

# Dionex\_COMPASS\_TEMPEST

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## Daily Set up

Read in and Format the raw data - change wd & file names (Make sure files are UTF-8 encoded)

```
# SULFATE DATA:

## Read in raw data file from Dionex - copied and saved as a txt
Sdat <- read.table("Raw Data/COMPASS_TEMPEST_20230608_S04.txt",sep='\t' , header=T, skip=3)
head(Sdat)

##      X          X.1          X.2 IC.S04 IC.S04.1 IC.S04.2 IC.S04.3 IC.S04.4
## 1 1 Lab Blank Unknown    n.a.    n.a.    n.a.    n.a.    n.a.
## 2 2 Lab Blank Unknown    n.a.    n.a.    n.a.    n.a.    n.a.
## 3 3 Lab Blank Unknown    n.a.    n.a.    n.a.    n.a.    n.a.
## 4 4 Standard 1 Calibration Standard 4.343   0.4565   6.17   0.5324   2.53
## 5 5 Standard 2 Calibration Standard 4.343   0.9668   6.58   1.1275   5.32
## 6 6 Standard 3 Calibration Standard 4.330   1.9262   7.25   2.2463  10.35
##      IC.S04.5
## 1      n.a.
## 2      n.a.
## 3      n.a.
## 4      MB
## 5      MB
## 6      M

## Only keep the columns that we need
Sdat <- Sdat[,c(2,5,7)] # dont need this here
head(Sdat)

##          X.1 IC.S04.1 IC.S04.3
## 1  Lab Blank    n.a.    n.a.
## 2  Lab Blank    n.a.    n.a.
## 3  Lab Blank    n.a.    n.a.
## 4 Standard 1  0.4565  0.5324
## 5 Standard 2  0.9668  1.1275
## 6 Standard 3  1.9262  2.2463
```

```

## Name the columns correctly
colnames(Sdat) <- c( "Sample_ID", "S04_ppm", "S04_Area")
Sdat$Sample_ID <- as.factor(Sdat$Sample_ID)
Sdat$S04_ppm <- as.numeric(Sdat$S04_ppm)

## Warning: NAs introduced by coercion

Sdat$S04_Area <- as.numeric(Sdat$S04_Area)

## Warning: NAs introduced by coercion

Sdat <- as.data.frame(Sdat)
head(Sdat)

##      Sample_ID S04_ppm S04_Area
## 1    Lab Blank      NA       NA
## 2    Lab Blank      NA       NA
## 3    Lab Blank      NA       NA
## 4 Standard 1   0.4565   0.5324
## 5 Standard 2   0.9668   1.1275
## 6 Standard 3   1.9262   2.2463

#Chloride data
## Read in raw data file from Dionex - copied and saved as a txt
Cldat <- read.table("Raw Data/COMPASS_TEMPEST_20230608_Cl.txt",sep='\t' , header=T, skip=3)
head(Cldat)

##      X          X.1          X.2  IC.Cl  IC.Cl.1 IC.Cl.2  IC.Cl.3 IC.Cl.4
## 1 1 Lab Blank Unknown  n.a.     n.a.     n.a.     n.a.     n.a.
## 2 2 Lab Blank Unknown  n.a.     n.a.     n.a.     n.a.     n.a.
## 3 3 Lab Blank Unknown  n.a.     n.a.     n.a.     n.a.     n.a.
## 4 4 Standard 1 Calibration Standard 3.417   5.7300   93.70   8.0833   45.41
## 5 5 Standard 2 Calibration Standard 3.417  11.3295   93.26  15.9825   88.56
## 6 6 Standard 3 Calibration Standard 3.413  20.3441   92.64  28.6992  159.19
##      IC.Cl.5
## 1      n.a.
## 2      n.a.
## 3      n.a.
## 4      BM
## 5      M
## 6      BM

## Only keep the columns that we need
Cldat <- Cldat[,c(2,5,7)]
head(Cldat)

##          X.1  IC.Cl.1  IC.Cl.3
## 1  Lab Blank     n.a.     n.a.
## 2  Lab Blank     n.a.     n.a.
## 3  Lab Blank     n.a.     n.a.
## 4 Standard 1   5.7300   8.0833
## 5 Standard 2  11.3295  15.9825
## 6 Standard 3  20.3441  28.6992

```

```

## Name the columns correctly
colnames(Cldat) <- c( "Sample_ID", "Cl_ppm", "Cl_Area")
Cldat$Sample_ID <- as.factor(Cldat$Sample_ID)
Cldat$Cl_ppm <- as.numeric(Cldat$Cl_ppm)

```

```

## Warning: NAs introduced by coercion

```

```

Cldat$Cl_Area <- as.numeric(Cldat$Cl_Area)

```

```

## Warning: NAs introduced by coercion

```

```

Cldat <- as.data.frame(Cldat)
head(Cldat)

```

```

##   Sample_ID  Cl_ppm Cl_Area
## 1 Lab Blank      NA      NA
## 2 Lab Blank      NA      NA
## 3 Lab Blank      NA      NA
## 4 Standard 1  5.7300  8.0833
## 5 Standard 2 11.3295 15.9825
## 6 Standard 3 20.3441 28.6992

```

```

## Bring the data back together:
all_dat <- merge(Sdat, Cldat, by="Sample_ID")
head(all_dat)

```

```

##   Sample_ID  S04_ppm S04_Area    Cl_ppm  Cl_Area
## 1           581.2849  23.7212 4345.0138 286.6714
## 2           581.2849  23.7212 1400.2957  69.8002
## 3           581.2849  23.7212     0.0058  0.0082
## 4           581.2849  23.7212 1605.3399  61.2697
## 5           581.2849  23.7212       NA      NA
## 6          202.4819   8.0517 4345.0138 286.6714

```

```

## Remove empty lines
all_dat <- all_dat[!(is.na(all_dat$Sample_ID) | all_dat$Sample_ID=="") , ]
head(all_dat)

```

```

##   Sample_ID  S04_ppm S04_Area  Cl_ppm  Cl_Area
## 26 Lab Blank      NA      NA      NA      NA
## 27 Lab Blank      NA      NA      NA      NA
## 28 Lab Blank      NA      NA      NA      NA
## 29 Lab Blank      NA      NA      NA      NA
## 30 Lab Blank      NA  0.0196  0.0277
## 31 Lab Blank      NA  0.0100  0.0141

```

```

all_dat <- droplevels(all_dat[!all_dat$Sample_ID == 'Lab Blank',])
head(all_dat)

```

```

##      Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area
## 170 Standard 1  0.4565   0.5324 5.7300  8.0833
## 171 Standard 1  0.4565   0.5324 5.8701  8.2810
## 172 Standard 1  0.4565   0.5324 5.7836  8.1589
## 173 Standard 1  0.4826   0.5628 5.7300  8.0833
## 174 Standard 1  0.4826   0.5628 5.8701  8.2810
## 175 Standard 1  0.4826   0.5628 5.7836  8.1589

```

```
all_dat[is.na(all_dat)] <- 0
```

Pull out standards - could do some checks here if we want

```

stds <- all_dat[grep("Standard", all_dat$Sample_ID),]
head(stds)

```

```

##      Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area
## 170 Standard 1  0.4565   0.5324 5.7300  8.0833
## 171 Standard 1  0.4565   0.5324 5.8701  8.2810
## 172 Standard 1  0.4565   0.5324 5.7836  8.1589
## 173 Standard 1  0.4826   0.5628 5.7300  8.0833
## 174 Standard 1  0.4826   0.5628 5.8701  8.2810
## 175 Standard 1  0.4826   0.5628 5.7836  8.1589

```

```

stds_chk_S <- stds %>%
  group_by(Sample_ID) %>%
  summarise(mean = mean(S04_ppm), sd = sd(S04_ppm))
stds_chk_S$cv <- (stds_chk_S$sd/stds_chk_S$mean)*100
stds_chk_S$flag <- ifelse(stds_chk_S$cv <2, 'YES', 'NO, rerun')
head(stds_chk_S)

```

```

## # A tibble: 5 x 5
##   Sample_ID     mean       sd     cv flag
##   <fct>        <dbl>     <dbl>  <dbl> <chr>
## 1 Standard 1  0.472  0.0118  2.50  NO, rerun
## 2 Standard 2  0.985  0.0218  2.21  NO, rerun
## 3 Standard 3  1.92   0.00999 0.520 YES
## 4 Standard 4 10.1    0.121   1.21  YES
## 5 Standard 5 20.2    0.183   0.904 YES

```

```

stds_chk_Cl <- stds %>%
  group_by(Sample_ID) %>%
  summarise(mean = mean(Cl_ppm), sd = sd(Cl_ppm))
stds_chk_Cl$cv <- (stds_chk_Cl$sd/stds_chk_Cl$mean)*100
stds_chk_Cl$flag <- ifelse(stds_chk_Cl$cv <2, 'YES', 'NO, rerun')
head(stds_chk_Cl)

```

```

## # A tibble: 5 x 5
##   Sample_ID     mean       sd     cv flag
##   <fct>        <dbl>     <dbl>  <dbl> <chr>
## 1 Standard 1  5.79   0.0612  1.06  YES

