

# Tutorial

This tutorial explains how to use the **QuantyFey** application, step by step. It covers everything you need to get started and use it efficiently. QuantyFey is designed to quantify targeted LC-MS/MS data using external calibration, but can also be used with other data formats that include intensity and retention time values.

## Installation

The standalone version of this application runs on **Windows** and **Linux**. You can also run it directly from **R**, **RStudio**, or **VS Code**, which makes it compatible with **macOS** as well.

## Prerequisites

### Windows

- **RTools 4.2** is required:
  - **Option A:** Install from the [official CRAN page](#).
  - **Option B:** Use the included Portable R:
    - Navigate to **R-portable/bin/** and launch **R.exe**
    - Run the following in the R console:

```
install.packages("installr", repos = "https://cloud.r-project.org/")
installr::install.Rtools()
```

- Do not update the R version if prompted.
- Follow the installer instructions. You can ignore any non-critical error during the final steps.

### Linux

Make sure all system dependencies are installed:

```
sudo apt install -y cmake libcurl4-openssl-dev libssl-dev libfontconfig1-dev
libfreetype6-dev \
libharfbuzz-dev libfribidi-dev libpng-dev libjpeg-dev libtiff5-dev default-jdk
libtirpc-dev \
build-essential pkg-config

sudo R CMD javareconf
```

## Standalone Installation

- **Download** the latest version of [QuantyFey](#)
- **Unzip** the folder to a destination of your choice.

- **Run** the batch (Windows) or shell (Linux) file to start the App *Note: you may need to approve the execution of the script on Windows*
- A console will open and first all required packages will be installed automatically

### Installation for launching the app using RStudio, VS Code etc.

- **Download** the GitHub repository.
- **Unzip** the files to a destination of your choosing.
- **Install** [RStudio](#) or [VS Code](#)
- **Install** [pandoc](#)
- **Install** prerequisites as mentioned above.
- **Open** `app.R` and **run** the script

*Note: On first launch, the app uses the `renv` package to restore the required R package environment. This ensures compatibility by installing the correct package versions. This process may take up to 20 minutes.*

### Notes before using it on your own data

The application comes with several test datasets based on LC-MS/MS data. These are included to help you get started and explore the app's features.

**!! Important:** The app is preconfigured to work with these test datasets. This includes predefined **Calibration Standard Names** and their **Concentrations**. Before using your own data, you'll need to update this setup.

To adjust the configuration:

1. open the file at `Dependencies/templates.xlsx`
2. Add a new sheet or edit an existing one to define your own analysis setup.

**Template Structure Requirements** The template must follow this structure exactly:

- **Cal.Name:** The first column must be named **Cal.Name** (spelling and case-sensitive). It should list the **names of the Calibration Standards** used in your sequence.
- **Concentration:** The second column should specify the **concentration** for each corresponding **Calibration Standard**. You may also define concentrations at the **transition level** (e.g., different concentrations for every single transition). In that case:
- Every individual **transition name must be included in the template**.


The app uses this file to **map the concentrations** of the Calibration standards to their corresponding names in the dataset. The concentrations are used for compound quantification.

*Note: Standard names in the template must exactly match those in the sequence (case-sensitive). Note: if any standard that is defined in your data (Sample.Type column) is not included in this template, it will be appended with a concentration of 0 - but will not be included in the quantification process.*

	A	B
1	Cal.Name	Concentration
2	Cal 7	0.001
3	Cal 6	0.003
4	Cal 5	0.01
5	Cal 4	0.03
6	Cal 3	0.1
7	Cal 2	0.3
8	Cal 1	1

## Application Structure

The app is divided into multiple tabs, each with a specific function. This layout helps guide users through the analysis process step by step.

 Overview of main tabs of the QuantyFey Application

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### Data Upload

The **Data Upload** tab lets you import the two required data files for analysis. Two files must be uploaded in **CSV**, **TXT**, or **XLSX** format using standard delimiters ( , , ; , : , or tab \t ):

<

Upload Peak Area Data:

Choose File

Exar

Upload complete

Upload Retention Time Data:

Choose File

Exar

Upload complete

☒ Use Project Name

Project Name:

Results\_20250429

Reset App

## 1. Peak Table

This file contains **peak intensity data** (e.g., *Peak Areas* or *Peak Heights*) for the compounds of interest. The required structure is as follows:

- **Sample.Name**: Identifier for each sample.
- **Sample.Type**: Must be one of the following (case-sensitive):
  - **Sample** – Experimental samples.
  - **Standard** or **Cal** – Calibration standards.
  - **Blank** – Blank samples for background correction.
  - **QC** – Quality control samples.
- **Classification** (*optional*): Defines distinct sequence blocks for **custom bracketing analysis**.
  - Calibration standards must follow the pattern **Cal n** (e.g., **Cal 1**, **Cal 2**, ...).
  - Sample blocks may be named freely (e.g., **Sample Block 1**, **Block A**).
  - **Note**: If this column is missing, it will be auto-generated:
    - Calibration curves are identified as three or more consecutive standards.
    - Sample blocks are defined between these curves.
    - Samples before the first calibration are labeled **Pre 1**.

### !! Important

- Missing or incorrectly formatted columns will lead to an error message and upload will be halted.
- If internal standards are not recognized by the default pattern, a warning message will appear and internal standard correction features will be disabled. By updating the internal standard pattern these features can be restored.

### Example Table: Peak Table Format

Example file: [Example\\_Datasets/Example1\\_Drift\\_Areas.csv](#)

