

Dionex_COMPASS_October2022

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Daily Set up

Read in and Format the raw data - change wd & file names (Make sure files are UTF-8 encoded)

```
# SULFATE DATA:

## Read in raw data file from Dionex - copied and saved as a txt
Sdat <- read.table("Raw Data/COMPASS_Synoptic_CB_MonMon_202210_S04.txt",sep='\t' , header=T, skip=3)
head(Sdat)

##      X          X.1          X.2 IC.S04 IC.S04.1 IC.S04.2 IC.S04.3 IC.S04.4
## 1 1 Lab Blank Unknown    n.a.    n.a.    n.a.    n.a.    n.a.
## 2 2 Lab Blank Unknown    n.a.    n.a.    n.a.    n.a.    n.a.
## 3 3 Standard 1 Unknown 4.400   0.5687   6.39   0.4728   2.96
## 4 4     Blank Unknown    n.a.    n.a.    n.a.    n.a.    n.a.
## 5 5 Standard 1 Calibration Standard 4.397   0.5680   6.40   0.4722   2.98
## 6 6 Standard 2 Calibration Standard 4.400   1.1543   6.46   0.9595   5.88

## Only keep the columns that we need
Sdat <- Sdat[,c(2,5,7)] # dont need this here
head(Sdat)

##          X.1 IC.S04.1 IC.S04.3
## 1  Lab Blank    n.a.    n.a.
## 2  Lab Blank    n.a.    n.a.
## 3 Standard 1  0.5687  0.4728
## 4     Blank    n.a.    n.a.
## 5 Standard 1  0.5680  0.4722
## 6 Standard 2  1.1543  0.9595

## Name the columns correctly
colnames(Sdat) <- c( "Sample_ID", "S04_ppm", "S04_Area")
Sdat$Sample_ID <- as.factor(Sdat$Sample_ID)
Sdat$S04_ppm <- as.numeric(Sdat$S04_ppm)

## Warning: NAs introduced by coercion
```

```

Sdat$SO4_Area <- as.numeric(Sdat$SO4_Area)

## Warning: NAs introduced by coercion

Sdat <- as.data.frame(Sdat)
head(Sdat)

##      Sample_ID SO4_ppm SO4_Area
## 1    Lab Blank      NA       NA
## 2    Lab Blank      NA       NA
## 3 Standard 1   0.5687   0.4728
## 4     Blank      NA       NA
## 5 Standard 1   0.5680   0.4722
## 6 Standard 2   1.1543   0.9595

#Chloride data
## Read in raw data file from Dionex - copied and saved as a txt
Cldat <- read.table("Raw Data/COMPASS_Synoptic_CB_MonMon_202210_Cl.txt",sep='\t' , header=T, skip=3)
head(Cldat)

##      X        X.1          X.2  IC.Cl  IC.Cl.1 IC.Cl.2  IC.Cl.3 IC.Cl.4
## 1 1  Lab Blank Unknown 3.273  0.0471 100.00  0.0531  0.37
## 2 2  Lab Blank Unknown 3.280  0.0472 100.00  0.0531  0.38
## 3 3 Standard 1 Unknown 3.280  6.0990  92.87  6.8696  47.28
## 4 4     Blank      Unknown 3.273  0.0480 100.00  0.0541  0.39
## 5 5 Standard 1 Calibration Standard 3.280  6.0838  92.90  6.8525  47.38
## 6 6 Standard 2 Calibration Standard 3.280 12.2230  92.75 13.7674  94.28

## Only keep the columns that we need
Cldat <- Cldat[,c(2,5,7)]
head(Cldat)

##      X.1  IC.Cl.1  IC.Cl.3
## 1  Lab Blank  0.0471  0.0531
## 2  Lab Blank  0.0472  0.0531
## 3 Standard 1  6.0990  6.8696
## 4     Blank    0.0480  0.0541
## 5 Standard 1  6.0838  6.8525
## 6 Standard 2 12.2230 13.7674

## Name the columns correctly
colnames(Cldat) <- c( "Sample_ID", "Cl_ppm", "Cl_Area")
Cldat$Sample_ID <- as.factor(Cldat$Sample_ID)
Cldat$Cl_ppm <- as.numeric(Cldat$Cl_ppm)

## Warning: NAs introduced by coercion

Cldat$Cl_Area <- as.numeric(Cldat$Cl_Area)

## Warning: NAs introduced by coercion

