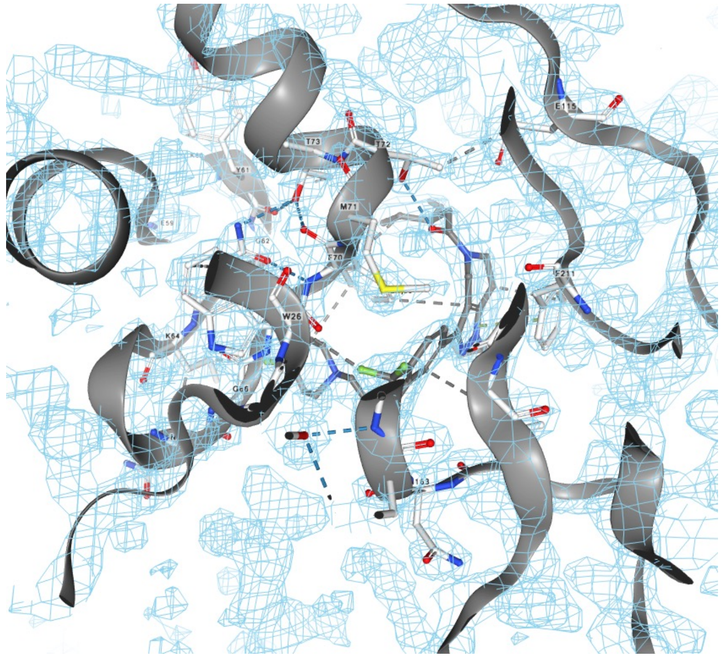
Glutamine (Q)



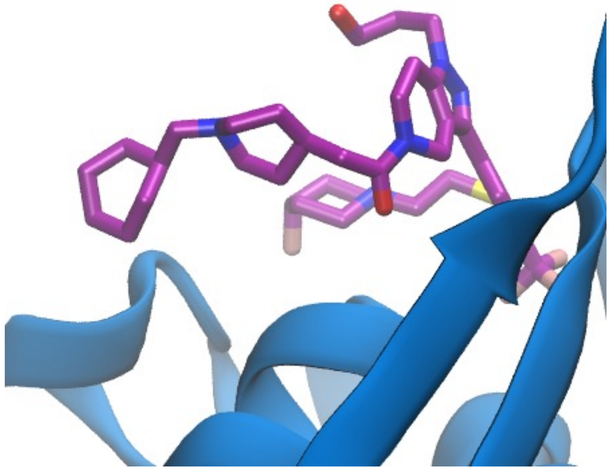
Histidine (H)

Ligand densities can also have creative input from the crystallographer

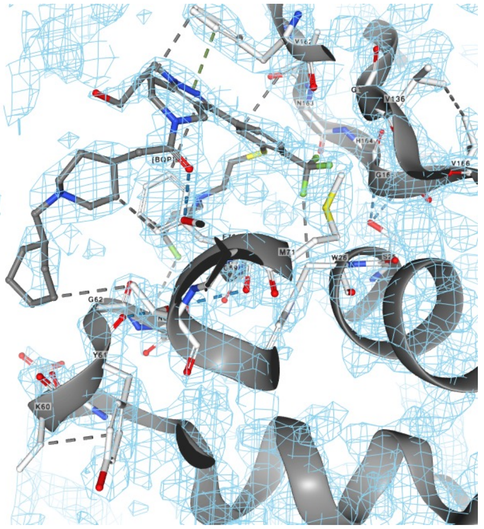
Cathepsin S



Cathepsin S
with ligand

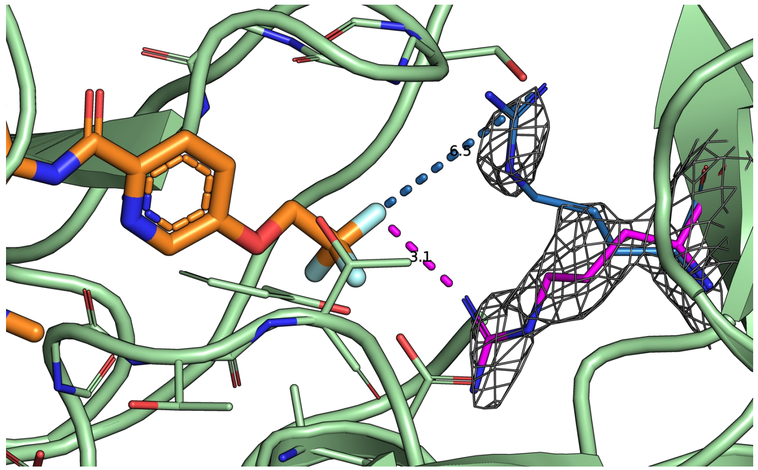


boat/chair?

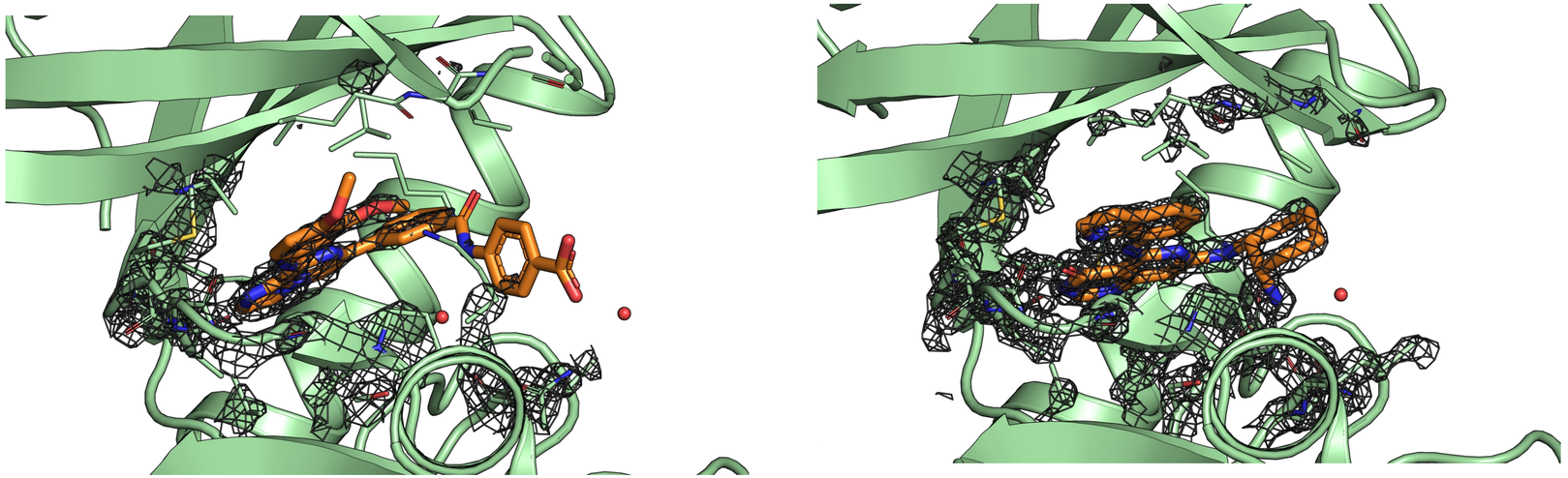


Picking the best crystal structure requires care

BACE (Hunt)



spleen tyrosine kinase

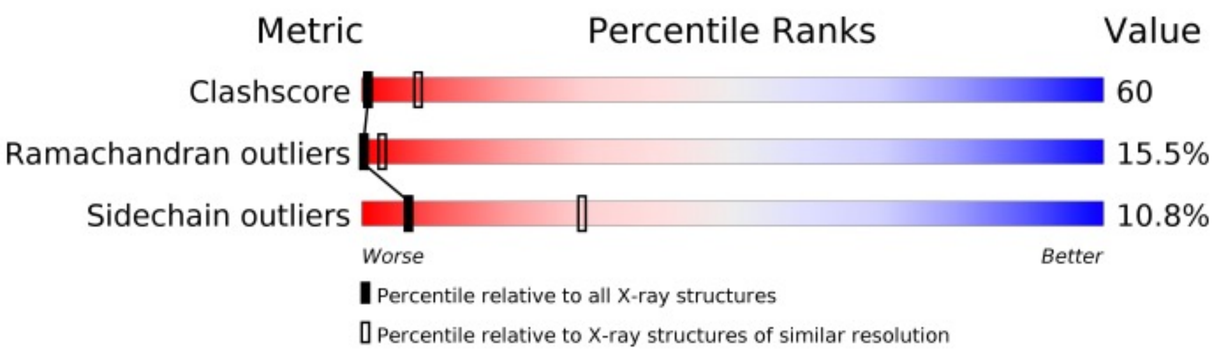


Which protein structure?