	A	B	C	D	E	F	G	H	I
1	Sample.Name	Sample.Type	Sample.ID	Classification	M243_N.Me thyl.Alanin e_104_42_q ual	M243_N.Me thyl.Alanin e_104_58_q uant	T245_Trime thylamine_ 60.1_60.1_q uant	T246_TMAO _76.1_58_q ual	T246_TMAO _76.1_59.1_ quant
2	MeOH 1	Blank	NA	Pre 1	10255.5464	108959.009	3667474.81	5426.59198	43691.4053
3	blank = water	Blank	NA	Pre 1	5794.61209	272365.6	3091312.6	10225.7659	26038.2619
4	IS Mix 1	Blank	NA	Pre 1	4706.36492	26872.7415	3737873.66	3597.72681	33083.9459
5	Cal 7	Standard	NA	Cal 1	7438.23749	N/A	3930873.07	31899.8731	21732.6687
6	Cal 6	Standard	NA	Cal 1	12207.8578	N/A	2898491.3	111190.963	82370.8499
7	Cal 5	Standard	NA	Cal 1	36543.4196	75488.7533	3404711.09	305090.393	224935.503
8	Cal 4	Standard	NA	Cal 1	99724.1491	98562.4676	3605492.73	1062254.52	705611.917
9	Cal 3	Standard	NA	Cal 1	210026.008	271913.567	3261862.73	2271598.58	1827749.52
10	Cal 2	Standard	NA	Cal 1	610552.72	778302.887	2974493.06	7395806.51	5287018.35
11	Cal 1	Standard	NA	Cal 1	953131.675	1529100.97	4584337.16	13016067.9	9596643.72
12	blank = water	Blank	NA	Block 1.1	5363.1439	191511.319	2571467.32	7558.07531	35154.444
13	MeOH 2	Blank	NA	Block 1.1	6201.44666	93052.577	3245695.64	3616.24053	20567.3801
14	QC 1	QC	NA	Block 1.1	199221.315	170507.221	3561925.44	308367.77	187637.449
15	P78	Sample	NA	Block 1.1	226582.464	222706.867	3341864.37	81202.8218	50567.8587
16	P69	Sample	NA	Block 1.1	319484.197	336831.342	3827932.76	567678.735	359390.637
17	P40	Sample	NA	Block 1.1	202608.311	179986.134	3660065.3	235778.611	132486.92
18	P64	Sample	NA	Block 1.1	128200.212	136070.709	4486660.79	58355.8542	22917.1177
19	P41	Sample	NA	Block 1.1	195994.381	169799.386	3288155.07	588918.429	412641.997
20	P79	Sample	NA	Block 1.1	188640.597	212238.701	2993025.08	352120.909	191910.208
21	P43	Sample	NA	Block 1.1	167790.648	179064.338	3015652.22	446134.004	359386.363
22	P72	Sample	NA	Block 1.1	200395.312	223152.849	2929443.8	429402.408	284692.865
23	Cal 3	Standard	NA	Block 1.2	175858.607	264131.234	2778859.68	1824967.15	1222004.31
24	IS Mix 2	Blank	NA	Block 1.2	4072.41018	N/A	2853684.28	N/A	19913.9868
25	P77	Sample	NA	Block 1.2	166659.254	226842.212	1996351.51	359183.028	255466.264
26	P52	Sample	NA	Block 1.2	218662.68	232794.99	2915251.61	94931.4427	74001.3363
27	P60	Sample	NA	Block 1.2	194972.314	168947.506	2895212.8	34268.6446	9819.30281
28	P73	Sample	NA	Block 1.2	239703.755	195228.407	1588943.29	116288.951	67118.6296
29	P53	Sample	NA	Block 1.2	131636.69	140929.188	2155004.4	58286.1371	36953.8689
30	P68	Sample	NA	Block 1.2	170315.432	165608.355	2508369.94	223913.326	110380.734
31	P66	Sample	NA	Block 1.2	155628.292	166203.918	3036155.16	153714.067	100077.969
32	P82	Sample	NA	Block 1.2	191681.894	200862.579	2559000.9	122620.964	62419.8588
33	Cal 3	Standard	NA	Block 1.3	157891.215	268496.178	2406119.29	1544396.39	966874.039

## 2. Retention Time Table

This file provides **retention time (RT)** data for the compounds.

- Must include a **Sample.Name** column identical to that in the Peak Table.
- Only compounds present in the Peak Table will be considered.
- Upload the RT Table **after** the Peak Table.

*Note:*\* the app will ignore any additional columns that aren't needed.

### Example Table: Retention Time Table Format

Example file: [Example\\_Datasets/Example1\\_Drift\\_RT.csv](#)

	A	B	C	D	E	F	G	H
1	Sample.Name	Sample.Type	Sample.ID	M243_N.Me thyl.Alanin e_104_42_q ual	M243_N.Me thyl.Alanin e_104_58_q uant	T245_Trime thylamine_ 60.1_60.1_q uant	T246_TMAO _76.1_58_q ual	T246_TMAO _76.1_59.1_ quant
2	MeOH 1	Unknown	NA	0.25926921	0.20881508	0.18447828	0.16684883	0.17938056
3	blank = water	Unknown	NA	0.25708283	0.2445533	0.37306067	0.20626313	0.66467404
4	IS Mix 1	Unknown	NA	0.26012177	0.22631998	0.1835997	0.151439	0.19140657
5	Cal 7	Unknown	NA	0.255808	N/A	0.1817455	0.25991771	0.26471666
6	Cal 6	Unknown	NA	0.25895029	N/A	0.18890418	0.26368089	0.26336981
7	Cal 5	Unknown	NA	0.2562293	0.23400076	0.18400052	0.26279085	0.26494092
8	Cal 4	Unknown	NA	0.25997888	0.25843157	0.18454699	0.26324391	0.26202178
9	Cal 3	Unknown	NA	0.25895655	0.26190355	0.17634002	0.26274974	0.26388911
10	Cal 2	Unknown	NA	0.25851859	0.26391956	0.18691936	0.26305417	0.26361763
11	Cal 1	Unknown	NA	0.26068388	0.26858329	0.22075668	0.26486155	0.26525941
12	blank = water	Unknown	NA	0.2581849	0.24304482	0.17426189	0.25472888	0.16397883
13	MeOH 2	Unknown	NA	0.26267049	0.24608549	0.18265215	0.14628303	0.15966025
14	QC 1	Unknown	NA	0.26252305	0.2642646	0.17816692	0.27094407	0.26978924
15	P78	Unknown	NA	0.26454516	0.26593551	0.1782392	0.27221292	0.27172804
16	P69	Unknown	NA	0.26540968	0.26666039	0.17498445	0.27239636	0.271831
17	P40	Unknown	NA	0.26177872	0.26213872	0.18077401	0.2716874	0.26891411
18	P64	Unknown	NA	0.25936649	0.2619704	0.16059253	0.26707984	0.27217436
19	P41	Unknown	NA	0.25778883	0.2586956	0.1793193	0.2699452	0.26931156
20	P79	Unknown	NA	0.26179437	0.26470812	0.17846981	0.27086952	0.27043151
21	P43	Unknown	NA	0.26218013	0.2648908	0.18096375	0.26915383	0.27066371
22	P72	Unknown	NA	0.26426757	0.26604387	0.17655518	0.26868131	0.26849529
23	Cal 3	Unknown	NA	0.2592905	0.26222526	0.17326083	0.26330201	0.26391545
24	IS Mix 2	Unknown	NA	0.2612305	N/A	0.16513071	N/A	0.16886235
25	P77	Unknown	NA	0.26380933	0.26635102	0.18759601	0.27008585	0.27117233
26	P52	Unknown	NA	0.26361245	0.26459516	0.17305974	0.27086855	0.26999416
27	P60	Unknown	NA	0.26301135	0.26270627	0.18134947	0.27032375	0.26485716
28	P73	Unknown	NA	0.26402593	0.26480116	0.19025931	0.27041182	0.26907487
29	P53	Unknown	NA	0.26265458	0.26458979	0.18097702	0.27016662	0.26981335
30	P68	Unknown	NA	0.26118892	0.26223379	0.17228428	0.26910705	0.27016214
31	P66	Unknown	NA	0.26423223	0.26664551	0.16902463	0.27201638	0.27055547
32	P82	Unknown	NA	0.26492555	0.26489323	0.17105257	0.26994948	0.27056762

