

# Dionex\_COMPASS\_TEMPEST

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## Daily Set up

Read in and Format the raw data - change wd & file names (Make sure files are UTF-8 encoded)

```
# SULFATE DATA:
```

```
## Read in raw data file from Dionex - copied and saved as a txt
```

```
Sdat <- read.table("Raw Data/COMPASS_TEMPEST_20230609_S04.txt", sep='\t' , header=T, skip=3)
head(Sdat)
```

```
##      X          X.1          X.2 IC.S04 IC.S04.1 IC.S04.2 IC.S04.3 IC.S04.4
## 1 1 Lab Blank      Unknown    n.a.    n.a.    n.a.    n.a.    n.a.
## 2 2 Lab Blank      Unknown    n.a.    n.a.    n.a.    n.a.    n.a.
## 3 3 Lab Blank      Unknown    n.a.    n.a.    n.a.    n.a.    n.a.
## 4 4 Standard 1 Calibration Standard 4.400 0.4517 6.08 0.5351 2.55
## 5 5 Standard 2 Calibration Standard 4.397 0.9638 6.53 1.1415 5.33
## 6 6 Standard 3 Calibration Standard 4.393 1.9094 7.18 2.2616 10.33
##      IC.S04.5
## 1      n.a.
## 2      n.a.
## 3      n.a.
## 4      M
## 5      M
## 6      M
```

```
## Only keep the columns that we need
```

```
Sdat <- Sdat[ ,c(2,5,7)] # dont need this here
head(Sdat)
```

```
##          X.1 IC.S04.1 IC.S04.3
## 1 Lab Blank    n.a.    n.a.
## 2 Lab Blank    n.a.    n.a.
## 3 Lab Blank    n.a.    n.a.
## 4 Standard 1 0.4517 0.5351
## 5 Standard 2 0.9638 1.1415
## 6 Standard 3 1.9094 2.2616
```

```
## Name the columns correctly
colnames(Sdat) <- c( "Sample_ID", "S04_ppm", "S04_Area")
Sdat$Sample_ID <- as.factor(Sdat$Sample_ID)
Sdat$S04_ppm <- as.numeric(Sdat$S04_ppm)
```

```
## Warning: NAs introduced by coercion
```

```
Sdat$S04_Area <- as.numeric(Sdat$S04_Area)
```

```
## Warning: NAs introduced by coercion
```

```
Sdat <- as.data.frame(Sdat)
head(Sdat)
```

```
##      Sample_ID S04_ppm S04_Area
## 1 Lab Blank      NA      NA
## 2 Lab Blank      NA      NA
## 3 Lab Blank      NA      NA
## 4 Standard 1  0.4517  0.5351
## 5 Standard 2  0.9638  1.1415
## 6 Standard 3  1.9094  2.2616
```

```
#Chloride data
```

```
## Read in raw data file from Dionex - copied and saved as a txt
```

```
Cldat <- read.table("Raw Data/COMPASS_TEMPEST_20230609_Cl.txt", sep='\t' , header=T, skip=3)
head(Cldat)
```

```
##      X      X.1      X.2 IC.Cl IC.Cl.1 IC.Cl.2 IC.Cl.3 IC.Cl.4
## 1 1 Lab Blank      Unknown 3.470  0.0141  22.51  0.0202  0.10
## 2 2 Lab Blank      Unknown  n.a.    n.a.    n.a.    n.a.    n.a.
## 3 3 Lab Blank      Unknown 3.467  0.0076  17.96  0.0109  0.06
## 4 4 Standard 1 Calibration Standard 3.470  5.7586  93.79  8.2504  45.07
## 5 5 Standard 2 Calibration Standard 3.467  11.3682  93.12  16.2873  88.33
## 6 6 Standard 3 Calibration Standard 3.467  20.3417  92.57  29.1437  156.75
##      IC.Cl.5
## 1      MB
## 2     n.a.
## 3      MB
## 4      BM
## 5      BM
## 6      BM
```

```
## Only keep the columns that we need
```

```
Cldat <- Cldat[ ,c(2,5,7)]
head(Cldat)
```

```
##      X.1 IC.Cl.1 IC.Cl.3
## 1 Lab Blank 0.0141 0.0202
## 2 Lab Blank  n.a.   n.a.
## 3 Lab Blank 0.0076 0.0109
## 4 Standard 1 5.7586 8.2504
## 5 Standard 2 11.3682 16.2873
## 6 Standard 3 20.3417 29.1437
```

```
## Name the columns correctly
colnames(Cldat) <- c( "Sample_ID", "Cl_ppm", "Cl_Area")
Cldat$Sample_ID <- as.factor(Cldat$Sample_ID)
Cldat$Cl_ppm <- as.numeric(Cldat$Cl_ppm)
```

```
## Warning: NAs introduced by coercion
```

```
Cldat$Cl_Area <- as.numeric(Cldat$Cl_Area)
```

```
## Warning: NAs introduced by coercion
```

```
Cldat <- as.data.frame(Cldat)
head(Cldat)
```

```
##      Sample_ID Cl_ppm Cl_Area
## 1 Lab Blank  0.0141  0.0202
## 2 Lab Blank      NA      NA
## 3 Lab Blank  0.0076  0.0109
## 4 Standard 1  5.7586  8.2504
## 5 Standard 2 11.3682 16.2873
## 6 Standard 3 20.3417 29.1437
```

```
## Bring the data back together:
all_dat <- merge(Sdat, Cldat, by="Sample_ID")
head(all_dat)
```

```
##      Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area
## 1          457.3600  28.7072 3706.9076 287.1185
## 2          457.3600  28.7072 1073.4812  62.2928
## 3          457.3600  28.7072   0.0076   0.0109
## 4          457.3600  28.7072 1469.0566  71.5412
## 5          457.3600  28.7072      NA      NA
## 6          142.4999   9.0068 3706.9076 287.1185
```

```
## Remove empty lines
all_dat <- all_dat[!(is.na(all_dat$Sample_ID) | all_dat$Sample_ID==""), ]
head(all_dat)
```

```
##      Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area
## 26 Lab Blank      NA      NA 0.0141  0.0202
## 27 Lab Blank      NA      NA      NA      NA
## 28 Lab Blank      NA      NA 0.0076  0.0109
## 29 Lab Blank      NA      NA 0.0091  0.0130
## 30 Lab Blank      NA      NA 0.0160  0.0229
## 31 Lab Blank      NA      NA 0.0224  0.0321
```

```
all_dat <- droplevels(all_dat[!all_dat$Sample_ID == 'Lab Blank',])
head(all_dat)
```

```
##      Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area
## 107 Standard 1  0.4633  0.5488 5.7703 8.2671
## 108 Standard 1  0.4633  0.5488 5.7586 8.2504
## 109 Standard 1  0.4633  0.5488 5.7883 8.2930
## 110 Standard 1  0.4517  0.5351 5.7703 8.2671
## 111 Standard 1  0.4517  0.5351 5.7586 8.2504
## 112 Standard 1  0.4517  0.5351 5.7883 8.2930
```

```
all_dat[is.na(all_dat)] <- 0
```