```

```

Cldat <- as.data.frame(Cldat)
head(Cldat)

##      Sample_ID Cl_ppm Cl_Area
## 1   Lab Blank  0.0471  0.0531
## 2   Lab Blank  0.0472  0.0531
## 3 Standard 1  6.0990  6.8696
## 4     Blank    0.0480  0.0541
## 5 Standard 1  6.0838  6.8525
## 6 Standard 2 12.2230 13.7674

## Bring the data back together:
all_dat <- merge(Sdat, Cldat, by="Sample_ID")
head(all_dat)

##      Sample_ID S04_ppm S04_Area      Cl_ppm Cl_Area
## 1           2418.1928  17.1467 15120.9494 224.2274
## 2           2418.1928  17.1467  4224.4183  60.4754
## 3           2418.1928  17.1467       0.0212  0.0239
## 4           2418.1928  17.1467  4473.1255  50.9915
## 5           2418.1928  17.1467        NA       NA
## 6           586.9775   5.9183 15120.9494 224.2274

## Remove empty lines
all_dat <- all_dat[!(is.na(all_dat$Sample_ID) | all_dat$Sample_ID=="") , ]
head(all_dat)

##                               Sample_ID S04_ppm S04_Area      Cl_ppm Cl_Area
## 26      1_202210_Gcrew_UP_LysA_20cm  4.1491  0.0690  14.4387  0.3253
## 27      10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338
## 28 10_202210_GCrew_TR_LysB_20cm_spk 165.6846  2.7546  38.8350  0.8748
## 29      100_202210_MSM_UP_RHZ_SF_1  31.3728  0.5216  671.7957 15.1336
## 30     100_202210_MSM_UP_RHZ_SF_1_spk 203.4807  3.3830 1340.3550 30.1943
## 31      101_202210_MSM_UP_RHZ_SF_2  65.5675  1.0901  933.5416 21.0300

all_dat <- droplevels(all_dat[!all_dat$Sample_ID == 'Lab Blank',])
head(all_dat)

##                               Sample_ID S04_ppm S04_Area      Cl_ppm Cl_Area
## 26      1_202210_Gcrew_UP_LysA_20cm  4.1491  0.0690  14.4387  0.3253
## 27      10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338
## 28 10_202210_GCrew_TR_LysB_20cm_spk 165.6846  2.7546  38.8350  0.8748
## 29      100_202210_MSM_UP_RHZ_SF_1  31.3728  0.5216  671.7957 15.1336
## 30     100_202210_MSM_UP_RHZ_SF_1_spk 203.4807  3.3830 1340.3550 30.1943
## 31      101_202210_MSM_UP_RHZ_SF_2  65.5675  1.0901  933.5416 21.0300

```

Pull out standards - could do some checks here if we want

```

stds <- all_dat[grep("Standard", all_dat$Sample_ID),]
head(stds)

##      Sample_ID S04_ppm S04_Area Cl_ppm Cl_Area
## 1424 Standard 1  0.5615   0.4668 6.1419  6.9180
## 1425 Standard 1  0.5615   0.4668 6.1360  6.9113
## 1426 Standard 1  0.5615   0.4668 6.0990  6.8696
## 1427 Standard 1  0.5615   0.4668 6.1169  6.8898
## 1428 Standard 1  0.5615   0.4668 6.0921  6.8619
## 1429 Standard 1  0.5615   0.4668 6.0838  6.8525

stds_chk_S <- stds %>%
  group_by(Sample_ID) %>%
  summarise(mean = mean(S04_ppm), sd = sd(S04_ppm))
stds_chk_S$cv <- (stds_chk_S$sd/stds_chk_S$mean)*100
stds_chk_S$flag <- ifelse(stds_chk_S$cv <2, 'YES', 'NO, rerun')
head(stds_chk_S)

## # A tibble: 5 x 5
##   Sample_ID     mean       sd     cv flag
##   <fct>     <dbl>    <dbl>  <dbl> <chr>
## 1 Standard 1  0.568 0.00410 0.722 YES
## 2 Standard 2  1.16 0.00688 0.595 YES
## 3 Standard 3  2.30 0.00792 0.345 YES
## 4 Standard 4 10.4  0.0278  0.268 YES
## 5 Standard 5 19.9  0.0475  0.239 YES

stds_chk_Cl <- stds %>%
  group_by(Sample_ID) %>%
  summarise(mean = mean(Cl_ppm), sd = sd(Cl_ppm))
stds_chk_Cl$cv <- (stds_chk_Cl$sd/stds_chk_Cl$mean)*100
stds_chk_Cl$flag <- ifelse(stds_chk_Cl$cv <2, 'YES', 'NO, rerun')
head(stds_chk_Cl)

## # A tibble: 5 x 5
##   Sample_ID     mean       sd     cv flag
##   <fct>     <dbl>    <dbl>  <dbl> <chr>
## 1 Standard 1   6.11 0.0210 0.344 YES
## 2 Standard 2  12.2 0.0218 0.178 YES
## 3 Standard 3  23.6 0.0444 0.189 YES
## 4 Standard 4 104.   0.233 0.224 YES
## 5 Standard 5 199.   0.313 0.157 YES

lmS <- lm(stds$S04_Area ~ stds$S04_ppm)
S04_sum <- summary(lmS)
S04_Slope <- S04_sum$coefficients[2, 1]
S04_Int <- S04_sum$coefficients[1, 1]

lmCl <- lm(stds$Cl_Area ~ stds$Cl_ppm)
Cl_sum <- summary(lmCl)

```

```

Cl_Slope <- Cl_sum$coefficients[2, 1]
Cl_Int <- Cl_sum$coefficients[1, 1]