```

```

## 2 Standard 2 11.4 0.0937 0.819 YES
## 3 Standard 3 20.5 0.159 0.777 YES
## 4 Standard 4 101. 1.14 1.12 YES
## 5 Standard 5 201. 1.98 0.982 YES

```

```

lmS <- lm(stds$S04_Area ~ stds$S04_ppm)
S04_sum <- summary(lmS)
S04_Slope <- S04_sum$coefficients[2, 1]
S04_Int <- S04_sum$coefficients[1, 1]

lmCl <- lm(stds$Cl_Area ~ stds$Cl_ppm)
Cl_sum <- summary(lmCl)
Cl_Slope <- Cl_sum$coefficients[2, 1]
Cl_Int <- Cl_sum$coefficients[1, 1]

```

## Calculate mmol/L concentrations

```

#remove standards from sample dataframe
sampledat <- all_dat[!grepl("Standard", all_dat$Sample_ID),]
head(sampledat)

##                                     Sample_ID  S04_ppm  S04_Area    Cl_ppm  Cl_Area
## 205 TEMPEST_ESTUARY_SOURCE_20230607_0710 581.2849 13.5575 4335.7195 122.3274
## 206 TEMPEST_ESTUARY_SOURCE_20230607_1346 547.2306 12.7632 4145.0144 116.9468
## 207 TEMPEST_ESTUARY_SOURCE_20230607_1700 553.9434 12.9198 4153.6146 117.1895
## 208      TEMPEST_FW_B4_20230607_0800 11.1494 11.8200 27.9934 35.9001
## 209      TEMPEST_FW_B4_20230607_1600  2.4624  2.2973 206.4674 233.0096
## 210      TEMPEST_FW_C3_20230607_0800 10.7750 11.4231 23.9976 30.7756

# Constants needed for calculations:
clmw <- 35.45      #molecular weight of Chloride: 35.45
smw <- 32.06        #molecular weight of sulfur: 32.06

# Convert ppm to mmol/L
sampledat$S04_mM <- (sampledat$S04_ppm / smw)
sampledat$Cl_mM <- (sampledat$Cl_ppm / clmw)

# Calculate Salinity
# calculated using the Knudsen equation
# Salinity = 0.03 + 1.8050 * Chlorinity
# Ref: A Practical Handbook of Seawater Analysis by Strickland & Parsons (P. 11)
# =((1.807*Cl_ppm)+0.026)/1000
sampledat$salinity <- ((1.807 * sampledat$Cl_ppm) + 0.026) / 1000

head(sampledat)

##                                     Sample_ID  S04_ppm  S04_Area    Cl_ppm  Cl_Area
## 205 TEMPEST_ESTUARY_SOURCE_20230607_0710 581.2849 13.5575 4335.7195 122.3274
## 206 TEMPEST_ESTUARY_SOURCE_20230607_1346 547.2306 12.7632 4145.0144 116.9468
## 207 TEMPEST_ESTUARY_SOURCE_20230607_1700 553.9434 12.9198 4153.6146 117.1895
## 208      TEMPEST_FW_B4_20230607_0800 11.1494 11.8200 27.9934 35.9001
## 209      TEMPEST_FW_B4_20230607_1600  2.4624  2.2973 206.4674 233.0096
## 210      TEMPEST_FW_C3_20230607_0800 10.7750 11.4231 23.9976 30.7756

```

```

## 205 TEMPEST_ESTUARY_SOURCE_20230607_0710 581.2849 13.5575 4335.7195 122.3274
## 206 TEMPEST_ESTUARY_SOURCE_20230607_1346 547.2306 12.7632 4145.0144 116.9468
## 207 TEMPEST_ESTUARY_SOURCE_20230607_1700 553.9434 12.9198 4153.6146 117.1895
## 208 TEMPEST_FW_B4_20230607_0800 11.1494 11.8200 27.9934 35.9001
## 209 TEMPEST_FW_B4_20230607_1600 2.4624 2.2973 206.4674 233.0096
## 210 TEMPEST_FW_C3_20230607_0800 10.7750 11.4231 23.9976 30.7756
## S04_mM Cl_mM salinity
## 205 18.13115721 122.3052045 7.83467114
## 206 17.06895197 116.9256530 7.49006702
## 207 17.27833437 117.1682539 7.50560758
## 208 0.34776669 0.7896587 0.05061007
## 209 0.07680599 5.8241862 0.37311259
## 210 0.33608858 0.6769422 0.04338966

```

## Pull out dups and check with percent difference

```

#Show me the data that we have from the calculations
head(sampledat)

```

```

## Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area
## 205 TEMPEST_ESTUARY_SOURCE_20230607_0710 581.2849 13.5575 4335.7195 122.3274
## 206 TEMPEST_ESTUARY_SOURCE_20230607_1346 547.2306 12.7632 4145.0144 116.9468
## 207 TEMPEST_ESTUARY_SOURCE_20230607_1700 553.9434 12.9198 4153.6146 117.1895
## 208 TEMPEST_FW_B4_20230607_0800 11.1494 11.8200 27.9934 35.9001
## 209 TEMPEST_FW_B4_20230607_1600 2.4624 2.2973 206.4674 233.0096
## 210 TEMPEST_FW_C3_20230607_0800 10.7750 11.4231 23.9976 30.7756
## S04_mM Cl_mM salinity
## 205 18.13115721 122.3052045 7.83467114
## 206 17.06895197 116.9256530 7.49006702
## 207 17.27833437 117.1682539 7.50560758
## 208 0.34776669 0.7896587 0.05061007
## 209 0.07680599 5.8241862 0.37311259
## 210 0.33608858 0.6769422 0.04338966

```

```

#pull out any rows that have "dup" in the SampleID column
dups <- sampledat %>%
  filter(str_detect(Sample_ID, "_dup"))      #have to change this to match data
head(dups)

```

```

## Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area
## 1 TEMPEST_FW_SOURCE_20230607_1519_dup 574.2564 13.3935 0.000 0.0000
## 2 TEMPEST_SW_B4_20230607_0800_dup 403.8132 9.4182 3303.890 93.2155
## 3 TEMPEST_SW_B4_20230607_1600_dup 440.2724 10.2686 3510.899 99.0560
## 4 TEMPEST_SW_D5_20230607_0800_dup 292.2655 6.8166 2343.111 66.1082
## 5 TEMPEST_SW_I5_20230607_0800_dup 273.6440 6.3823 2744.173 77.4237
## S04_mM Cl_mM salinity
## 1 17.911928 0.00000 0.000026
## 2 12.595546 93.19858 5.970155
## 3 13.732764 99.03805 6.344220
## 4 9.116204 66.09623 4.234028
## 5 8.535371 77.40967 4.958746

```

```

#remove these from sample dataframe in a new dataframe
sampledat2 <- sampledat %>%
  filter(!str_detect(Sample_ID, "_dup")) %>%
  filter(!str_detect(Sample_ID, "_spk"))
head(sampledat2)

##                                     Sample_ID  SO4_ppm SO4_Area    Cl_ppm  Cl_Area
## 1 TEMPEST_ESTUARY_SOURCE_20230607_0710 581.2849 13.5575 4335.7195 122.3274
## 2 TEMPEST_ESTUARY_SOURCE_20230607_1346 547.2306 12.7632 4145.0144 116.9468
## 3 TEMPEST_ESTUARY_SOURCE_20230607_1700 553.9434 12.9198 4153.6146 117.1895
## 4          TEMPEST_FW_B4_20230607_0800 11.1494 11.8200 27.9934 35.9001
## 5          TEMPEST_FW_B4_20230607_1600   2.4624  2.2973 206.4674 233.0096
## 6          TEMPEST_FW_C3_20230607_0800 10.7750 11.4231 23.9976 30.7756
##           SO4_mM      Cl_mM  salinity
## 1 18.13115721 122.3052045 7.83467114
## 2 17.06895197 116.9256530 7.49006702
## 3 17.27833437 117.1682539 7.50560758
## 4 0.34776669  0.7896587 0.05061007
## 5 0.07680599  5.8241862 0.37311259
## 6 0.33608858  0.6769422 0.04338966

#remove the dup from these IDs so we will have duplicate sample names
dups$Sample_ID<-gsub("_dup","",as.character(dups$Sample_ID))
dups <- dups[, -c(2:5)]
colnames(dups) <- c('Sample_ID', 'SO4_mM_dup', 'Cl_mM_dup', "salinity_dup")
head(dups)

##                                     Sample_ID SO4_mM_dup Cl_mM_dup salinity_dup
## 1 TEMPEST_FW_SOURCE_20230607_1519 17.911928  0.00000  0.000026
## 2          TEMPEST_SW_B4_20230607_0800 12.595546  93.19858  5.970155
## 3          TEMPEST_SW_B4_20230607_1600 13.732764  99.03805  6.344220
## 4          TEMPEST_SW_D5_20230607_0800  9.116204  66.09623  4.234028
## 5          TEMPEST_SW_I5_20230607_0800  8.535371  77.40967  4.958746

#put it back together with the old data set and look for duplicates
QAdups <- merge(sampledat2, dups)
head(QAdups)

##                                     Sample_ID  SO4_ppm SO4_Area    Cl_ppm  Cl_Area     SO4_mM
## 1 TEMPEST_FW_SOURCE_20230607_1519 581.1936 13.5553     0.000  0.0000 18.128309
## 2          TEMPEST_SW_B4_20230607_0800 377.6524  8.8081 3087.801 87.1188 11.779551
## 3          TEMPEST_SW_B4_20230607_1600 440.3089 10.2694 3507.065 98.9478 13.733902
## 4          TEMPEST_SW_D5_20230607_0800 293.4522  6.8443 2353.532 66.4022  9.153219
## 5          TEMPEST_SW_I5_20230607_0800 284.5868  6.6375 2842.504 80.1980  8.876694
##           Cl_mM salinity SO4_mM_dup Cl_mM_dup salinity_dup
## 1 0.00000 0.000026 17.911928  0.00000  0.000026
## 2 87.10298 5.579682 12.595546  93.19858  5.970155
## 3 98.92990 6.337292 13.732764  99.03805  6.344220
## 4 66.39019 4.252859  9.116204  66.09623  4.234028
## 5 80.18346 5.136430  8.535371  77.40967  4.958746

