

Dionex_COMPASS_October2022

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Daily Set up

Read in and Format the raw data - change wd & file names (Make sure files are UTF-8 encoded)

```
# SULFATE DATA:
```

```
## Read in raw data file from Dionex - copied and saved as a txt
```

```
Sdat <- read.table("Raw Data/COMPASS_Synoptic_CB_MonMon_202210_S04.txt", sep='\t' , header=T, skip=3)  
head(Sdat)
```

```
##      X          X.1          X.2 IC.S04 IC.S04.1 IC.S04.2 IC.S04.3 IC.S04.4  
## 1 1 Lab Blank      Unknown  n.a.      n.a.      n.a.      n.a.      n.a.  
## 2 2 Lab Blank      Unknown  n.a.      n.a.      n.a.      n.a.      n.a.  
## 3 3 Standard 1      Unknown  4.400    0.5687    6.39    0.4728    2.96  
## 4 4 Blank          Unknown  n.a.      n.a.      n.a.      n.a.      n.a.  
## 5 5 Standard 1 Calibration Standard 4.397    0.5680    6.40    0.4722    2.98  
## 6 6 Standard 2 Calibration Standard 4.400    1.1543    6.46    0.9595    5.88
```

```
## Only keep the columns that we need
```

```
Sdat <- Sdat[ ,c(2,5,7)] # dont need this here  
head(Sdat)
```

```
##      X.1 IC.S04.1 IC.S04.3  
## 1 Lab Blank      n.a.      n.a.  
## 2 Lab Blank      n.a.      n.a.  
## 3 Standard 1 0.5687    0.4728  
## 4 Blank      n.a.      n.a.  
## 5 Standard 1 0.5680    0.4722  
## 6 Standard 2 1.1543    0.9595
```

```
## Name the columns correctly
```

```
colnames(Sdat) <- c( "Sample_ID", "S04_ppm", "S04_Area")  
Sdat$Sample_ID <- as.factor(Sdat$Sample_ID)  
Sdat$S04_ppm <- as.numeric(Sdat$S04_ppm)
```

```
## Warning: NAs introduced by coercion
```

```
Sdat$S04_Area <- as.numeric(Sdat$S04_Area)
```

```
## Warning: NAs introduced by coercion
```

```
Sdat <- as.data.frame(Sdat)
head(Sdat)
```

```
##   Sample_ID S04_ppm S04_Area
## 1 Lab Blank      NA        NA
## 2 Lab Blank      NA        NA
## 3 Standard 1 0.5687 0.4728
## 4 Blank        NA        NA
## 5 Standard 1 0.5680 0.4722
## 6 Standard 2 1.1543 0.9595
```

```
#Chloride data
```

```
## Read in raw data file from Dionex - copied and saved as a txt
```

```
Cldat <- read.table("Raw Data/COMPASS_Synoptic_CB_MonMon_202210_Cl.txt", sep='\t' , header=T, skip=3)
head(Cldat)
```

```
##   X      X.1          X.2 IC.Cl IC.Cl.1 IC.Cl.2 IC.Cl.3 IC.Cl.4
## 1 1 Lab Blank      Unknown 3.273 0.0471 100.00 0.0531 0.37
## 2 2 Lab Blank      Unknown 3.280 0.0472 100.00 0.0531 0.38
## 3 3 Standard 1      Unknown 3.280 6.0990 92.87 6.8696 47.28
## 4 4 Blank          Unknown 3.273 0.0480 100.00 0.0541 0.39
## 5 5 Standard 1 Calibration Standard 3.280 6.0838 92.90 6.8525 47.38
## 6 6 Standard 2 Calibration Standard 3.280 12.2230 92.75 13.7674 94.28
```

```
## Only keep the columns that we need
```

```
Cldat <- Cldat[,c(2,5,7)]
head(Cldat)
```

```
##      X.1 IC.Cl.1 IC.Cl.3
## 1 Lab Blank 0.0471 0.0531
## 2 Lab Blank 0.0472 0.0531
## 3 Standard 1 6.0990 6.8696
## 4 Blank     0.0480 0.0541
## 5 Standard 1 6.0838 6.8525
## 6 Standard 2 12.2230 13.7674
```

```
## Name the columns correctly
```

```
colnames(Cldat) <- c("Sample_ID", "Cl_ppm", "Cl_Area")
Cldat$Sample_ID <- as.factor(Cldat$Sample_ID)
Cldat$Cl_ppm <- as.numeric(Cldat$Cl_ppm)
```

```
## Warning: NAs introduced by coercion
```

```
Cldat$Cl_Area <- as.numeric(Cldat$Cl_Area)
```

```
## Warning: NAs introduced by coercion
```

```
Cldat <- as.data.frame(Cldat)
head(Cldat)
```

```
##      Sample_ID Cl_ppm Cl_Area
## 1 Lab Blank  0.0471  0.0531
## 2 Lab Blank  0.0472  0.0531
## 3 Standard 1  6.0990  6.8696
## 4      Blank  0.0480  0.0541
## 5 Standard 1  6.0838  6.8525
## 6 Standard 2 12.2230 13.7674
```

```
## Bring the data back together:
all_dat <- merge(Sdat, Cldat, by="Sample_ID")
head(all_dat)
```

```
##      Sample_ID  SO4_ppm SO4_Area      Cl_ppm Cl_Area
## 1           2418.1928  17.1467 15120.9494 224.2274
## 2           2418.1928  17.1467  4224.4183  60.4754
## 3           2418.1928  17.1467    0.0212   0.0239
## 4           2418.1928  17.1467 4473.1255  50.9915
## 5           2418.1928  17.1467         NA        NA
## 6           586.9775   5.9183 15120.9494 224.2274
```

```
## Remove empty lines
all_dat <- all_dat[!(is.na(all_dat$Sample_ID) | all_dat$Sample_ID==""), ]
head(all_dat)
```

```
##              Sample_ID  SO4_ppm SO4_Area      Cl_ppm Cl_Area
## 26      1_202210_Gcrew_UP_LysA_20cm  4.1491  0.0690  14.4387  0.3253
## 27     10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338
## 28 10_202210_GCrew_TR_LysB_20cm_spk 165.6846  2.7546  38.8350  0.8748
## 29      100_202210_MSM_UP_RHZ_SF_1  31.3728  0.5216 671.7957 15.1336
## 30 100_202210_MSM_UP_RHZ_SF_1_spk 203.4807  3.3830 1340.3550 30.1943
## 31     101_202210_MSM_UP_RHZ_SF_2  65.5675  1.0901  933.5416 21.0300
```

```
all_dat <- droplevels(all_dat[!all_dat$Sample_ID == 'Lab Blank',])
head(all_dat)
```

```
##              Sample_ID  SO4_ppm SO4_Area      Cl_ppm Cl_Area
## 26      1_202210_Gcrew_UP_LysA_20cm  4.1491  0.0690  14.4387  0.3253
## 27     10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338
## 28 10_202210_GCrew_TR_LysB_20cm_spk 165.6846  2.7546  38.8350  0.8748
## 29      100_202210_MSM_UP_RHZ_SF_1  31.3728  0.5216 671.7957 15.1336
## 30 100_202210_MSM_UP_RHZ_SF_1_spk 203.4807  3.3830 1340.3550 30.1943
## 31     101_202210_MSM_UP_RHZ_SF_2  65.5675  1.0901  933.5416 21.0300
```

Pull out standards - could do some checks here if we want

```
stds <- all_dat[grep1("Standard", all_dat$Sample_ID),]
head(stds)
```

```
##      Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area
## 1424 Standard 1  0.5615  0.4668 6.1419  6.9180
## 1425 Standard 1  0.5615  0.4668 6.1360  6.9113
## 1426 Standard 1  0.5615  0.4668 6.0990  6.8696
## 1427 Standard 1  0.5615  0.4668 6.1169  6.8898
## 1428 Standard 1  0.5615  0.4668 6.0921  6.8619
## 1429 Standard 1  0.5615  0.4668 6.0838  6.8525
```

```
stds_chk_S <- stds %>%
  group_by(Sample_ID) %>%
  summarise(mean = mean(S04_ppm), sd = sd(S04_ppm))
stds_chk_S$cv <- (stds_chk_S$sd/stds_chk_S$mean)*100
stds_chk_S$flag <- ifelse(stds_chk_S$cv < 2, 'YES', 'NO, rerun')
head(stds_chk_S)
```

```
## # A tibble: 5 x 5
##   Sample_ID   mean      sd    cv flag
##   <fct>      <dbl>   <dbl> <dbl> <chr>
## 1 Standard 1  0.568 0.00410 0.722 YES
## 2 Standard 2  1.16 0.00688 0.595 YES
## 3 Standard 3  2.30 0.00792 0.345 YES
## 4 Standard 4 10.4  0.0278  0.268 YES
## 5 Standard 5 19.9  0.0475  0.239 YES
```

```
stds_chk_Cl <- stds %>%
  group_by(Sample_ID) %>%
  summarise(mean = mean(Cl_ppm), sd = sd(Cl_ppm))
stds_chk_Cl$cv <- (stds_chk_Cl$sd/stds_chk_Cl$mean)*100
stds_chk_Cl$flag <- ifelse(stds_chk_Cl$cv < 2, 'YES', 'NO, rerun')
head(stds_chk_Cl)
```

```
## # A tibble: 5 x 5
##   Sample_ID   mean      sd    cv flag
##   <fct>      <dbl>   <dbl> <dbl> <chr>
## 1 Standard 1   6.11 0.0210 0.344 YES
## 2 Standard 2  12.2 0.0218 0.178 YES
## 3 Standard 3  23.6 0.0444 0.189 YES
## 4 Standard 4 104.  0.233  0.224 YES
## 5 Standard 5 199.  0.313  0.157 YES
```

```
lmS <- lm(stds$S04_Area ~ stds$S04_ppm)
S04_sum <- summary(lmS)
S04_Slope <- S04_sum$coefficients[2, 1]
S04_Int <- S04_sum$coefficients[1, 1]
```

```
lmCl <- lm(stds$Cl_Area ~ stds$Cl_ppm)
Cl_sum <- summary(lmCl)
```

```
Cl_Slope <- Cl_sum$coefficients[2, 1]
Cl_Int <- Cl_sum$coefficients[1, 1]
```