```

Calculate mmol/L concentrations

```

#remove standards from sample dataframe
sampledat <- all_dat[!grepl("Standard", all_dat$Sample_ID),]
head(sampledat)

##                                     Sample_ID  SO4_ppm  SO4_Area      Cl_ppm  Cl_Area
## 26      1_202210_Gcrew_UP_LysA_20cm   4.1491   0.0690   14.4387  0.3253
## 27      10_202210_GCrew_TR_LysB_20cm  28.5838   0.4752   41.4508  0.9338
## 28     10_202210_GCrew_TR_LysB_20cm_spk 165.6846   2.7546   38.8350  0.8748
## 29      100_202210_MSM_UP_RHZ_SF_1    31.3728   0.5216   671.7957 15.1336
## 30     100_202210_MSM_UP_RHZ_SF_1_spk 203.4807   3.3830 1340.3550 30.1943
## 31     101_202210_MSM_UP_RHZ_SF_2    65.5675   1.0901  933.5416 21.0300

# Constants needed for calculations:
clmw <- 35.45      #molecular weight of Chloride: 35.45
smw <- 32.06        #molecular weight of sulfur: 32.06

# Convert ppm to mmol/L
sampledat$SO4_mM <- (sampledat$SO4_ppm / smw)
sampledat$Cl_mM <- (sampledat$Cl_ppm / clmw)

# Calculate Salinity
# calculated using the Knudsen equation
# Salinity = 0.03 + 1.8050 * Chlorinity
# Ref: A Practical Handbook of Seawater Analysis by Strickland & Parsons (P. 11)
# =((1.807*Cl_ppm)+0.026)/1000
sampledat$salinity <- ((1.8070 * sampledat$Cl_ppm) + 0.026) / 1000

head(sampledat)

##                                     Sample_ID  SO4_ppm  SO4_Area      Cl_ppm  Cl_Area
## 26      1_202210_Gcrew_UP_LysA_20cm   4.1491   0.0690   14.4387  0.3253
## 27      10_202210_GCrew_TR_LysB_20cm  28.5838   0.4752   41.4508  0.9338
## 28     10_202210_GCrew_TR_LysB_20cm_spk 165.6846   2.7546   38.8350  0.8748
## 29      100_202210_MSM_UP_RHZ_SF_1    31.3728   0.5216   671.7957 15.1336
## 30     100_202210_MSM_UP_RHZ_SF_1_spk 203.4807   3.3830 1340.3550 30.1943
## 31     101_202210_MSM_UP_RHZ_SF_2    65.5675   1.0901  933.5416 21.0300
##          SO4_mM      Cl_mM  salinity
## 26 0.1294167  0.4072976 0.02611673
## 27 0.8915721  1.1692750 0.07492760
## 28 5.1679538  1.0954866 0.07020084
## 29 0.9785652 18.9505134 1.21396083
## 30 6.3468715 37.8097320 2.42204748
## 31 2.0451497 26.3340367 1.68693567

```

Pull out dups and check with percent difference

```
#Show me the data that we have from the calculations
head(sampledat)
```

```
##                               Sample_ID  S04_ppm  S04_Area      Cl_ppm  Cl_Area
## 26      1_202210_Gcrew_UP_LysA_20cm  4.1491  0.0690  14.4387  0.3253
## 27      10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338
## 28     10_202210_GCrew_TR_LysB_20cm_spk 165.6846  2.7546  38.8350  0.8748
## 29      100_202210_MSM_UP_RHZ_SF_1   31.3728  0.5216  671.7957 15.1336
## 30     100_202210_MSM_UP_RHZ_SF_1_spk 203.4807  3.3830 1340.3550 30.1943
## 31      101_202210_MSM_UP_RHZ_SF_2   65.5675  1.0901  933.5416 21.0300
##           S04_mM      Cl_mM  salinity
## 26 0.1294167  0.4072976 0.02611673
## 27 0.8915721  1.1692750 0.07492760
## 28 5.1679538  1.0954866 0.07020084
## 29 0.9785652 18.9505134 1.21396083
## 30 6.3468715 37.8097320 2.42204748
## 31 2.0451497 26.3340367 1.68693567
```

```
#pull out any rows that have "dup" in the SampleID column
dups <- sampledat %>%
  filter(str_detect(Sample_ID, "dup"))      #have to change this to match data
head(dups)
```

```
##                               Sample_ID  S04_ppm  S04_Area      Cl_ppm  Cl_Area
## 1      109_202210_MSM_PPR_UP_2_dup  57.3788  0.9540 1055.464 23.7765
## 2      119_202210_MSM_PPR_UP_12_dup 140.2629  2.3320 2355.659 53.0661
## 3      129_202210_MSM_PPR_TR_10_dup 466.9523  7.7634 3585.616 80.7734
## 4     19_202210_GCrew_WC_SipB_20cm_dup 436.3971  3.6273 5767.606 64.9571
## 5      29_202210_GWI_UP_LysA_45cm_dup 651.2403  5.4131 5494.783 61.8845
## 6      39_202210_GWI_TR_LysB_45cm_dup 1084.1925  9.0118 9592.738 108.0373
##           S04_mM      Cl_mM  salinity
## 1    1.789732 29.77331 1.907249
## 2    4.375012 66.45018 4.256702
## 3   14.564950 101.14572 6.479233
## 4  13.611887 162.69693 10.422090
## 5 20.313172 155.00093 9.929099
## 6 33.817608 270.59911 17.334104
```

```
#remove these from sample dataframe in a new dataframe
sampledat2 <- sampledat %>%
  filter(!str_detect(Sample_ID, "dup")) %>%
  filter(!str_detect(Sample_ID, "spk"))
head(sampledat2)
```

```
##                               Sample_ID  S04_ppm  S04_Area      Cl_ppm  Cl_Area      S04_mM
## 1 1_202210_Gcrew_UP_LysA_20cm  4.1491  0.0690  14.4387  0.3253 0.1294167
## 2 10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338 0.8915721
## 3 100_202210_MSM_UP_RHZ_SF_1   31.3728  0.5216  671.7957 15.1336 0.9785652
## 4 101_202210_MSM_UP_RHZ_SF_2   65.5675  1.0901  933.5416 21.0300 2.0451497
```

```

## 5 102_202210_MSM_UP_RHZ_SF_3 105.7117 1.7575 1230.2357 27.7136 3.2973082
## 6 103_202210_MSM_UP_RHZ_SF_4 54.7246 0.9098 1060.9960 23.9011 1.7069432
## Cl_mM salinity
## 1 0.4072976 0.02611673
## 2 1.1692750 0.07492760
## 3 18.9505134 1.21396083
## 4 26.3340367 1.68693567
## 5 34.7034048 2.22306191
## 6 29.9293653 1.91724577