Pull out standards - could do some checks here if we want

```
stds <- all_dat[grepl("Standard", all_dat$Sample_ID),]
head(stds)
```

```
##      Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area
## 107 Standard 1  0.4633  0.5488 5.7703 8.2671
## 108 Standard 1  0.4633  0.5488 5.7586 8.2504
## 109 Standard 1  0.4633  0.5488 5.7883 8.2930
## 110 Standard 1  0.4517  0.5351 5.7703 8.2671
## 111 Standard 1  0.4517  0.5351 5.7586 8.2504
## 112 Standard 1  0.4517  0.5351 5.7883 8.2930
```

```
stds_chk_S <- stds %>%
  group_by(Sample_ID) %>%
  summarise(mean = mean(S04_ppm), sd = sd(S04_ppm))
stds_chk_S$cv <- (stds_chk_S$sd/stds_chk_S$mean)*100
stds_chk_S$flag <- ifelse(stds_chk_S$cv < 2, 'YES', 'NO, rerun')
head(stds_chk_S)
```

```
## # A tibble: 5 x 5
##   Sample_ID mean      sd      cv flag
##   <fct>     <dbl>   <dbl> <dbl> <chr>
## 1 Standard 1  0.456 0.00532  1.17 YES
## 2 Standard 2  0.963 0.000635  0.0659 YES
## 3 Standard 3  1.91  0.00318  0.167 YES
## 4 Standard 4  8.22  2.02    24.6 NO, rerun
## 5 Standard 5 20.0  0.0280   0.140 YES
```

```
stds_chk_Cl <- stds %>%
  group_by(Sample_ID) %>%
  summarise(mean = mean(Cl_ppm), sd = sd(Cl_ppm))
stds_chk_Cl$cv <- (stds_chk_Cl$sd/stds_chk_Cl$mean)*100
stds_chk_Cl$flag <- ifelse(stds_chk_Cl$cv < 2, 'YES', 'NO, rerun')
head(stds_chk_Cl)
```

```
## # A tibble: 5 x 5
##   Sample_ID mean      sd      cv flag
##   <fct>     <dbl>   <dbl> <dbl> <chr>
## 1 Standard 1  5.77 0.0130  0.224 YES
```

```
## 2 Standard 2 11.4 0.00803 0.0706 YES
## 3 Standard 3 20.4 0.0289 0.142 YES
## 4 Standard 4 100. 0.0318 0.0317 YES
## 5 Standard 5 200. 0.381 0.191 YES
```

```
lmS <- lm(stds$S04_Area ~ stds$S04_ppm)
S04_sum <- summary(lmS)
S04_Slope <- S04_sum$coefficients[2, 1]
S04_Int <- S04_sum$coefficients[1, 1]
```

```
lmCl <- lm(stds$Cl_Area ~ stds$Cl_ppm)
Cl_sum <- summary(lmCl)
Cl_Slope <- Cl_sum$coefficients[2, 1]
Cl_Int <- Cl_sum$coefficients[1, 1]
```

## Calculate mmol/L concentrations

```
#remove standards from sample dataframe
sampledat <- all_dat[!grepl("Standard", all_dat$Sample_ID),]
head(sampledat)
```

```
##           Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area
## 132 TEMEPST_FW_H6_20230608 10.0574 10.8297 0.0000 0.0000
## 133 TEMEPST_SW_C3_20230608 436.0629 10.3300 3597.9994 103.0974
## 134 TEMPEST_FW_B4_20230608 28.2839 28.7072 0.0000 0.0000
## 135 TEMPEST_FW_C3_20230608 11.6026 12.4935 24.1648 31.4737
## 136 TEMPEST_FW_C3_20230608_dup 11.5730 12.4616 24.3234 31.6803
## 137 TEMPEST_FW_D5_20230608 3.1147 3.3538 27.7577 36.1532
```

```
# Constants needed for calculations:
clmw <- 35.45 #molecular weight of Chloride: 35.45
smw <- 32.06 #molecular weight of sulfur: 32.06

# Convert ppm to mmol/L
sampledat$S04_mM <- (sampledat$S04_ppm / smw)
sampledat$Cl_mM <- (sampledat$Cl_ppm / clmw)

# Calculate Salinity
# calculated using the Knudsen equation
# Salinity = 0.03 + 1.8050 * Chlorinity
# Ref: A Practical Handbook of Seawater Analysis by Strickland & Parsons (P. 11)
# = ((1.807*Cl_ppm)+0.026)/1000
sampledat$salinity <- ((1.8070 * sampledat$Cl_ppm) + 0.026) / 1000

head(sampledat)
```

```
##           Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area S04_mM
```

```
## 132      TEMEPST_FW_H6_20230608 10.0574 10.8297 0.0000 0.0000 0.31370555
## 133      TEMEPST_SW_C3_20230608 436.0629 10.3300 3597.9994 103.0974 13.60146288
## 134      TEMPEST_FW_B4_20230608 28.2839 28.7072 0.0000 0.0000 0.88221772
## 135      TEMPEST_FW_C3_20230608 11.6026 12.4935 24.1648 31.4737 0.36190268
## 136 TEMPEST_FW_C3_20230608_dup 11.5730 12.4616 24.3234 31.6803 0.36097941
## 137      TEMPEST_FW_D5_20230608 3.1147 3.3538 27.7577 36.1532 0.09715221
##          Cl_mM      salinity
## 132 0.0000000 0.00002600
## 133 101.4950465 6.50161092
## 134 0.0000000 0.00002600
## 135 0.6816587 0.04369179
## 136 0.6861326 0.04397838
## 137 0.7830099 0.05018416
```