Calculate mmol/L concentrations

```
#remove standards from sample dataframe
sampledat <- all_dat[!grepl("Standard", all_dat$Sample_ID),]
head(sampledat)
```

```
##
##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area
## 26      1_202210_Gcrew_UP_LysA_20cm    4.1491    0.0690   14.4387  0.3253
## 27     10_202210_GCrew_TR_LysB_20cm   28.5838    0.4752   41.4508  0.9338
## 28 10_202210_GCrew_TR_LysB_20cm_spk 165.6846    2.7546   38.8350  0.8748
## 29     100_202210_MSM_UP_RHZ_SF_1    31.3728    0.5216  671.7957 15.1336
## 30    100_202210_MSM_UP_RHZ_SF_1_spk 203.4807    3.3830 1340.3550 30.1943
## 31     101_202210_MSM_UP_RHZ_SF_2    65.5675    1.0901  933.5416 21.0300
```

```
# Constants needed for calculations:
clmw <- 35.45      #molecular weight of Chloride: 35.45
smw <- 32.06      #molecular weight of sulfur: 32.06

# Convert ppm to mmol/L
sampledat$S04_mM <- (sampledat$S04_ppm / smw)
sampledat$Cl_mM <- (sampledat$Cl_ppm / clmw)

# Calculate Salinity
# calculated using the Knudsen equation
# Salinity = 0.03 + 1.8050 * Chlorinity
# Ref: A Practical Handbook of Seawater Analysis by Strickland & Parsons (P. 11)
# =((1.807*Cl_ppm)+0.026)/1000
sampledat$salinity <- ((1.8070 * sampledat$Cl_ppm) + 0.026) / 1000

head(sampledat)
```

```
##
##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area
## 26      1_202210_Gcrew_UP_LysA_20cm    4.1491    0.0690   14.4387  0.3253
## 27     10_202210_GCrew_TR_LysB_20cm   28.5838    0.4752   41.4508  0.9338
## 28 10_202210_GCrew_TR_LysB_20cm_spk 165.6846    2.7546   38.8350  0.8748
## 29     100_202210_MSM_UP_RHZ_SF_1    31.3728    0.5216  671.7957 15.1336
## 30    100_202210_MSM_UP_RHZ_SF_1_spk 203.4807    3.3830 1340.3550 30.1943
## 31     101_202210_MSM_UP_RHZ_SF_2    65.5675    1.0901  933.5416 21.0300
##           S04_mM      Cl_mM  salinity
## 26 0.1294167  0.4072976 0.02611673
## 27 0.8915721  1.1692750 0.07492760
## 28 5.1679538  1.0954866 0.07020084
## 29 0.9785652 18.9505134 1.21396083
## 30 6.3468715 37.8097320 2.42204748
## 31 2.0451497 26.3340367 1.68693567
```

Pull out dups and check with percent difference

```
#Show me the data that we have from the calculations
head(sampledat)
```

```
##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area
## 26      1_202210_Gcrew_UP_LysA_20cm    4.1491    0.0690   14.4387  0.3253
## 27     10_202210_GCrew_TR_LysB_20cm   28.5838    0.4752   41.4508  0.9338
## 28 10_202210_GCrew_TR_LysB_20cm_spk  165.6846    2.7546   38.8350  0.8748
## 29      100_202210_MSM_UP_RHZ_SF_1    31.3728    0.5216  671.7957 15.1336
## 30   100_202210_MSM_UP_RHZ_SF_1_spk  203.4807    3.3830 1340.3550 30.1943
## 31      101_202210_MSM_UP_RHZ_SF_2    65.5675    1.0901  933.5416 21.0300
##      S04_mM      Cl_mM      salinity
## 26 0.1294167  0.4072976 0.02611673
## 27 0.8915721  1.1692750 0.07492760
## 28 5.1679538  1.0954866 0.07020084
## 29 0.9785652 18.9505134 1.21396083
## 30 6.3468715 37.8097320 2.42204748
## 31 2.0451497 26.3340367 1.68693567
```

```
#pull out any rows that have "dup" in the SampleID column
dups <- sampledat %>%
  filter(str_detect(Sample_ID, "dup")) #have to change this to match data
head(dups)
```

```
##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area
## 1      109_202210_MSM_PPR_UP_2_dup    57.3788    0.9540 1055.464 23.7765
## 2      119_202210_MSM_PPR_UP_12_dup   140.2629    2.3320 2355.659 53.0661
## 3      129_202210_MSM_PPR_TR_10_dup   466.9523    7.7634 3585.616 80.7734
## 4 19_202210_GCrew_WC_SipB_20cm_dup   436.3971    3.6273 5767.606 64.9571
## 5      29_202210_GWI_UP_LysA_45cm_dup   651.2403    5.4131 5494.783 61.8845
## 6      39_202210_GWI_TR_LysB_45cm_dup  1084.1925    9.0118 9592.738 108.0373
##      S04_mM      Cl_mM      salinity
## 1  1.789732  29.77331  1.907249
## 2  4.375012  66.45018  4.256702
## 3 14.564950 101.14572  6.479233
## 4 13.611887 162.69693 10.422090
## 5 20.313172 155.00093  9.929099
## 6 33.817608 270.59911 17.334104
```

```
#remove these from sample dataframe in a new dataframe
sampledat2 <- sampledat %>%
  filter(!str_detect(Sample_ID, "dup")) %>%
  filter(!str_detect(Sample_ID, "spk"))
head(sampledat2)
```

```
##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 1  1_202210_Gcrew_UP_LysA_20cm    4.1491    0.0690   14.4387  0.3253 0.1294167
## 2 10_202210_GCrew_TR_LysB_20cm   28.5838    0.4752   41.4508  0.9338 0.8915721
## 3   100_202210_MSM_UP_RHZ_SF_1    31.3728    0.5216  671.7957 15.1336 0.9785652
## 4   101_202210_MSM_UP_RHZ_SF_2    65.5675    1.0901  933.5416 21.0300 2.0451497
```

```
## 5 102_202210_MSM_UP_RHZ_SF_3 105.7117 1.7575 1230.2357 27.7136 3.2973082
## 6 103_202210_MSM_UP_RHZ_SF_4 54.7246 0.9098 1060.9960 23.9011 1.7069432
## Cl_mM salinity
## 1 0.4072976 0.02611673
## 2 1.1692750 0.07492760
## 3 18.9505134 1.21396083
## 4 26.3340367 1.68693567
## 5 34.7034048 2.22306191
## 6 29.9293653 1.91724577
```

```
#remove the dup from these IDs so we will have duplicate sample names
dups$Sample_ID<-gsub("_dup","",as.character(dups$Sample_ID))
dups <- dups[ , -c(2:5)]
colnames(dups) <- c('Sample_ID', 'SO4_mM_dup', 'Cl_mM_dup', 'salinity_dup')
head(dups)
```

```
## Sample_ID SO4_mM_dup Cl_mM_dup salinity_dup
## 1 109_202210_MSM_PPR_UP_2 1.789732 29.77331 1.907249
## 2 119_202210_MSM_PPR_UP_12 4.375012 66.45018 4.256702
## 3 129_202210_MSM_PPR_TR_10 14.564950 101.14572 6.479233
## 4 19_202210_GCrew_WC_SipB_20cm 13.611887 162.69693 10.422090
## 5 29_202210_GWI_UP_LysA_45cm 20.313172 155.00093 9.929099
## 6 39_202210_GWI_TR_LysB_45cm 33.817608 270.59911 17.334104
```

```
#put it back together with the old data set and look for duplicates
QAdups <- merge(sampled2, dups)
head(QAdups)
```

```
## Sample_ID SO4_ppm SO4_Area Cl_ppm Cl_Area SO4_mM
## 1 109_202210_MSM_PPR_UP_2 56.9315 0.9465 1042.804 23.4913 1.775780
## 2 119_202210_MSM_PPR_UP_12 141.9209 2.3595 2376.423 53.5338 4.426728
## 3 129_202210_MSM_PPR_TR_10 462.9205 7.6964 3552.531 80.0281 14.439192
## 4 19_202210_GCrew_WC_SipB_20cm 441.4171 3.6691 5832.465 65.6876 13.768468
## 5 29_202210_GWI_UP_LysA_45cm 655.8744 5.4516 5534.610 62.3330 20.457717
## 6 39_202210_GWI_TR_LysB_45cm 1109.8531 9.2251 9834.080 110.7554 34.618001
## Cl_mM salinity SO4_mM_dup Cl_mM_dup salinity_dup
## 1 29.41620 1.884373 1.789732 29.77331 1.907249
## 2 67.03591 4.294222 4.375012 66.45018 4.256702
## 3 100.21243 6.419449 14.564950 101.14572 6.479233
## 4 164.52650 10.539290 13.611887 162.69693 10.422090
## 5 156.12440 10.001066 20.313172 155.00093 9.929099
## 6 277.40705 17.770208 33.817608 270.59911 17.334104
```

```
QAdups$SO4_dups_chk <- ((abs(QAdups$SO4_mM-QAdups$SO4_mM_dup))/((QAdups$SO4_mM+QAdups$SO4_mM_dup)/2))*100
QAdups$SO4_dups_flag <- ifelse(QAdups$SO4_dups_chk <10, 'YES', 'NO, rerun')

QAdups$Cl_dups_chk <- ((abs(QAdups$Cl_mM-QAdups$Cl_mM_dup))/((QAdups$Cl_mM+QAdups$Cl_mM_dup)/2))*100
QAdups$Cl_dups_flag <- ifelse(QAdups$Cl_dups_chk <10, 'YES', 'NO, rerun')

head(QAdups)
```

```
## Sample_ID SO4_ppm SO4_Area Cl_ppm Cl_Area SO4_mM
```