### 3. Project Name (optional)

You may optionally enter a **Project Name**, which will be used to label output folders.

- Output will be saved in the user's **Documents** folder under:

`Documents/QuantyFey/<ProjectName>/`

### 4. Reset the Application

Click **Reset App** to restart the session and clear all uploaded data — useful if the wrong files were selected or you want to begin a new analysis.

Summary: Minimum Upload Requirements

File	Required Columns	Notes
Peak Table	Sample.Name, Sample.Type	Optional: Classification; supports multiple transitions.
RT Table	Sample.Name, transitions from Peak Table	Must match Peak Table sample names.

Configure Settings:

Selecting Quantification Template

To perform quantification, you need to select the appropriate template. Use the dropdown menu in the app to choose from available sheets in the templates.xlsx file. Each sheet represents a separate template. When selected, the left pane will be updated with the respective template. If the template encompasses all transition names, it will show the concentrations of the selected compound.

\*Make sure the correct template is selected - quantification will not work if the template does not match your data or the wrong sheet is chosen.

Change patterns

QuantyFey automatically detects Quantifier, Qualifier, and Internal Standard (IS) transitions by searching for predefined patterns in the column names of the Peak Table. These patterns can be customized to match your dataset's naming. Patterns can be changed by selecting the Change Patterns Checkbox. Then three inputs will be shown, that can be adjusted.

**Tip:** You can set default values by modifying the default\_settings.R file, or override them directly within the app interface.

Default Pattern Setup (in default\_settings.R)

Below are the default settings used by the app. You can change them in the default\_settings.R file.

Setting	Default Value	Description
quant_pattern	_quant	Identifies quantifier transition columns
qual_pattern	_qual	Identifies qualifier transition columns
IS_pattern	IS	Identifies internal standard transition columns
conc_unit*	µg/mL	Unit used for concentration in plots and tables
int_unit*	counts	Unit used for intensity values in outputs
rt_unit*	min	Unit used for retention time
Template_name	Example1	Default name of the template used for quantification setup



\* These unit values can only be adjusted in the default\_settings.R file and are not editable from within the app interface.

Code (as seen in `default_settings.R`)

```
## Setup Default Settings for QuantyFey

# Default Template name
Template_name = "Example1"
# Pattern for Quant Transition:
quant_pattern = "_quant"
# Pattern for Qual Transition:
qual_pattern = "_qual"
# Pattern for IS Transition:
IS_pattern = "IS"

#### ---- Units ---- ####
# Set to NULL if you don't want to show the unit in the plot
# Concentration
conc_unit = "µg/mL"
# Intensity Unit
int_unit = "counts"
# RT Unit
rt_unit = "min"
```

*Note: Updating the units helps ensure consistency between your dataset and the visualization/output displays in the app.*

## Pattern for Quantifier Transitions

This pattern identifies **quantifier** transitions from Peak Table column names.

- Supports **regular expressions** (*Regex*)
- Only columns matching this pattern (and **not** matching the IS pattern) will be used for quantification.

### Examples:

- `^Compound1_` → Columns starting with `Compound1_`
- `_quant$` → Columns ending with `_quant`
- `Compound[0-9]+_` → Matches names like `Compound1_`, `Compound2_`, etc.
- `.*_qual` → Columns containing `_qual`
- `^Cal.*ppb$` → Columns starting with `Cal` and ending with `ppb`

### !! Important: Understanding Regular Expressions

- `.` matches any single character.
- `*` means zero or more of the previous character.
- `^` anchors the match to the start of the text.



- `$` anchors the match to the end of the text.
- `.*` matches any number of any character.
- To match a literal `.` or `*`, escape it with a backslash: `\.` or `\*`.
- For more information check out this (Regex Quick Reference)[https://hypebright.nl/en/r-en/ultimate-cheatsheet-for-regex-in-r-2/]

## Pattern for Qualifier Transitions

Defines how **qualifier** transitions are detected.

- Also uses **regular expressions**
- Matches are determined based on the **prefix** of the corresponding quantifier transition.

### Prefix Matching Logic:

- The **prefix** is the string before the first underscore (`_`)
- Example formats:
  - `CompoundID_Q1_Q3_CE_quant`
  - `CompoundID.Q1_Q3_CE_qual`

If no matching qualifier transitions are found, the **Qualifier/Quantifier Ratio Analysis** tab will be hidden.

## Pattern for IS Transitions

This pattern identifies **internal standard** transitions.

- Supports **regular expressions**
- Matching columns will:
  - be **excluded** from quantification
  - be **used** for IS correction (if enabled)

If no IS transitions are detected, IS correction will be disabled automatically and a warning message will appear.

 Overview of the Configure Settings graphical output

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## Compound Quantification

The **Compound Quantification** tab is the main workspace for **visualization**, **drift correction**, **regression model optimization**, and **quantification**.