```

```

QAdups$S04_dups_chk <- ((abs(QAdups$S04_mM-QAdups$S04_mM_dup))/((QAdups$S04_mM+QAdups$S04_mM_dup)/2))*100
QAdups$S04_dups_flag <- ifelse(QAdups$S04_dups_chk <10, 'YES', 'NO, rerun')

QAdups$Cl_dups_chk <- ((abs(QAdups$Cl_mM-QAdups$Cl_mM_dup))/((QAdups$Cl_mM+QAdups$Cl_mM_dup)/2))*100
QAdups$Cl_dups_flag <- ifelse(QAdups$Cl_dups_chk <10, 'YES', 'NO, rerun')

head(QAdups)

##                                     Sample_ID  S04_ppm  S04_Area   Cl_ppm  Cl_Area   S04_mM
## 1 TEMPEST_FW_SOURCE_20230607_1519 581.1936 13.5553    0.000  0.0000 18.128309
## 2 TEMPEST_SW_B4_20230607_0800 377.6524  8.8081 3087.801 87.1188 11.779551
## 3 TEMPEST_SW_B4_20230607_1600 440.3089 10.2694 3507.065 98.9478 13.733902
## 4 TEMPEST_SW_D5_20230607_0800 293.4522  6.8443 2353.532 66.4022  9.153219
## 5 TEMPEST_SW_I5_20230607_0800 284.5868  6.6375 2842.504 80.1980  8.876694
##          Cl_mM salinity  S04_mM_dup  Cl_mM_dup salinity_dup  S04_dups_chk
## 1  0.00000 0.000026 17.911928  0.00000  0.000026 1.200778917
## 2 87.10298 5.579682 12.595546 93.19858  5.970155 6.695317107
## 3 98.92990 6.337292 13.732764 99.03805  6.344220 0.008289978
## 4 66.39019 4.252859  9.116204 66.09623  4.234028 0.405212272
## 5 80.18346 5.136430  8.535371 77.40967  4.958746 3.920528928
##      S04_dups_flag  Cl_dups_chk  Cl_dups_flag
## 1        YES       NaN      <NA>
## 2        YES     6.7615540     YES
## 3        YES     0.1092568     YES
## 4        YES     0.4437552     YES
## 5        YES     3.5201930     YES

#plot dups output as a bar graph to easily check - want any over 10% to be red need to work on this
Sdupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = S04_dups_chk, fill=S04_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="Difference Between S04 Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
                                             color = "black", size=1)

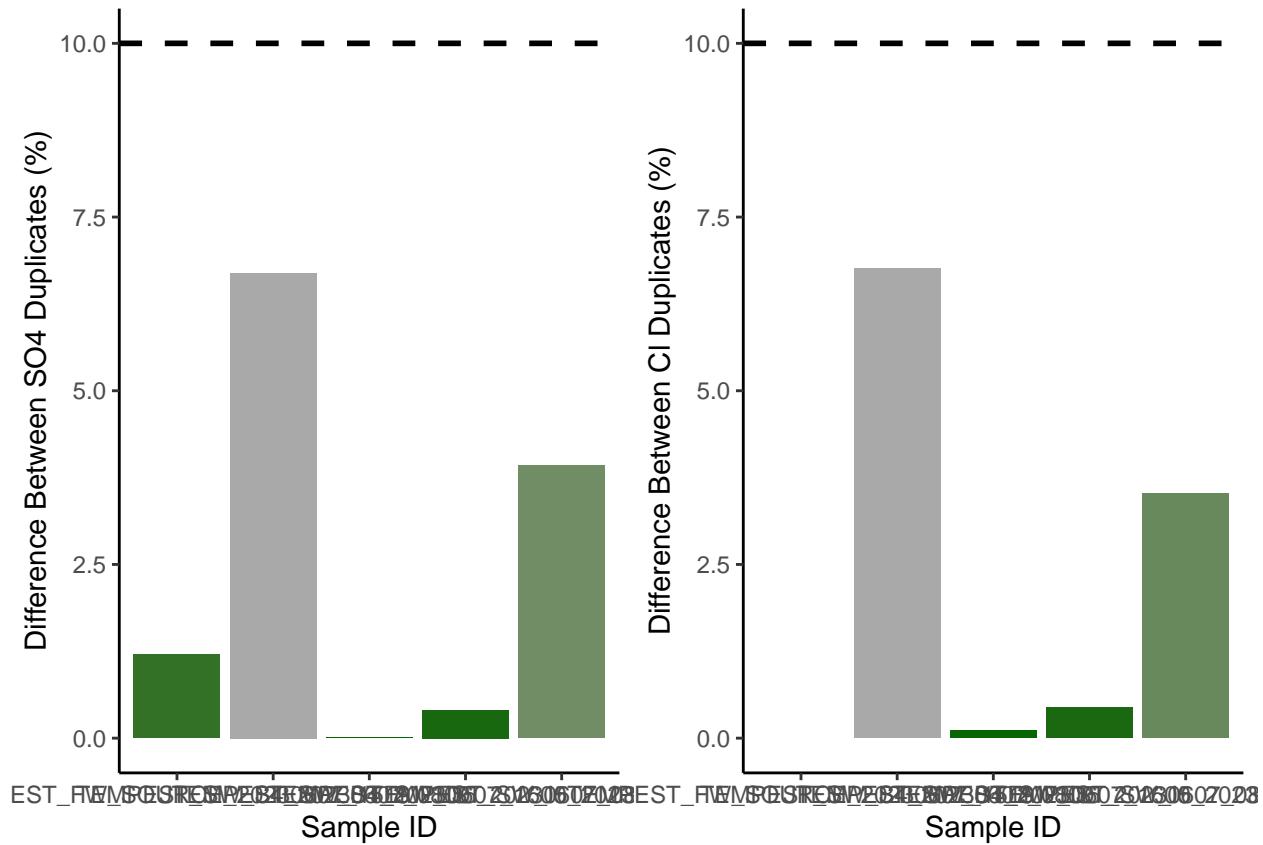
## Warning: Using 'size' aesthetic for lines was deprecated in ggplot2 3.4.0.
## i Please use 'linewidth' instead.
## This warning is displayed once every 8 hours.
## Call `lifecycle::last_lifecycle_warnings()` to see where this warning was
## generated.

Cldupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = Cl_dups_chk, fill=Cl_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="Difference Between Cl Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
                                             color = "black", size=1)

ggarrange(Sdupsbar, Cldupsbar, ncol=2, nrow=1)

## Warning: Removed 1 row containing missing values or values outside the scale range
## ('geom_bar()').

```



```
#check for percent of no, reruns to see if it would warrant reruns
Perc_dups <- QAdups %>%
  group_by(SO4_dups_flag) %>%
  summarise(S_no_rows = length(SO4_dups_flag))
Perc_dups1 <- QAdups %>%
  group_by(Cl_dups_flag) %>%
  summarise(Cl_no_rows = length(Cl_dups_flag))
colnames(Perc_dups) <- c("Flag", "S_no_rows")
colnames(Perc_dups1) <- c("Flag", "Cl_no_rows")
Perc_dups2 <- cbind(Perc_dups, Perc_dups1)

Perc_dups2$Total <- length(QAdups$SO4_dups_flag)
Perc_dups2$S_Percent <- (Perc_dups2$S_no_rows / Perc_dups2$Total)*100
Perc_dups2$Cl_Percent <- (Perc_dups2$Cl_no_rows / Perc_dups2$Total)*100
head(Perc_dups2)
```

```
##   Flag S_no_rows Flag Cl_no_rows Total S_Percent Cl_Percent
## 1 YES      5 YES      4     5    100     80       80
## 2 YES      5 <NA>     1     5    100     20       20
```