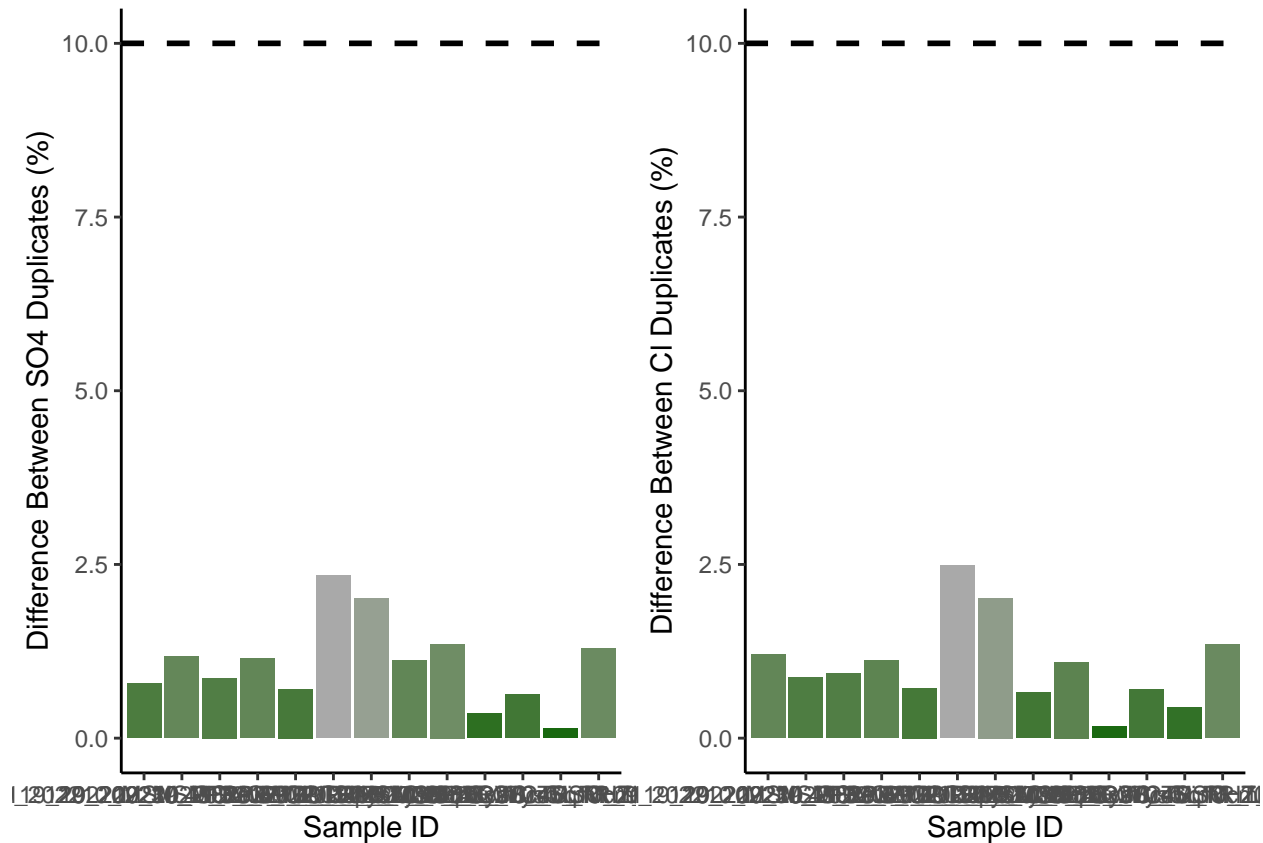
```
## 1      109_202210_MSM_PPR_UP_2    56.9315    0.9465 1042.804    23.4913    1.775780
## 2      119_202210_MSM_PPR_UP_12  141.9209    2.3595 2376.423    53.5338    4.426728
## 3      129_202210_MSM_PPR_TR_10  462.9205    7.6964 3552.531    80.0281   14.439192
## 4 19_202210_GCrew_WC_SipB_20cm  441.4171    3.6691 5832.465    65.6876   13.768468
## 5      29_202210_GWI_UP_LysA_45cm  655.8744    5.4516 5534.610    62.3330   20.457717
## 6      39_202210_GWI_TR_LysB_45cm 1109.8531    9.2251 9834.080   110.7554   34.618001
##      Cl_mM    salinity S04_mM_dup Cl_mM_dup    salinity_dup    S04_dups_chk
## 1 29.41620    1.884373    1.789732    29.77331        1.907249        0.7826066
## 2 67.03591    4.294222    4.375012    66.45018        4.256702        1.1751206
## 3 100.21243    6.419449   14.564950   101.14572        6.479233        0.8671724
## 4 164.52650   10.539290   13.611887   162.69693       10.422090        1.1437500
## 5 156.12440   10.001066   20.313172   155.00093        9.929099        0.7090579
## 6 277.40705   17.770208   33.817608   270.59911       17.334104        2.3391127
##      S04_dups_flag Cl_dups_chk Cl_dups_flag
## 1              YES    1.2066713              YES
## 2              YES    0.8775842              YES
## 3              YES    0.9269914              YES
## 4              YES    1.1182432              YES
## 5              YES    0.7222011              YES
## 6              YES    2.4846220              YES
```

```
#plot dups output as a bar graph to easily check - want any over 10% to be red need to work on this
Sdupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = S04_dups_chk, fill=S04_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="Difference Between S04 Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
    color = "black", size=1)
```

```
## Warning: Using 'size' aesthetic for lines was deprecated in ggplot2 3.4.0.
## i Please use 'linewidth' instead.
## This warning is displayed once every 8 hours.
## Call 'lifecycle::last_lifecycle_warnings()' to see where this warning was
## generated.
```

```
Cldupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = Cl_dups_chk, fill=Cl_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="Difference Between Cl Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
    color = "black", size=1)

ggarrange(Sdupsbar, Cldupsbar, ncol=2, nrow=1)
```

```
#check for percent of no, reruns to see if it would warrant reruns
Perc_dups <- QAdups %>%
  group_by(SO4_dups_flag) %>%
  summarise(S_no_rows = length(SO4_dups_flag))
Perc_dups1 <- QAdups %>%
  group_by(Cl_dups_flag) %>%
  summarise(Cl_no_rows = length(Cl_dups_flag))
colnames(Perc_dups) <- c("Flag", "S_no_rows")
colnames(Perc_dups1) <- c("Flag", "Cl_no_rows")
Perc_dups2 <- cbind(Perc_dups, Perc_dups1)

Perc_dups2$Total <- length(QAdups$SO4_dups_flag)
Perc_dups2$S_Percent <- (Perc_dups2$S_no_rows / Perc_dups2$Total)*100
Perc_dups2$Cl_Percent <- (Perc_dups2$Cl_no_rows / Perc_dups2$Total)*100
head(Perc_dups2)
```

```
##   Flag S_no_rows Flag Cl_no_rows Total S_Percent Cl_Percent
## 1  YES         13  YES         13    13         100         100
```