Pull out dups and check with percent difference

```
#Show me the data that we have from the calculations
head(sampledat)
```

```
##          Sample_ID S04_ppm S04_Area  Cl_ppm Cl_Area      S04_mM
## 132      TEMEPST_FW_H6_20230608 10.0574 10.8297 0.0000 0.0000 0.31370555
## 133      TEMEPST_SW_C3_20230608 436.0629 10.3300 3597.9994 103.0974 13.60146288
## 134      TEMPEST_FW_B4_20230608 28.2839 28.7072 0.0000 0.0000 0.88221772
## 135      TEMPEST_FW_C3_20230608 11.6026 12.4935 24.1648 31.4737 0.36190268
## 136 TEMPEST_FW_C3_20230608_dup 11.5730 12.4616 24.3234 31.6803 0.36097941
## 137      TEMPEST_FW_D5_20230608 3.1147 3.3538 27.7577 36.1532 0.09715221
##          Cl_mM      salinity
## 132 0.0000000 0.00002600
## 133 101.4950465 6.50161092
## 134 0.0000000 0.00002600
## 135 0.6816587 0.04369179
## 136 0.6861326 0.04397838
## 137 0.7830099 0.05018416
```

```
#pull out any rows that have "dup" in the SampleID column
dups <- sampledat %>%
  filter(str_detect(Sample_ID, "_dup"))      #have to change this to match data
head(dups)
```

```
##          Sample_ID S04_ppm S04_Area  Cl_ppm Cl_Area
## 1      TEMPEST_FW_C3_20230608_dup 11.5730 12.4616 24.3234 31.6803
## 2 TEMPEST_SW_I5_20230607_0800_dup 0.0000 0.0000 0.0000 0.0000
## 3      TEMPEST_SW_I5_20230608_dup 420.3553 9.9579 3651.4021 104.6276
##          S04_mM      Cl_mM      salinity
## 1 0.3609794 0.6861326 0.04397838
## 2 0.0000000 0.0000000 0.00002600
## 3 13.1115190 103.0014697 6.59810959
```

```
#remove these from sample dataframe in a new dataframe
sampledat2 <- sampledat %>%
  filter(!str_detect(Sample_ID, "_dup")) %>%
```

```
filter(!str_detect(Sample_ID, "_spk"))
head(sampledat2)
```

```
##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 1 TEMEPST_FW_H6_20230608  10.0574  10.8297   0.0000   0.0000  0.31370555
## 2 TEMEPST_SW_C3_20230608 436.0629  10.3300 3597.9994 103.0974 13.60146288
## 3 TEMPEST_FW_B4_20230608  28.2839  28.7072   0.0000   0.0000  0.88221772
## 4 TEMPEST_FW_C3_20230608  11.6026  12.4935  24.1648  31.4737  0.36190268
## 5 TEMPEST_FW_D5_20230608   3.1147   3.3538  27.7577  36.1532  0.09715221
## 6 TEMPEST_FW_E3_20230608 23.9154  24.2733   0.0000   0.0000  0.74595758
##           Cl_mM  salinity
## 1   0.0000000 0.00002600
## 2 101.4950465 6.50161092
## 3   0.0000000 0.00002600
## 4   0.6816587 0.04369179
## 5   0.7830099 0.05018416
## 6   0.0000000 0.00002600
```

```
#remove the dup from these IDs so we will have duplicate sample names
dups$Sample_ID<-gsub("_dup","",as.character(dups$Sample_ID))
dups <- dups[, -c(2:5)]
colnames(dups) <- c('Sample_ID', 'S04_mM_dup', 'Cl_mM_dup', 'salinity_dup')
head(dups)
```

```
##           Sample_ID S04_mM_dup  Cl_mM_dup salinity_dup
## 1 TEMPEST_FW_C3_20230608  0.3609794  0.6861326  0.04397838
## 2 TEMPEST_SW_I5_20230607_0800 0.0000000  0.0000000  0.00002600
## 3 TEMPEST_SW_I5_20230608 13.1115190 103.0014697  6.59810959
```

```
#put it back together with the old data set and look for duplicates
QAdups <- merge(sampledat2, dups)
head(QAdups)
```

```
##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 1 TEMPEST_FW_C3_20230608  11.6026  12.4935  24.1648  31.4737  0.3619027
## 2 TEMPEST_SW_I5_20230608 413.5339   9.7963 3579.4092 102.5647 12.8987492
##           Cl_mM  salinity S04_mM_dup  Cl_mM_dup salinity_dup
## 1   0.6816587 0.04369179  0.3609794  0.6861326  0.04397838
## 2 100.9706403 6.46801842 13.1115190 103.0014697  6.59810959
```

```
QAdups$S04_dups_chk <- ((abs(QAdups$S04_mM-QAdups$S04_mM_dup))/((QAdups$S04_mM+QAdups$S04_mM_dup)/2))*100
QAdups$S04_dups_flag <- ifelse(QAdups$S04_dups_chk <10, 'YES', 'NO, rerun')
```

```
QAdups$Cl_dups_chk <- ((abs(QAdups$Cl_mM-QAdups$Cl_mM_dup))/((QAdups$Cl_mM+QAdups$Cl_mM_dup)/2))*100
QAdups$Cl_dups_flag <- ifelse(QAdups$Cl_dups_chk <10, 'YES', 'NO, rerun')
```

```
head(QAdups)
```

```
##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 1 TEMPEST_FW_C3_20230608  11.6026  12.4935  24.1648  31.4737  0.3619027
## 2 TEMPEST_SW_I5_20230608 413.5339   9.7963 3579.4092 102.5647 12.8987492
```

```
##           Cl_mM    salinity S04_mM_dup    Cl_mM_dup salinity_dup S04_dups_chk
## 1    0.6816587 0.04369179  0.3609794    0.6861326    0.04397838    0.2554411
## 2 100.9706403 6.46801842 13.1115190 103.0014697    6.59810959    1.6360447
##   S04_dups_flag Cl_dups_chk Cl_dups_flag
## 1             YES    0.6541798         YES
## 2             YES    1.9912814         YES
```

