

The background of the slide features a complex network graph. It consists of numerous clusters of small, light-blue circular nodes. These clusters are interconnected by a web of thin, light-grey lines, creating a star-like or radial pattern from each cluster. The overall structure is dense and spread across the dark blue background.

Clustering amplicons

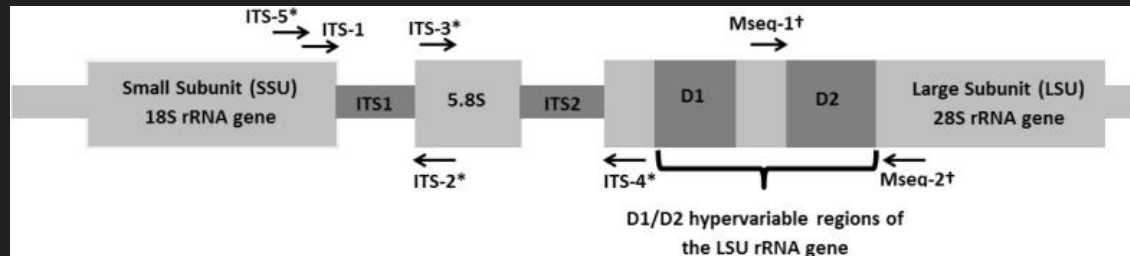
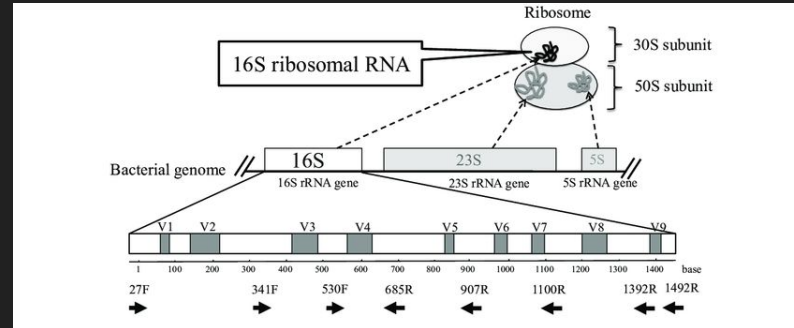
Acer VanWalleendael

Outline

1. Amplicon sequencing background
2. Bioinformatic workflow
 - a. QC
 - b. Clustering
 - c. Understanding outputs
3. Amplicon clustering tutorial

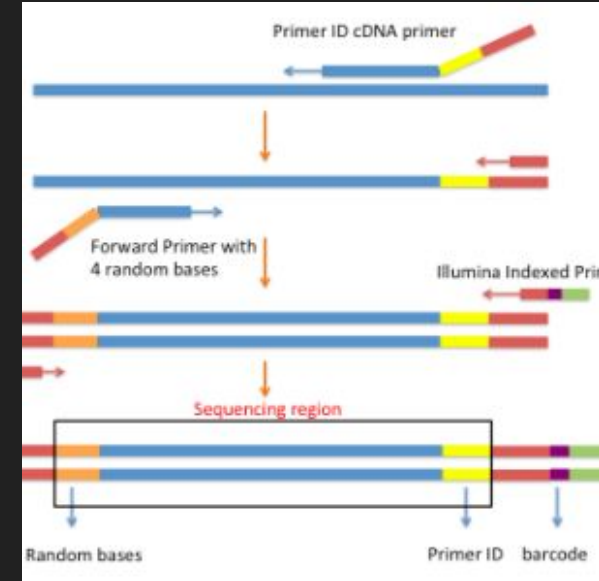
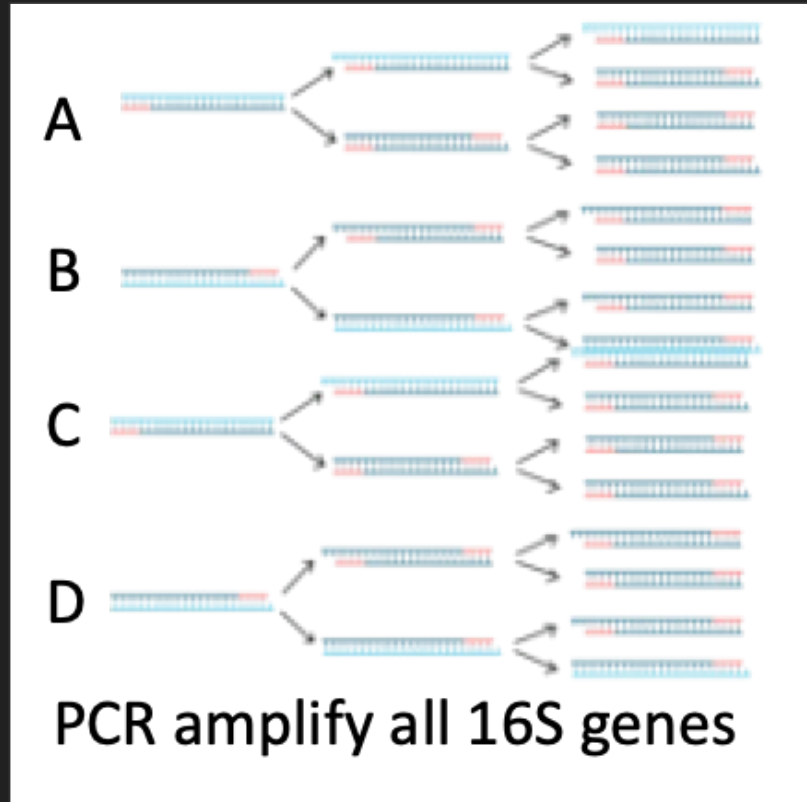
So you want to do a microbiome.