Pull out dups and check with cv

```
#the cv
# calculate the sd bewteen the two duplicates then divide by the mean and multiply by 100
```

```
df2 <- as.data.frame(QAdups$S04_mM)
df2$dups <- QAdups$S04_mM_dup

df2$sds <- apply(df2,1,sd)

QAdups$S04_dups_cv <- (df2$sds)/((QAdups$S04_mM+QAdups$S04_mM_dup)/2) * 100
QAdups$S04_dups_cv_flag <- ifelse(QAdups$S04_dups_cv <11, 'YES', 'NO, rerun')

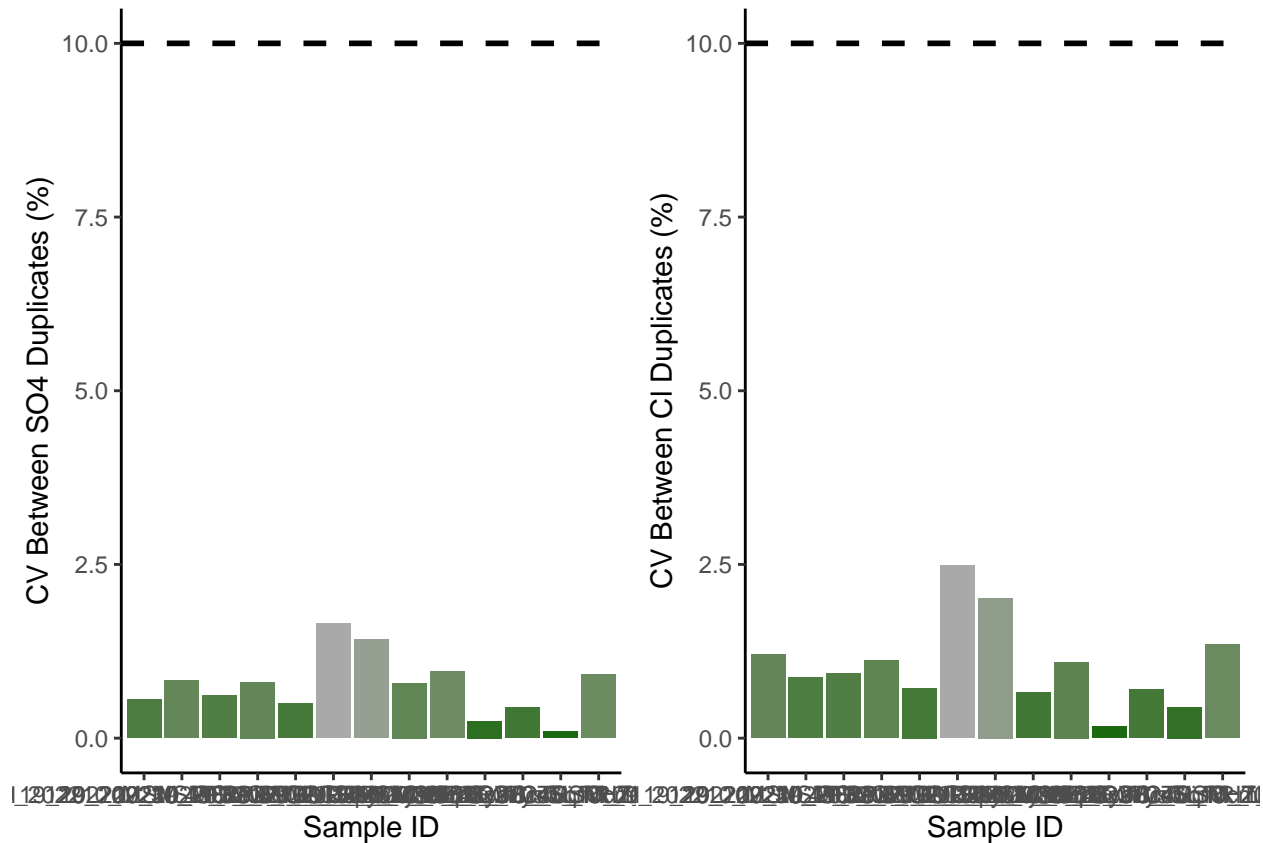
head(QAdups)
```

```
##           Sample_ID  S04_ppm S04_Area  Cl_ppm  Cl_Area  S04_mM
## 1      109_202210_MSM_PPR_UP_2  56.9315  0.9465 1042.804  23.4913  1.775780
## 2      119_202210_MSM_PPR_UP_12 141.9209  2.3595 2376.423  53.5338  4.426728
## 3      129_202210_MSM_PPR_TR_10  462.9205  7.6964 3552.531  80.0281 14.439192
## 4 19_202210_GCrew_WC_SipB_20cm  441.4171  3.6691 5832.465  65.6876 13.768468
## 5      29_202210_GWI_UP_LysA_45cm  655.8744  5.4516 5534.610  62.3330 20.457717
## 6      39_202210_GWI_TR_LysB_45cm 1109.8531  9.2251 9834.080 110.7554 34.618001
##           Cl_mM  salinity S04_mM_dup Cl_mM_dup  salinity_dup S04_dups_chk
## 1  29.41620  1.884373  1.789732  29.77331  1.907249  0.7826066
## 2  67.03591  4.294222  4.375012  66.45018  4.256702  1.1751206
## 3 100.21243  6.419449 14.564950 101.14572  6.479233  0.8671724
## 4 164.52650 10.539290 13.611887 162.69693 10.422090  1.1437500
## 5 156.12440 10.001066 20.313172 155.00093  9.929099  0.7090579
## 6 277.40705 17.770208 33.817608 270.59911 17.334104  2.3391127
##      S04_dups_flag Cl_dups_chk Cl_dups_flag S04_dups_cv S04_dups_cv_flag
## 1             YES  1.2066713             YES  0.5533865             YES
## 2             YES  0.8775842             YES  0.8309358             YES
## 3             YES  0.9269914             YES  0.6131835             YES
## 4             YES  1.1182432             YES  0.8087534             YES
## 5             YES  0.7222011             YES  0.5013796             YES
## 6             YES  2.4846220             YES  1.6540025             YES
```

```
#plot dups output as a bar graph to easily check - want any over 10% to be red need to work on this
Sdupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = S04_dups_cv, fill=S04_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="CV Between S04 Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
  color = "black", size=1)

Cldupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = Cl_dups_chk, fill=Cl_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="CV Between Cl Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
  color = "black", size=1)

ggarrange(Sdupsbar, Cldupsbar,ncol=2, nrow=1)
```



```
#check for percent of no, reruns to see if it would warrant reruns
Perc_dups <- QAdups %>%
  group_by(S04_dups_flag) %>%
  summarise(S_no_rows = length(S04_dups_flag))
Perc_dups1 <- QAdups %>%
  group_by(Cl_dups_flag) %>%
  summarise(Cl_no_rows = length(Cl_dups_flag))
colnames(Perc_dups) <- c("Flag", "S_no_rows")
colnames(Perc_dups1) <- c("Flag", "Cl_no_rows")
Perc_dups2 <- cbind(Perc_dups, Perc_dups1)

Perc_dups2$Total <- length(QAdups$S04_dups_flag)
Perc_dups2$S_Percent <- (Perc_dups2$S_no_rows / Perc_dups2$Total)*100
Perc_dups2$Cl_Percent <- (Perc_dups2$Cl_no_rows / Perc_dups2$Total)*100
head(Perc_dups2)
```

```
##   Flag S_no_rows Flag Cl_no_rows Total S_Percent Cl_Percent
## 1  YES         13  YES         13    13         100         100
```