Pull out dups and check with cv

```

#the cv
# calculate the sd bewteen the two duplicates then divide by the mean and multiply by 100

df2 <- as.data.frame(QAdups$S04_mM)
df2$dups <- QAdups$S04_mM_dup

df2$sds <- apply(df2,1,sd)

QAdups$S04_dups_cv <- (df2$sds)/((QAdups$S04_mM+QAdups$S04_mM_dup)/2) * 100
QAdups$S04_dups_cv_flag <- ifelse(QAdups$S04_dups_cv <11, 'YES', 'NO, rerun')

head(QAdups)

##                                     Sample_ID   S04_ppm   S04_Area    Cl_ppm   Cl_Area    S04_mM
## 1 TEMPEST_FW_SOURCE_20230607_1519 581.1936  13.5553    0.000  0.0000 18.128309
## 2 TEMPEST_SW_B4_20230607_0800 377.6524   8.8081 3087.801 87.1188 11.779551
## 3 TEMPEST_SW_B4_20230607_1600 440.3089   10.2694 3507.065 98.9478 13.733902
## 4 TEMPEST_SW_D5_20230607_0800 293.4522   6.8443 2353.532 66.4022  9.153219
## 5 TEMPEST_SW_I5_20230607_0800 284.5868   6.6375 2842.504 80.1980  8.876694
##          Cl_mM salinity S04_mM_dup Cl_mM_dup salinity_dup S04_dups_chk
## 1 0.00000 0.000026 17.911928 0.00000 0.000026 1.200778917
## 2 87.10298 5.579682 12.595546 93.19858 5.970155 6.695317107
## 3 98.92990 6.337292 13.732764 99.03805 6.344220 0.008289978
## 4 66.39019 4.252859 9.116204 66.09623 4.234028 0.405212272
## 5 80.18346 5.136430 8.535371 77.40967 4.958746 3.920528928
##      S04_dups_flag Cl_dups_chk Cl_dups_flag S04_dups_cv S04_dups_cv_flag
## 1      YES       NaN     <NA>  0.8490789      YES
## 2      YES     6.7615540     YES  4.7343041      YES
## 3      YES     0.1092568     YES  0.0058619      YES
## 4      YES     0.4437552     YES  0.2865283      YES
## 5      YES     3.5201930     YES  2.7722326      YES

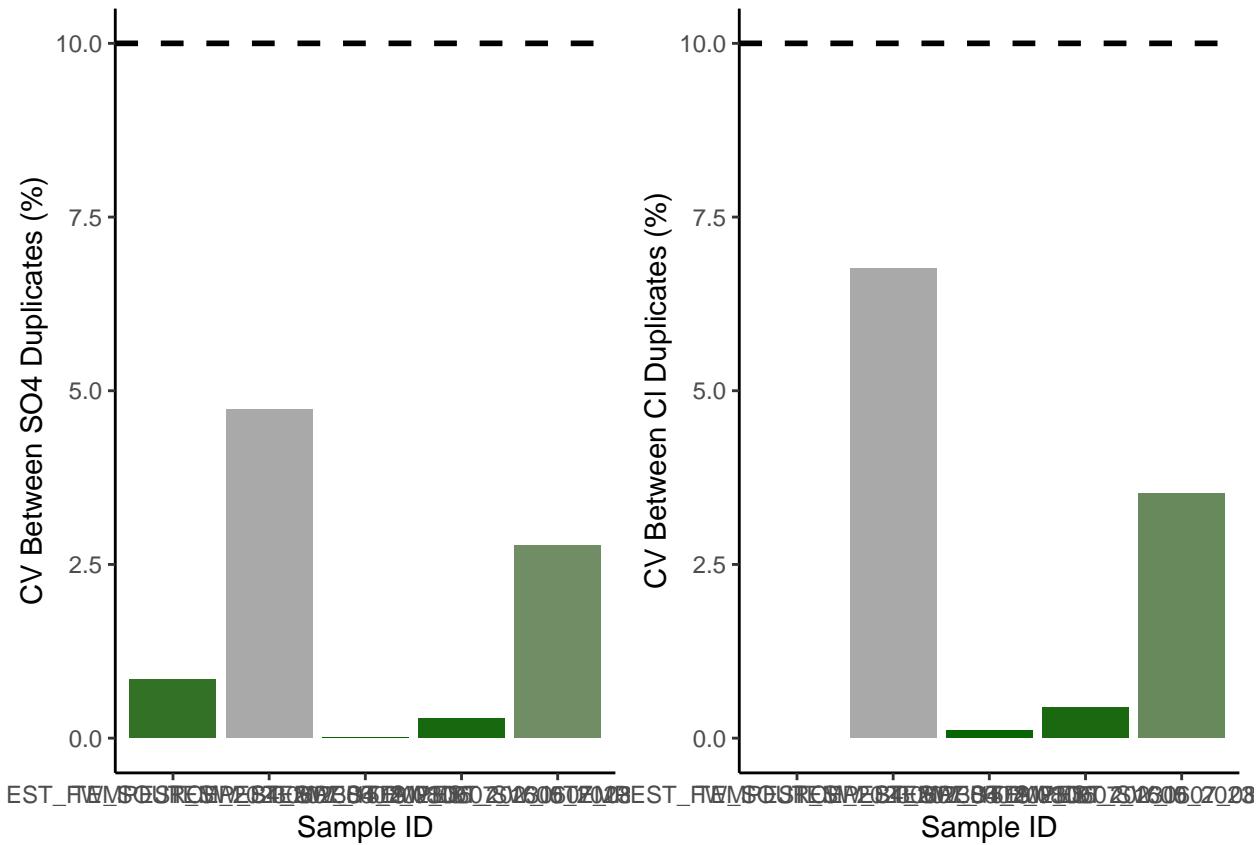
#plot dups output as a bar graph to easily check - want any over 10% to be red need to work on this
Sdupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = S04_dups_cv, fill=S04_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="CV Between S04 Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
                                              color = "black", size=1)

Cldupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = Cl_dups_chk, fill=Cl_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="CV Between Cl Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
                                              color = "black", size=1)

ggarrange(Sdupsbar, Cldupsbar, ncol=2, nrow=1)

## Warning: Removed 1 row containing missing values or values outside the scale range
## ('geom_bar()').

```



```
#check for percent of no, reruns to see if it would warrant reruns
Perc_dups <- QAdups %>%
  group_by(SO4_dups_flag) %>%
  summarise(S_no_rows = length(SO4_dups_flag))
Perc_dups1 <- QAdups %>%
  group_by(Cl_dups_flag) %>%
  summarise(Cl_no_rows = length(Cl_dups_flag))
colnames(Perc_dups) <- c("Flag", "S_no_rows")
colnames(Perc_dups1) <- c("Flag", "Cl_no_rows")
Perc_dups2 <- cbind(Perc_dups, Perc_dups1)

Perc_dups2$Total <- length(QAdups$SO4_dups_flag)
Perc_dups2$S_Percent <- (Perc_dups2$S_no_rows / Perc_dups2$Total)*100
Perc_dups2$Cl_Percent <- (Perc_dups2$Cl_no_rows / Perc_dups2$Total)*100
head(Perc_dups2)
```

```
##   Flag S_no_rows Flag Cl_no_rows Total S_Percent Cl_Percent
## 1 YES      5 YES      4     5    100     80       80
## 2 YES      5 <NA>      1     5    100      20       20
```