### Setup

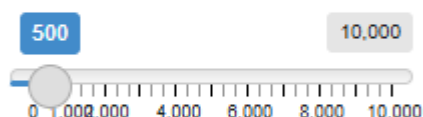
Compound:

M243\_N.Methyl.Alanine\_10

Internal Standard:

IS\_L.Histidine\_296.1\_115.1

Intensity Threshold:



Comment:

Save

☐ Generate Report

In the left-side panel, users can configure the following:

- **Compound:** Select the quantification transition.
- **Comment:** Add notes for the quantification process.
- **Save:**
  - Output is saved to users **Documents** folder
  - If the default path fails (e.g., due to permissions), results are saved to the local QuantyFey directory.
  - Files are placed inside a folder names **Results\_<date>** unless a different name is specified in the **Data Upload** tab.
  - Two output files are created and updated with each save:
  - A **concentration table** (**Results\_quant.csv**) per compound.
  - A **summary file** (**Quant\_summary.xlsx**) listing all parameters and settings, with one sheet per save.
- **Generate Report:** Optional checkbox. If enabled, a PDF report will be created containing all plots and a summary of the quantification.


Not on File Management: To avoid overfitting while limiting file clutter:


- **Concentration tables:** The app checks for differences between the existing file and the new data. If all data from the existing file matches data from the new data, the file is overwritten. If differences exist, a new file is created with a time stamp.
- **Summary File:** With every save, data is appended to the file.
- **PDF Report:** Reports may be overwritten if the app is restarted and outputs are saved to the same directory. Suggestion: After launching the app, create a new output folder by renaming the project name in the **Data Upload** tab. This helps prevent conflicts and ensures all outputs are saved cleanly.


## Main Tabs


The main panel contains five tabs:

 **Data Visualization**

 **Drift Correction**

 **IS Correction**


 **Bracketing**

 **Quantitation**

Each tab will be explained in detail in the following section.

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## Data Visualization

 Overview of the Data Visualization Tab showing the Qualifier Quantifier Ratio Plot

This tab provides an overview of the data for the selected transitions, including **Retention Time (RT)**, **Ion Ratio Analysis**, and **Blank Analysis**.

- **Retention Time:** Interactive plot of RT values for selected sample types. Hover over any point to view sample-specific details.
- **Ion Ratio Analysis:** Interactive plot of Qualifier-to-Quantifier Ion ratios for selected sample types. Hover over any point to view sample-specific details.

Note: This section is only visible if a matching Qualifier was found for the selected Quantifier transition.

- **Blank Analysis:** Boxplots comparing blank and sample signal ratios. Blank ratios are calculated as:  $\text{sample intensity} / \text{mean blank intensity}$ . This helps distinguish background noise from true signals.

This visualization tab is designed to help **verify compound identity** by comparing **RT**, and **Ion Ratios** between Samples and Standards and to compare signal intensities from Samples to Blanks to assess **carry**

## over and background signals.

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### \*\* Correcting for Intensity Drift\*\*

The following four tabs are used to set up different drift correction strategies. For best results, it's recommended to configure all applicable methods **before** performing quantification.

#### IS Correction

This module performs internal standard correction by calculating intensity ratios:

$$\text{IS ratio} = \text{Peak Area of Quantifier} / \text{Peak Area of IS}$$

These ratios are then used instead of raw intensities for both the regression model and concentration calculations.

 Overview of the Correction Factors Table for the IS Correction

- **Plots:**
  - **Raw Intensity:** Bars indicate raw intensities; red dots indicate IS intensities.
  - **IS Ratios:** Displays IS ratios.

**Note:** For IS intensities falling below 0.1 % of the median value, the median value is used for correction.

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#### Drift Correction

This module corrects signal drift based on QC samples or other repeated injections. It builds a model based on the injection order and intensity values. This model is used to correct for intensity drift by normalizing intensities to the model.

 Overview of the Drift Correction Parameters

- **Models:**
  - **Linear Model (lm):** Simple linear regression.
  - **Loess:** Non-linear locally estimated scatterplot smoothing.
  - **Spline:** Flexible non-linear regression using spline curves.
- **Sample for Drift Correction:**
  - Select a sample (e.g., QC) injected regularly throughout the sequence. Only non-blank samples injected **3+ times** can be selected.
- **Span Width** (for loess only): Controls smoothness of the loess fit.
- **Degree** (for spline only): Specifies the degree of the spline function.

The main panel displays:

- **Raw Intensity Plot:** Intensity values before correction.
- **Corrected Intensity Plot:** Intensity values after drift correction.

**Note:** Loess models cannot extrapolate: Edge values are estimated using the nearest available data point. Span width must be above 0.4. Smaller values may cause errors. If an error appears, adjust the

parameters and retry. Span width cahnges are only applied after clicking outside the input box.

Custom Bracketing

This tab allows you to manually assign **calibration blocks** to specific sections of your sequence. These are derived from the **Classification** column in the **Peak Table**.

Bracketing

Class	Cal 1	Cal 2	Cal 3
Pre 1	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Cal 1	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Block 1.1	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
Block 1.2	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>
Block 1.3	<input type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>
Cal 2	<input type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>
Block 2.1	<input type="checkbox"/>	<input checked="" type="checkbox"/>	<input type="checkbox"/>
Block 2.2	<input type="checkbox"/>	<input checked="" type="checkbox"/>	<input checked="" type="checkbox"/>
Block 2.3	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/>
Cal 3	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/>

Bracketing Table:

- Rows = Sample blocks (from **Classification**)
- Columns = Calibration blocks (also from **Classification**)
- Checkboxes let you assign calibration blocks to sample blocks.

**Note:** Every sample block must be linked to **at least one** calibration block for bracketing to be used in quantification.

Weighted Bracketing

This module builds a separate **regression model for each injection**, based on its position between two calibration blocks. The weight assigned to each calibration point is determined by the selected model type:

- In the **linear model**, weights increase or decrease **linearly** across the injection sequence.
- In the **non-linear models** (requires technical replicates over the sequence), a model is fitted to their intensities over the measurement sequence. The **derivative** of this model is then used to define how

the weights change across the sequence.

### Settings:

- **Model Type**
  - **linear** - linear weight increase/decrease (*does not require QC samples*)
  - **non-linear** - use of technical replicates.
    - loess: adjustable **span width**
    - spline: adjustable **degree**

!(Overview of weighted bracketing in QuantyFey)[images/weighted\_bracketing.png]

**Note:** Weights using the linear model are just increasing/decreasing weights from one to the next calibration block. They don't require QC injections. Non-linear models require QC samples.

## Quantification

The **Quantification** tab is where the actual quantification of the selected transition takes place. The left panel allows configuration of key model parameters, while the main area provides tools for visualization, diagnostics, and results review.