16S (bacteria) and ITS (fungi) regions of the ribosome evolve just slowly enough that primer regions are conserved, but species are different.



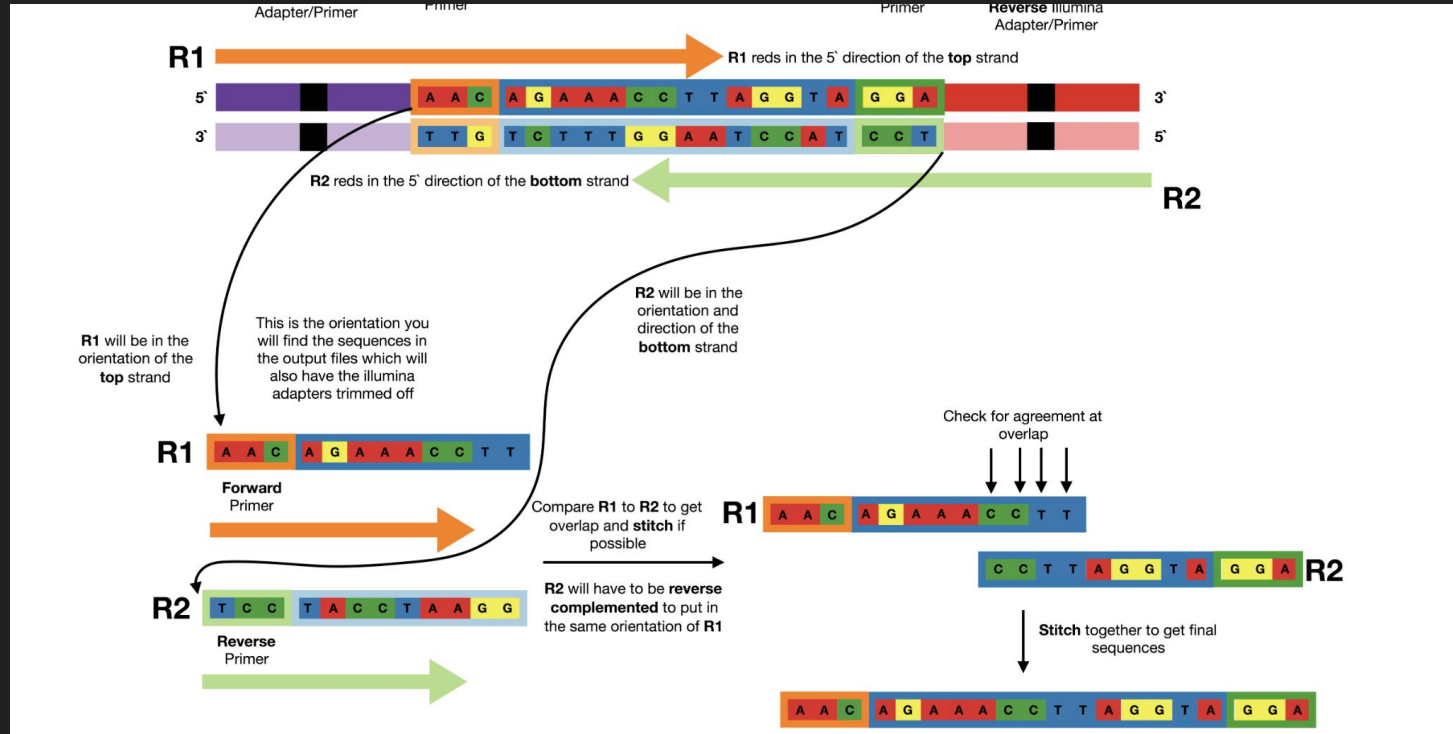
Sequencing ITS or 16S: Library prep

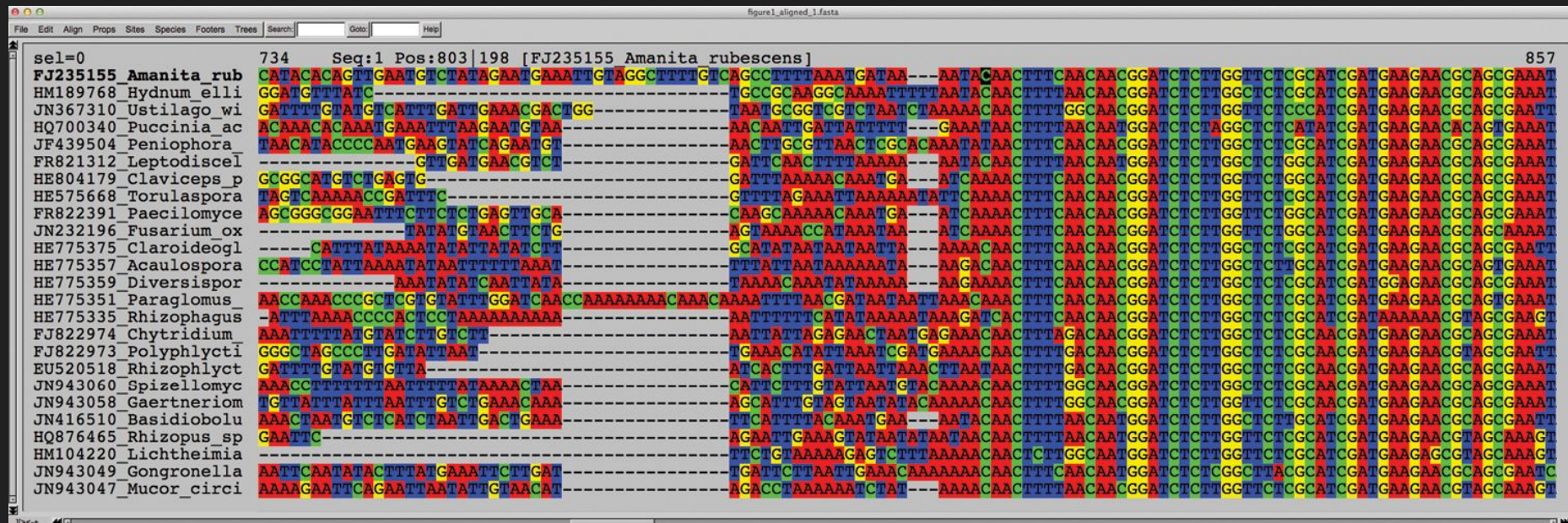
Extract, clean,
and normalize
DNA from
sample