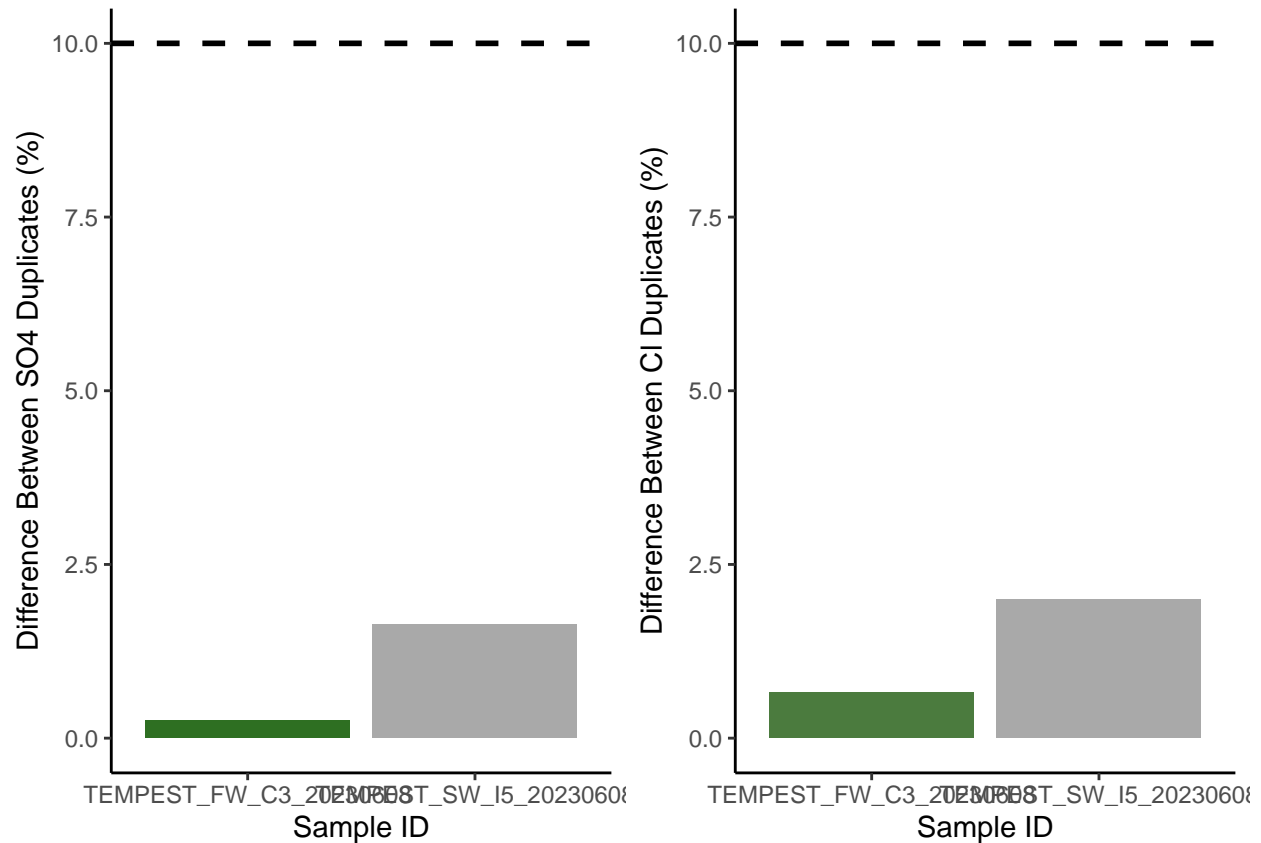
*#plot dups output as a bar graph to easily check - want any over 10% to be red need to work on this*

```
Sdupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = S04_dups_chk, fill=S04_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="Difference Between S04 Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
    color = "black", size=1)
```

```
## Warning: Using 'size' aesthetic for lines was deprecated in ggplot2 3.4.0.
## i Please use 'linewidth' instead.
## This warning is displayed once every 8 hours.
## Call 'lifecycle::last_lifecycle_warnings()' to see where this warning was
## generated.
```

```
Cldupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = Cl_dups_chk, fill=Cl_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="Difference Between Cl Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
    color = "black", size=1)

ggarrange(Sdupsbar, Cldupsbar, ncol=2, nrow=1)
```



```
#check for percent of no, reruns to see if it would warrant reruns
Perc_dups <- QAdups %>%
  group_by(SO4_dups_flag) %>%
  summarise(S_no_rows = length(SO4_dups_flag))
Perc_dups1 <- QAdups %>%
  group_by(Cl_dups_flag) %>%
  summarise(Cl_no_rows = length(Cl_dups_flag))
colnames(Perc_dups) <- c("Flag", "S_no_rows")
colnames(Perc_dups1) <- c("Flag", "Cl_no_rows")
Perc_dups2 <- cbind(Perc_dups, Perc_dups1)

Perc_dups2$Total <- length(QAdups$SO4_dups_flag)
Perc_dups2$S_Percent <- (Perc_dups2$S_no_rows / Perc_dups2$Total)*100
Perc_dups2$Cl_Percent <- (Perc_dups2$Cl_no_rows / Perc_dups2$Total)*100
head(Perc_dups2)
```

```
##   Flag S_no_rows Flag Cl_no_rows Total S_Percent Cl_Percent
## 1  YES         2  YES         2      2        100        100
```

Pull out dups and check with cv

```
#the cv
# calculate the sd bewteen the two duplicates then divide by the mean and multiply by 100
```

```
df2 <- as.data.frame(QAdups$S04_mM)
df2$dups <- QAdups$S04_mM_dup

df2$sds <- apply(df2,1,sd)

QAdups$S04_dups_cv <- (df2$sds)/((QAdups$S04_mM+QAdups$S04_mM_dup)/2) * 100
QAdups$S04_dups_cv_flag <- ifelse(QAdups$S04_dups_cv <11, 'YES', 'NO, rerun')

head(QAdups)
```

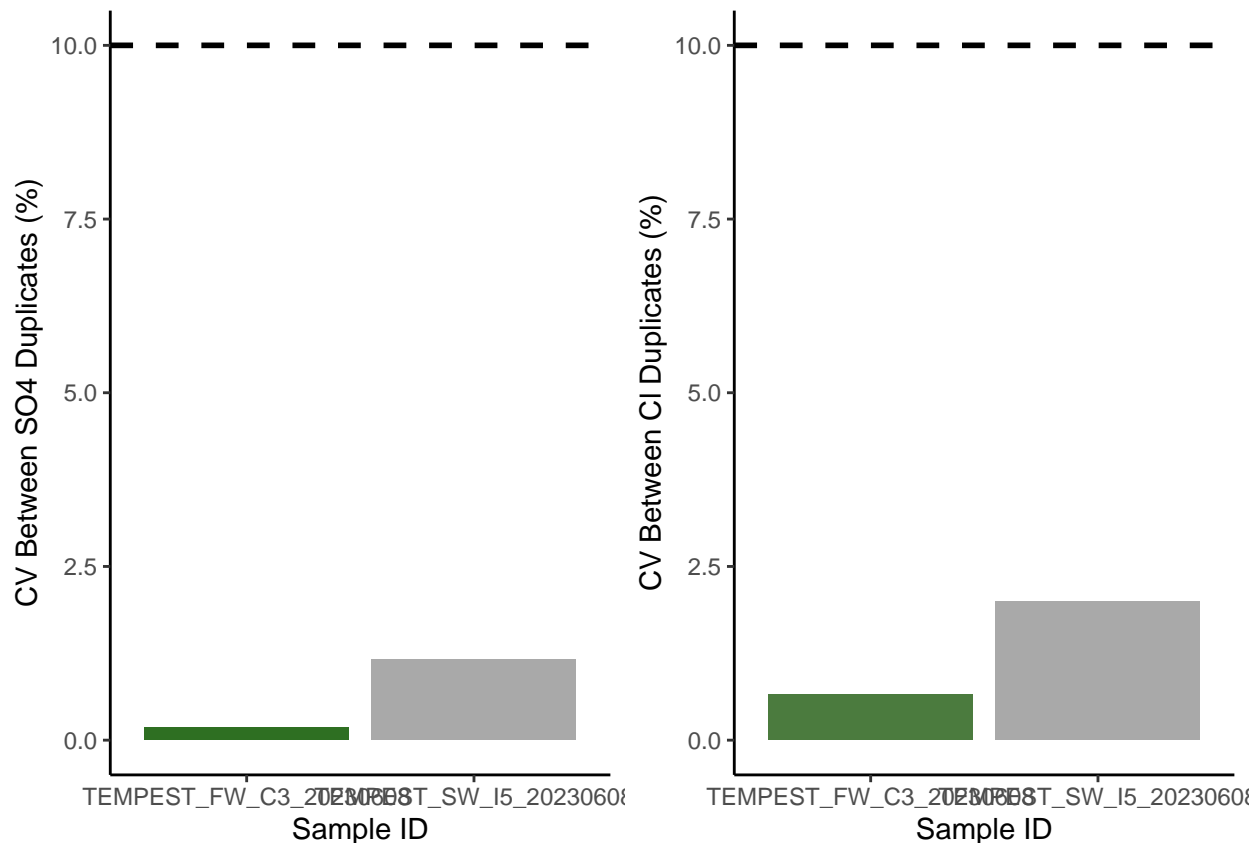
```
##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 1 TEMPEST_FW_C3_20230608  11.6026  12.4935   24.1648  31.4737  0.3619027
## 2 TEMPEST_SW_I5_20230608 413.5339   9.7963 3579.4092 102.5647 12.8987492
##           Cl_mM  salinity S04_mM_dup   Cl_mM_dup salinity_dup S04_dups_chk
## 1   0.6816587 0.04369179  0.3609794   0.6861326   0.04397838   0.2554411
## 2 100.9706403 6.46801842 13.1115190 103.0014697   6.59810959   1.6360447
##   S04_dups_flag Cl_dups_chk Cl_dups_flag S04_dups_cv S04_dups_cv_flag
## 1             YES  0.6541798             YES  0.1806241             YES
## 2             YES  1.9912814             YES  1.1568583             YES
```