Pull out spikes and check with dionex output conc.

```
#Show me the data that we have from the calculations
head(sampledat)
```

```
##           Sample_ID  S04_ppm S04_Area  Cl_ppm Cl_Area
## 26      1_202210_GCrew_UP_LysA_20cm    4.1491    0.0690   14.4387   0.3253
## 27     10_202210_GCrew_TR_LysB_20cm   28.5838    0.4752   41.4508   0.9338
## 28 10_202210_GCrew_TR_LysB_20cm_spk  165.6846    2.7546   38.8350   0.8748
## 29      100_202210_MSM_UP_RHZ_SF_1    31.3728    0.5216  671.7957  15.1336
## 30   100_202210_MSM_UP_RHZ_SF_1_spk  203.4807    3.3830 1340.3550  30.1943
## 31      101_202210_MSM_UP_RHZ_SF_2    65.5675    1.0901  933.5416  21.0300
##      S04_mM      Cl_mM      salinity
## 26 0.1294167  0.4072976  0.02611673
## 27 0.8915721  1.1692750  0.07492760
## 28 5.1679538  1.0954866  0.07020084
## 29 0.9785652  18.9505134  1.21396083
## 30 6.3468715  37.8097320  2.42204748
## 31 2.0451497  26.3340367  1.68693567
```

```
#pull out any rows that have "spk" in the SampleID column
spks <- sampledats %>%
  filter(str_detect(Sample_ID, "spk")) #have to change this to match data
head(spks)
```

```
##           Sample_ID  S04_ppm S04_Area  Cl_ppm Cl_Area  S04_mM
## 1 10_202210_GCrew_TR_LysB_20cm_spk  165.6846    2.7546   38.835  0.8748  5.167954
## 2   100_202210_MSM_UP_RHZ_SF_1_spk  203.4807    3.3830 1340.355  30.1943  6.346871
## 3    110_202210_MSM_PPR_UP_3_spk  206.8264    3.4386 1094.155  24.6481  6.451229
## 4    120_202210_MSM_PPR_TR_1_spk  374.0176    6.2183 2572.971  57.9615 11.666176
## 5    130_202210_MSM_PPR_TR_11_spk  577.9479    9.6088 3459.534  77.9332 18.027071
## 6 20_202210_GCrew_WC_SipB_45cm_spk  698.3096    5.8043 5526.222  62.2386 21.781335
##      Cl_mM      salinity
## 1  1.095487  0.07020084
## 2  37.809732  2.42204748
## 3  30.864725  1.97716318
## 4  72.580279  4.64938442
## 5  97.589109  6.25140376
## 6 155.887797  9.98590988
```

```
#remove the dup from these IDs so we will have duplicate sample names
spks$Sample_ID<-gsub("_spk","",as.character(spks$Sample_ID))
spks <- spks[ , -c(2:5,7,8)]
colnames(spks) <- c('Sample_ID', 'S04_mM_spk')
head(spks)
```

```
##           Sample_ID S04_mM_spk
## 1 10_202210_GCrew_TR_LysB_20cm    5.167954
## 2   100_202210_MSM_UP_RHZ_SF_1    6.346871
## 3    110_202210_MSM_PPR_UP_3    6.451229
## 4    120_202210_MSM_PPR_TR_1   11.666176
## 5    130_202210_MSM_PPR_TR_11   18.027071
## 6 20_202210_GCrew_WC_SipB_45cm   21.781335
```

```
#put it back together with the old data set and look for duplicates
QAspks <- merge(sampledats, spks)
head(QAspks)
```

```
##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 1 10_202210_GCrew_TR_LysB_20cm 28.5838 0.4752 41.4508 0.9338 0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1 31.3728 0.5216 671.7957 15.1336 0.9785652
## 3 110_202210_MSM_PPR_UP_3 70.1598 1.1665 1104.8402 24.8888 2.1883905
## 4 120_202210_MSM_PPR_TR_1 234.3596 3.8964 2538.4834 57.1846 7.3100312
## 5 130_202210_MSM_PPR_TR_11 439.7246 7.3107 3454.7595 77.8256 13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420 3.5063 5626.6706 63.3699 13.1578915
##           Cl_mM   salinity S04_mM_spk
## 1 1.169275 0.0749276 5.167954
## 2 18.950513 1.2139608 6.346871
## 3 31.166155 1.9964722 6.451229
## 4 71.607430 4.5870655 11.666176
## 5 97.454429 6.2427764 18.027071
## 6 158.721315 10.1674198 21.781335
```

```
#now we need to calculate the spike concentration and calculate the spike recovery
spkconc <- (250/smw)      # in mM
spkvol <- 10              # in uL
spkvol <- spkvol/1000000
#spike for these samples was 10uL of the 250mM standard
QAspks$S04_spk_Conc <- (spkconc)*spkvol      # mmoles of SO4
head(QAspks)
```

```
##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 1 10_202210_GCrew_TR_LysB_20cm 28.5838 0.4752 41.4508 0.9338 0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1 31.3728 0.5216 671.7957 15.1336 0.9785652
## 3 110_202210_MSM_PPR_UP_3 70.1598 1.1665 1104.8402 24.8888 2.1883905
## 4 120_202210_MSM_PPR_TR_1 234.3596 3.8964 2538.4834 57.1846 7.3100312
## 5 130_202210_MSM_PPR_TR_11 439.7246 7.3107 3454.7595 77.8256 13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420 3.5063 5626.6706 63.3699 13.1578915
##           Cl_mM   salinity S04_mM_spk S04_spk_Conc
## 1 1.169275 0.0749276 5.167954 7.797879e-05
## 2 18.950513 1.2139608 6.346871 7.797879e-05
## 3 31.166155 1.9964722 6.451229 7.797879e-05
## 4 71.607430 4.5870655 11.666176 7.797879e-05
## 5 97.454429 6.2427764 18.027071 7.797879e-05
## 6 158.721315 10.1674198 21.781335 7.797879e-05
```

```
#need to determine dilution factors and initial amount of sample added
#if your samples are all the same dilution just use the first line
#for Steph / COMPASS this depends on the site so...
QAspks$Dilution <- 1
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 200, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$Dilution)
```

```

#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$Dilution)

#Set Sample volumes in uL
QAspks$SampleVol <- 1
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 1462, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$SampleVol)

#change sample volume to L
QAspks$SampleVol <- QAspks$SampleVol/1000000
head(QAspks)

```

```

##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 1 10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338  0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1 31.3728  0.5216  671.7957 15.1336  0.9785652
## 3 110_202210_MSM_PPR_UP_3 70.1598  1.1665 1104.8402 24.8888  2.1883905
## 4 120_202210_MSM_PPR_TR_1 234.3596  3.8964 2538.4834 57.1846  7.3100312
## 5 130_202210_MSM_PPR_TR_11 439.7246  7.3107 3454.7595 77.8256 13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420  3.5063 5626.6706 63.3699 13.1578915
##           Cl_mM  salinity S04_mM_spk S04_spk_Conc Dilution SampleVol
## 1 1.169275 0.0749276 5.167954 7.797879e-05 50 0.001501
## 2 18.950513 1.2139608 6.346871 7.797879e-05 50 0.001501
## 3 31.166155 1.9964722 6.451229 7.797879e-05 1 0.000001
## 4 71.607430 4.5870655 11.666176 7.797879e-05 1 0.000001
## 5 97.454429 6.2427764 18.027071 7.797879e-05 1 0.000001
## 6 158.721315 10.1674198 21.781335 7.797879e-05 100 0.001475

```

```

#gives us the total SO4 in the sample in mmoles
QAspks$S04_Total_unspkd <- (QAspks$S04_mM/QAspks$Dilution)*(QAspks$SampleVol)

##total SO4 in spiked sample in mmoles
QAspks$S04_Total_spkd <- (QAspks$S04_mM_spk/QAspks$Dilution)*(QAspks$SampleVol+spkvol)

QAspks$S04_expctd_spkd <- (QAspks$S04_Total_unspkd + QAspks$S04_spk_Conc)
QAspks$spk_recovery <- (QAspks$S04_Total_spkd/QAspks$S04_expctd_spkd)*100
QAspks$S04_spks_flag <- ifelse(QAspks$spk_recovery <=120 & QAspks$spk_recovery >=80 , 'YES', 'NO, rerun')

head(QAspks)

```

```

##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 1 10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338  0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1 31.3728  0.5216  671.7957 15.1336  0.9785652
## 3 110_202210_MSM_PPR_UP_3 70.1598  1.1665 1104.8402 24.8888  2.1883905

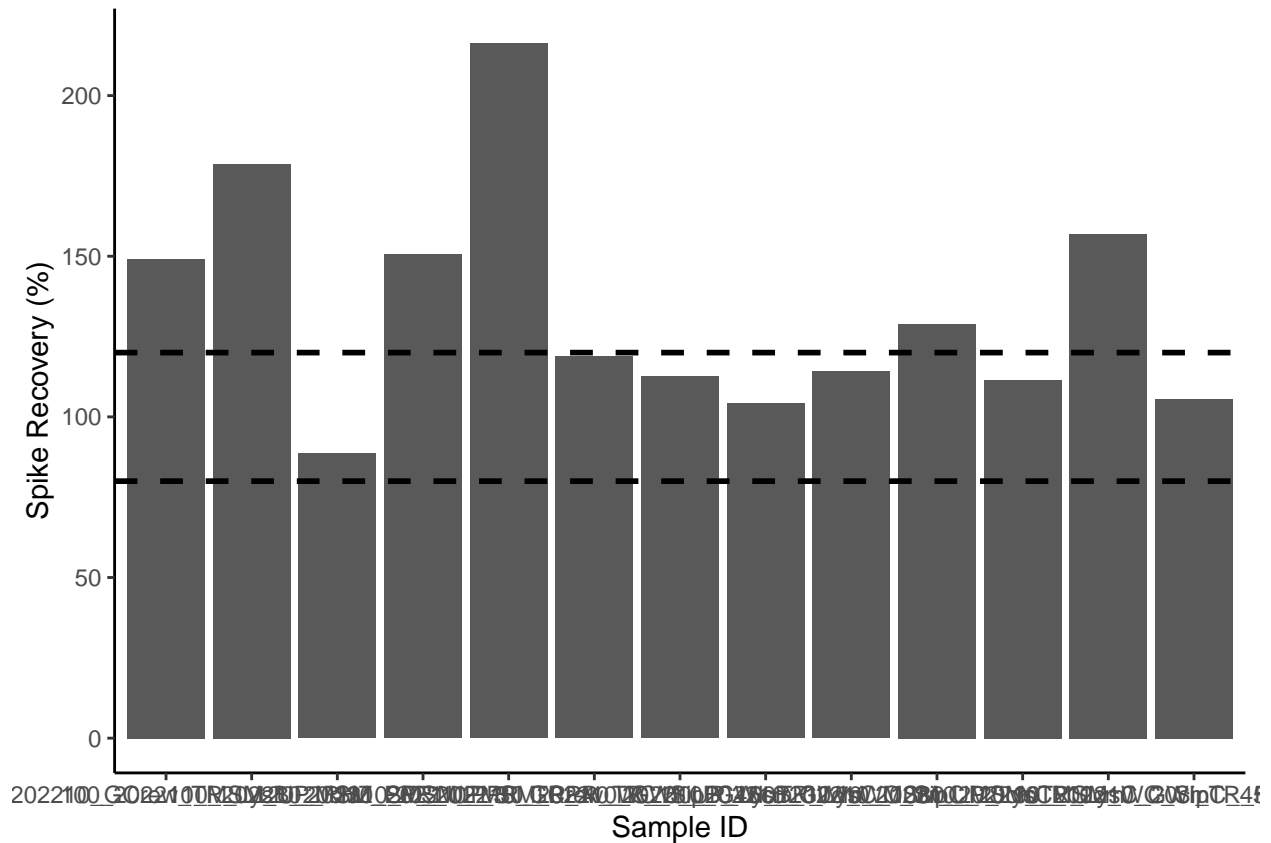
```

```
## 4      120_202210_MSM_PPR_TR_1 234.3596    3.8964 2538.4834 57.1846    7.3100312
## 5      130_202210_MSM_PPR_TR_11 439.7246    7.3107 3454.7595 77.8256   13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420    3.5063 5626.6706 63.3699   13.1578915
##      Cl_mM    salinity S04_mM_spk S04_spk_Conc Dilution SampleVol
## 1    1.169275  0.0749276   5.167954 7.797879e-05      50  0.001501
## 2   18.950513  1.2139608   6.346871 7.797879e-05      50  0.001501
## 3   31.166155  1.9964722   6.451229 7.797879e-05      1  0.000001
## 4   71.607430  4.5870655  11.666176 7.797879e-05      1  0.000001
## 5   97.454429  6.2427764  18.027071 7.797879e-05      1  0.000001
## 6  158.721315 10.1674198  21.781335 7.797879e-05     100  0.001475
##      S04_Total_unspkd S04_Total_spkd S04_expctd_spkd spk_recovery S04_spks_flag
## 1    2.676499e-05    1.561756e-04    1.047438e-04    149.10247      NO, rerun
## 2    2.937653e-05    1.918025e-04    1.073553e-04    178.66135      NO, rerun
## 3    2.188391e-06    7.096352e-05    8.016718e-05     88.51941        YES
## 4    7.310031e-06    1.283279e-04    8.528882e-05    150.46278      NO, rerun
## 5    1.371568e-05    1.982978e-04    9.169447e-05    216.25927      NO, rerun
## 6    1.940789e-04    3.234528e-04    2.720577e-04    118.89126        YES
```