```

```

#remove the dup from these IDs so we will have duplicate sample names
dups$Sample_ID<-gsub("_dup","",as.character(dups$Sample_ID))
dups <- dups[, -c(2:5)]
colnames(dups) <- c('Sample_ID', 'SO4_mM_dup', 'Cl_mM_dup', "salinity_dup")
head(dups)

```

```

##                               Sample_ID SO4_mM_dup Cl_mM_dup salinity_dup
## 1      109_202210_MSM_PPR_UP_2   1.789732  29.77331     1.907249
## 2      119_202210_MSM_PPR_UP_12   4.375012  66.45018     4.256702
## 3      129_202210_MSM_PPR_TR_10  14.564950 101.14572     6.479233
## 4 19_202210_GCrew_WC_SipB_20cm 13.611887 162.69693    10.422090
## 5 29_202210_GWI_UP_LysA_45cm  20.313172 155.00093     9.929099
## 6 39_202210_GWI_TR_LysB_45cm  33.817608 270.59911    17.334104

```

```

#put it back together with the old data set and look for duplicates
QAdups <- merge(sampledat2, dups)
head(QAdups)

```

```

##                               Sample_ID   SO4_ppm SO4_Area   Cl_ppm Cl_Area   SO4_mM
## 1      109_202210_MSM_PPR_UP_2 56.9315  0.9465 1042.804 23.4913 1.775780
## 2      119_202210_MSM_PPR_UP_12 141.9209  2.3595 2376.423 53.5338 4.426728
## 3      129_202210_MSM_PPR_TR_10 462.9205  7.6964 3552.531 80.0281 14.439192
## 4 19_202210_GCrew_WC_SipB_20cm 441.4171  3.6691 5832.465 65.6876 13.768468
## 5 29_202210_GWI_UP_LysA_45cm  655.8744  5.4516 5534.610 62.3330 20.457717
## 6 39_202210_GWI_TR_LysB_45cm 1109.8531  9.2251 9834.080 110.7554 34.618001
## Cl_mM salinity SO4_mM_dup Cl_mM_dup salinity_dup
## 1 29.41620 1.884373  1.789732  29.77331     1.907249
## 2 67.03591 4.294222  4.375012  66.45018     4.256702
## 3 100.21243 6.419449 14.564950 101.14572     6.479233
## 4 164.52650 10.539290 13.611887 162.69693    10.422090
## 5 156.12440 10.001066 20.313172 155.00093     9.929099
## 6 277.40705 17.770208 33.817608 270.59911    17.334104

```

```

QAdups$SO4_dups_chk <- ((abs(QAdups$SO4_mM-QAdups$SO4_mM_dup))/((QAdups$SO4_mM+QAdups$SO4_mM_dup)/2))*100
QAdups$SO4_dups_flag <- ifelse(QAdups$SO4_dups_chk <10, 'YES', 'NO, rerun')

```

```

QAdups$Cl_dups_chk <- ((abs(QAdups$Cl_mM-QAdups$Cl_mM_dup))/((QAdups$Cl_mM+QAdups$Cl_mM_dup)/2))*100
QAdups$Cl_dups_flag <- ifelse(QAdups$Cl_dups_chk <10, 'YES', 'NO, rerun')

```

```

head(QAdups)

```

```

##                               Sample_ID   SO4_ppm SO4_Area   Cl_ppm Cl_Area   SO4_mM