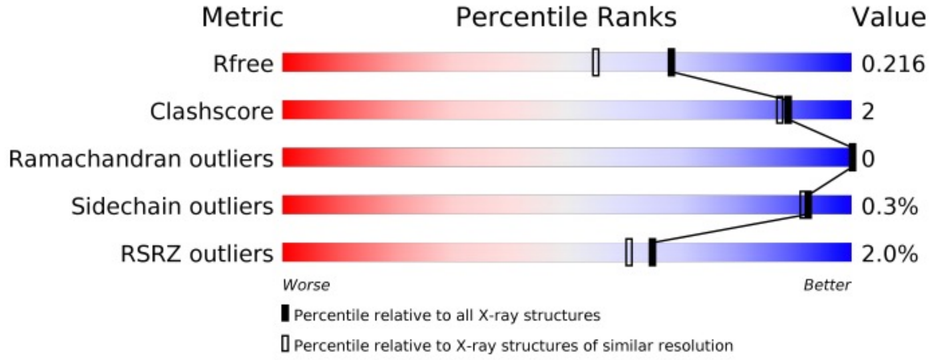
- Alternative side chain conformations need to be assessed carefully
- Active site residue densities are important for choosing the right crystal structure

The RCSB PDB report can help with choosing structures

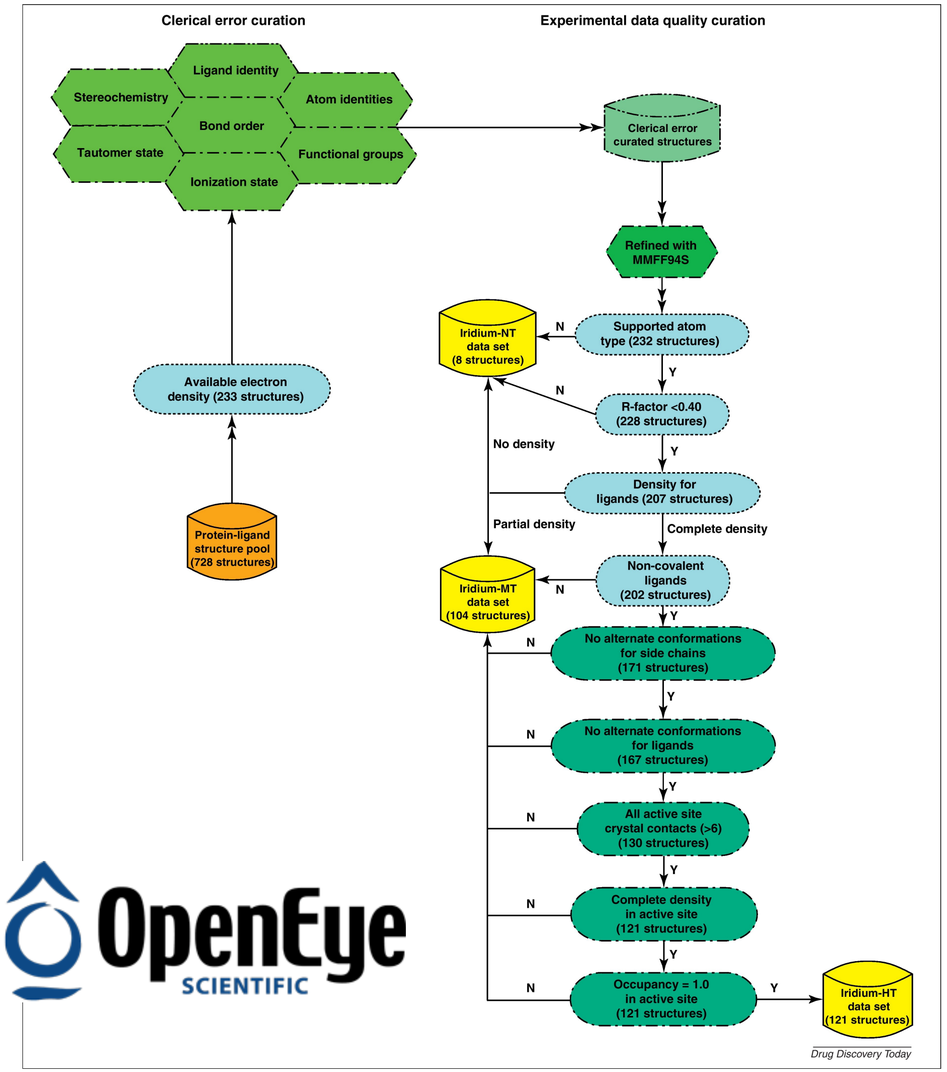
Jnk1 - 2GMX



Jnk1 - 3ELJ



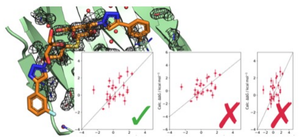
The Iridium score can help assess the trustworthiness of an X-ray structure



Warren et al. *Drug Discovery Today* (2012) **17** 270-1281

Best Practices for Constructing, Preparing, and Evaluating Protein-Ligand Binding Affinity Benchmarks [Article v1.0]

David F. Hahn
Computational Chemistry, Janssen Research & Development, Turnhoutseweg 30, Beerse B-2340, Belgium
https://orcid.org/0000-0003-2830-6880



Jnk1[74, 86]	2GMX (NT, -)[f]	0.77	6D09 (HT, 0.35)[b]	3V3V (MT, 1.5)[b, h, e]	21	3.4
	4HW3 (HT, 0.41)	0.26	3ELJ (MT, 0.31)[a]	4WMU (HT, 0.41)[b]	42	4.2
MCL1[74, 87]			6O6F (HT, 0.30)	4ZBI (MT, 0.45)[b]		
			4ZBF (HT, 0.35)[b]			
			3WIX (HT, 0.37)[a, c]			

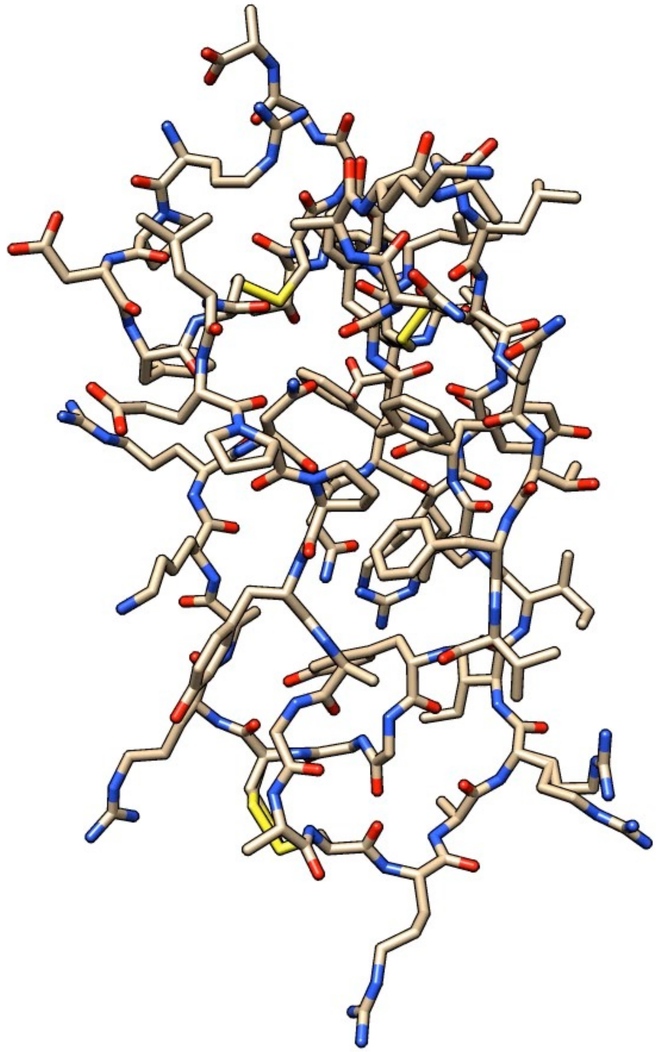
12 Features to inform the Iridium Score:

- R-free value
- Resolution
- Density coverage of ligand heavy atoms
- Active site density coverage
- Alternative locations active site/ligands
- […]

	HT	MT	NT
Ligand	> 0.9	< 0.9 and > 0.5	< 0.5
Active Site	> 0.95	< 0.95 and > 0.5	< 0.5

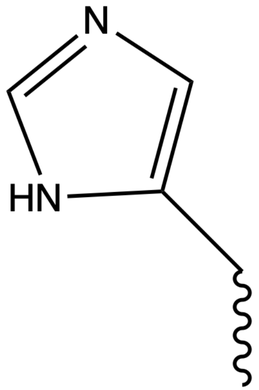
Missing information in structures: Hydrogen atoms

- Missing from most crystal structures
- Needed for molecular modelling
- Most MD packages (AMBER, CHARMM, GROMACS, etc.) include tools to “automatically” add H-atoms.
- Any issues?

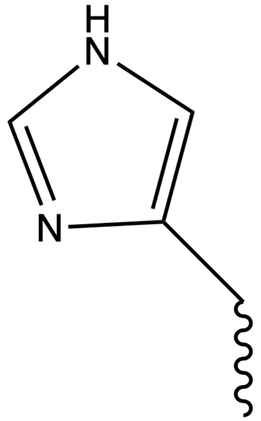


How to choose the right tautomer?