#plot spk recoveries output as a bar graph to easily check - want any over 10% to be red need to work o

```
spksbar <- ggplot(data = QAspks, aes(x = Sample_ID, y = spk_recovery)) +
  geom_bar(stat = 'identity') +
  scale_fill_manual(values=c("lightblue4")) +
  theme_classic() + labs(x= "Sample ID", y="Spike Recovery (%)") +
  theme(legend.position="none") +
  geom_hline(yintercept=80, linetype="dashed", color = "black", size=1) +
  geom_hline(yintercept=120, linetype="dashed", color = "black", size=1)
```

spksbar



```
#check for percent of no, reruns to see if it would warrant reruns
Perc_spks <- QAspks %>%
  group_by(S04_spks_flag) %>%
  summarise(no_rows = length(S04_spks_flag))
Perc_spks$Total <- length(QAspks$S04_spks_flag)
Perc_spks$Percent <- (Perc_spks$no_rows / Perc_spks$Total)*100
head(Perc_spks)
```

```
## # A tibble: 2 x 4
##   S04_spks_flag no_rows Total Percent
##   <chr>         <int> <int>   <dbl>
## 1 NO, rerun      6    13    46.2
## 2 YES           7    13    53.8
```

Pull out spikes and check with area calc

```
#Show me the data that we have from the calculations
head(sampledat)
```

```
##           Sample_ID S04_ppm S04_Area   Cl_ppm Cl_Area
## 26  1_202210_Gcrew_UP_LysA_20cm  4.1491  0.0690  14.4387  0.3253
## 27  10_202210_Gcrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338
## 28 10_202210_Gcrew_TR_LysB_20cm_spk 165.6846  2.7546  38.8350  0.8748
```



```
## 29      100_202210_MSM_UP_RHZ_SF_1 31.3728 0.5216 671.7957 15.1336
## 30    100_202210_MSM_UP_RHZ_SF_1_spk 203.4807 3.3830 1340.3550 30.1943
## 31      101_202210_MSM_UP_RHZ_SF_2 65.5675 1.0901 933.5416 21.0300
##      S04_mM      Cl_mM      salinity
## 26 0.1294167 0.4072976 0.02611673
## 27 0.8915721 1.1692750 0.07492760
## 28 5.1679538 1.0954866 0.07020084
## 29 0.9785652 18.9505134 1.21396083
## 30 6.3468715 37.8097320 2.42204748
## 31 2.0451497 26.3340367 1.68693567
```

```
#now use area of all samples and calculate undilution corrected concentrations in ug/mL
sampledat$S04_ugmL <- ((sampledat$S04_Area)-S04_Int)/S04_Slope
sampledat$Cl_ugmL <- (sampledat$Cl_Area-Cl_Int)/Cl_Slope
head(sampledat)
```

```
##      Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area
## 26    1_202210_Gcrew_UP_LysA_20cm 4.1491 0.0690 14.4387 0.3253
## 27    10_202210_GCrew_TR_LysB_20cm 28.5838 0.4752 41.4508 0.9338
## 28 10_202210_GCrew_TR_LysB_20cm_spk 165.6846 2.7546 38.8350 0.8748
## 29      100_202210_MSM_UP_RHZ_SF_1 31.3728 0.5216 671.7957 15.1336
## 30    100_202210_MSM_UP_RHZ_SF_1_spk 203.4807 3.3830 1340.3550 30.1943
## 31      101_202210_MSM_UP_RHZ_SF_2 65.5675 1.0901 933.5416 21.0300
##      S04_mM      Cl_mM      salinity S04_ugmL Cl_ugmL
## 26 0.1294167 0.4072976 0.02611673 0.08300035 0.2888063
## 27 0.8915721 1.1692750 0.07492760 0.57164426 0.8290452
## 28 5.1679538 1.0954866 0.07020084 3.31368003 0.7766638
## 29 0.9785652 18.9505134 1.21396083 0.62746178 13.4359223
## 30 6.3468715 37.8097320 2.42204748 4.06962251 26.8071243
## 31 2.0451497 26.3340367 1.68693567 1.31134672 18.6708686
```

```
#pull out any rows that have "spk" in the SampleID column
spks <- sampledat %>%
  filter(str_detect(Sample_ID, "spk")) #have to change this to match data
head(spks)
```

```
##      Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area S04_mM
## 1 10_202210_GCrew_TR_LysB_20cm_spk 165.6846 2.7546 38.835 0.8748 5.167954
## 2 100_202210_MSM_UP_RHZ_SF_1_spk 203.4807 3.3830 1340.355 30.1943 6.346871
## 3 110_202210_MSM_PPR_UP_3_spk 206.8264 3.4386 1094.155 24.6481 6.451229
## 4 120_202210_MSM_PPR_TR_1_spk 374.0176 6.2183 2572.971 57.9615 11.666176
## 5 130_202210_MSM_PPR_TR_11_spk 577.9479 9.6088 3459.534 77.9332 18.027071
## 6 20_202210_GCrew_WC_SipB_45cm_spk 698.3096 5.8043 5526.222 62.2386 21.781335
##      Cl_mM      salinity S04_ugmL Cl_ugmL
## 1 1.095487 0.07020084 3.313680 0.7766638
## 2 37.809732 2.42204748 4.069623 26.8071243
## 3 30.864725 1.97716318 4.136507 21.8830928
## 4 72.580279 4.64938442 7.480386 51.4594206
## 5 97.589109 6.25140376 11.559035 69.1907103
## 6 155.887797 9.98590988 6.982359 55.2567187
```

```
## Only keep the columns that we need
spks <- spks[,c(1,9,10)] # dont need this here
head(spks)
```

```
##           Sample_ID  S04_ugmL  Cl_ugmL
## 1 10_202210_GCrew_TR_LysB_20cm_spk  3.313680  0.7766638
## 2 100_202210_MSM_UP_RHZ_SF_1_spk  4.069623 26.8071243
## 3 110_202210_MSM_PPR_UP_3_spk  4.136507 21.8830928
## 4 120_202210_MSM_PPR_TR_1_spk  7.480386 51.4594206
## 5 130_202210_MSM_PPR_TR_11_spk 11.559035 69.1907103
## 6 20_202210_GCrew_WC_SipB_45cm_spk  6.982359 55.2567187
```

```
#remove the spk from these IDs so we will have duplicate sample names
spks$Sample_ID<-gsub("_spk","",as.character(spks$Sample_ID))
spks <- spks[,c(1,2)]
colnames(spks) <- c('Sample_ID', 'S04_ugmL_spk')
head(spks)
```

```
##           Sample_ID S04_ugmL_spk
## 1 10_202210_GCrew_TR_LysB_20cm      3.313680
## 2 100_202210_MSM_UP_RHZ_SF_1      4.069623
## 3 110_202210_MSM_PPR_UP_3      4.136507
## 4 120_202210_MSM_PPR_TR_1      7.480386
## 5 130_202210_MSM_PPR_TR_11     11.559035
## 6 20_202210_GCrew_WC_SipB_45cm      6.982359
```

```
#put it back together with the old data set and look for duplicates
QAspks <- merge(sampledats, spks)
head(QAspks)
```

```
##           Sample_ID  S04_ppm S04_Area  Cl_ppm Cl_Area  S04_mM
## 1 10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338  0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1 31.3728  0.5216  671.7957 15.1336  0.9785652
## 3 110_202210_MSM_PPR_UP_3 70.1598  1.1665 1104.8402 24.8888  2.1883905
## 4 120_202210_MSM_PPR_TR_1 234.3596  3.8964 2538.4834 57.1846  7.3100312
## 5 130_202210_MSM_PPR_TR_11 439.7246  7.3107 3454.7595 77.8256 13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420  3.5063 5626.6706 63.3699 13.1578915
##           Cl_mM  salinity S04_ugmL  Cl_ugmL S04_ugmL_spk
## 1 1.169275  0.0749276 0.5716443  0.8290452      3.313680
## 2 18.950513  1.2139608 0.6274618 13.4359223      4.069623
## 3 31.166155  1.9964722 1.4032532 22.0967913      4.136507
## 4 71.607430  4.5870655 4.6872241 50.7696727      7.480386
## 5 97.454429  6.2427764 8.7945036 69.0951807     11.559035
## 6 158.721315 10.1674198 4.2179479 56.2611103      6.982359
```

```
#now we need to calculate the spike concentration and calculate the spike recovery
spkconc <- (250) # in ug
spkvol <- 10 # in uL
spkvol <- spkvol/1000
#spike for these samples was 10uL of the 250mM standard
QAspks$S04_spk_Conc <- (spkconc)*spkvol # mmoles of S04
head(QAspks)
```