```

```

## 1      109_202210_MSM_PPR_UP_2   56.9315  0.9465 1042.804 23.4913 1.775780
## 2      119_202210_MSM_PPR_UP_12  141.9209  2.3595 2376.423 53.5338 4.426728
## 3      129_202210_MSM_PPR_TR_10  462.9205  7.6964 3552.531 80.0281 14.439192
## 4 19_202210_GCrew_WC_SipB_20cm  441.4171  3.6691 5832.465 65.6876 13.768468
## 5      29_202210_GWI_UP_LysA_45cm 655.8744  5.4516 5534.610 62.3330 20.457717
## 6      39_202210_GWI_TR_LysB_45cm 1109.8531  9.2251 9834.080 110.7554 34.618001
##      Cl_mM  salinity SO4_mM_dup Cl_mM_dup salinity_dup SO4_dups_chk
## 1  29.41620  1.884373  1.789732 29.77331  1.907249  0.7826066
## 2  67.03591  4.294222  4.375012 66.45018  4.256702  1.1751206
## 3 100.21243  6.419449 14.564950 101.14572  6.479233  0.8671724
## 4 164.52650 10.539290 13.611887 162.69693 10.422090 1.1437500
## 5 156.12440 10.001066 20.313172 155.00093  9.929099  0.7090579
## 6 277.40705 17.770208 33.817608 270.59911 17.334104  2.3391127
##      SO4_dups_flag Cl_dups_chk Cl_dups_flag
## 1          YES    1.2066713      YES
## 2          YES    0.8775842      YES
## 3          YES    0.9269914      YES
## 4          YES    1.1182432      YES
## 5          YES    0.7222011      YES
## 6          YES    2.4846220      YES

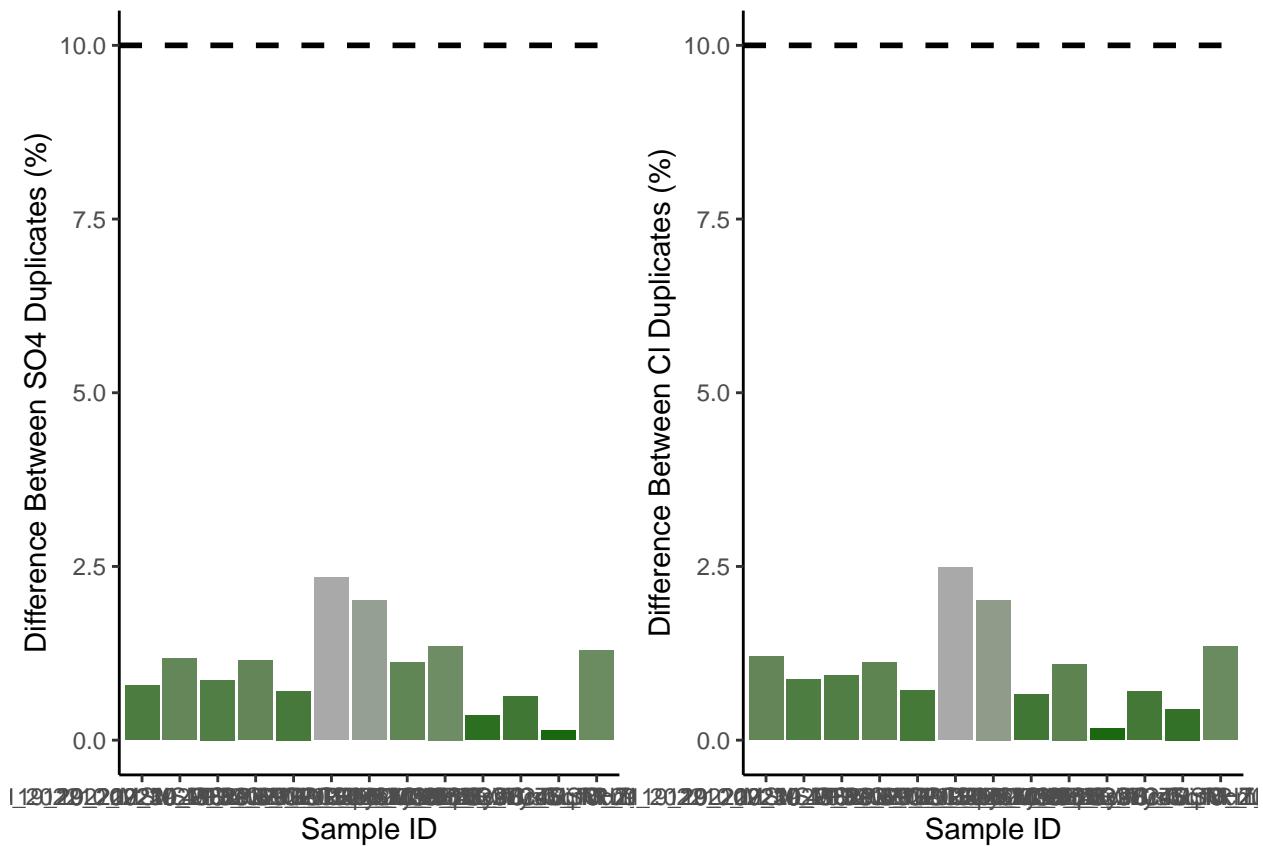
#plot dups output as a bar graph to easily check - want any over 10% to be red need to work on this
Sdupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = SO4_dups_chk, fill=SO4_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="Difference Between SO4 Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
                                             color = "black", size=1)

## Warning: Using 'size' aesthetic for lines was deprecated in ggplot2 3.4.0.
## i Please use 'linewidth' instead.
## This warning is displayed once every 8 hours.
## Call 'lifecycle::last_lifecycle_warnings()' to see where this warning was
## generated.

Cldupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = Cl_dups_chk, fill=Cl_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="Difference Between Cl Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
                                             color = "black", size=1)

ggarrange(Sdupsbar, Cldupsbar, ncol=2, nrow=1)

```



```
#check for percent of no, reruns to see if it would warrant reruns
Perc_dups <- QAdups %>%
  group_by(S04_dups_flag) %>%
  summarise(S_no_rows = length(S04_dups_flag))
Perc_dups1 <- QAdups %>%
  group_by(Cl_dups_flag) %>%
  summarise(Cl_no_rows = length(Cl_dups_flag))
colnames(Perc_dups) <- c("Flag", "S_no_rows")
colnames(Perc_dups1) <- c("Flag", "Cl_no_rows")
Perc_dups2 <- cbind(Perc_dups, Perc_dups1)

Perc_dups2$Total <- length(QAdups$S04_dups_flag)
Perc_dups2$S_Percent <- (Perc_dups2$S_no_rows / Perc_dups2$Total)*100
Perc_dups2$Cl_Percent <- (Perc_dups2$Cl_no_rows / Perc_dups2$Total)*100
head(Perc_dups2)
```

```
##   Flag S_no_rows Flag Cl_no_rows Total S_Percent Cl_Percent
## 1  YES      13  YES      13     13     100      100
```