### Parameters

Degree:

1

Block to Visualize

all

Limit of Quantification

0,001

Method for Weighing:

1/x

Method for Quantification

IS Correction

☒ Show Samples

Apply Cal Levels to All

Apply LLOQ to All

Optimize Model

- **Regression Model:** Choose between **linear** and **quadratic** regression models.
- **Limit of Quantification (LLOQ):** Defines the lower reporting threshold for concentration values.
  - Defaults to the lowest calibration standard.
  - Does not affect the plot, but any value below the LLOQ will be marked as "< LLOQ" in the output.
- **Weighting Method:** Specifies the regression weighting:
  - **1/x:** Weight = 1 / Concentration.
  - **1/x<sup>2</sup>:** Weight = 1 / Concentration<sup>2</sup>.
  - **1/y:** Weight = 1 / PeakArea
  - **1/y<sup>2</sup>:** Weight = 1 / PeakArea<sup>2</sup>
  - **1/x force 0:** Weight = 1 / Concentration and goes through 0|0
  - **1/y force 0:** Weight = 1 / PeakArea and goes through 0|0
  - **None:** No weighting applied.



**Note:** Values with PeakArea = 0 will automatically have a weight of 0 and will not be included in the regression.

- **Quantitation Method:** Selects the quantification approach:
  - **IS Correction:** One calibration function over all calibration blocks using IS ratios
  - **Drift Correction:** One calibration function over all calibration blocks using drift corrected peak areas.
  - **Custom Bracketing:** Individual calibration function assigned to each sample block.
  - **Weighted Bracketing:** Individual calibration function for all injection weighted according to the position in the sequence.
  - **Default Bracketing:** One regression function over all calibration blocks.
- **Show Samples:** Toggles sample visibility in plots.

#### Bracketing specific settings

(Visible only if Custom or Weighted Bracketing is selected as the quantitation method)

- **Block to Visualize:** Select which classification block to display in the plot. **Note:** in Weighted Bracketing, each sample is treated as its own block.
- **Apply Cal Levels to All:** Apply the currently selected calibration level settings to **all blocks**.

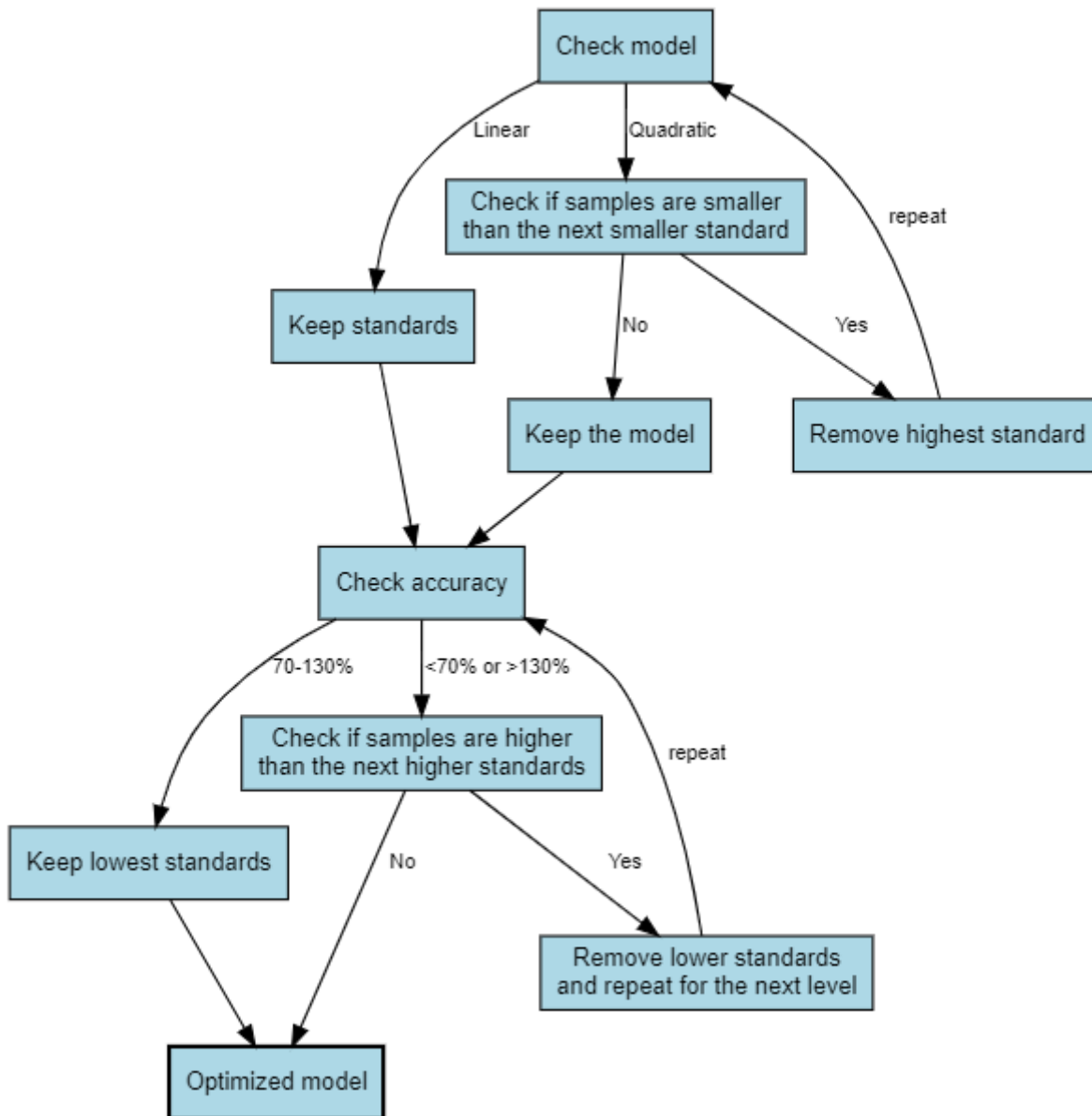
**Note:** If a calibration level has been removed in one block, this action will remove that level across all blocks.

- **Apply LLOQ to All:** Applies the defined LLOQ settings to all blocks.

An automatic optimization Button allows the user to do a generic optimization of the regression model.

- **Optimize Model:** Automates model optimization:
  - Removes higher standards for quadratic models if samples are lower.
  - Removes lower standards if accuracy falls outside 70–130% and samples are higher.
  - Selects linear or quadratic models based on a lack-of-fit test.

**Note:** Optimization may fail or produce poor fits for low-quality data. Always verify the model manually before saving.