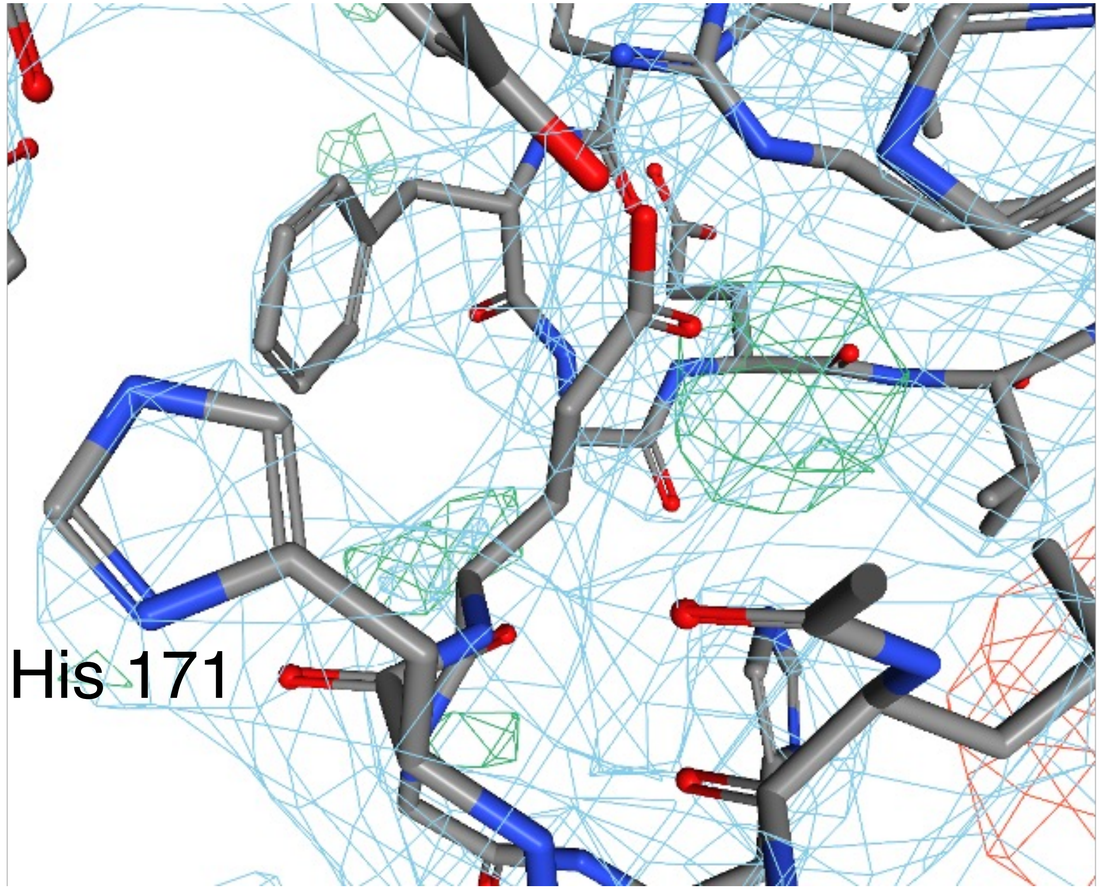
Which tautomer?



δ-tautomer

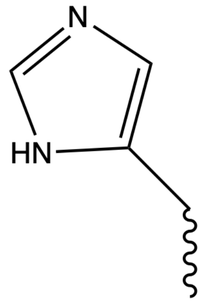


ε-tautomer

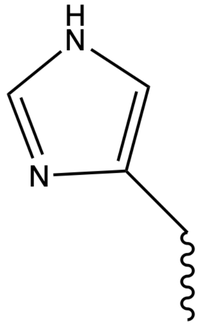


How to choose the right tautomer and protonation state?

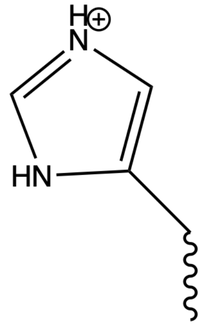
Which tautomer and protonation?



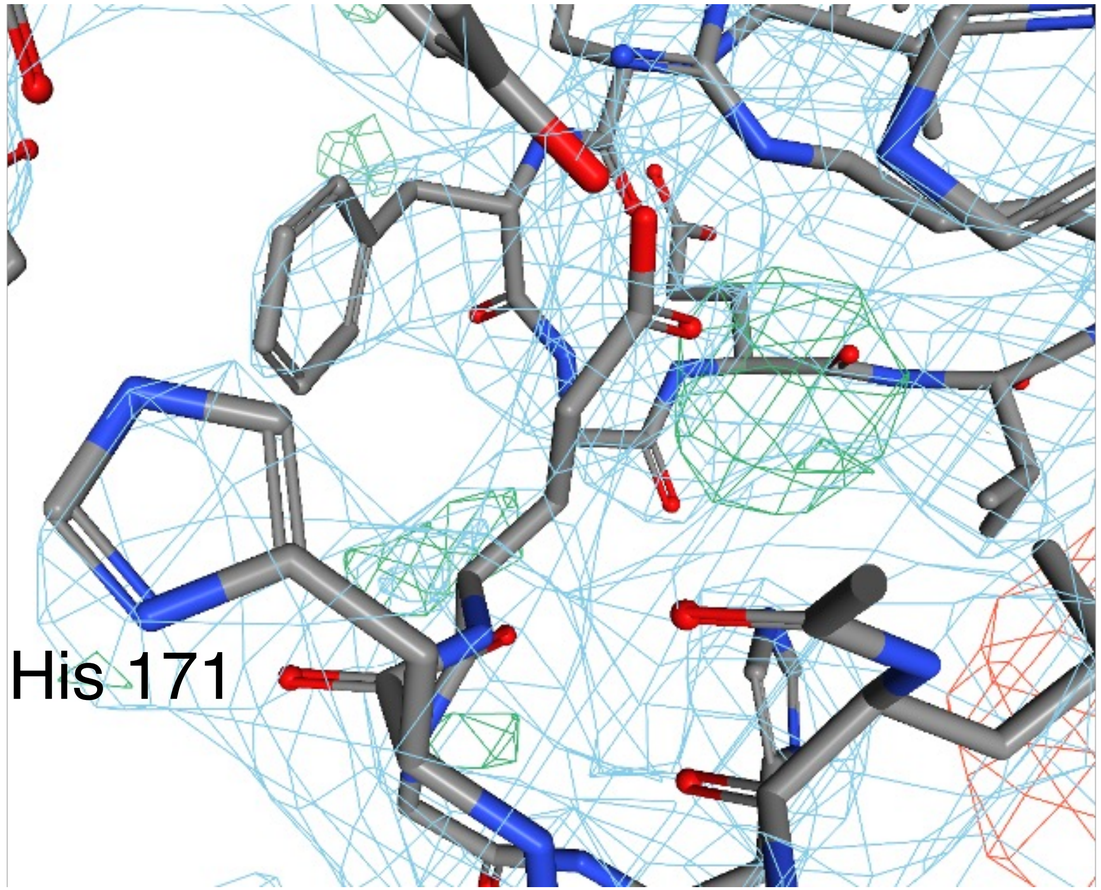
δ-tautomer



ε-tautomer

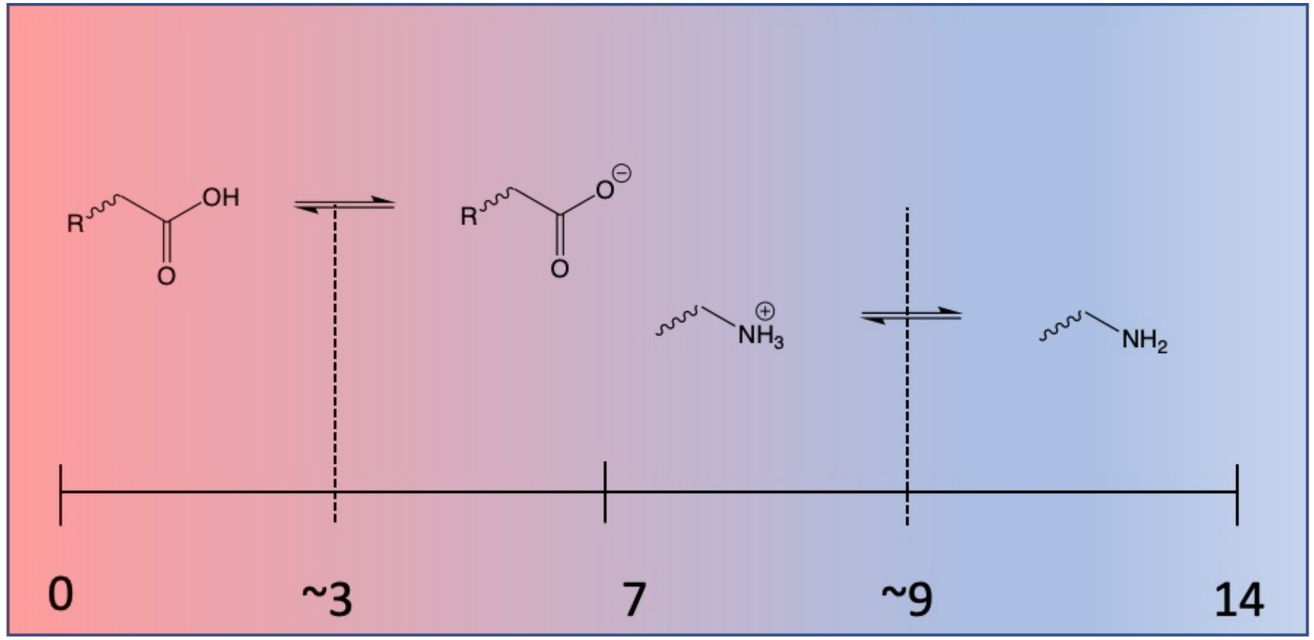


Προτονατεδ



We need to worry about pKa to decide protonation

- pKa: pH at which an acidic/basic group is 50% protonated/deprotonated.
- pKas are not fixed things!
- “Standard” values refer to the situation when the group is in dilute aqueous solution.
- Groups buried in the centre of hydrophobic proteins or close to other charged groups can show large pKa shifts.