Pull out dups and check with cv

```
#the cv
# calculate the sd bewteen the two duplicates then divide by the mean and multiply by 100
```

```

df2 <- as.data.frame(QAdups$SO4_mM)
df2$dups <- QAdups$SO4_mM_dup

df2$sds <- apply(df2, 1, sd)

QAdups$SO4_dups_cv <- (df2$sds) / ((QAdups$SO4_mM + QAdups$SO4_mM_dup) / 2) * 100
QAdups$SO4_dups_cv_flag <- ifelse(QAdups$SO4_dups_cv < 11, 'YES', 'NO, rerun')

head(QAdups)

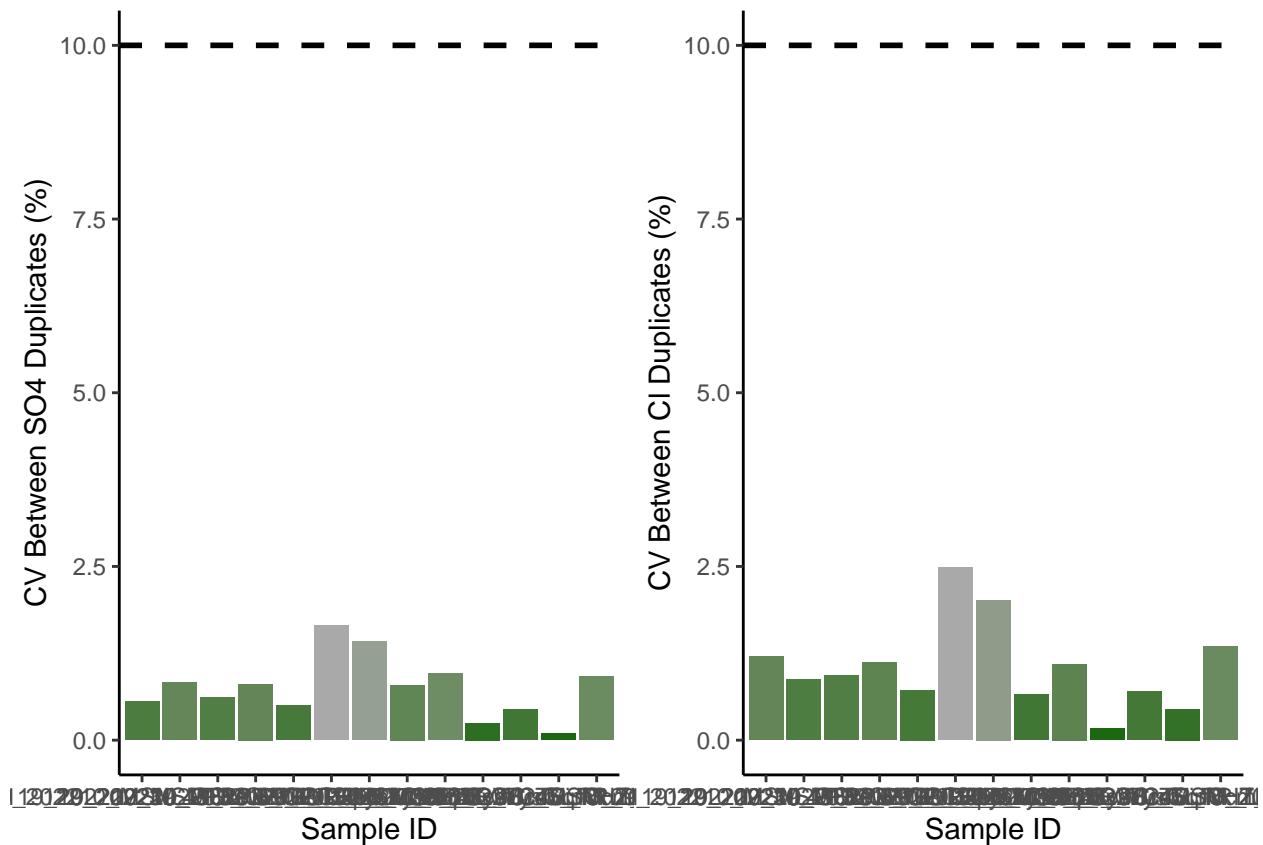
##                               Sample_ID   SO4_ppm SO4_Area   Cl_ppm Cl_Area   SO4_mM
## 1      109_202210_MSM_PPR_UP_2 56.9315  0.9465 1042.804 23.4913 1.775780
## 2      119_202210_MSM_PPR_UP_12 141.9209  2.3595 2376.423 53.5338 4.426728
## 3      129_202210_MSM_PPR_TR_10 462.9205  7.6964 3552.531 80.0281 14.439192
## 4 19_202210_GCrew_WC_SipB_20cm 441.4171  3.6691 5832.465 65.6876 13.768468
## 5      29_202210_GWI_UP_LysA_45cm 655.8744  5.4516 5534.610 62.3330 20.457717
## 6      39_202210_GWI_TR_LysB_45cm 1109.8531  9.2251 9834.080 110.7554 34.618001
##          Cl_mM salinity SO4_mM_dup Cl_mM_dup salinity_dup SO4_dups_chk
## 1 29.41620  1.884373  1.789732 29.77331    1.907249  0.7826066
## 2 67.03591  4.294222  4.375012 66.45018    4.256702  1.1751206
## 3 100.21243 6.419449 14.564950 101.14572    6.479233  0.8671724
## 4 164.52650 10.539290 13.611887 162.69693   10.422090  1.1437500
## 5 156.12440 10.001066 20.313172 155.00093   9.929099  0.7090579
## 6 277.40705 17.770208 33.817608 270.59911   17.334104  2.3391127
##          SO4_dups_flag Cl_dups_chk Cl_dups_flag SO4_dups_cv SO4_dups_cv_flag
## 1             YES     1.2066713      YES  0.5533865           YES
## 2             YES     0.8775842      YES  0.8309358           YES
## 3             YES     0.9269914      YES  0.6131835           YES
## 4             YES     1.1182432      YES  0.8087534           YES
## 5             YES     0.7222011      YES  0.5013796           YES
## 6             YES     2.4846220      YES  1.6540025           YES

#plot dups output as a bar graph to easily check - want any over 10% to be red need to work on this
Sdupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = SO4_dups_cv, fill=SO4_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="CV Between SO4 Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
                                              color = "black", size=1)

Cldupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = Cl_dups_chk, fill=Cl_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="CV Between Cl Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
                                              color = "black", size=1)

ggarrange(Sdupsbar, Cldupsbar, ncol=2, nrow=1)