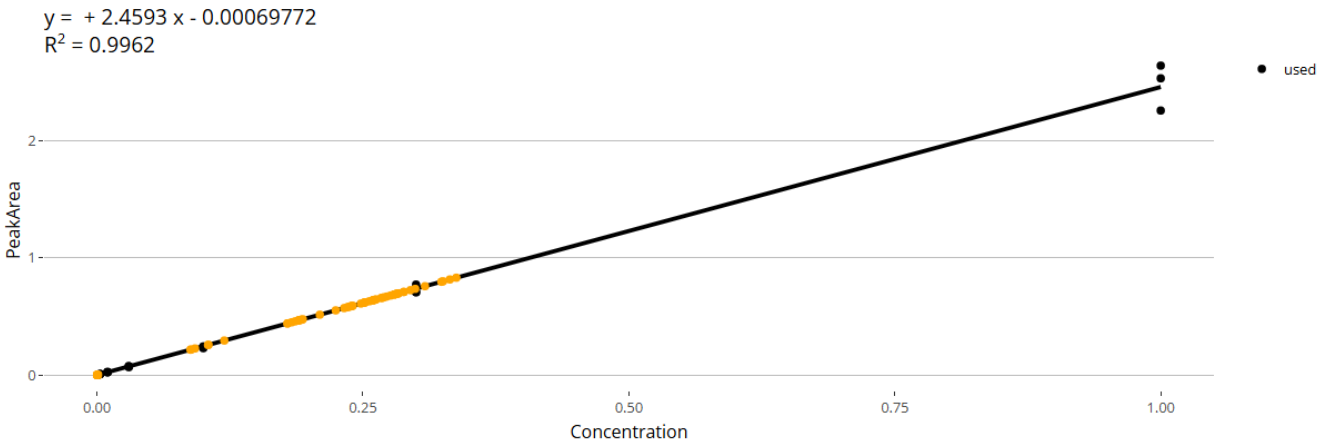
### Interactive Features

- **Exclude Standards:** Left-click on a standard to exclude it from the calibration. Click again to restore.
- **Toggle Standards:** By using one of the select tools in the upper corner, multiple standards can be removed/added to the model by selecting them and approving the message.

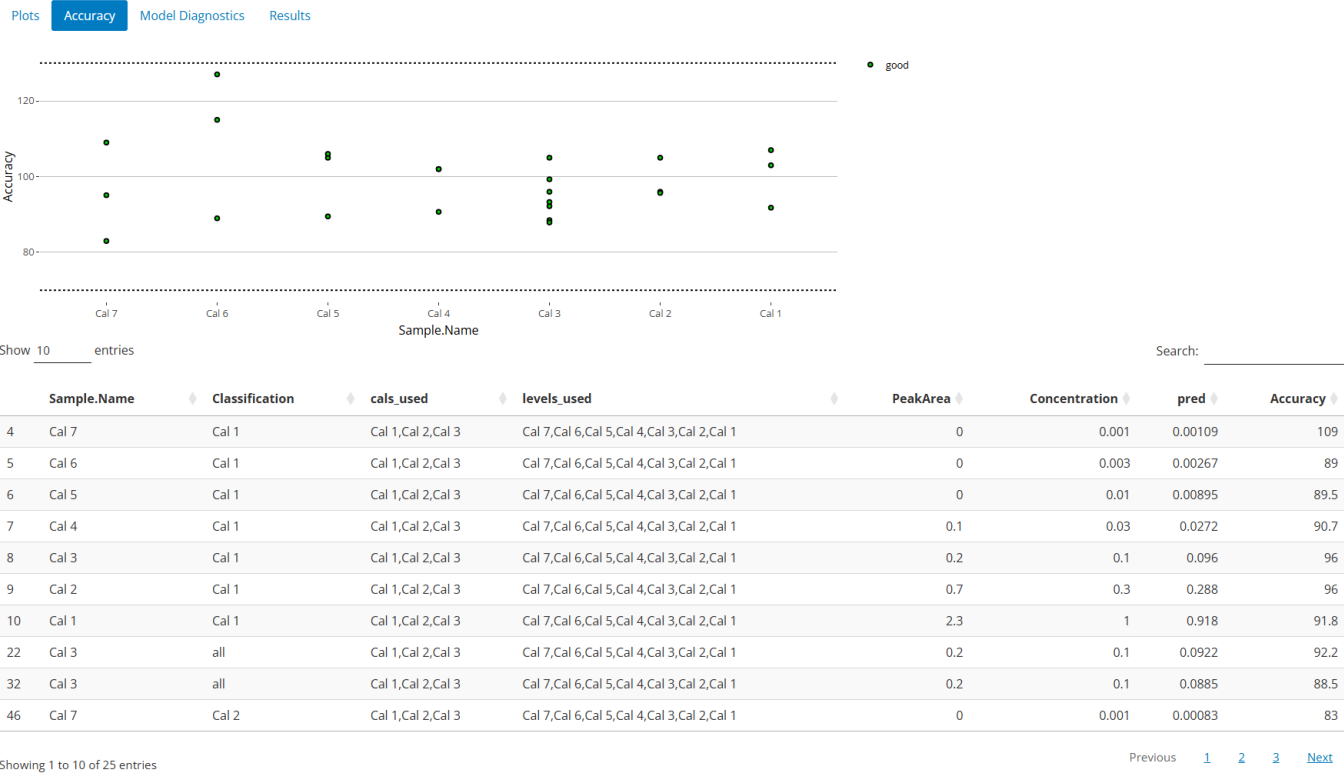
### Main Tabs

1. **Plots:** Interactive regression plot. Calibration points can be excluded/restored in real time.