	Sample_ID	S04_ppm	S04_Area	Cl_ppm	Cl_Area	S04_mM
## 1	10_202210_GCrew_TR_LysB_20cm	28.5838	0.4752	41.4508	0.9338	0.8915721
## 2	100_202210_MSM_UP_RHZ_SF_1	31.3728	0.5216	671.7957	15.1336	0.9785652
## 3	110_202210_MSM_PPR_UP_3	70.1598	1.1665	1104.8402	24.8888	2.1883905
## 4	120_202210_MSM_PPR_TR_1	234.3596	3.8964	2538.4834	57.1846	7.3100312
## 5	130_202210_MSM_PPR_TR_11	439.7246	7.3107	3454.7595	77.8256	13.7156769
## 6	20_202210_GCrew_WC_SipB_45cm	421.8420	3.5063	5626.6706	63.3699	13.1578915

	Cl_mM	salinity	S04_ugmL	Cl_ugmL	S04_ugmL_spk	S04_spk_Conc
## 1	1.169275	0.0749276	0.5716443	0.8290452	3.313680	2.5
## 2	18.950513	1.2139608	0.6274618	13.4359223	4.069623	2.5
## 3	31.166155	1.9964722	1.4032532	22.0967913	4.136507	2.5
## 4	71.607430	4.5870655	4.6872241	50.7696727	7.480386	2.5
## 5	97.454429	6.2427764	8.7945036	69.0951807	11.559035	2.5
## 6	158.721315	10.1674198	4.2179479	56.2611103	6.982359	2.5

#need to determine dilution factors and initial amount of sample added

#if your samples are all the same dilution just use the first line

#for Steph / COMPASS this depends on the site so...

#QAspks\$Dilution <- 1

#QAspks\$Dilution <- ifelse(str_detect(QAspks\$Sample_ID, "MSM_UP"), 50, QAspks\$Dilution)

##Dilution <- ifelse(str_detect(QAspks\$Sample_ID, "MSM_TR"), 50, QAspks\$Dilution)

#QAspks\$Dilution <- ifelse(str_detect(QAspks\$Sample_ID, "MSM_WC"), 100, QAspks\$Dilution)

##Dilution <- ifelse(str_detect(QAspks\$Sample_ID, "GCrew_UP"), 50, QAspks\$Dilution)

#QAspks\$Dilution <- ifelse(str_detect(QAspks\$Sample_ID, "GCrew_TR"), 50, QAspks\$Dilution)

#QAspks\$Dilution <- ifelse(str_detect(QAspks\$Sample_ID, "GCrew_WC"), 100, QAspks\$Dilution)

#QAspks\$Dilution <- ifelse(str_detect(QAspks\$Sample_ID, "GWI_UP"), 100, QAspks\$Dilution)

##Dilution <- ifelse(str_detect(QAspks\$Sample_ID, "GWI_TR"), 100, QAspks\$Dilution)

#QAspks\$Dilution <- ifelse(str_detect(QAspks\$Sample_ID, "GWI_WC"), 200, QAspks\$Dilution)

#QAspks\$Dilution <- ifelse(str_detect(QAspks\$Sample_ID, "SWH_UP"), 100, QAspks\$Dilution)

#QAspks\$Dilution <- ifelse(str_detect(QAspks\$Sample_ID, "SWH_TR"), 100, QAspks\$Dilution)

#QAspks\$Dilution <- ifelse(str_detect(QAspks\$Sample_ID, "SWH_WC"), 100, QAspks\$Dilution)

#Set Sample volumes in uL

QAspks\$SampleVol <- 1

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "MSM_UP"), 1501, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "MSM_TR"), 1501, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "MSM_WC"), 1475, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "MSM_PPR_UP"), 1501, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "MSM_PPR_TR"), 1501, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "MSM_PPR_WC"), 1475, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "GCrew_UP"), 1501, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "GCrew_TR"), 1501, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "GCrew_WC"), 1475, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "GWI_UP"), 1475, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "GWI_TR"), 1475, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "GWI_WC"), 1462, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "SWH_UP"), 100, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "SWH_TR"), 100, QAspks\$SampleVol)

QAspks\$SampleVol <- ifelse(str_detect(QAspks\$Sample_ID, "SWH_WC"), 100, QAspks\$SampleVol)

#change sample volume to L

QAspks\$SampleVol <- QAspks\$SampleVol/1000

head(QAspks)

```
##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 1 10_202210_GCrew_TR_LysB_20cm 28.5838 0.4752 41.4508 0.9338 0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1 31.3728 0.5216 671.7957 15.1336 0.9785652
## 3 110_202210_MSM_PPR_UP_3 70.1598 1.1665 1104.8402 24.8888 2.1883905
## 4 120_202210_MSM_PPR_TR_1 234.3596 3.8964 2538.4834 57.1846 7.3100312
## 5 130_202210_MSM_PPR_TR_11 439.7246 7.3107 3454.7595 77.8256 13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420 3.5063 5626.6706 63.3699 13.1578915
##           Cl_mM  salinity  S04_ugmL   Cl_ugmL S04_ugmL_spk S04_spk_Conc
## 1 1.169275 0.0749276 0.5716443 0.8290452 3.313680 2.5
## 2 18.950513 1.2139608 0.6274618 13.4359223 4.069623 2.5
## 3 31.166155 1.9964722 1.4032532 22.0967913 4.136507 2.5
## 4 71.607430 4.5870655 4.6872241 50.7696727 7.480386 2.5
## 5 97.454429 6.2427764 8.7945036 69.0951807 11.559035 2.5
## 6 158.721315 10.1674198 4.2179479 56.2611103 6.982359 2.5
## SampleVol
## 1 1.501
## 2 1.501
## 3 1.501
## 4 1.501
## 5 1.501
## 6 1.475
```

#gives us the total SO4 in the sample in mmoles

```
QAspks$S04_Total_unspkd <- QAspks$S04_ugmL*QAspks$SampleVol
```

##total SO4 in spiked sample in mmoles

```
QAspks$S04_Total_spkd <- (QAspks$S04_ugmL_spk)*(QAspks$SampleVol+spkvol)
```

```
QAspks$S04_expctd_spkd <- (QAspks$S04_Total_unspkd + QAspks$S04_spk_Conc)
```

```
QAspks$spk_recovery <- (QAspks$S04_Total_spkd/QAspks$S04_expctd_spkd)*100
```

```
QAspks$S04_spks_flag <- ifelse(QAspks$spk_recovery <=120 & QAspks$spk_recovery >=80 , 'YES', 'NO, rerun
```

```
head(QAspks)
```