```



```
#check for percent of no, reruns to see if it would warrant reruns
Perc_dups <- QAdups %>%
  group_by(SO4_dups_flag) %>%
  summarise(S_no_rows = length(SO4_dups_flag))
Perc_dups1 <- QAdups %>%
  group_by(Cl_dups_flag) %>%
  summarise(Cl_no_rows = length(Cl_dups_flag))
colnames(Perc_dups) <- c("Flag", "S_no_rows")
colnames(Perc_dups1) <- c("Flag", "Cl_no_rows")
Perc_dups2 <- cbind(Perc_dups, Perc_dups1)

Perc_dups2$Total <- length(QAdups$SO4_dups_flag)
Perc_dups2$S_Percent <- (Perc_dups2$S_no_rows / Perc_dups2$Total)*100
Perc_dups2$Cl_Percent <- (Perc_dups2$Cl_no_rows / Perc_dups2$Total)*100
head(Perc_dups2)
```

	Flag	S_no_rows	Flag	Cl_no_rows	Total	S_Percent	Cl_Percent
## 1	YES	13	YES	13	13	100	100

Pull out spikes and check with dionex output conc.

```
#Show me the data that we have from the calculations
head(sampledat)
```

```

##                               Sample_ID  SO4_ppm SO4_Area      Cl_ppm Cl_Area
## 26      1_202210_Gcrew_UP_LysA_20cm   4.1491  0.0690    14.4387 0.3253
## 27      10_202210_GCrew_TR_LysB_20cm  28.5838  0.4752    41.4508 0.9338
## 28 10_202210_GCrew_TR_LysB_20cm_spk 165.6846  2.7546    38.8350 0.8748
## 29      100_202210_MSM_UP_RHZ_SF_1   31.3728  0.5216   671.7957 15.1336
## 30     100_202210_MSM_UP_RHZ_SF_1_spk 203.4807  3.3830 1340.3550 30.1943
## 31      101_202210_MSM_UP_RHZ_SF_2   65.5675  1.0901  933.5416 21.0300
##                               SO4_mM   Cl_mM  salinity
## 26 0.1294167 0.4072976 0.02611673
## 27 0.8915721 1.1692750 0.07492760
## 28 5.1679538 1.0954866 0.07020084
## 29 0.9785652 18.9505134 1.21396083
## 30 6.3468715 37.8097320 2.42204748
## 31 2.0451497 26.3340367 1.68693567

```

```

#pull out any rows that have "spk" in the SampleID column
spks <- sampledat %>%
  filter(str_detect(Sample_ID, "spk"))      #have to change this to match data
head(spks)

```

```

##                               Sample_ID  SO4_ppm SO4_Area      Cl_ppm Cl_Area      SO4_mM
## 1 10_202210_GCrew_TR_LysB_20cm_spk 165.6846  2.7546    38.835 0.8748 5.167954
## 2 100_202210_MSM_UP_RHZ_SF_1_spk 203.4807  3.3830 1340.355 30.1943 6.346871
## 3 110_202210_MSM_PPR_UP_3_spk 206.8264  3.4386 1094.155 24.6481 6.451229
## 4 120_202210_MSM_PPR_TR_1_spk 374.0176  6.2183 2572.971 57.9615 11.666176
## 5 130_202210_MSM_PPR_TR_11_spk 577.9479  9.6088 3459.534 77.9332 18.027071
## 6 20_202210_GCrew_WC_SipB_45cm_spk 698.3096  5.8043 5526.222 62.2386 21.781335
##                               Cl_mM  salinity
## 1 1.095487 0.07020084
## 2 37.809732 2.42204748
## 3 30.864725 1.97716318
## 4 72.580279 4.64938442
## 5 97.589109 6.25140376
## 6 155.887797 9.98590988

```

```

#remove the dup from these IDs so we will have duplicate sample names
spks$Sample_ID<-gsub("_spk","",as.character(spks$Sample_ID))
spks <- spks[, -c(2:5,7,8)]
colnames(spks) <- c('Sample_ID', 'SO4_mM_spk')
head(spks)

```

```

##                               Sample_ID SO4_mM_spk
## 1 10_202210_GCrew_TR_LysB_20cm 5.167954
## 2 100_202210_MSM_UP_RHZ_SF_1 6.346871
## 3 110_202210_MSM_PPR_UP_3 6.451229
## 4 120_202210_MSM_PPR_TR_1 11.666176
## 5 130_202210_MSM_PPR_TR_11 18.027071
## 6 20_202210_GCrew_WC_SipB_45cm 21.781335

```

```

#put it back together with the old data set and look for duplicates
QAspks <- merge(sampledat, spks)
head(QAspks)