*#plot dups output as a bar graph to easily check - want any over 10% to be red need to work on this*

```
Sdupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = S04_dups_cv, fill=S04_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="CV Between S04 Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
  color = "black", size=1)

Cldupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = Cl_dups_chk, fill=Cl_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="CV Between Cl Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
  color = "black", size=1)

ggarrange(Sdupsbar, Cldupsbar,ncol=2, nrow=1)
```



```
#check for percent of no, reruns to see if it would warrant reruns
Perc_dups <- QAdups %>%
  group_by(SO4_dups_flag) %>%
  summarise(S_no_rows = length(SO4_dups_flag))
Perc_dups1 <- QAdups %>%
  group_by(Cl_dups_flag) %>%
  summarise(Cl_no_rows = length(Cl_dups_flag))
colnames(Perc_dups) <- c("Flag", "S_no_rows")
colnames(Perc_dups1) <- c("Flag", "Cl_no_rows")
Perc_dups2 <- cbind(Perc_dups, Perc_dups1)

Perc_dups2$Total <- length(QAdups$SO4_dups_flag)
Perc_dups2$S_Percent <- (Perc_dups2$S_no_rows / Perc_dups2$Total)*100
Perc_dups2$Cl_Percent <- (Perc_dups2$Cl_no_rows / Perc_dups2$Total)*100
head(Perc_dups2)
```

```
##   Flag S_no_rows Flag Cl_no_rows Total S_Percent Cl_Percent
## 1  YES          2  YES          2      2        100        100
```

Pull out spikes and check with dionex output conc.

```
#Show me the data that we have from the calculations
head(sampledat)
```

```
##           Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area S04_mM
## 132 TEMEPST_FW_H6_20230608 10.0574 10.8297 0.0000 0.0000 0.31370555
## 133 TEMEPST_SW_C3_20230608 436.0629 10.3300 3597.9994 103.0974 13.60146288
## 134 TEMPEST_FW_B4_20230608 28.2839 28.7072 0.0000 0.0000 0.88221772
## 135 TEMPEST_FW_C3_20230608 11.6026 12.4935 24.1648 31.4737 0.36190268
## 136 TEMPEST_FW_C3_20230608_dup 11.5730 12.4616 24.3234 31.6803 0.36097941
## 137 TEMPEST_FW_D5_20230608 3.1147 3.3538 27.7577 36.1532 0.09715221
##           Cl_mM salinity
## 132 0.0000000 0.00002600
## 133 101.4950465 6.50161092
## 134 0.0000000 0.00002600
## 135 0.6816587 0.04369179
## 136 0.6861326 0.04397838
## 137 0.7830099 0.05018416
```

```
#pull out any rows that have "spk" in the SampleID column
spks <- sampledats %>%
  filter(str_detect(Sample_ID, "spk")) #have to change this to match data
head(spks)
```

```
##           Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area S04_mM
## 1 TEMPEST_SW_E3_20230608_spk 320.6901 7.5969 2356.323 67.5183 10.00281
##           Cl_mM salinity
## 1 66.46891 4.257901
```

```
#remove the dup from these IDs so we will have duplicate sample names
spks$Sample_ID<-gsub("_spk","",as.character(spks$Sample_ID))
spks <- spks[ , -c(2,3,4 ,5, 7,8)]
colnames(spks) <- c('Sample_ID', 'S04_mM_spk')
head(spks)
```

```
##           Sample_ID S04_mM_spk
## 1 TEMPEST_SW_E3_20230608 10.00281
```

```
#put it back together with the old data set and look for duplicates
QAspks <- merge(sampledat, spks)
head(QAspks)
```

```
##           Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area S04_mM Cl_mM
## 1 TEMPEST_SW_E3_20230608 243.9557 5.7791 2299.434 65.8882 7.609348 64.86415
## salinity S04_mM_spk
## 1 4.155103 10.00281
```

```
#now we need to calculate the spike concentration and calculate the spike recovery
spkconc <- (250/smw) # in mM
spkvol <- 10 # in uL
spkvol <- spkvol/1000000
#spike for these samples was 10uL of the 250mM standard
QAspks$S04_spk_Conc <- (spkconc)*spkvol # mmoles of S04
head(QAspks)
```

```
##           Sample_ID  S04_ppm S04_Area  Cl_ppm Cl_Area  S04_mM  Cl_mM
## 1 TEMPEST_SW_E3_20230608 243.9557  5.7791 2299.434 65.8882 7.609348 64.86415
##   salinity S04_mM_spk S04_spk_Conc
## 1 4.155103  10.00281 7.797879e-05
```

```
#need to determine dilution factors and initial amount of sample added
#if your samples are all the same dilution just use the first line
#for Steph / COMPASS this depends on the site so...
QAspks$Dilution <- 1
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 200, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$Dilution)

#Set Sample volumes in uL
QAspks$SampleVol <- 1
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 1462, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$SampleVol)

#change sample volume to L
QAspks$SampleVol <- QAspks$SampleVol/1000000
head(QAspks)
```

```
##           Sample_ID  S04_ppm S04_Area  Cl_ppm Cl_Area  S04_mM  Cl_mM
## 1 TEMPEST_SW_E3_20230608 243.9557  5.7791 2299.434 65.8882 7.609348 64.86415
##   salinity S04_mM_spk S04_spk_Conc Dilution SampleVol
## 1 4.155103  10.00281 7.797879e-05          1      1e-06
```

```
#gives us the total SO4 in the sample in mmoles
QAspks$SO4_Total_unspkd <- (QAspks$S04_mM/QAspks$Dilution)*(QAspks$SampleVol)

##total SO4 in spiked sample in mmoles
QAspks$SO4_Total_spkd <- (QAspks$S04_mM_spk/QAspks$Dilution)*(QAspks$SampleVol+spkvol)

QAspks$SO4_expctd_spkd <- (QAspks$SO4_Total_unspkd + QAspks$S04_spk_Conc)
```

```

QAspks$spk_recovery <- (QAspks$S04_Total_spkd/QAspks$S04_expctd_spkd)*100
QAspks$S04_spks_flag <- ifelse(QAspks$spk_recovery <=120 & QAspks$spk_recovery >=80 , 'YES', 'NO, rerun')

head(QAspks)