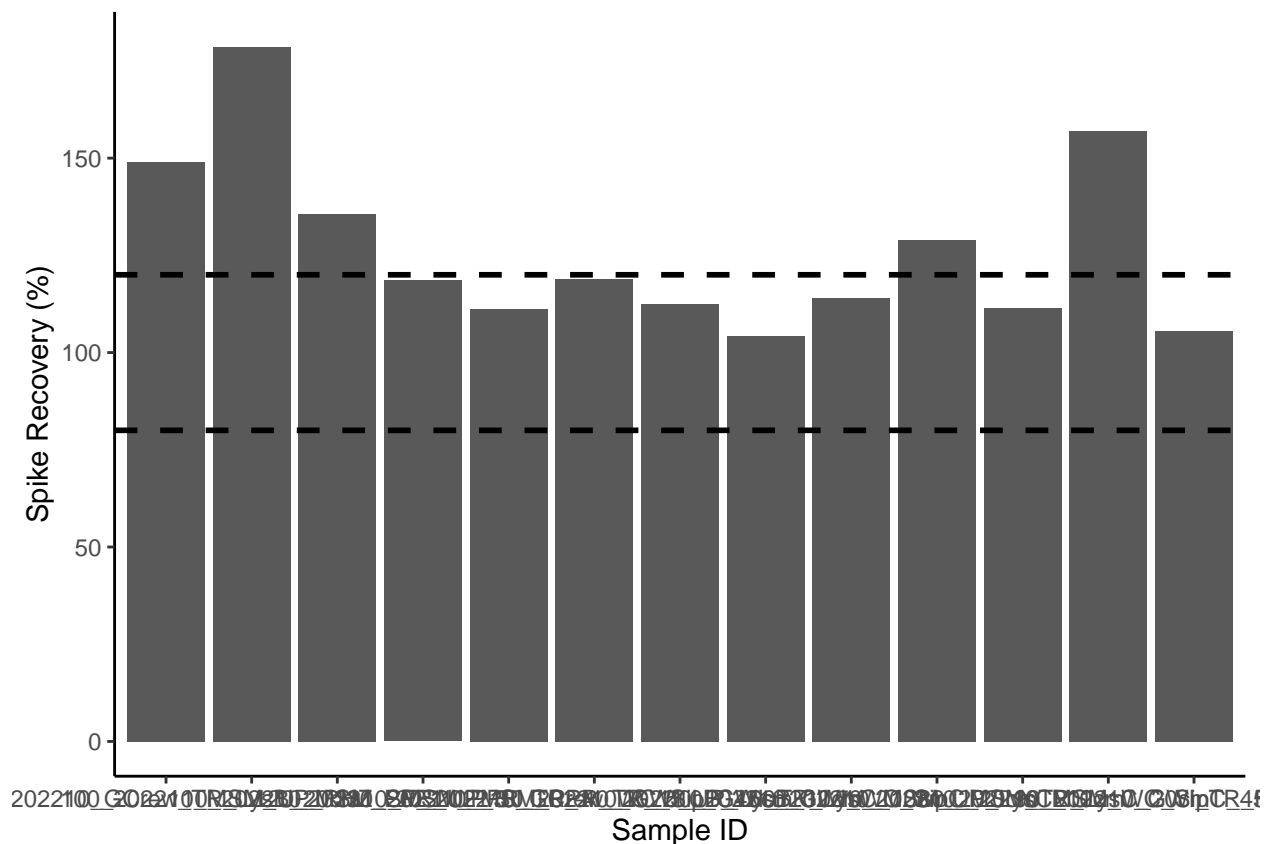
```
##           Sample_ID  S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 1 10_202210_GCrew_TR_LysB_20cm 28.5838 0.4752 41.4508 0.9338 0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1 31.3728 0.5216 671.7957 15.1336 0.9785652
## 3 110_202210_MSM_PPR_UP_3 70.1598 1.1665 1104.8402 24.8888 2.1883905
## 4 120_202210_MSM_PPR_TR_1 234.3596 3.8964 2538.4834 57.1846 7.3100312
## 5 130_202210_MSM_PPR_TR_11 439.7246 7.3107 3454.7595 77.8256 13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420 3.5063 5626.6706 63.3699 13.1578915
##           Cl_mM  salinity  S04_ugmL   Cl_ugmL S04_ugmL_spk S04_spk_Conc
## 1 1.169275 0.0749276 0.5716443 0.8290452 3.313680 2.5
## 2 18.950513 1.2139608 0.6274618 13.4359223 4.069623 2.5
## 3 31.166155 1.9964722 1.4032532 22.0967913 4.136507 2.5
## 4 71.607430 4.5870655 4.6872241 50.7696727 7.480386 2.5
## 5 97.454429 6.2427764 8.7945036 69.0951807 11.559035 2.5
## 6 158.721315 10.1674198 4.2179479 56.2611103 6.982359 2.5
## SampleVol S04_Total_unspkd S04_Total_spkd S04_expctd_spkd spk_recovery
## 1 1.501 0.8580380 5.006971 3.358038 149.1040
## 2 1.501 0.9418201 6.149200 3.441820 178.6613
## 3 1.501 2.1062830 6.250263 4.606283 135.6899
## 4 1.501 7.0355234 11.302863 9.535523 118.5343
## 5 1.501 13.2005499 17.465702 15.700550 111.2426
```

```
## 6      1.475      6.2214732      10.368803      8.721473      118.8882
## S04_spks_flag
## 1      NO, rerun
## 2      NO, rerun
## 3      NO, rerun
## 4              YES
## 5              YES
## 6              YES
```

#plot spk recoveries output as a bar graph to easily check - want any over 10% to be red need to work on

```
spksbar <- ggplot(data = QAspks, aes(x = Sample_ID, y = spk_recovery)) +
  geom_bar(stat = 'identity') +
  scale_fill_manual(values=c("lightblue4")) +
  theme_classic() + labs(x= "Sample ID", y="Spike Recovery (%)") +
  theme(legend.position="none") +
  geom_hline(yintercept=80, linetype="dashed", color = "black", size=1) +
  geom_hline(yintercept=120, linetype="dashed", color = "black", size=1)
```

spksbar



#check for percent of no, reruns to see if it would warrant reruns

```
Perc_spks <- QAspks %>%
  group_by(S04_spks_flag) %>%
  summarise(no_rows = length(S04_spks_flag))
Perc_spks$Total <- length(QAspks$S04_spks_flag)
```

```
Perc_spks$Percent <- (Perc_spks$no_rows / Perc_spks$Total)*100
head(Perc_spks)
```

```
## # A tibble: 2 x 4
##   S04_spks_flag no_rows Total Percent
##   <chr>         <int> <int>   <dbl>
## 1 NO, rerun      5     13   38.5
## 2 YES           8     13   61.5
```

Make final dataframe with IDs

```
#for steph <- pull out identifiers of the sample names
#pull the sample ID and separate it by the underscores
IDs <- data.frame(do.call('rbind', strsplit(as.character(sampledat2$Sample_ID), '_ ', fixed=TRUE)))
```

```
## Warning in rbind(c("1", "202210", "Gcrew", "UP", "LysA", "20cm"), c("10", :
## number of columns of result is not a multiple of vector length (arg 1)
```

```
colnames(IDs) <- c("Analysis_No" , "Date", "Site", "Zone", "Replicate", "Depth", "Tree")
head(IDs)
```

```
##   Analysis_No   Date Site Zone Replicate Depth Tree
## 1           1 202210 Gcrew  UP      LysA  20cm   1
## 2          10 202210 GCrew  TR      LysB  20cm  10
## 3         100 202210  MSM  UP       RHZ    SF   1
## 4         101 202210  MSM  UP       RHZ    SF   2
## 5         102 202210  MSM  UP       RHZ    SF   3
## 6         103 202210  MSM  UP       RHZ    SF   4
```

```
#rejoin them to the dataframe
alldat <- cbind(IDs, sampledat2)
head(alldat)
```

```
##   Analysis_No   Date Site Zone Replicate Depth Tree
## 1           1 202210 Gcrew  UP      LysA  20cm   1
## 2          10 202210 GCrew  TR      LysB  20cm  10
## 3         100 202210  MSM  UP       RHZ    SF   1
## 4         101 202210  MSM  UP       RHZ    SF   2
## 5         102 202210  MSM  UP       RHZ    SF   3
## 6         103 202210  MSM  UP       RHZ    SF   4
##           Sample_ID S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 1 1_202210_Gcrew_UP_LysA_20cm  4.1491  0.0690  14.4387  0.3253 0.1294167
## 2 10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338 0.8915721
## 3 100_202210_MSM_UP_RHZ_SF_1  31.3728  0.5216 671.7957 15.1336 0.9785652
## 4 101_202210_MSM_UP_RHZ_SF_2  65.5675  1.0901 933.5416 21.0300 2.0451497
## 5 102_202210_MSM_UP_RHZ_SF_3 105.7117  1.7575 1230.2357 27.7136 3.2973082
## 6 103_202210_MSM_UP_RHZ_SF_4  54.7246  0.9098 1060.9960 23.9011 1.7069432
##           Cl_mM   salinity
## 1 0.4072976 0.02611673
```

```
## 2  1.1692750 0.07492760
## 3 18.9505134 1.21396083
## 4 26.3340367 1.68693567
## 5 34.7034048 2.22306191
## 6 29.9293653 1.91724577
```

Make final dataframe with IDs

```
write.csv(alldat, file="Processed Data/COMPASS_SynopticCB_PW_Processed_Cl_S04_202210.csv") #Change file
```

END