Pull out spikes and check with dionex output conc.

```

##Merge sample data with dilution data
Dilutions <- read.csv("Raw Data/TMP 2023 Sample Dilutions from Dionex.csv")
sampledat_dils <- merge(sampledat, Dilutions, by = "Sample_ID")
unique(sampledat_dils$Dilution)

## [1] 50.00 1.10 1.25 56.51 52.74

#Show me the data that we have from the calculations
head(sampledat_dils)

##                                     Sample_ID S04_ppm S04_Area   Cl_ppm Cl_Area
## 1 TEMPEST_ESTUARY_SOURCE_20230607_0710 581.2849 13.5575 4335.7195 122.3274
## 2 TEMPEST_ESTUARY_SOURCE_20230607_1346 547.2306 12.7632 4145.0144 116.9468
## 3 TEMPEST_ESTUARY_SOURCE_20230607_1700 553.9434 12.9198 4153.6146 117.1895
## 4          TEMPEST_FW_B4_20230607_0800 11.1494 11.8200 27.9934 35.9001
## 5          TEMPEST_FW_B4_20230607_1600  2.4624  2.2973 206.4674 233.0096
## 6          TEMPEST_FW_C3_20230607_0800 10.7750 11.4231 23.9976 30.7756
##           S04_mM      Cl_mM  salinity Dilution
## 1 18.13115721 122.3052045 7.83467114    50.00
## 2 17.06895197 116.9256530 7.49006702    50.00
## 3 17.27833437 117.1682539 7.50560758    50.00
## 4  0.34776669   0.7896587 0.05061007     1.10
## 5  0.07680599   5.8241862 0.37311259     1.25
## 6  0.33608858   0.6769422 0.04338966     1.10

#pull out any rows that have "spk" in the SampleID column
spks <- sampledat_dils %>%
  filter(str_detect(Sample_ID, "spk"))      #have to change this to match data
head(spks)

##                                     Sample_ID S04_ppm S04_Area   Cl_ppm Cl_Area
## 1 TEMPEST_FW_SOURCE_20230607_0611_spk 16.0473 17.0125 23.1723 29.7173
## 2          TEMPEST_SW_E3_20230607_1600_spk 332.1500 7.7468 2355.1999 66.4493
## 3          TEMPEST_SW_I5_20230607_1600_spk 468.0219 10.9158 3179.1880 89.6971
## 4 TEMPEST_SW_SOURCE_20230607_0621_spk 563.1599 13.1347 4002.2094 112.9177
## 5  TEMPEST_SW_SOURCE_20230607_1515spk 535.8099 12.4968 4345.0138 122.5896
##           S04_mM      Cl_mM  salinity Dilution
## 1 0.5005396   0.6536615 0.04189835     1.1
## 2 10.3602620   66.4372327 4.25587222    50.0
## 3 14.5983125   89.6809027 5.74481872    50.0
## 4 17.5658110 112.8973032 7.23201839    50.0
## 5 16.7127230 122.5673850 7.85146594    50.0

#remove the dup from these IDs so we will have duplicate sample names
spks$Sample_ID<-gsub("_spk","",as.character(spks$Sample_ID))
spks <- spks[,c(1,6)]
colnames(spks) <- c('Sample_ID', 'S04_mM_spk')
head(spks)

##                                     Sample_ID S04_mM_spk
```

```

## 1      TEMPEST_FW_SOURCE_20230607_0611  0.5005396
## 2      TEMPEST_SW_E3_20230607_1600 10.3602620
## 3      TEMPEST_SW_I5_20230607_1600 14.5983125
## 4      TEMPEST_SW_SOURCE_20230607_0621 17.5658110
## 5 TEMPEST_SW_SOURCE_20230607_1515spk 16.7127230

#put it back together with the old data set and look for duplicates
QAspks <- merge(sampledat_dils, spks)
head(QAspks)

##                                     Sample_ID  SO4_ppm SO4_Area   Cl_ppm  Cl_Area
## 1      TEMPEST_FW_SOURCE_20230607_0611 12.7760 13.5445 22.8178 29.2627
## 2      TEMPEST_SW_E3_20230607_1600 259.0745  6.0425 2327.3879 65.6646
## 3      TEMPEST_SW_I5_20230607_1600 336.2264  7.8419 3120.4635 88.0403
## 4      TEMPEST_SW_SOURCE_20230607_0621 571.0464 13.3187 4300.3781 121.3302
## 5 TEMPEST_SW_SOURCE_20230607_1515spk 535.8099 12.4968 4345.0138 122.5896
##           SO4_mM     Cl_mM    salinity Dilution SO4_mM_spk
## 1  0.3985028  0.6436615 0.04125776       1.1  0.5005396
## 2  8.0809264  65.6526911 4.20561594      50.0 10.3602620
## 3 10.4874111  88.0243583 5.63870354      50.0 14.5983125
## 4 17.8118029 121.3082680 7.77080923      50.0 17.5658110
## 5 16.7127230 122.5673850 7.85146594      50.0 16.7127230

#now we need to calculate the spike concentration and calculate the spike recovery
spkconc <- (250/smw)      # in mM
spkvolt <- 10              # in uL
spkvolt <- spkvolt/1000000
#spike for these samples was 10uL of the 250mM standard
QAspks$SO4_spk_Conc <- (spkconc)*spkvolt          # mmoles of SO4
head(QAspks)

##                                     Sample_ID  SO4_ppm SO4_Area   Cl_ppm  Cl_Area
## 1      TEMPEST_FW_SOURCE_20230607_0611 12.7760 13.5445 22.8178 29.2627
## 2      TEMPEST_SW_E3_20230607_1600 259.0745  6.0425 2327.3879 65.6646
## 3      TEMPEST_SW_I5_20230607_1600 336.2264  7.8419 3120.4635 88.0403
## 4      TEMPEST_SW_SOURCE_20230607_0621 571.0464 13.3187 4300.3781 121.3302
## 5 TEMPEST_SW_SOURCE_20230607_1515spk 535.8099 12.4968 4345.0138 122.5896
##           SO4_mM     Cl_mM    salinity Dilution SO4_mM_spk SO4_spk_Conc
## 1  0.3985028  0.6436615 0.04125776       1.1  0.5005396 7.797879e-05
## 2  8.0809264  65.6526911 4.20561594      50.0 10.3602620 7.797879e-05
## 3 10.4874111  88.0243583 5.63870354      50.0 14.5983125 7.797879e-05
## 4 17.8118029 121.3082680 7.77080923      50.0 17.5658110 7.797879e-05
## 5 16.7127230 122.5673850 7.85146594      50.0 16.7127230 7.797879e-05

#need to determine dilution factors and initial amount of sample added
#if your samples are all the same dilution just use the first line
#for Steph / COMPASS this depends on the site so...
# QAspks$Dilution <- 1
# QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "TEMPEST"), 50, QAspks$Dilution)
# QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 50, QAspks$Dilution)
# QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 50, QAspks$Dilution)
# QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 100, QAspks$Dilution)

```

```

# QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 50, QAspks$Dilution)
# QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 50, QAspks$Dilution)
# QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 100, QAspks$Dilution)
# QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 100, QAspks$Dilution)
# QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 100, QAspks$Dilution)
# QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 200, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$Dilution)

#Set Sample volumes in uL
unique(QAspks$Dilution)

## [1] 1.1 50.0

QAspks$SampleVol <- 1
QAspks$SampleVol <- ifelse(QAspks$Dilution == 50, 1483, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 56.85, 1483, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 51.16, 1483, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 49.27, 1483, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 56.51, 1483, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 5.15, 1500, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 1.25, 1500, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 1.1, 1500, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 52.74, 1483, QAspks$SampleVol)

# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "TEMPEST"), 1483, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 1501, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 1501, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 1475, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 1501, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 1501, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 1475, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 1475, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 1475, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 1462, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$SampleVol)

#change sample volume to L
QAspks$SampleVol <- QAspks$SampleVol/1000000
head(QAspks)

##                                     Sample_ID  SO4_ppm  SO4_Area      Cl_ppm  Cl_Area
## 1 TEMPEST_FW_SOURCE_20230607_0611 12.7760  13.5445  22.8178 29.2627
## 2 TEMPEST_SW_E3_20230607_1600 259.0745   6.0425 2327.3879 65.6646
## 3 TEMPEST_SW_I5_20230607_1600 336.2264   7.8419 3120.4635 88.0403
## 4 TEMPEST_SW_SOURCE_20230607_0621 571.0464  13.3187 4300.3781 121.3302
## 5 TEMPEST_SW_SOURCE_20230607_1515spk 535.8099  12.4968 4345.0138 122.5896
##          SO4_mM      Cl_mM  salinity Dilution SO4_mM_spk SO4_spk_Conc SampleVol
## 1 0.3985028 0.6436615 0.04125776 1.1 0.5005396 7.797879e-05 0.001500