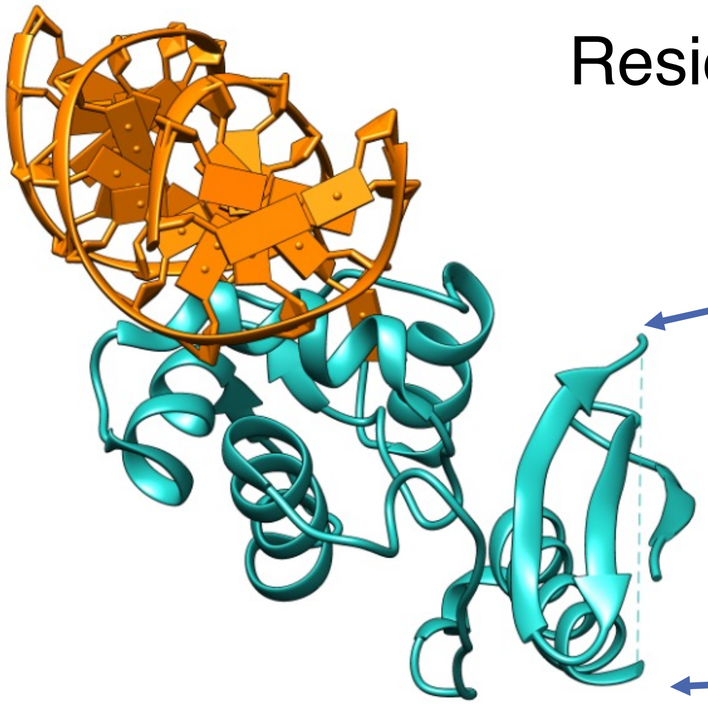


Amino acids that need protonation state consideration

Amino acid	pKa	options	Significance*
Aspartic acid	3.65	-COOH instead of COO^-?	possible
Glutamic acid	4.25	-COOH instead of COO^-?	possible
Histidine	6.00	protonated instead of neutral?	very possible
Cysteine	8.18	$-S^-$ instead of SH?	very possible
Tyrosine	10.07	$-O^-$ instead of OH?	possible
Lysine	10.53	$-NH_2$ instead of NH_3^+?	possible
Arginine	12.48	neutral instead of protonated?	unlikely

*for a simulation around physiological pH

What if the best structure has missing residues?



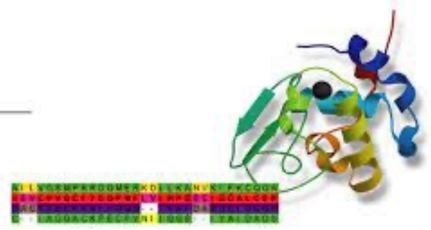
Residues 36 to 55 missing

```
REMARK 465 MISSING RESIDUES
REMARK 465 THE FOLLOWING RESIDUES WERE NOT LOCATED IN THE
REMARK 465 EXPERIMENT. (M=MODEL NUMBER; RES=RESIDUE NAME; C=CHAIN
REMARK 465 IDENTIFIER; SSSEQ=SEQUENCE NUMBER; I=INSERTION CODE.)
REMARK 465
REMARK 465   M RES C SSSEQI
```

```
REMARK 465     LYS A    36
REMARK 465     GLY A    37
REMARK 465     THR A    38
REMARK 465     SER A    39
REMARK 465     ALA A    40
REMARK 465     ALA A    41
REMARK 465     ASP A    42
REMARK 465     ALA A    43
REMARK 465     VAL A    44
REMARK 465     GLU A    45
REMARK 465     VAL A    46
REMARK 465     PRO A    47
REMARK 465     ALA A    48
REMARK 465     PRO A    49
REMARK 465     ALA A    50
REMARK 465     ALA A    51
REMARK 465     VAL A    52
REMARK 465     LEU A    53
REMARK 465     GLY A    54
REMARK 465     GLY A    55
```

Modeller

Program for Comparative Protein Structure Modelling by Satisfaction of Spatial Restraints

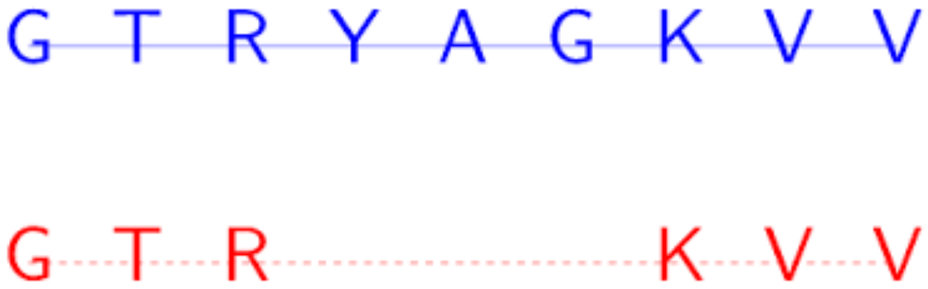


PDB FIXER

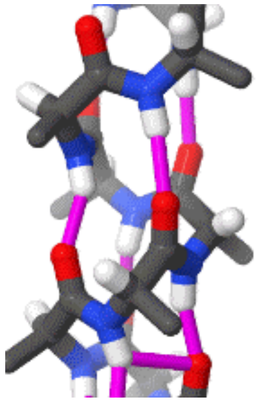
AlphaFold

AlphaFold2 structures have no missing residues or atoms

AlphaFold



No missing residues



Has hydrogens

AlphaFold2 structures can have unreliable areas

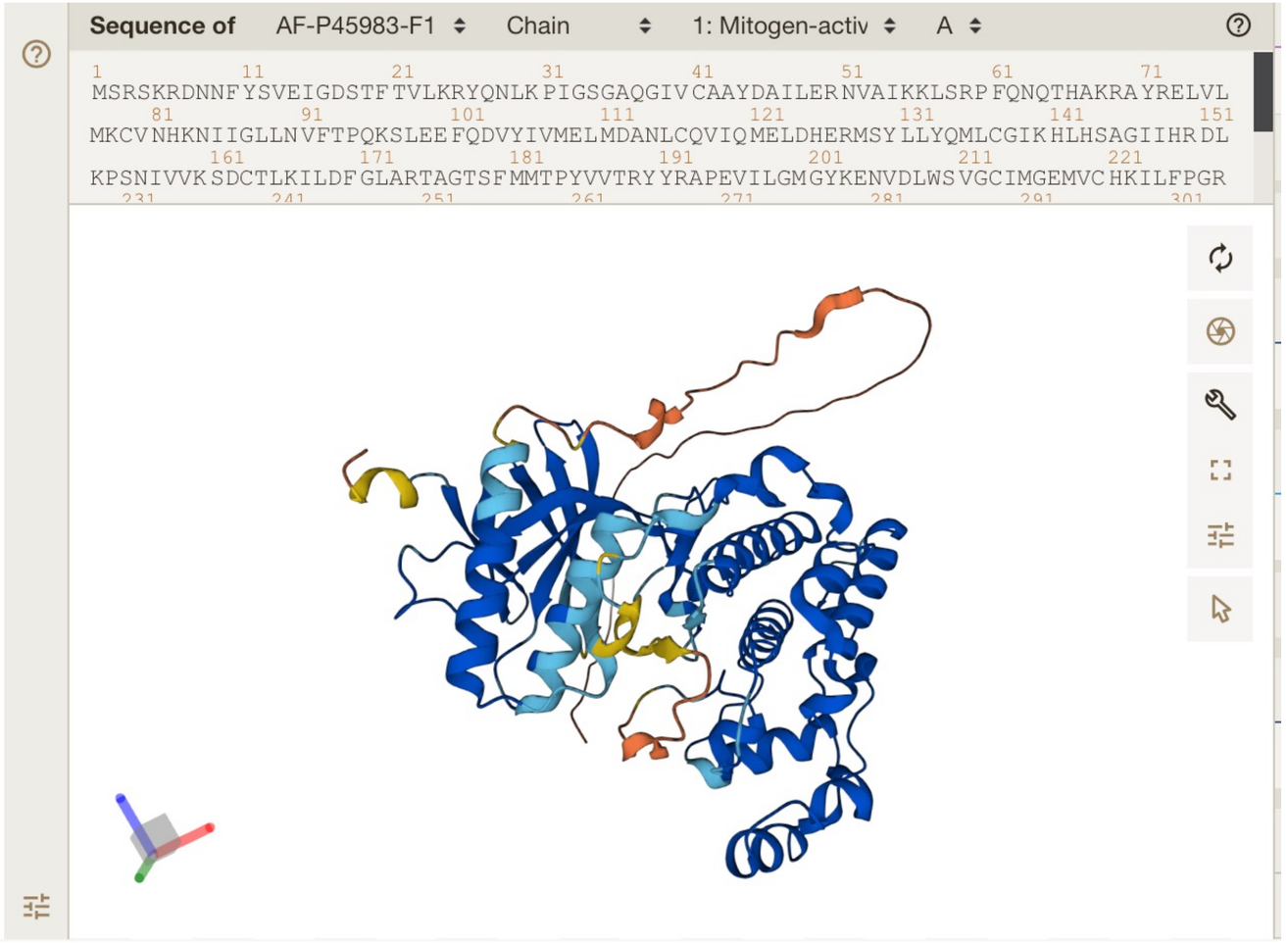
JNK1 — again

3D viewer

Model Confidence:

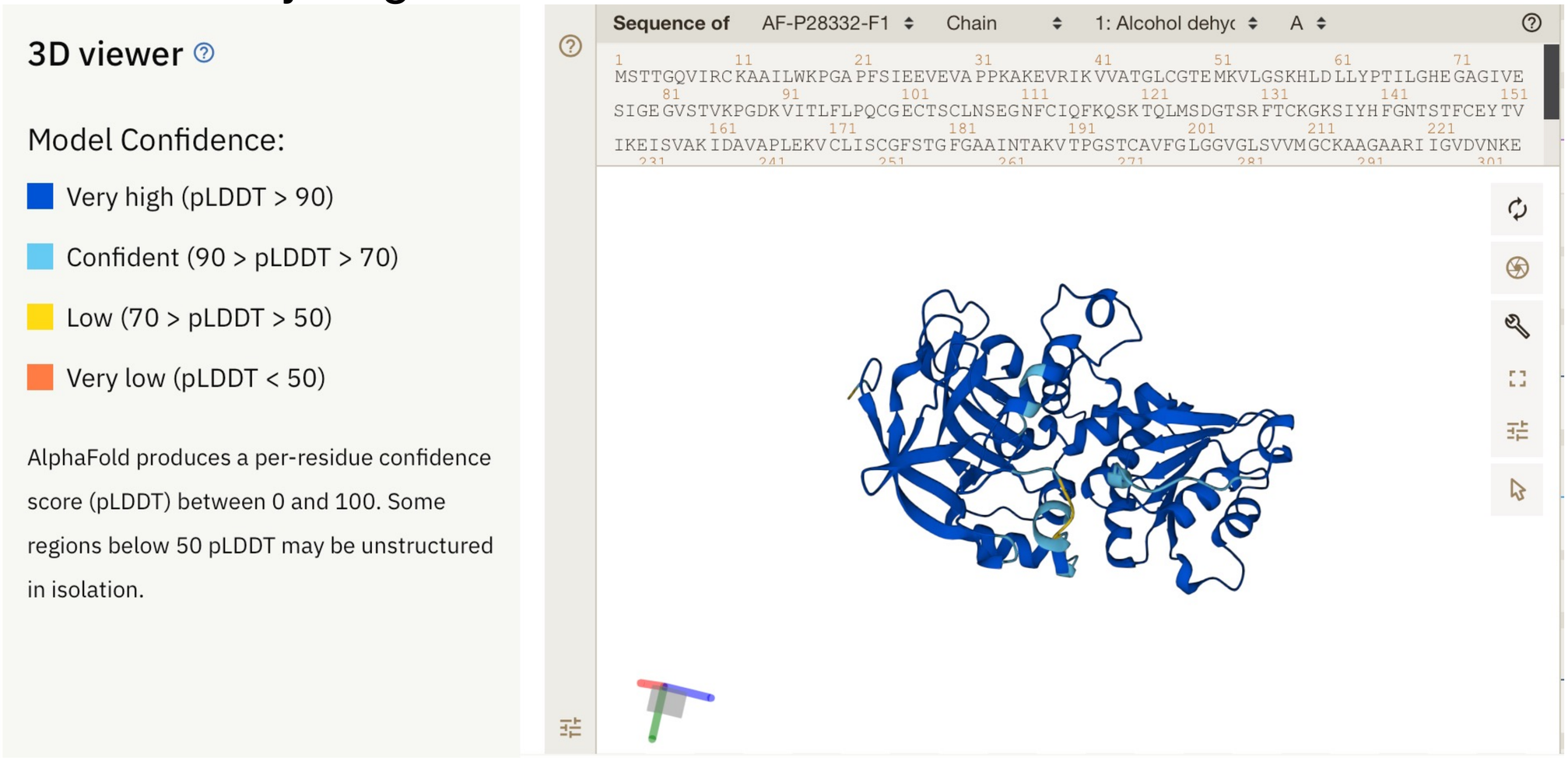
- Very high (pLDDT > 90)
- Confident (90 > pLDDT > 70)
- Low (70 > pLDDT > 50)
- Very low (pLDDT < 50)

AlphaFold produces a per-residue confidence score (pLDDT) between 0 and 100. Some regions below 50 pLDDT may be unstructured in isolation.



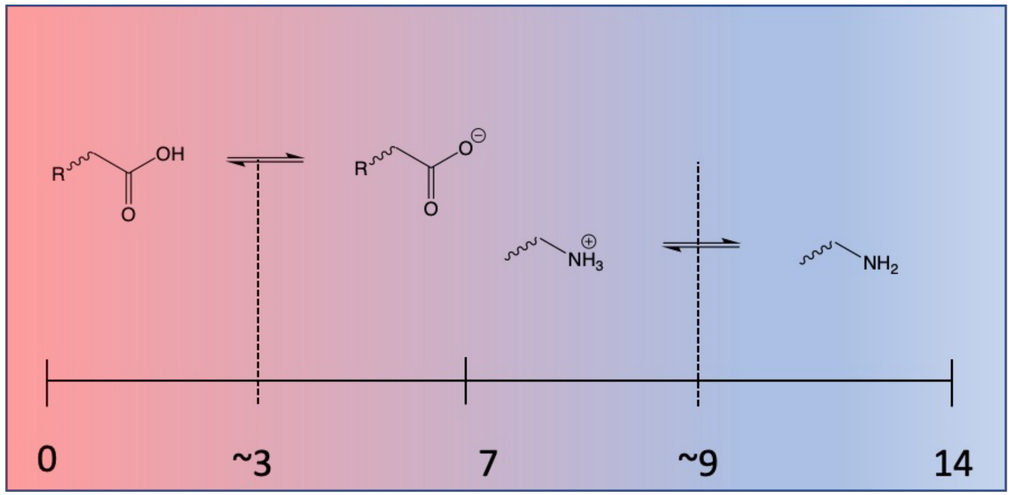
What is missing in this very good structure?

Alcohol dehydrogenase

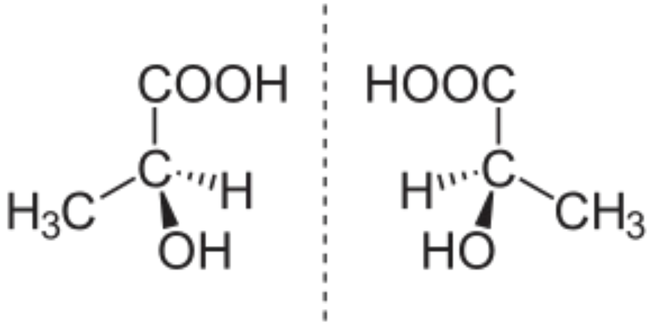


What about ligands and co-factors?

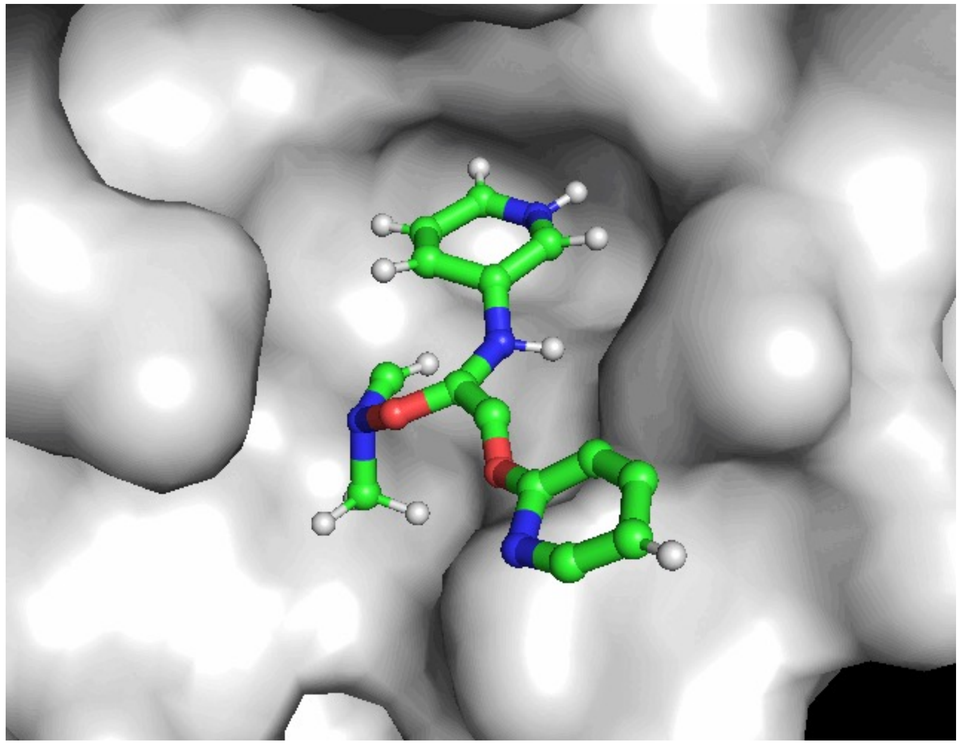
pKa for ligands is also important!



Choosing the right enantiomer



How do you get a ligand position when there is no crystal structure?



Trypsin: 1TRN

Task 1:
Your PhD supervisor has told you to go and simulate Trypsin and gave you 1TRN. Think about how to make choices in modelling

- Have a look at the protein 1TRN in the pdb and note down what you see and choices you make
- Stoichiometry (monomer or dimer protein?)
- PTMs (Does it have post translational modifications?)
- Non protein molecules (simulate or remove?)
- Disulphides (does it have disulphides?)
- Histidines (What will be the protonation state of histidines?)

https://www.rcsb.org/structure/1TRN

RCSB PDB | Deposit | Search | Visualize | Analyze | Download | Learn | About | Documentation | Careers | COVID-19 | MyPDB | Contact us

Biological Assembly 1

Explore in 3D: Structure | Sequence Annotations | Electron Density | Validation Report | Ligand Interaction (ISP)

Global Symmetry: Asymmetric - C1

Global Stoichiometry: Homo 2-mer - A2

Display Files | Download Files | Data API

1TRN

CRYSTAL STRUCTURE OF HUMAN TRYPSIN 1: UNEXPECTED PHOSPHORYLATION OF TYROSINE 151

PDB DOI: https://doi.org/10.2210/pdb1TRN/pdb

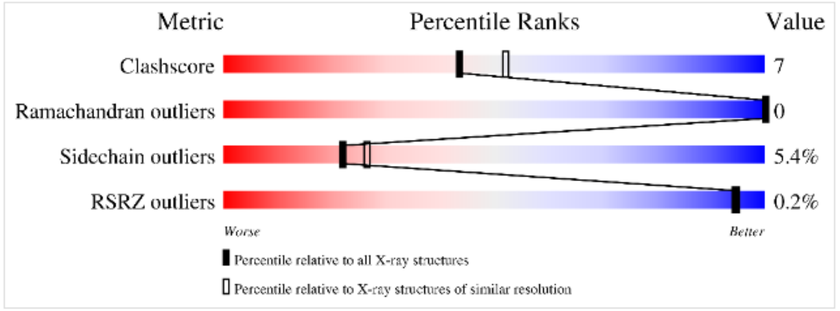
Classification: HYDROLASE (SERINE PROTEINASE)
Organism(s): Homo sapiens
Mutation(s): No

Deposited: 1995-03-16 **Released:** 1995-06-03
Deposition Author(s): Gaboriaud, C., Fontecilla-Camps, J.C.