Plots Accuracy Model Diagnostics Results

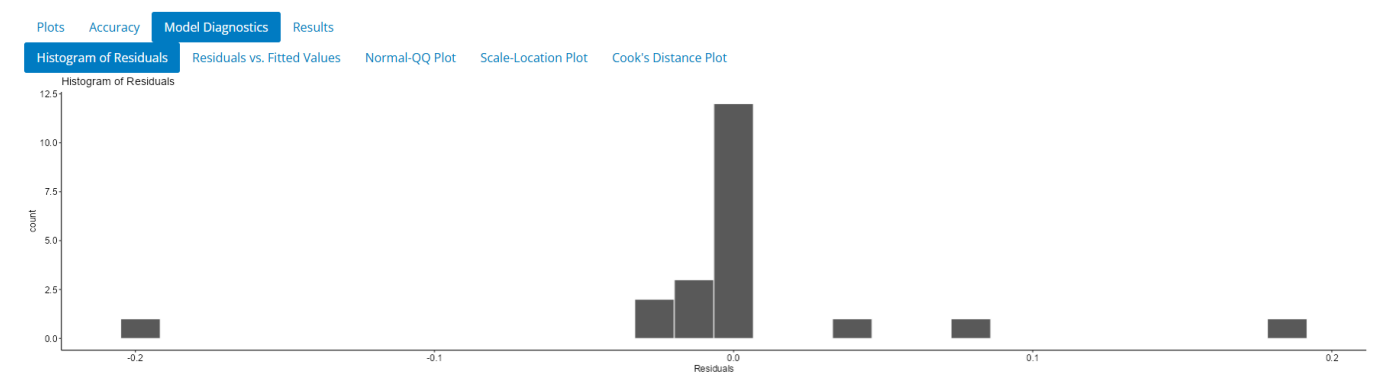


2. **Accuracy:** Visual and numeric summary of model accuracy. Points can also be removed here by clicking on them.



3. **Model Diagnostics:** Provides diagnostic plots (via **lindia** package):
- Histogram of Residuals
  - Residuals vs. Fitted Values
  - Normal Q-Q Plot
  - Scale-Location Plot
  - Residuals vs. Leverage
  - Cook's Distance

**Note:** For more information please look at the documentation/vignette fo the **lindia** package.



4. **Results:** Displays quantification results for the selected transition.

[Plots](#)[Accuracy](#)[Model Diagnostics](#)[Results](#)Show 10 entriesSearch: 

	Sample.Name ♦	Classification ♦	cals_used ♦	PeakArea ♦	pred ♦
1	MeOH 1	all	Cal 1, Cal 2, Cal 3	0	0.000385
2	blank = water	all	Cal 1, Cal 2, Cal 3	0	0.000354
3	IS Mix 1	all	Cal 1, Cal 2, Cal 3	0	0.000402
11	blank = water	all	Cal 1, Cal 2, Cal 3	0	0.00109
12	MeOH 2	all	Cal 1, Cal 2, Cal 3	0	0.000435
13	QC 1	all	Cal 1, Cal 2, Cal 3	0.7	0.279
14	P78	all	Cal 1, Cal 2, Cal 3	0.7	0.288
15	P69	all	Cal 1, Cal 2, Cal 3	0.6	0.235
16	P40	all	Cal 1, Cal 2, Cal 3	0.3	0.104
17	P64	all	Cal 1, Cal 2, Cal 3	0.6	0.239

Showing 1 to 10 of <sup>Previous</sup> 73 entries    [1](#)   [2](#)   [3](#)   [4](#)   [5](#)   ...   [8](#)   [Next](#)

### Saving Results

Once the quantitation method is selected, and the model is optimized, results can be saved by clicking on the **save** button.:

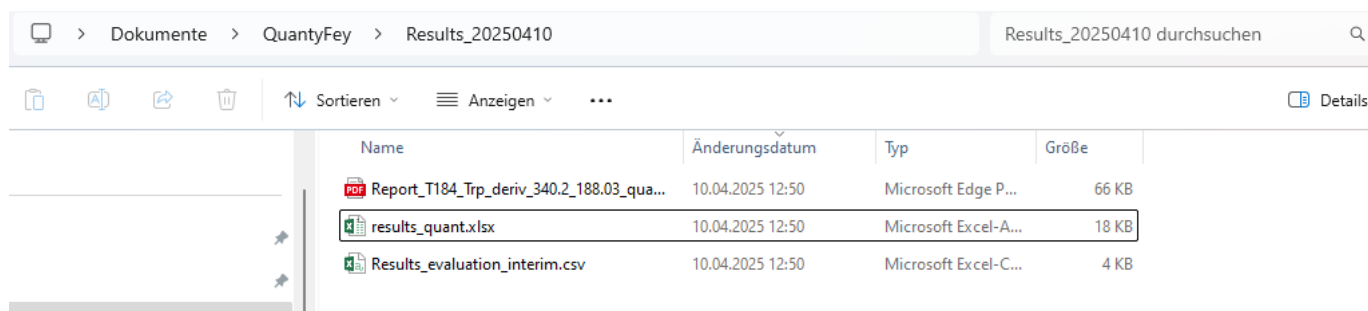
- **Comment:** Add notes for the compound.

- **Save:** Saves the data and generates in the documents folder

`Documents/QuantyFey/Results_<date>/` (folder name can be customized by setting a Project Name in the **Data Upload** tab):

- `Results_quant.csv`: Contains concentrations for all quantified compounds. Values below LLOQ are labeled as "<".
- `Quant_summary.xlsx`: Summarizes all parameters and results for each saved compound, with one sheet per quantification.
- **Generate Report** (optional): Creates a PDF report with relevant plots and details.

**Notes:** To prevent overwriting, files are timestamped if duplicates exist. Reports may be overwritten - when the app is restarted with the same project name; rename or move them to avoid conflicts. All files are saved in the "QuantyFey" folder in the user's Documents directory.



Name	Änderungsdatum	Typ	Größe
Report_T184_Trp_deriv_340.2_188.03_qua...	10.04.2025 12:50	Microsoft Edge P...	66 KB
results_quant.xlsx	10.04.2025 12:50	Microsoft Excel-A...	18 KB
Results_evaluation_interim.csv	10.04.2025 12:50	Microsoft Excel-C...	4 KB

### Quant\_summary.xlsx

This excel file contains all information necessary for the repetition of the exact concentrations calculated for the quantified compounds. For each saved transition, a new sheet is appended.







Figure 1: overview of Retention Time per Sample Type

1



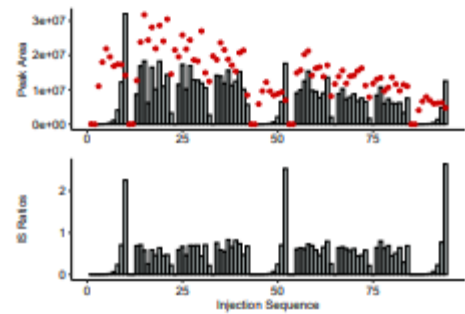
Figure 2: Overview of Sample to Blank

2

Method for Quantification

For this compound the method IS Correction was used.

The red dots mark the Peak Area of the Internal Standard compounds and the Blue represent the Peak Area of the Compound over the whole Measurement Sequence. The lower plot represents the ratio from the respective Sample after each Peak Area from the Compound was divided by the Peak Area from the selected Internal Standard.