```

```

## 2 8.0809264 65.6526911 4.20561594      50.0 10.3602620 7.797879e-05 0.001483
## 3 10.4874111 88.0243583 5.63870354     50.0 14.5983125 7.797879e-05 0.001483
## 4 17.8118029 121.3082680 7.77080923     50.0 17.5658110 7.797879e-05 0.001483
## 5 16.7127230 122.5673850 7.85146594     50.0 16.7127230 7.797879e-05 0.001483

#gives us the total SO4 in the sample in mmoles
QAspks$SO4_Total_unspkd <- (QAspks$SO4_mM/QAspks$Dilution)*(QAspks$SampleVol)

##total SO4 in spiked sample in mmoles
QAspks$SO4_Total_spkd <- (QAspks$SO4_mM_spk/QAspks$Dilution)*(QAspks$SampleVol+spkvol)

QAspks$SO4_expctd_spkd <- (QAspks$SO4_Total_unspkd + QAspks$SO4_spk_Conc)
QAspks$spk_recovery <- (QAspks$SO4_Total_spkd/QAspks$SO4_expctd_spkd)*100
QAspks$SO4_spks_flag <- ifelse(QAspks$spk_recovery <=120 & QAspks$spk_recovery >=80 , 'YES', 'NO, rerun'

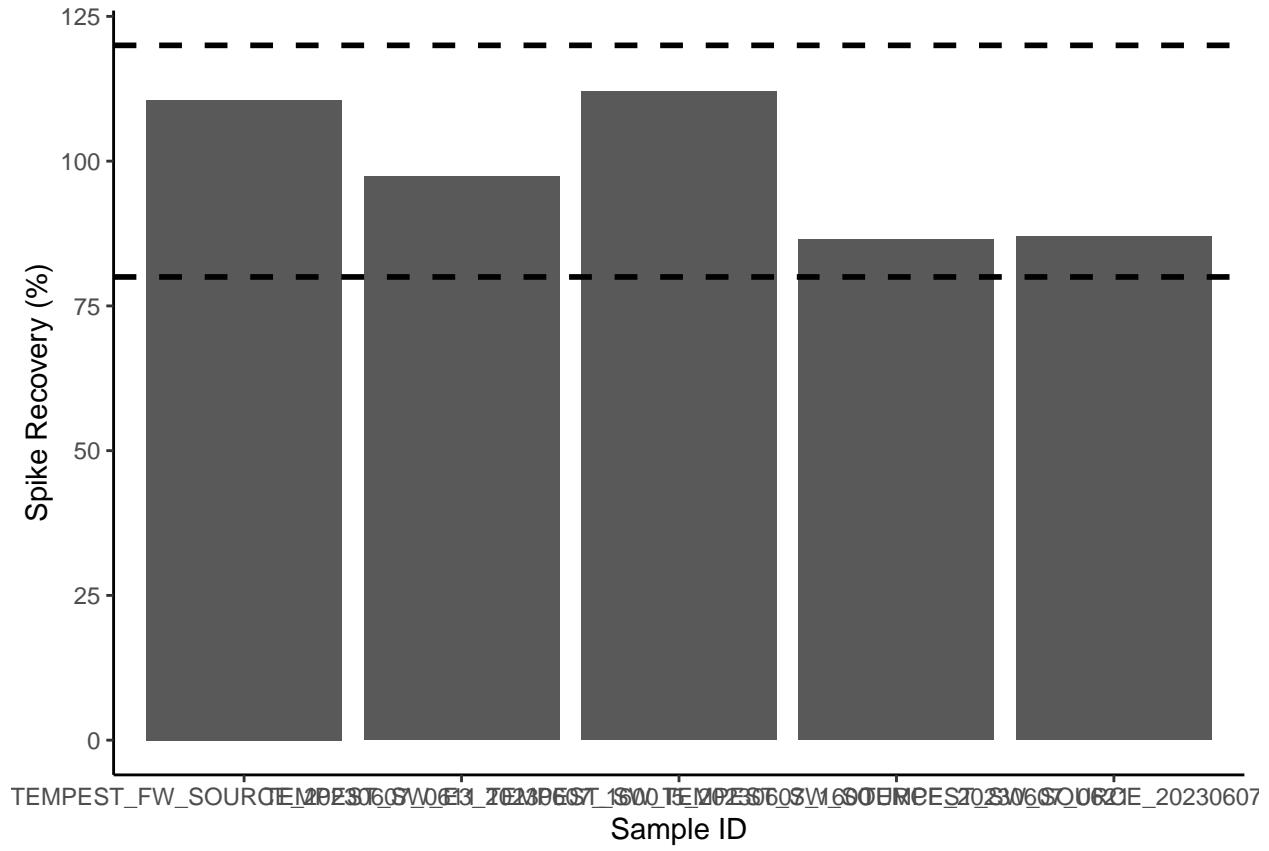
head(QAspks)

##                                     Sample_ID  SO4_ppm SO4_Area      Cl_ppm  Cl_Area
## 1 TEMPEST_FW_SOURCE_20230607_0611 12.7760 13.5445 22.8178 29.2627
## 2 TEMPEST_SW_E3_20230607_1600 259.0745  6.0425 2327.3879 65.6646
## 3 TEMPEST_SW_I5_20230607_1600 336.2264  7.8419 3120.4635 88.0403
## 4 TEMPEST_SW_SOURCE_20230607_0621 571.0464 13.3187 4300.3781 121.3302
## 5 TEMPEST_SW_SOURCE_20230607_1515spk 535.8099 12.4968 4345.0138 122.5896
##          SO4_mM      Cl_mM  salinity Dilution SO4_mM_spk SO4_spk_Conc SampleVol
## 1 0.3985028 0.6436615 0.04125776      1.1 0.5005396 7.797879e-05 0.001500
## 2 8.0809264 65.6526911 4.20561594      50.0 10.3602620 7.797879e-05 0.001483
## 3 10.4874111 88.0243583 5.63870354     50.0 14.5983125 7.797879e-05 0.001483
## 4 17.8118029 121.3082680 7.77080923     50.0 17.5658110 7.797879e-05 0.001483
## 5 16.7127230 122.5673850 7.85146594     50.0 16.7127230 7.797879e-05 0.001483
##   SO4_Total_unspkd SO4_Total_spkd SO4_expctd_spkd spk_recovery SO4_spks_flag
## 1 0.0005434129 0.0006871044 0.0006213917 110.57508 YES
## 2 0.0002396803 0.0003093574 0.0003176591 97.38662 YES
## 3 0.0003110566 0.0004359056 0.0003890354 112.04780 YES
## 4 0.0005282981 0.0005245151 0.0006062769 86.51412 YES
## 5 0.0004956994 0.0004990419 0.0005736782 86.98987 YES

#plot spk recoveries output as a bar graph to easily check - want any over 10% to be red need to work on
spksbar <- ggplot(data = QAspks, aes(x = Sample_ID, y = spk_recovery)) +
  geom_bar(stat = 'identity') +
  scale_fill_manual(values=c("lightblue4")) +
  theme_classic() + labs(x= "Sample ID", y="Spike Recovery (%)") +
  theme(legend.position="none") +
  geom_hline(yintercept=80, linetype="dashed", color = "black", size=1) +
  geom_hline(yintercept=120, linetype="dashed", color = "black", size=1)

spksbar

```



```
#check for percent of no, reruns to see if it would warrant reruns
Perc_spks <- QAspks %>%
  group_by(S04_spks_flag) %>%
  summarise(no_rows = length(S04_spks_flag))
Perc_spks$Total <- length(QAspks$S04_spks_flag)
Perc_spks$Percent <- (Perc_spks$no_rows / Perc_spks$Total)*100
head(Perc_spks)
```

```
## # A tibble: 1 x 4
##   S04_spks_flag no_rows Total Percent
##   <chr>          <int> <int>   <dbl>
## 1 YES              5      5     100
```

Pull out spikes and check with area calc

```
##Merge sample data with dilution data
Dilutions <- read.csv("Raw Data/TMP 2023 Sample Dilutions from Dionex.csv")
sampledat_dils <- merge(sampledat, Dilutions, by = "Sample_ID")
unique(sampledat_dils$Dilution)

## [1] 50.00 1.10 1.25 56.51 52.74
```

```

#Show me the data that we have from the calculations
head(sampledat_dils)

##                                     Sample_ID  S04_ppm S04_Area      Cl_ppm  Cl_Area
## 1 TEMPEST_ESTUARY_SOURCE_20230607_0710 581.2849 13.5575 4335.7195 122.3274
## 2 TEMPEST_ESTUARY_SOURCE_20230607_1346 547.2306 12.7632 4145.0144 116.9468
## 3 TEMPEST_ESTUARY_SOURCE_20230607_1700 553.9434 12.9198 4153.6146 117.1895
## 4          TEMPEST_FW_B4_20230607_0800 11.1494 11.8200 27.9934 35.9001
## 5          TEMPEST_FW_B4_20230607_1600  2.4624  2.2973 206.4674 233.0096
## 6          TEMPEST_FW_C3_20230607_0800 10.7750 11.4231 23.9976 30.7756
##           S04_mM      Cl_mM  salinity Dilution
## 1 18.13115721 122.3052045 7.83467114    50.00
## 2 17.06895197 116.9256530 7.49006702    50.00
## 3 17.27833437 117.1682539 7.50560758    50.00
## 4  0.34776669   0.7896587 0.05061007    1.10
## 5  0.07680599   5.8241862 0.37311259    1.25
## 6  0.33608858   0.6769422 0.04338966    1.10

#now use area of all samples and calculate undilution corrected concentrations in ug/mL
sampledat_dils$S04_ugmL <- ((sampledat_dils$S04_Area-S04_Int)/S04_Slope
sampledat_dils$Cl_ugmL <- (sampledat_dils$Cl_Area-Cl_Int)/Cl_Slope
head(sampledat_dils)

##                                     Sample_ID  S04_ppm S04_Area      Cl_ppm  Cl_Area
## 1 TEMPEST_ESTUARY_SOURCE_20230607_0710 581.2849 13.5575 4335.7195 122.3274
## 2 TEMPEST_ESTUARY_SOURCE_20230607_1346 547.2306 12.7632 4145.0144 116.9468
## 3 TEMPEST_ESTUARY_SOURCE_20230607_1700 553.9434 12.9198 4153.6146 117.1895
## 4          TEMPEST_FW_B4_20230607_0800 11.1494 11.8200 27.9934 35.9001
## 5          TEMPEST_FW_B4_20230607_1600  2.4624  2.2973 206.4674 233.0096
## 6          TEMPEST_FW_C3_20230607_0800 10.7750 11.4231 23.9976 30.7756
##           S04_mM      Cl_mM  salinity Dilution  S04_ugmL  Cl_ugmL
## 1 18.13115721 122.3052045 7.83467114    50.00 11.625713 86.71442
## 2 17.06895197 116.9256530 7.49006702    50.00 10.944591 82.90027
## 3 17.27833437 117.1682539 7.50560758    50.00 11.078877 83.07231
## 4  0.34776669   0.7896587 0.05061007    1.10 10.135786 25.44856
## 5  0.07680599   5.8241862 0.37311259    1.25 1.969955 165.17390
## 6  0.33608858   0.6769422 0.04338966    1.10 9.795439 21.81594

#pull out any rows that have "spk" in the SampleID column
spks <- sampledat_dils %>%
  filter(str_detect(Sample_ID, "spk"))      #have to change this to match data
head(spks)

##                                     Sample_ID  S04_ppm S04_Area      Cl_ppm  Cl_Area
## 1 TEMPEST_FW_SOURCE_20230607_0611_spk 16.0473 17.0125 23.1723 29.7173
## 2          TEMPEST_SW_E3_20230607_1600_spk 332.1500  7.7468 2355.1999 66.4493
## 3          TEMPEST_SW_I5_20230607_1600_spk 468.0219 10.9158 3179.1880 89.6971
## 4 TEMPEST_SW_SOURCE_20230607_0621_spk 563.1599 13.1347 4002.2094 112.9177
## 5  TEMPEST_SW_SOURCE_20230607_1515spk 535.8099 12.4968 4345.0138 122.5896
##           S04_mM      Cl_mM  salinity Dilution  S04_ugmL  Cl_ugmL
## 1  0.5005396  0.6536615 0.04189835     1.1 14.588417 21.06574
## 2 10.3602620  66.4372327 4.25587222    50.0  6.642967 47.10402