Experimental Data Snapshot

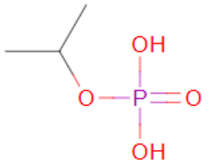
Method: X-RAY DIFFRACTION
Resolution: 2.20 Å
R-Value Work: 0.177
R-Value Observed: 0.177

wwPDB Validation | 3D Report | Full Report

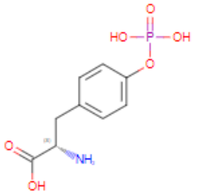


Small Molecules

Ligands 1 Unique

ID	Chains	Name / Formula / InChI Key	2D Diagram	3D Interactions
ISP Query on ISP Download Ideal Coordinates CCD File Download Instance Coordinates	C [auth A], D [auth B]	**PHOSPHORYLISOPROPANE** $C_3 H_9 O_4 P$ QPPQHRDVPBTVEV-UHFFFAOYSA-N		Interactions Interactions & Density

Modified Residues 1 Unique

ID	Chains	Type	Formula	2D Diagram	Parent
PTR Query on PTR	A, B	L-PEPTIDE LINKING	$C_9 H_{12} N O_6 P$		TYR

Find proteins for P07477 *(Homo sapiens)* Explore P07477 Go to UniProtKB: P07477

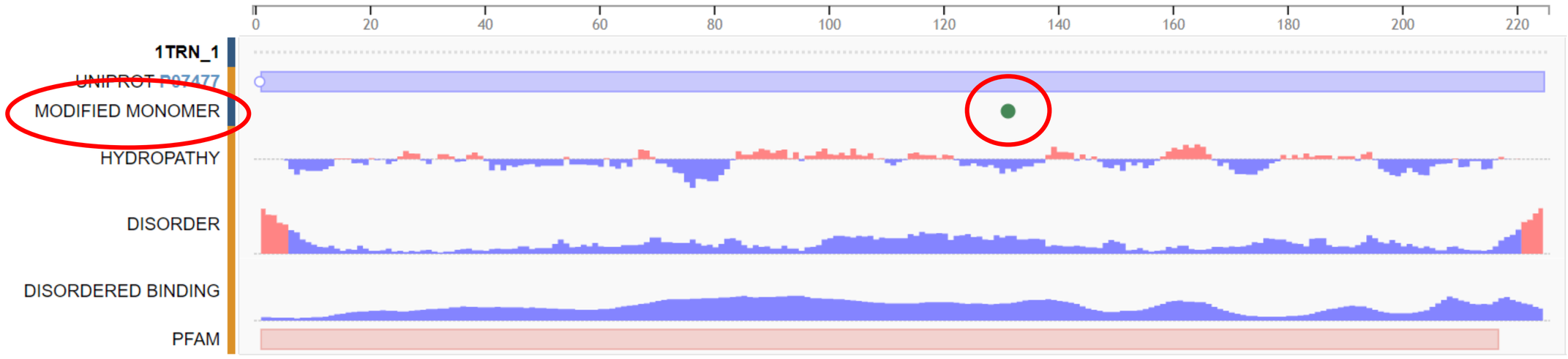
GTEx: ENSG00000204983

Entity Groups

Sequence Clusters	30% Identity 50% Identity 70% Identity 90% Identity 95% Identity 100% Identity
UniProt Group	P07477

Sequence Annotations Expand

Reference Sequence 1TRN_1



Protein	Serine protease 1	Amino acids	247 (go to sequence)
Gene	PRSS1	Protein existence	Evidence at protein level
Status	UniProtKB reviewed (Swiss-Prot)	Annotation score	5/5
Organism	Homo sapiens (Human)		

Function

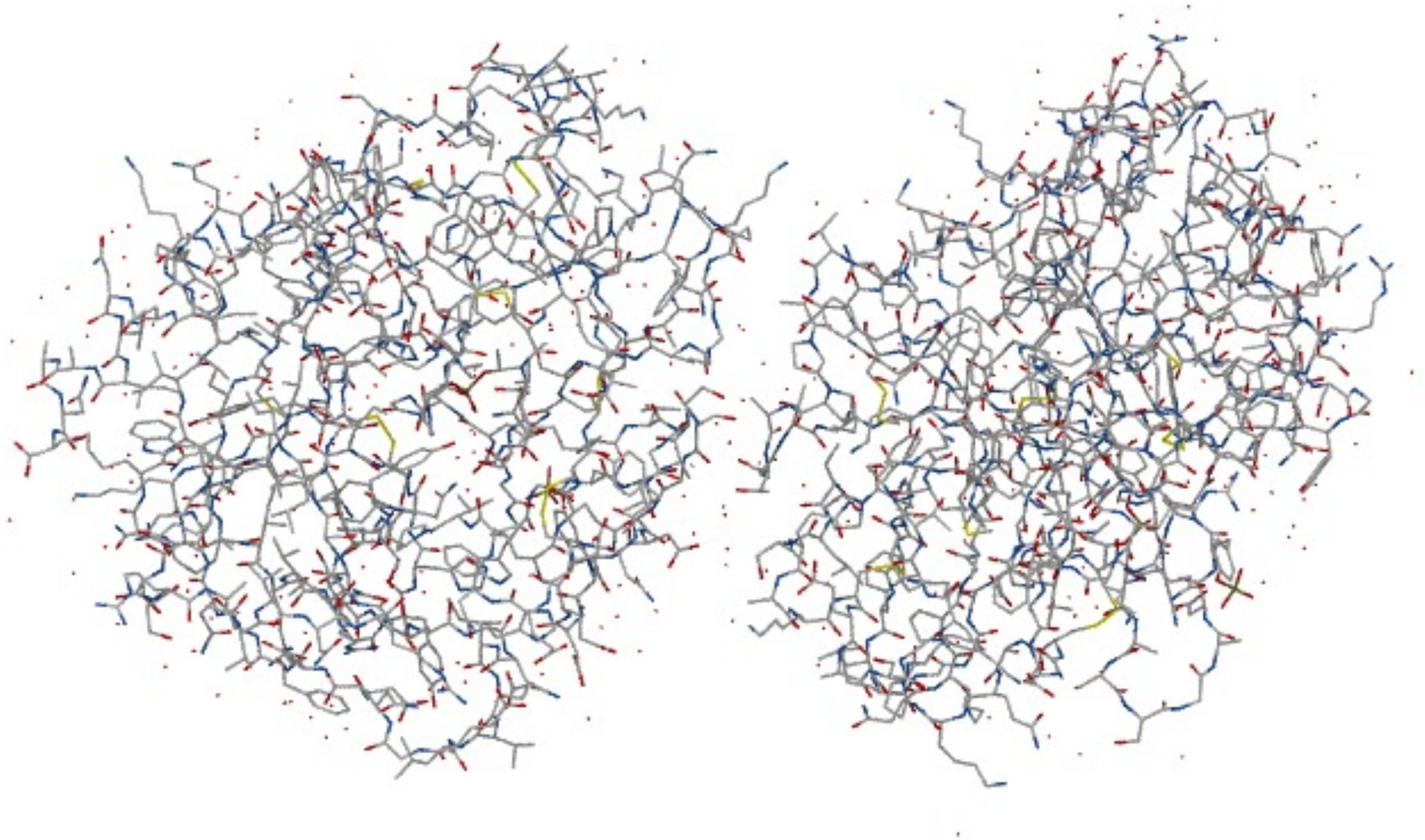
Has activity against the synthetic substrates Boc-Phe-Ser-Arg-Mec, Boc-Leu-Thr-Arg-Mec, Boc-Gln-Ala-Arg-Mec and Boc-Val-Pro-Arg-Mec. The single-chain form is more active than the two-chain form against all of these substrates.