The following table shows the correction factors used for the specific Sample Types. This is useful if different dilutions were used for the analysis.

Table 1: Information about the correction factors for each sample type

Sample Type	Correction Factors
Blank	1
Cal	1
QC	1
Sample	1

3

Quantitation Summary

Summary of the Calibration Functions and the Accuracy of the models

The following plots show for all separate Blocks the Calibration model used for the specific blocks.

$$y = + 2.4593 x - 0.00069772$$

$$R^2 = 0.9962$$

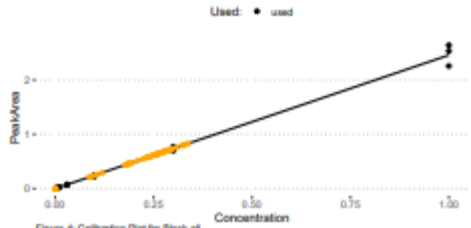


Figure 4: Calibration Plot for Block all

Table 1: Concentration levels

Cal Name	Concentration
Cal 7	0.001
Cal 6	0.003
Cal 5	0.010
Cal 4	0.030
Cal 3	0.100
Cal 2	0.300
Cal 1	1.000

Summary of Settings

Model used: linear  
Quantitation Method used: IS Correction  
Lower Limit of Quantification: 0.001  
Integration Method: 1/x  
IS Compound for Correction:  
IS-Tryptophan:SR1-1051-CE25-quant  
Comment added to this Compound:  
IS correction, look for

4

Table 2: Accuracy Table of Block all

Sample Name	Classification	PeakArea	Concentration	pred.	Accuracy
Cal 7	Cal 1	0.0	0.001	0.004000	109.0
Cal 6	Cal 1	0.0	0.003	0.002670	89.0
Cal 5	Cal 1	0.0	0.010	0.009950	89.5
Cal 4	Cal 1	0.1	0.030	0.027290	90.7
Cal 3	Cal 1	0.2	0.100	0.096600	96.0
Cal 2	Cal 1	0.7	0.300	0.288000	96.0
Cal 1	Cal 1	2.5	1.000	0.918000	91.8
Cal 3	all	0.2	0.100	0.092200	92.2
Cal 3	all	0.2	0.100	0.086500	86.5
Cal 7	Cal 2	0.0	0.001	0.008830	82.0
Cal 6	Cal 2	0.0	0.003	0.001140	115.0
Cal 5	Cal 2	0.0	0.010	0.010500	105.0
Cal 4	Cal 2	0.1	0.030	0.030500	102.0
Cal 3	Cal 2	0.2	0.100	0.090300	99.2
Cal 2	Cal 2	0.7	0.300	0.287000	95.7
Cal 1	Cal 2	2.5	1.000	1.030000	103.0
Cal 3	all	0.2	0.100	0.105000	105.0
Cal 3	all	0.2	0.100	0.097900	87.9
Cal 7	Cal 3	0.0	0.001	0.000951	95.1
Cal 6	Cal 3	0.0	0.003	0.003920	127.0
Cal 5	Cal 3	0.0	0.010	0.010600	106.0
Cal 4	Cal 3	0.1	0.030	0.030790	102.0
Cal 3	Cal 3	0.2	0.100	0.091300	93.2
Cal 2	Cal 3	0.8	0.300	0.314000	105.0
Cal 1	Cal 3	2.6	1.000	1.070000	107.0

5

## Results

This tab becomes active once **at least one compound** has been quantified and saved. It provides the overview of the concentration table and comments for the analysis - it reflects the contents of the `results_quant.csv` file.

**Note:** You can manually edit cells in this tab - for example, to change a **comment** - but **these changes will only take effect after the next save**. The update will not appear in the **PDF Report**. If you save a compound twice, it will add a prefix to it, and save it as an additional transition - a new column in `Quant_results.csv` will be added with the new transition name (e.g., `re_1_transition`), a new sheet, and a new report with the adjusted name will be generated.

Recommendation: When saving the same compound multiple times, make sure to use clear, informative comments. This helps you identifying which version you want to keep or report later on. All saved versions are kept - **nothing is overwritten** - but without good labeling, it can get confusing.

### Qty Fey

 Data Upload    Configure Settings    Compound Quantification    Results

Show All entries

	Sample.Name	Classification	T184_Trp_deriv_340.2_188.03_quant
1	Comment		IS correction, looks fine
2	MeOH 1	Pre 1	< 0.001
3	blank = water	Pre 1	< 0.001
4	IS Mix 1	Pre 1	< 0.001
5	Cal 7	Cal 1	0.0011
6	Cal 6	Cal 1	0.00267
7	Cal 5	Cal 1	0.00894
8	Cal 4	Cal 1	0.0272
9	Cal 3	Cal 1	0.0957
10	Cal 2	Cal 1	0.287
11	Cal 1	Cal 1	0.916
12	blank = water	Block 1.1	0.0011
13	MeOH 2	Block 1.1	< 0.001
14	QC 1	Block 1.1	0.279
15	P78	Block 1.1	0.288
16	P69	Block 1.1	0.234
17	P40	Block 1.1	0.104
18	P64	Block 1.1	0.238
19	P41	Block 1.1	0.189
20	P79	Block 1.1	0.259

## Summary

Once all settings are configured, the application supports a full evaluation of data quality and compound identification:

- **RT and Ion Ratios** help verify correct compound identification.
- **Blank Analysis** helps in distinguishing true signals from background noise.

Before quantification, users can apply **drift correction** strategies using the following methods:

- Internal standard correction
- QC-based drift correction

- Custom Bracketing
- Weighted Bracketing

Each method can be configured before quantification, and then methods can be compared, and the most appropriate method can be sued. The quantification interface includes tools for **interactive regression model optimization**, allowing users to:

- Adjust model parameters
- View diagnostic plots
- Assess model fit

An **automatic optimization feature** is available to suggest a possible regression fit based on a simple algorithm, but revision of the optimization is advised. **Overwriting safeguards** are in place to avoid accidental overwriting, while reducing file clutter.

## Troubleshooting

- **Package Installation Failure:**
  - Ensure RTools 4.2 is correctly installed.
  - Note: The application is compatible only with Windows systems.
- **Console Does Not Open:**
  - Relocate the application folder to a different directory.
  - Avoid running the application from the "Downloads" folder, as this may cause issues.