```

```

## 3 14.5983125 89.6809027 5.74481872      50.0 9.360423 63.58373
## 4 17.5658110 112.8973032 7.23201839     50.0 11.263157 80.04415
## 5 16.7127230 122.5673850 7.85146594     50.0 10.716150 86.90029

```

```

## Only keep the columns that we need
spks <- spks[,c(1,9,10)]
head(spks)

```

```

##                               Sample_ID Dilution SO4_ugmL
## 1 TEMPEST_FW_SOURCE_20230607_0611_spk      1.1 14.588417
## 2 TEMPEST_SW_E3_20230607_1600_spk        50.0 6.642967
## 3 TEMPEST_SW_I5_20230607_1600_spk        50.0 9.360423
## 4 TEMPEST_SW_SOURCE_20230607_0621_spk     50.0 11.263157
## 5 TEMPEST_SW_SOURCE_20230607_1515spk      50.0 10.716150

```

```

#remove the spk from these IDs so we will have duplicate sample names
spks$Sample_ID<-gsub("_spk","",as.character(spks$Sample_ID))
colnames(spks) <- c('Sample_ID', 'Dilution', 'SO4_ugmL_spk')
head(spks)

```

```

##                               Sample_ID Dilution SO4_ugmL_spk
## 1 TEMPEST_FW_SOURCE_20230607_0611      1.1 14.588417
## 2 TEMPEST_SW_E3_20230607_1600        50.0 6.642967
## 3 TEMPEST_SW_I5_20230607_1600        50.0 9.360423
## 4 TEMPEST_SW_SOURCE_20230607_0621     50.0 11.263157
## 5 TEMPEST_SW_SOURCE_20230607_1515spk  50.0 10.716150

```

```

#put it back together with the old data set and look for duplicates
QAspks <- merge(sampledat_dils, spks)
head(QAspks)

```

```

##                               Sample_ID Dilution SO4_ppm SO4_Area Cl_ppm
## 1 TEMPEST_FW_SOURCE_20230607_0611      1.1 12.7760 13.5445 22.8178
## 2 TEMPEST_SW_E3_20230607_1600        50.0 259.0745 6.0425 2327.3879
## 3 TEMPEST_SW_I5_20230607_1600        50.0 336.2264 7.8419 3120.4635
## 4 TEMPEST_SW_SOURCE_20230607_0621     50.0 571.0464 13.3187 4300.3781
## 5 TEMPEST_SW_SOURCE_20230607_1515spk  50.0 535.8099 12.4968 4345.0138
##             Cl_Area   SO4_mM   Cl_mM salinity SO4_ugmL Cl_ugmL SO4_ugmL_spk
## 1 29.2627 0.3985028 0.6436615 0.04125776 11.614565 20.74349 14.588417
## 2 65.6646 8.0809264 65.6526911 4.20561594 5.181510 46.54777 6.642967
## 3 88.0403 10.4874111 88.0243583 5.63870354 6.724517 62.40927 9.360423
## 4 121.3302 17.8118029 121.3082680 7.77080923 11.420939 86.00754 11.263157
## 5 122.5896 16.7127230 122.5673850 7.85146594 10.716150 86.90029 10.716150

```

```

#now we need to calculate the spike concentration and calculate the spike recovery
spkconc <- (250)      # in ug
spkvol <- 10           # in uL
spkvol <- spkvol/1000
#spike for these samples was 10uL of the 250mM standard
QAspks$SO4_spk_Conc <- (spkconc)*spkvol          # mmoles of SO4
head(QAspks)

```

```

##                                     Sample_ID Dilution S04_ppm S04_Area    Cl_ppm
## 1      TEMPEST_FW_SOURCE_20230607_0611      1.1 12.7760 13.5445 22.8178
## 2      TEMPEST_SW_E3_20230607_1600     50.0 259.0745  6.0425 2327.3879
## 3      TEMPEST_SW_I5_20230607_1600     50.0 336.2264  7.8419 3120.4635
## 4      TEMPEST_SW_SOURCE_20230607_0621     50.0 571.0464 13.3187 4300.3781
## 5 TEMPEST_SW_SOURCE_20230607_1515spk     50.0 535.8099 12.4968 4345.0138
##          Cl_Area     S04_mM      Cl_mM salinity S04_ugmL Cl_ugmL S04_ugmL_spk
## 1 29.2627 0.3985028 0.6436615 0.04125776 11.614565 20.74349 14.588417
## 2 65.6646 8.0809264 65.6526911 4.20561594 5.181510 46.54777 6.642967
## 3 88.0403 10.4874111 88.0243583 5.63870354 6.724517 62.40927 9.360423
## 4 121.3302 17.8118029 121.3082680 7.77080923 11.420939 86.00754 11.263157
## 5 122.5896 16.7127230 122.5673850 7.85146594 10.716150 86.90029 10.716150
##          S04_spk_Conc
## 1          2.5
## 2          2.5
## 3          2.5
## 4          2.5
## 5          2.5

#need to determine dilution factors and initial amount of sample added
#if your samples are all the same dilution just use the first line
#for Steph / COMPASS this depends on the site so...
#QAspks$Dilution <- 1
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 50, QAspks$Dilution)
##$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 50, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 100, QAspks$Dilution)
##$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 50, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 50, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 100, QAspks$Dilution)
##$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 200, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$Dilution)

#Set Sample volumes in uL
QAspks$SampleVol <- 1
QAspks$SampleVol <- ifelse(QAspks$Dilution == 50, 1483, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 1.1, 1500, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 5.15, 1500, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 4.53, 1500, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 56.51, 1483, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 1.25, 1500, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 1.1, 1500, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(QAspks$Dilution == 52.74, 1483, QAspks$SampleVol)

# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "TEMPEST"), 1501, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 1501, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 1501, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 1475, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_PPR_UP"), 1501, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_PPR_TR"), 1501, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_PPR_WC"), 1475, QAspks$SampleVol)

```

```

# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 1501, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 1501, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 1475, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 1475, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 1475, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 1462, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$SampleVol)
# QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$SampleVol)

#change sample volume to L
QAspks$SampleVol <- QAspks$SampleVol/1000
head(QAspks)

```

```

##                                     Sample_ID Dilution S04_ppm S04_Area Cl_ppm
## 1      TEMPEST_FW_SOURCE_20230607_0611      1.1 12.7760 13.5445 22.8178
## 2      TEMPEST_SW_E3_20230607_1600     50.0 259.0745  6.0425 2327.3879
## 3      TEMPEST_SW_I5_20230607_1600     50.0 336.2264  7.8419 3120.4635
## 4      TEMPEST_SW_SOURCE_20230607_0621    50.0 571.0464 13.3187 4300.3781
## 5 TEMPEST_SW_SOURCE_20230607_1515spk    50.0 535.8099 12.4968 4345.0138
##   Cl_Area     S04_mM     Cl_mM salinity S04_ugmL Cl_ugmL S04_ugmL_spk
## 1 29.2627 0.3985028 0.6436615 0.04125776 11.614565 20.74349 14.588417
## 2 65.6646 8.0809264 65.6526911 4.20561594  5.181510 46.54777 6.642967
## 3 88.0403 10.4874111 88.0243583 5.63870354  6.724517 62.40927 9.360423
## 4 121.3302 17.8118029 121.3082680 7.77080923 11.420939 86.00754 11.263157
## 5 122.5896 16.7127230 122.5673850 7.85146594 10.716150 86.90029 10.716150
##   S04_spk_Conc SampleVol
## 1          2.5      1.500
## 2          2.5      1.483
## 3          2.5      1.483
## 4          2.5      1.483
## 5          2.5      1.483

```

```

#gives us the total SO4 in the sample in mmoles
QAspks$S04_Total_unspkd <- QAspks$S04_ugmL*QAspks$SampleVol

```

```

##total SO4 in spiked sample in mmoles
QAspks$S04_Total_spkd <- (QAspks$S04_ugmL_spk)*(QAspks$SampleVol+spkvol)

```

```

QAspks$S04_expctd_spkd <- (QAspks$S04_Total_unspkd + QAspks$S04_spk_Conc)
QAspks$spk_recovery <- (QAspks$S04_Total_spkd/QAspks$S04_expctd_spkd)*100
QAspks$S04_spks_flag <- ifelse(QAspks$spk_recovery <=120 & QAspks$spk_recovery >=80 , 'YES', 'NO', rerun)

```

```

head(QAspks)