```

```

##           Sample_ID  S04_ppm S04_Area  Cl_ppm Cl_Area  S04_mM  Cl_mM
## 1 TEMPEST_SW_E3_20230608 243.9557   5.7791 2299.434 65.8882 7.609348 64.86415
##   salinity S04_mM_spk S04_spk_Conc Dilution SampleVol S04_Total_unspkd
## 1 4.155103  10.00281 7.797879e-05         1      1e-06    7.609348e-06
##   S04_Total_spkd S04_expctd_spkd spk_recovery S04_spks_flag
## 1  0.0001100309   8.558814e-05    128.5586      NO, rerun

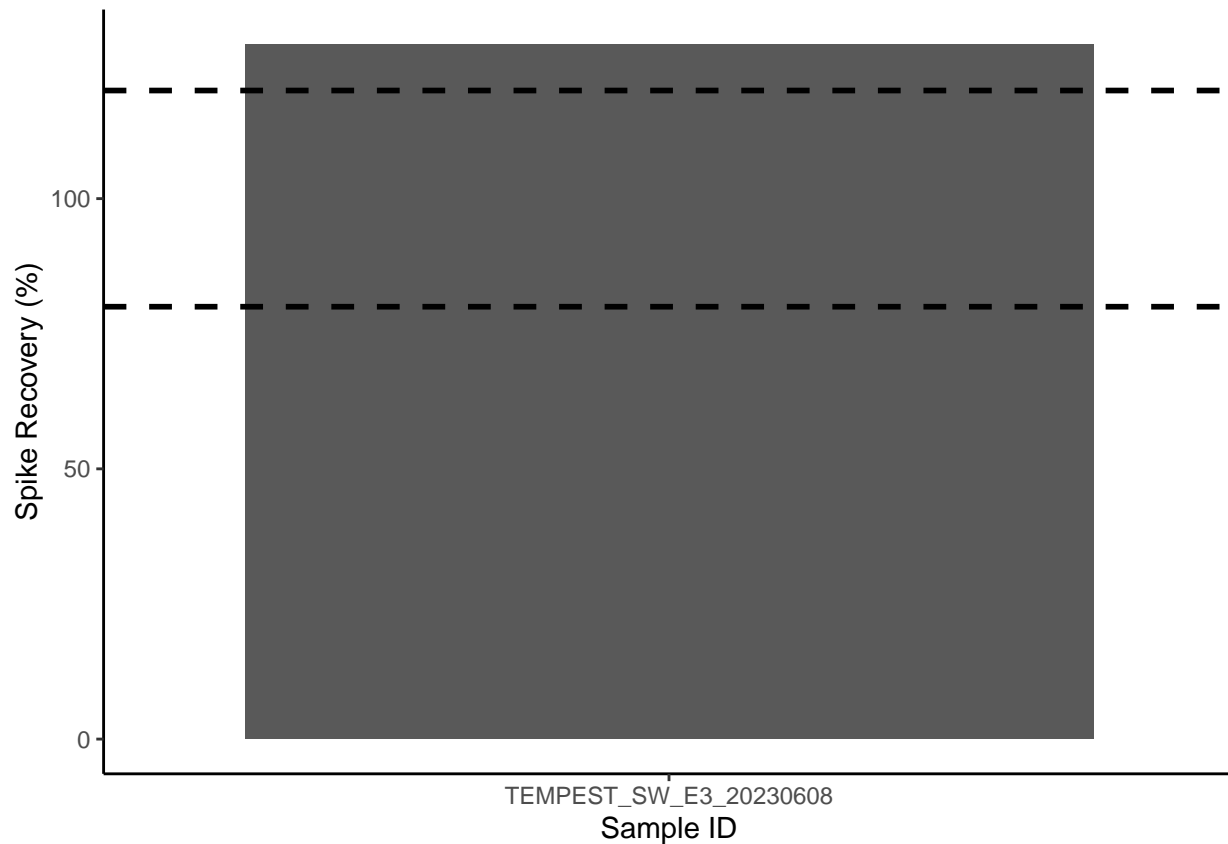
```

```

#plot spk recoveries output as a bar graph to easily check - want any over 10% to be red need to work on
spksbar <- ggplot(data = QAspks, aes(x = Sample_ID, y = spk_recovery)) +
  geom_bar(stat = 'identity') +
  scale_fill_manual(values=c("lightblue4")) +
  theme_classic() + labs(x= "Sample ID", y="Spike Recovery (%)") +
  theme(legend.position="none") +
  geom_hline(yintercept=80, linetype="dashed", color = "black", size=1) +
  geom_hline(yintercept=120, linetype="dashed", color = "black", size=1)

spksbar

```



```

#check for percent of no, reruns to see if it would warrant reruns
Perc_spks <- QAspks %>%
  group_by(S04_spks_flag) %>%
  summarise(no_rows = length(S04_spks_flag))
Perc_spks$Total <- length(QAspks$S04_spks_flag)
Perc_spks$Percent <- (Perc_spks$no_rows / Perc_spks$Total)*100
head(Perc_spks)

```

```

## # A tibble: 1 x 4
##   S04_spks_flag no_rows Total Percent
##   <chr>         <int> <int>   <dbl>
## 1 NO, rerun          1     1    100

```

## Pull out spikes and check with area calc

```

#Show me the data that we have from the calculations
head(sampledat)

```

```

##           Sample_ID S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 132  TEMEPST_FW_H6_20230608 10.0574 10.8297  0.0000  0.0000  0.31370555
## 133  TEMEPST_SW_C3_20230608 436.0629 10.3300 3597.9994 103.0974 13.60146288
## 134  TEMPEST_FW_B4_20230608 28.2839 28.7072  0.0000  0.0000  0.88221772
## 135  TEMPEST_FW_C3_20230608 11.6026 12.4935 24.1648 31.4737 0.36190268
## 136 TEMPEST_FW_C3_20230608_dup 11.5730 12.4616 24.3234 31.6803 0.36097941
## 137  TEMPEST_FW_D5_20230608  3.1147  3.3538 27.7577 36.1532 0.09715221
##           Cl_mM  salinity
## 132  0.0000000 0.00002600
## 133 101.4950465 6.50161092
## 134  0.0000000 0.00002600
## 135  0.6816587 0.04369179
## 136  0.6861326 0.04397838
## 137  0.7830099 0.05018416