1 Publication

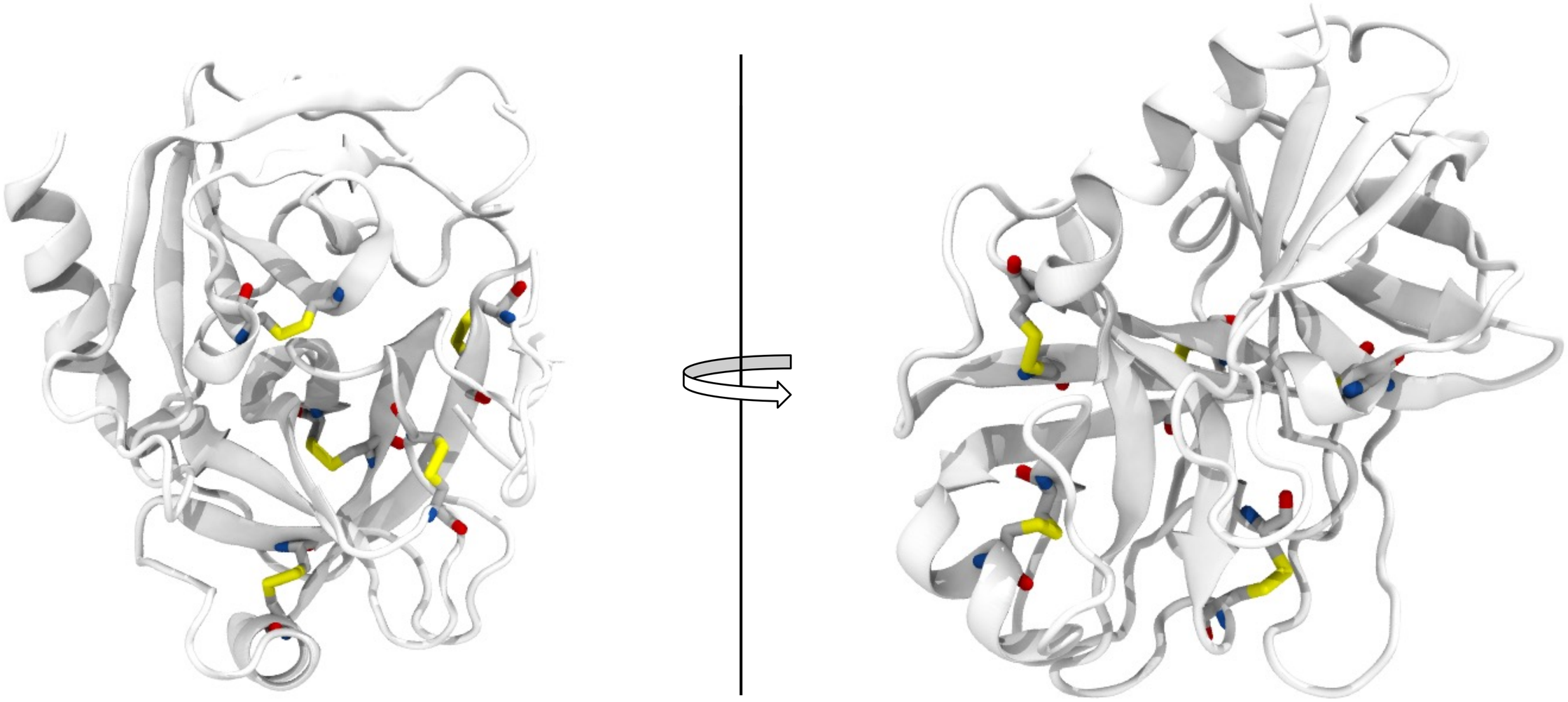
Caution

Tyr-154 was proposed to be phosphorylated (PubMed:8683601) but it has been shown (PubMed:17087724) to be sulfated instead. Phosphate and sulfate groups are similar in mass and size, and this can lead to erroneous interpretation of the results. 2 Publications