```

```

##                                     Sample_ID Dilution S04_ppm S04_Area Cl_ppm
## 1      TEMPEST_FW_SOURCE_20230607_0611      1.1 12.7760 13.5445 22.8178
## 2      TEMPEST_SW_E3_20230607_1600     50.0 259.0745  6.0425 2327.3879
## 3      TEMPEST_SW_I5_20230607_1600     50.0 336.2264  7.8419 3120.4635
## 4      TEMPEST_SW_SOURCE_20230607_0621    50.0 571.0464 13.3187 4300.3781
## 5 TEMPEST_SW_SOURCE_20230607_1515spk    50.0 535.8099 12.4968 4345.0138
##   Cl_Area     S04_mM     Cl_mM salinity S04_ugmL Cl_ugmL S04_ugmL_spk

```

```

## 1 29.2627 0.3985028 0.6436615 0.04125776 11.614565 20.74349 14.588417
## 2 65.6646 8.0809264 65.6526911 4.20561594 5.181510 46.54777 6.642967
## 3 88.0403 10.4874111 88.0243583 5.63870354 6.724517 62.40927 9.360423
## 4 121.3302 17.8118029 121.3082680 7.77080923 11.420939 86.00754 11.263157
## 5 122.5896 16.7127230 122.5673850 7.85146594 10.716150 86.90029 10.716150
##   S04_spk_Conc SampleVol S04_Total_unspkd S04_Total_spkd S04_exptd_spkd
## 1          2.5     1.500      17.421848     22.02851    19.92185
## 2          2.5     1.483      7.684179      9.91795    10.18418
## 3          2.5     1.483      9.972458     13.97511    12.47246
## 4          2.5     1.483     16.937252     16.81589    19.43725
## 5          2.5     1.483     15.892050     15.99921    18.39205
##   spk_recovery S04_spks_flag
## 1       110.57463      YES
## 2        97.38586      YES
## 3       112.04777      YES
## 4        86.51373      YES
## 5        86.98982      YES

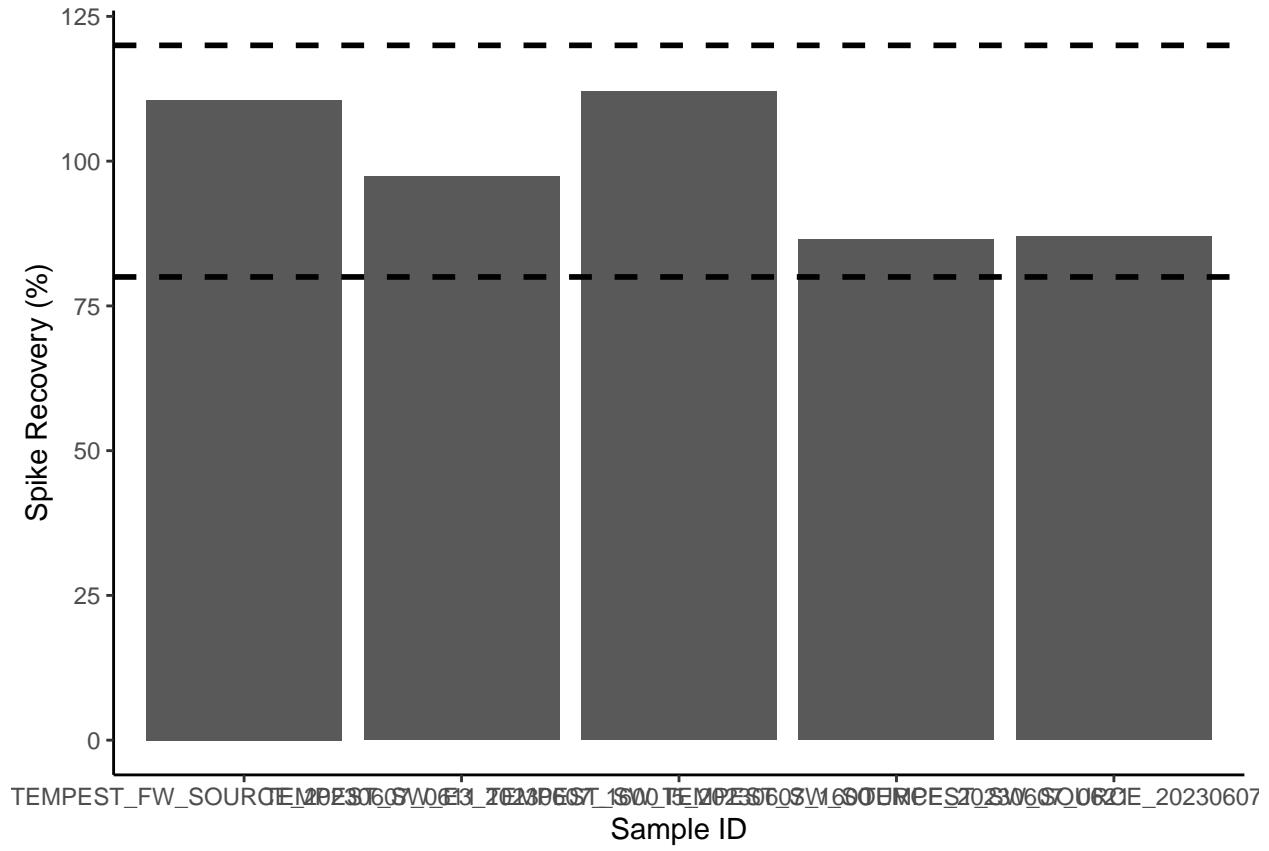
```

```

#plot spk recoveries output as a bar graph to easily check - want any over 10% to be red need to work on this
spksbar <- ggplot(data = QAspks, aes(x = Sample_ID, y = spk_recovery)) +
  geom_bar(stat = 'identity') +
  scale_fill_manual(values=c("lightblue4")) +
  theme_classic() + labs(x= "Sample ID", y="Spike Recovery (%)") +
  theme(legend.position="none") +
  geom_hline(yintercept=80, linetype="dashed", color = "black", size=1) +
  geom_hline(yintercept=120, linetype="dashed", color = "black", size=1)

```

```
spksbar
```



```
#check for percent of no, reruns to see if it would warrant reruns
Perc_spks <- QAspks %>%
  group_by(S04_spks_flag) %>%
  summarise(no_rows = length(S04_spks_flag))
Perc_spks$Total <- length(QAspks$S04_spks_flag)
Perc_spks$Percent <- (Perc_spks$no_rows / Perc_spks$Total)*100
head(Perc_spks)
```

```
## # A tibble: 1 x 4
##   S04_spks_flag no_rows Total Percent
##   <chr>          <int> <int>   <dbl>
## 1 YES              5      5     100
```

## Make final dataframe with IDs

```
#for steph <- pull out identifiers of the sample names
#pull the sample ID and separate it by the underscores
IDs <- data.frame(do.call('rbind', strsplit(as.character(sampledat2$Sample_ID), '_', fixed=TRUE)))
colnames(IDs) <- c("Project" , "Plot", "Grid", "Date", "Time")
head(IDs)
```

```
## Project Plot Grid Date Time
## 1 TEMPEST ESTUARY SOURCE 20230607 0710
```

```

## 2 TEMPEST ESTUARY SOURCE 20230607 1346
## 3 TEMPEST ESTUARY SOURCE 20230607 1700
## 4 TEMPEST      FW      B4 20230607 0800
## 5 TEMPEST      FW      B4 20230607 1600
## 6 TEMPEST      FW      C3 20230607 0800

```

```

#rejoin them to the dataframe
alldat <- cbind(IDs, sampledat2)
head(alldat)

```

	Project	Plot	Grid	Date	Time	Sample_ID	
## 1	TEMPEST	ESTUARY	SOURCE	20230607	0710	TEMPEST_ESTUARY_SOURCE_20230607_0710	
## 2	TEMPEST	ESTUARY	SOURCE	20230607	1346	TEMPEST_ESTUARY_SOURCE_20230607_1346	
## 3	TEMPEST	ESTUARY	SOURCE	20230607	1700	TEMPEST_ESTUARY_SOURCE_20230607_1700	
## 4	TEMPEST		FW	20230607	0800	TEMPEST_FW_B4_20230607_0800	
## 5	TEMPEST		FW	20230607	1600	TEMPEST_FW_B4_20230607_1600	
## 6	TEMPEST		FW	20230607	0800	TEMPEST_FW_C3_20230607_0800	
	S04_ppm	S04_Area	C1_ppm	C1_Area	S04_mM	C1_mM	salinity
## 1	581.2849	13.5575	4335.7195	122.3274	18.13115721	122.3052045	7.83467114
## 2	547.2306	12.7632	4145.0144	116.9468	17.06895197	116.9256530	7.49006702
## 3	553.9434	12.9198	4153.6146	117.1895	17.27833437	117.1682539	7.50560758
## 4	11.1494	11.8200	27.9934	35.9001	0.34776669	0.7896587	0.05061007
## 5	2.4624	2.2973	206.4674	233.0096	0.07680599	5.8241862	0.37311259
## 6	10.7750	11.4231	23.9976	30.7756	0.33608858	0.6769422	0.04338966

## Make final dataframe with IDs

```

write.csv(alldat, file="Processed Data/COMPASS_TEMPEST_Dionex_ClSo4_Final_20230608.csv")

```

END