```

```

#now use area of all samples and calculate undilution corrected concentrations in ug/mL
sampledat$S04_ugmL <- ((sampledat$S04_Area)-S04_Int)/S04_Slope
sampledat$Cl_ugmL <- (sampledat$Cl_Area-Cl_Int)/Cl_Slope
head(sampledat)

```

```

##           Sample_ID S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 132  TEMEPST_FW_H6_20230608 10.0574 10.8297  0.0000  0.0000  0.31370555
## 133  TEMEPST_SW_C3_20230608 436.0629 10.3300 3597.9994 103.0974 13.60146288
## 134  TEMPEST_FW_B4_20230608 28.2839 28.7072  0.0000  0.0000  0.88221772
## 135  TEMPEST_FW_C3_20230608 11.6026 12.4935 24.1648 31.4737 0.36190268
## 136 TEMPEST_FW_C3_20230608_dup 11.5730 12.4616 24.3234 31.6803 0.36097941
## 137  TEMPEST_FW_D5_20230608  3.1147  3.3538 27.7577 36.1532 0.09715221
##           Cl_mM  salinity S04_ugmL   Cl_ugmL
## 132  0.0000000 0.00002600  9.143118 -1.166337e-06
## 133 101.4950465 6.50161092  8.721239  7.195999e+01
## 134  0.0000000 0.00002600 24.236439 -1.166337e-06
## 135  0.6816587 0.04369179 10.547804  2.196803e+01

```

```
## 136 0.6861326 0.04397838 10.520872 2.211223e+01
## 137 0.7830099 0.05018416 2.831488 2.523423e+01
```

```
#pull out any rows that have "spk" in the SampleID column
spks <- sampledats %>%
  filter(str_detect(Sample_ID, "spk")) #have to change this to match data
head(spks)
```

```
##           Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area S04_mM
## 1 TEMPEST_SW_E3_20230608_spk 320.6901 7.5969 2356.323 67.5183 10.00281
##           Cl_mM salinity S04_ugmL Cl_ugmL
## 1 66.46891 4.257901 6.413783 47.12647
```

```
## Only keep the columns that we need
spks <- spks[,c(1,9,10)] # dont need this here
head(spks)
```

```
##           Sample_ID S04_ugmL Cl_ugmL
## 1 TEMPEST_SW_E3_20230608_spk 6.413783 47.12647
```

```
#remove the spk from these IDs so we will have duplicate sample names
spks$Sample_ID<-gsub("_spk","",as.character(spks$Sample_ID))
spks <- spks[,c(1,2)]
colnames(spks) <- c('Sample_ID', 'S04_ugmL_spk')
head(spks)
```

```
##           Sample_ID S04_ugmL_spk
## 1 TEMPEST_SW_E3_20230608 6.413783
```

```
#put it back together with the old data set and look for duplicates
QAspks <- merge(sampledat, spks)
head(QAspks)
```

```
##           Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area S04_mM Cl_mM
## 1 TEMPEST_SW_E3_20230608 243.9557 5.7791 2299.434 65.8882 7.609348 64.86415
##           salinity S04_ugmL Cl_ugmL S04_ugmL_spk
## 1 4.155103 4.879081 45.98869 6.413783
```

```
#now we need to calculate the spike concentration and calculate the spike recovery
spkconc <- (250) # in ug
spkvol <- 10 # in uL
spkvol <- spkvol/1000
#spike for these samples was 10uL of the 250mM standard
QAspks$S04_spk_Conc <- (spkconc)*spkvol # mmoles of S04
head(QAspks)
```

```
##           Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area S04_mM Cl_mM
## 1 TEMPEST_SW_E3_20230608 243.9557 5.7791 2299.434 65.8882 7.609348 64.86415
##           salinity S04_ugmL Cl_ugmL S04_ugmL_spk S04_spk_Conc
## 1 4.155103 4.879081 45.98869 6.413783 2.5
```

```

#need to determine dilution factors and initial amount of sample added
#if your samples are all the same dilution just use the first line
#for Steph / COMPASS this depends on the site so...
QAspks$Dilution <- 1
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 200, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$Dilution)

#Set Sample volumes in uL
QAspks$SampleVol <- 1
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_PPR_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_PPR_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_PPR_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 1462, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$SampleVol)

#change sample volume to L
QAspks$SampleVol <- QAspks$SampleVol/1000
head(QAspks)

```

```

##           Sample_ID  S04_ppm S04_Area  Cl_ppm Cl_Area  S04_mM  Cl_mM
## 1 TEMPEST_SW_E3_20230608 243.9557   5.7791 2299.434 65.8882 7.609348 64.86415
##  salinity S04_ugmL  Cl_ugmL S04_ugmL_spk S04_spk_Conc SampleVol
## 1 4.155103 4.879081 45.98869    6.413783        2.5    0.001

```

```

#gives us the total S04 in the sample in mmoles
QAspks$S04_Total_unspkd <- QAspks$S04_ugmL*QAspks$SampleVol

##total S04 in spiked sample in mmoles
QAspks$S04_Total_spkd <- (QAspks$S04_ugmL_spk)*(QAspks$SampleVol+spkvol)

QAspks$S04_expctd_spkd <- (QAspks$S04_Total_unspkd + QAspks$S04_spk_Conc)
QAspks$spk_recovery <- (QAspks$S04_Total_spkd/QAspks$S04_expctd_spkd)*100
QAspks$S04_spks_flag <- ifelse(QAspks$spk_recovery <=120 & QAspks$spk_recovery >=80 , 'YES', 'NO, rerun