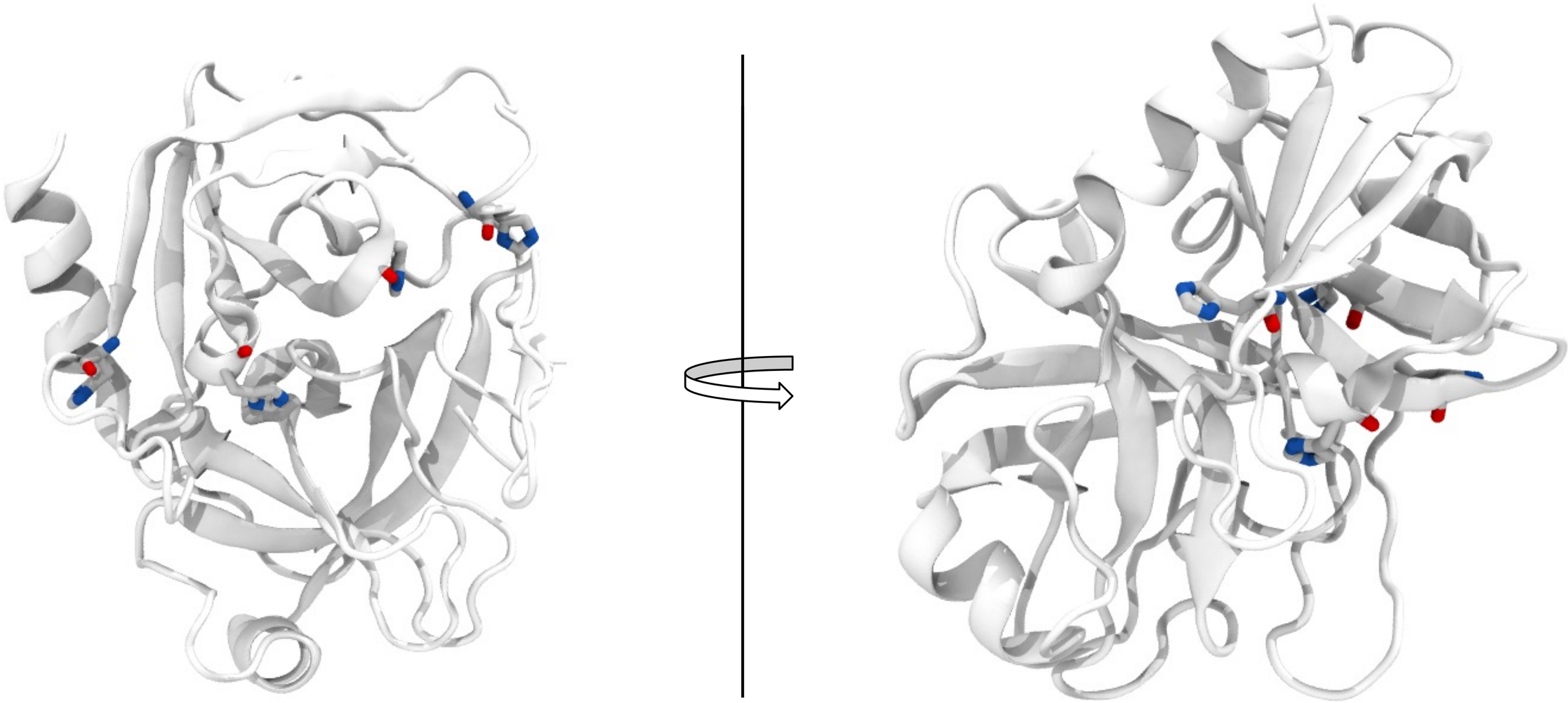
Non-protein molecules



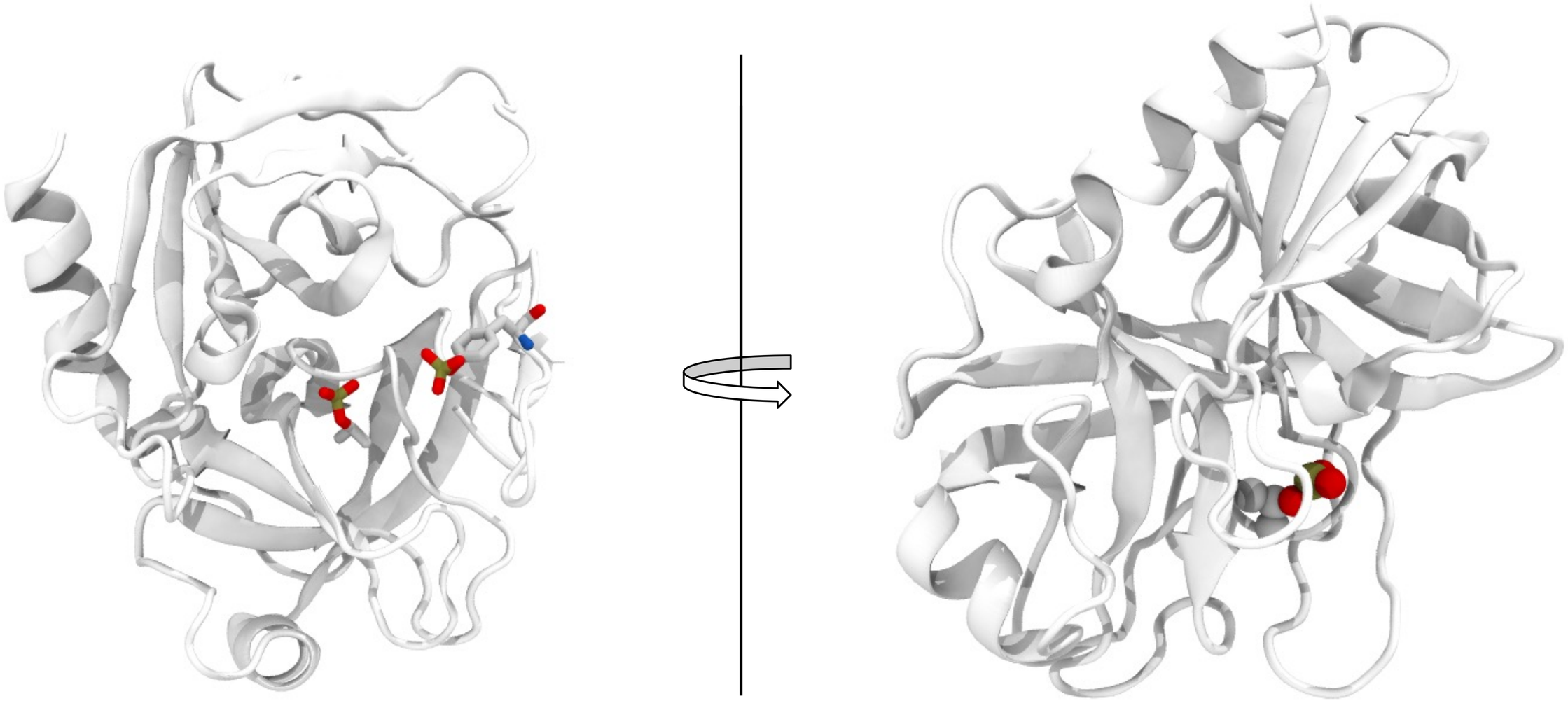
Disulphide bridges



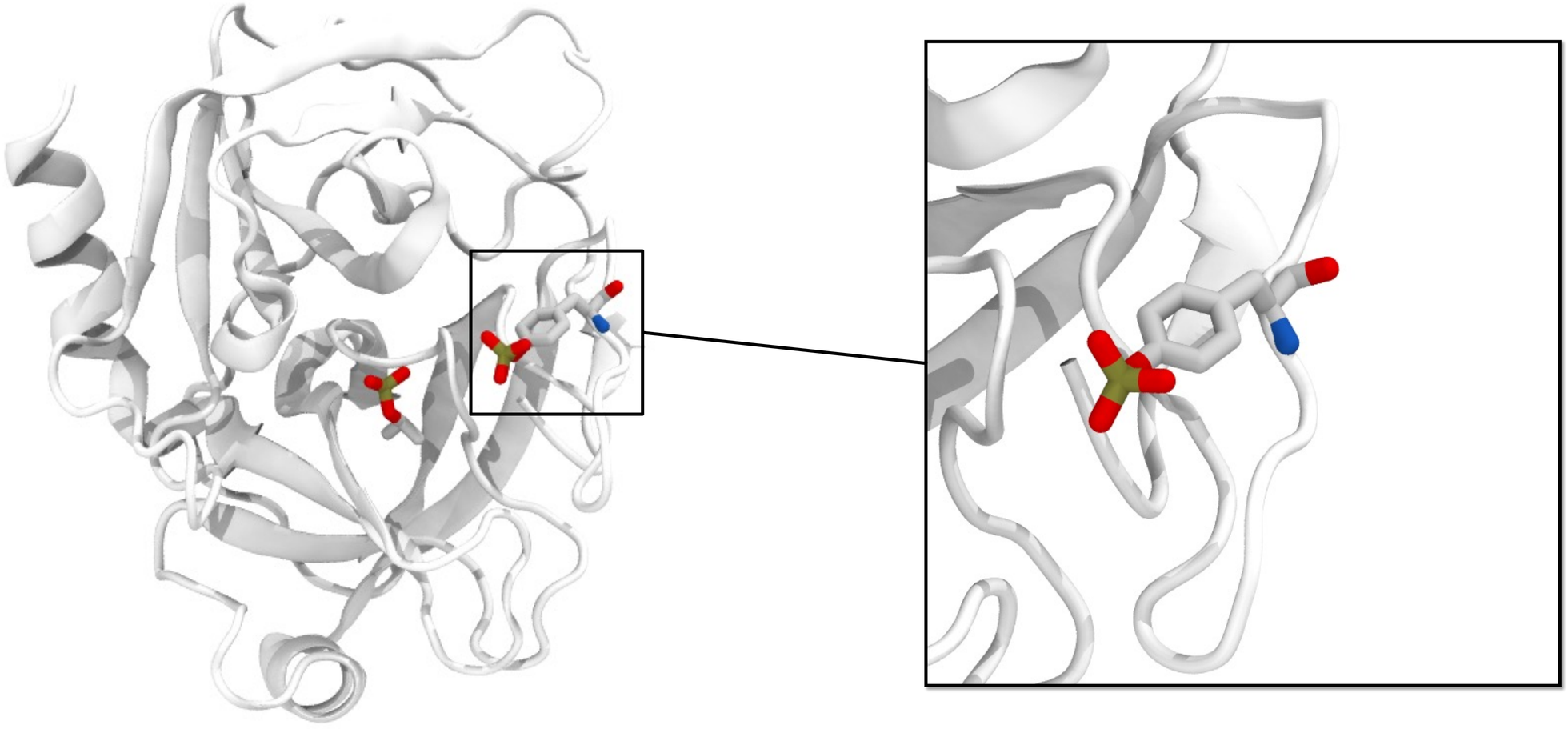
Histidines



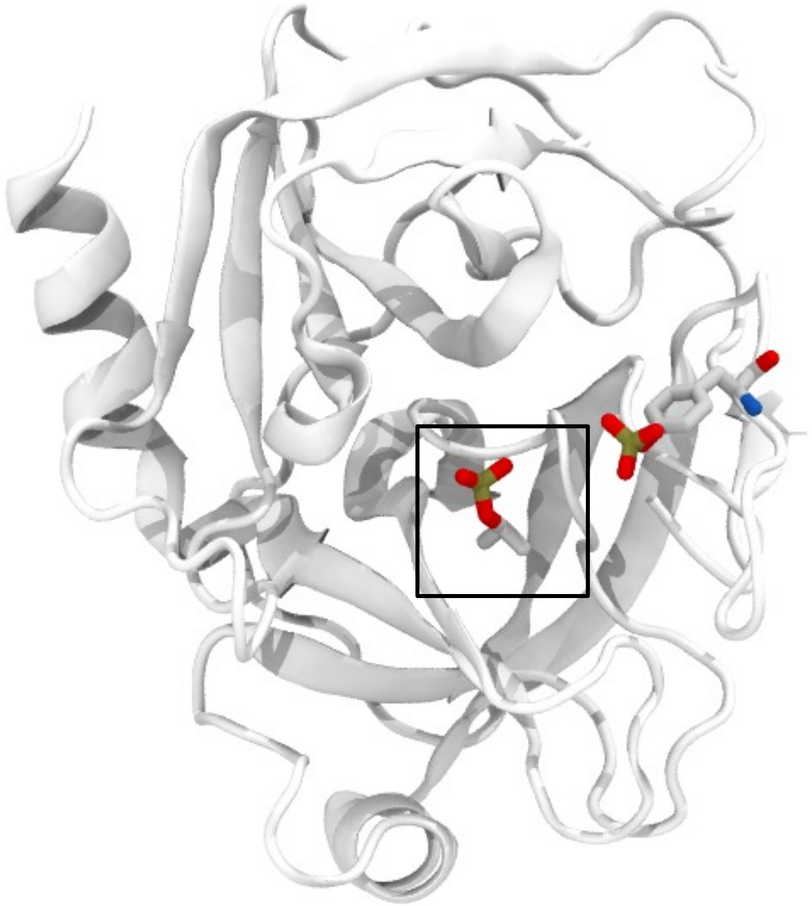
Amino acid modifications: TYR151



Amino acid modifications: TYR151



Ligand?



https://en.wikipedia.org/wiki/TAPS_(buffer)

WIKIPEDIA
The Free Encyclopedia

Create account Log in

TAPS (buffer)

8 languages

Article Talk | Read Edit View history Tools

From Wikipedia, the free encyclopedia

TAPS

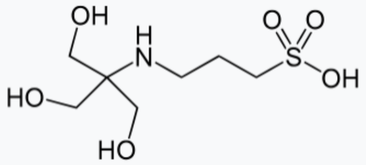
(**[tris(hydroxymethyl)methylamino]propanesulfonic acid**) is a chemical compound commonly used to make buffer solutions.

It can bind divalent cations, including Co(II) and Ni(II).[1]

TAPS is effective to make buffer solutions in the pH range 7.7–9.1, since it has a pK_a value of 8.44 (ionic strength I = 0, 25 °C).[2]

The pH (and pK_a at I ≠ 0) of the buffer solution changes with concentration and temperature, and this effect may be predicted e.g. using online calculators.[3]

TAPS



Names	
Preferred IUPAC name	3-{[1,3-Dihydroxy-2-(hydroxymethyl)propan-2-yl]amino}propane-1-sulfonic acid
Other names	*N*-Tris(hydroxymethyl)methyl-3-aminopropanesulfonic acid
Identifiers	
CAS Number	29915-38-6 ✓

Tools that will help with all of this

Most simulation packages contain tools to help add missing H-atoms. Not all work in the same way.

Where there is more than one possible answer, not all packages will **make the same decision**.

There are two independent, well-established tools designed specifically to look at these issues, and a webserver that puts them together:

Reduce

Adds hydrogens, and tests for 'NQH flips'

Download: http://kinemage.biochem.duke.edu/software/reduce.php

PropKa

Predicts amino acid protonation states

Download: https://github.com/jensengroup/propka-3.1

PDB2PQR server

Interface to both Reduce and proPka

Website: http://server.poissonboltzmann.org/pdb2pqr

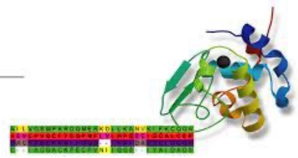
H++

OpenEye SCIENTIFIC

PDB FIXER

Modeller

Program for Comparative Protein Structure Modelling by Satisfaction of Spatial Restraints



...

Let’s try some protein prep out!

uniprot.org

rcsb.org

https://server.poissonboltzmann.org

Task 2:
Identify a good crystal structure for **Carboxypeptidase B2** and use the Poisson Boltzmann server to produce output for amber.
Starting structure PDB: 3D67 use uniprot to find out more!

Task 3:
Take a look at the AlphaFold 2 structure, what do you need to watch out for here?

Next: Docking