Ligate adapters &
barcodes for multiplex
sequencing

Illumina sequencing

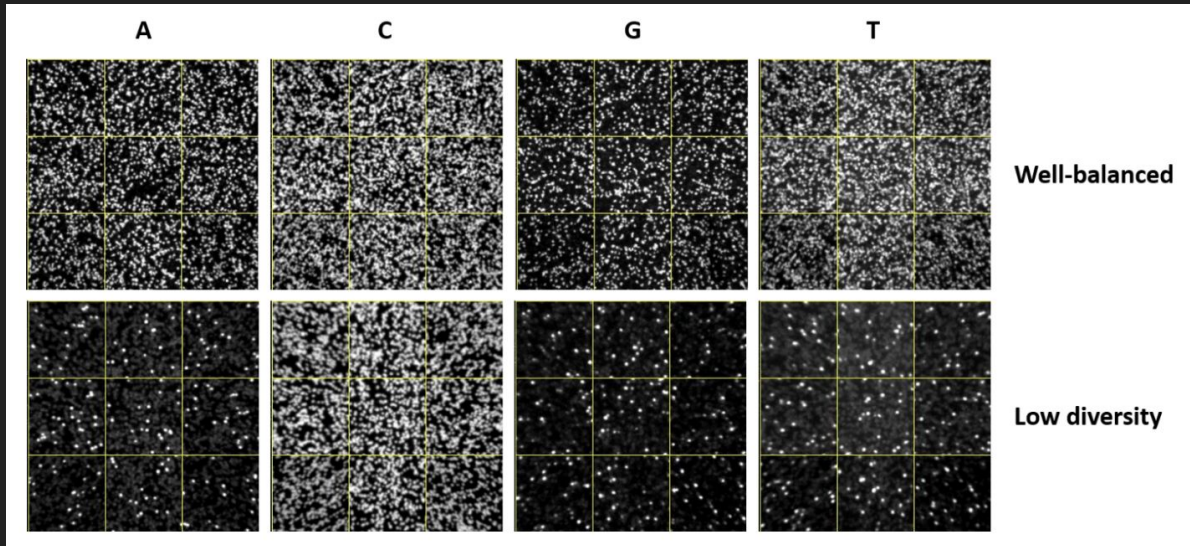




Sequencing controls

Field, Extraction, Library prep controls

PhiX control library = balanced proportions of ACGTs to correct for seq errors from low-diversity samples

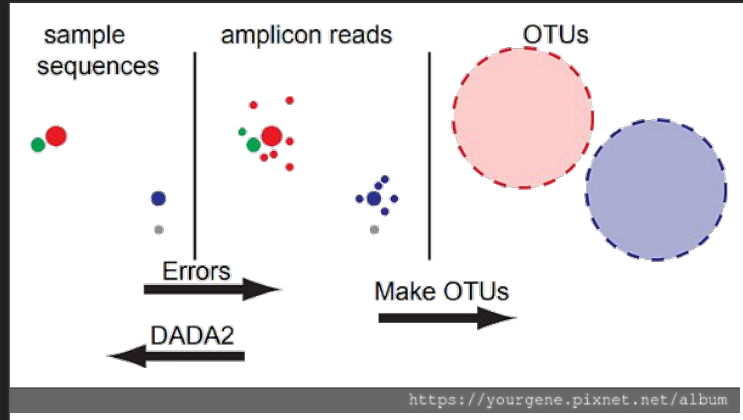


Bioinformatic workflow

1. Quality Check
2. Demultiplexing
3. Amplicon clean and QC
4. Amplicon clustering

Bioinformatic workflow

1. Quality check: were there major sequencing errors?
2. Demultiplexing: Use barcodes to distinguish samples
3. Amplicon clean and QC: Remove poor reads, primers, trim seqs, output stats
4. Amplicon clustering: Output OTUs (operational taxonomic units) or ASVs (Amplicon Sequence Variants)



Simplified Clustering Workflow

1. Remove PhiX
2. Trim primers
3. Strip variable region
4. Filter low quality
5. Identify unique reads
6. Cluster OTUs
7. Build OTU table

Simplified Clustering Workflow

1. Remove PhiX: *bowtie2*
2. Trim primers: *cutadapt*
3. Strip variable region: *usearch*
4. Filter low quality: *usearch*
5. Identify unique reads: *usearch*
6. Cluster OTUs: *usearch*
7. Build OTU table: *usearch*