```

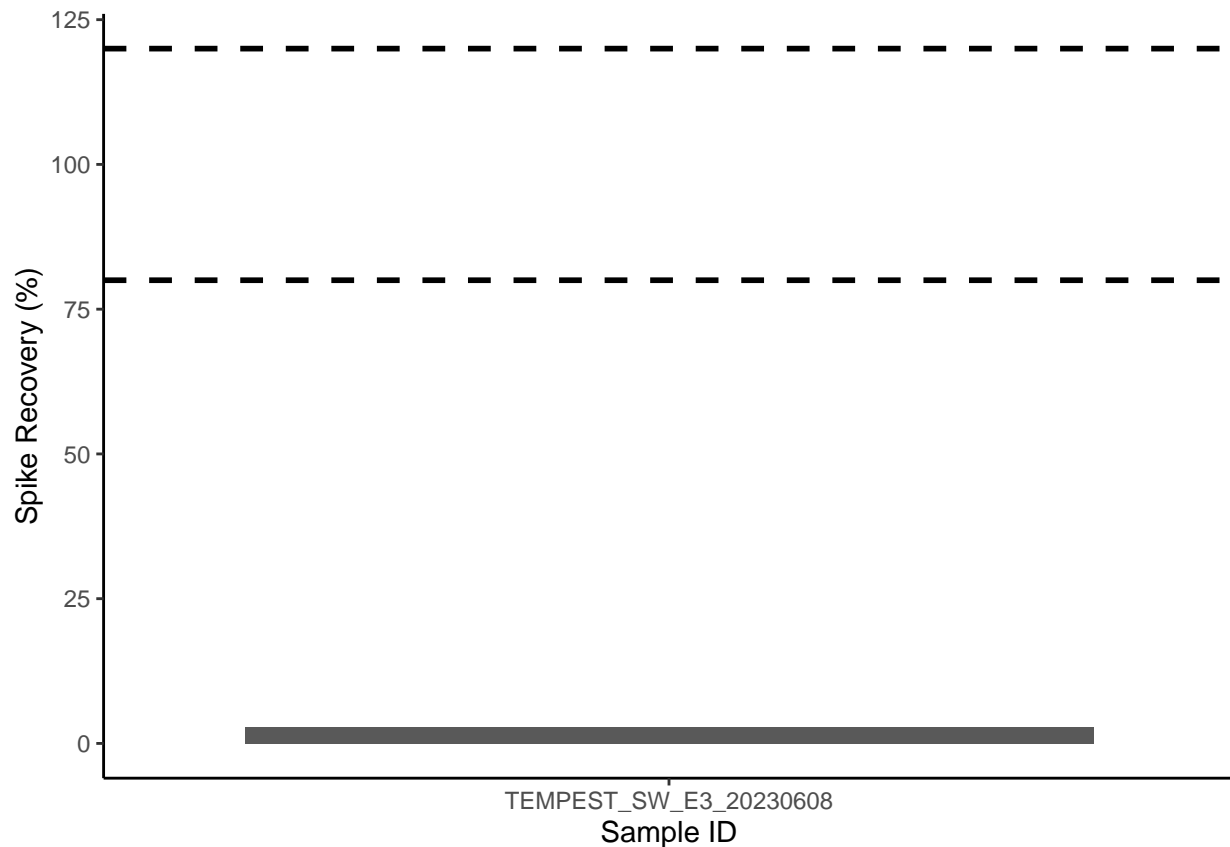
```
head(QAspks)
```

```
##           Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area S04_mM Cl_mM
## 1 TEMPEST_SW_E3_20230608 243.9557 5.7791 2299.434 65.8882 7.609348 64.86415
## salinity S04_ugmL Cl_ugmL S04_ugmL_spk S04_spk_Conc SampleVol
## 1 4.155103 4.879081 45.98869 6.413783 2.5 0.001
## S04_Total_unspkd S04_Total_spkd S04_expctd_spkd spk_recovery S04_spks_flag
## 1 0.004879081 0.07055161 2.504879 2.816568 NO, rerun
```

*#plot spk recoveries output as a bar graph to easily check - want any over 10% to be red need to work on*

```
spksbar <- ggplot(data = QAspks, aes(x = Sample_ID, y = spk_recovery)) +
  geom_bar(stat = 'identity') +
  scale_fill_manual(values=c("lightblue4")) +
  theme_classic() + labs(x= "Sample ID", y="Spike Recovery (%)") +
  theme(legend.position="none") +
  geom_hline(yintercept=80, linetype="dashed", color = "black", size=1) +
  geom_hline(yintercept=120, linetype="dashed", color = "black", size=1)
```

```
spksbar
```



*#check for percent of no, reruns to see if it would warrant reruns*

```
Perc_spks <- QAspks %>%
  group_by(S04_spks_flag) %>%
  summarise(no_rows = length(S04_spks_flag))
```

```
Perc_spks$Total <- length(QAspks$S04_spks_flag)
Perc_spks$Percent <- (Perc_spks$no_rows / Perc_spks$Total)*100
head(Perc_spks)
```

```
## # A tibble: 1 x 4
##   S04_spks_flag no_rows Total Percent
##   <chr>         <int> <int>   <dbl>
## 1 NO, rerun           1     1    100
```

## Make final dataframe with IDs

```
#for steph <- pull out identifiers of the sample names
#pull the sample ID and separate it by the underscores
IDs <- data.frame(do.call('rbind', strsplit(as.character(sampledat2$Sample_ID), '_ ', fixed=TRUE)))
colnames(IDs) <- c("Project" , "Plot", "Grid", "Date")
head(IDs)
```

```
##   Project Plot Grid    Date
## 1 TEMEPST  FW   H6 20230608
## 2 TEMEPST  SW   C3 20230608
## 3 TEMEPST  FW   B4 20230608
## 4 TEMEPST  FW   C3 20230608
## 5 TEMEPST  FW   D5 20230608
## 6 TEMEPST  FW   E3 20230608
```

```
#rejoin them to the dataframe
alldat <- cbind(IDs, sampledat2)
head(alldat)
```

```
##   Project Plot Grid    Date          Sample_ID  S04_ppm S04_Area  Cl_ppm
## 1 TEMEPST  FW   H6 20230608 TEMEPST_FW_H6_20230608  10.0574  10.8297   0.0000
## 2 TEMEPST  SW   C3 20230608 TEMEPST_SW_C3_20230608 436.0629  10.3300 3597.9994
## 3 TEMEPST  FW   B4 20230608 TEMEPST_FW_B4_20230608  28.2839  28.7072   0.0000
## 4 TEMEPST  FW   C3 20230608 TEMEPST_FW_C3_20230608  11.6026  12.4935  24.1648
## 5 TEMEPST  FW   D5 20230608 TEMEPST_FW_D5_20230608   3.1147   3.3538  27.7577
## 6 TEMEPST  FW   E3 20230608 TEMEPST_FW_E3_20230608  23.9154  24.2733   0.0000
##   Cl_Area      S04_mM      Cl_mM  salinity
## 1   0.0000  0.31370555  0.0000000 0.00002600
## 2 103.0974 13.60146288 101.4950465 6.50161092
## 3   0.0000  0.88221772  0.0000000 0.00002600
## 4  31.4737  0.36190268  0.6816587 0.04369179
## 5  36.1532  0.09715221  0.7830099 0.05018416
## 6   0.0000  0.74595758  0.0000000 0.00002600
```

## Make final dataframe with IDs

```
write.csv(alldat, file="Processed Data/COMPASS_TEMPEST_Dionex_ClSo4_Final_20230609.csv")
```

*#Cha*

END