```

```

##                               Sample_ID  S04_ppm S04_Area      Cl_ppm Cl_Area      S04_mM
## 1 10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338  0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1   31.3728  0.5216  671.7957 15.1336  0.9785652
## 3 110_202210_MSM_PPR_UP_3     70.1598  1.1665 1104.8402 24.8888  2.1883905
## 4 120_202210_MSM_PPR_TR_1    234.3596  3.8964 2538.4834 57.1846  7.3100312
## 5 130_202210_MSM_PPR_TR_11   439.7246  7.3107 3454.7595 77.8256  13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420  3.5063 5626.6706 63.3699  13.1578915
##                               Cl_mM  salinity S04_mM_spk
## 1    1.169275  0.0749276  5.167954
## 2   18.950513  1.2139608  6.346871
## 3   31.166155  1.9964722  6.451229
## 4   71.607430  4.5870655 11.666176
## 5   97.454429  6.2427764 18.027071
## 6 158.721315 10.1674198 21.781335

```

```

#now we need to calculate the spike concentration and calculate the spike recovery
spkconc <- (250/smw)      # in mM
spkvol <- 10                # in uL
spkvol <- spkvol/1000000
#spike for these samples was 10uL of the 250mM standard
QAspks$S04_spk_Conc <- (spkconc)*spkvol           # mmoles of S04
head(QAspks)

```

```

##                               Sample_ID  S04_ppm S04_Area      Cl_ppm Cl_Area      S04_mM
## 1 10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338  0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1   31.3728  0.5216  671.7957 15.1336  0.9785652
## 3 110_202210_MSM_PPR_UP_3     70.1598  1.1665 1104.8402 24.8888  2.1883905
## 4 120_202210_MSM_PPR_TR_1    234.3596  3.8964 2538.4834 57.1846  7.3100312
## 5 130_202210_MSM_PPR_TR_11   439.7246  7.3107 3454.7595 77.8256  13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420  3.5063 5626.6706 63.3699  13.1578915
##                               Cl_mM  salinity S04_mM_spk S04_spk_Conc
## 1    1.169275  0.0749276  5.167954 7.797879e-05
## 2   18.950513  1.2139608  6.346871 7.797879e-05
## 3   31.166155  1.9964722  6.451229 7.797879e-05
## 4   71.607430  4.5870655 11.666176 7.797879e-05
## 5   97.454429  6.2427764 18.027071 7.797879e-05
## 6 158.721315 10.1674198 21.781335 7.797879e-05

```

```

#need to determine dilution factors and initial amount of sample added
#if your samples are all the same dilution just use the first line
#for Steph / COMPASS this depends on the site so...
QAspks$Dilution <- 1
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 200, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$Dilution)

```

```

#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$Dilution)

#Set Sample volumes in uL
QAspks$SampleVol <- 1
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 1462, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$SampleVol)

#change sample volume to L
QAspks$SampleVol <- QAspks$SampleVol/1000000
head(QAspks)

##                               Sample_ID  SO4_ppm  SO4_Area   Cl_ppm Cl_Area    SO4_mM
## 1 10_202210_GCrew_TR_LysB_20cm  28.5838  0.4752  41.4508  0.9338  0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1   31.3728  0.5216  671.7957 15.1336  0.9785652
## 3 110_202210_MSM_PPR_UP_3     70.1598  1.1665 1104.8402 24.8888  2.1883905
## 4 120_202210_MSM_PPR_TR_1    234.3596  3.8964 2538.4834 57.1846  7.3100312
## 5 130_202210_MSM_PPR_TR_11   439.7246  7.3107 3454.7595 77.8256 13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420  3.5063 5626.6706 63.3699 13.1578915
##          Cl_mM  salinity SO4_mM_spk SO4_spk_Conc Dilution SampleVol
## 1  1.169275  0.0749276  5.167954 7.797879e-05      50  0.001501
## 2 18.950513  1.2139608  6.346871 7.797879e-05      50  0.001501
## 3 31.166155  1.9964722  6.451229 7.797879e-05      1  0.000001
## 4 71.607430  4.5870655 11.666176 7.797879e-05      1  0.000001
## 5 97.454429  6.2427764 18.027071 7.797879e-05      1  0.000001
## 6 158.721315 10.1674198 21.781335 7.797879e-05    100  0.001475

#gives us the total SO4 in the sample in mmoles
QAspks$SO4_Total_unspkd <- (QAspks$SO4_mM/QAspks$Dilution)*(QAspks$SampleVol)

##total SO4 in spiked sample in mmoles
QAspks$SO4_Total_spkd <- (QAspks$SO4_mM_spk/QAspks$Dilution)*(QAspks$SampleVol+spkvol)

QAspks$SO4_expctd_spkd <- (QAspks$SO4_Total_unspkd + QAspks$SO4_spk_Conc)
QAspks$spk_recovery <- (QAspks$SO4_Total_spkd/QAspks$SO4_expctd_spkd)*100
QAspks$SO4_spks_flag <- ifelse(QAspks$spk_recovery <=120 & QAspks$spk_recovery >=80 , 'YES', 'NO, rerun'

head(QAspks)

##                               Sample_ID  SO4_ppm  SO4_Area   Cl_ppm Cl_Area    SO4_mM
## 1 10_202210_GCrew_TR_LysB_20cm  28.5838  0.4752  41.4508  0.9338  0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1   31.3728  0.5216  671.7957 15.1336  0.9785652
## 3 110_202210_MSM_PPR_UP_3     70.1598  1.1665 1104.8402 24.8888  2.1883905

```

```

## 4      120_202210_MSM_PPR_TR_1 234.3596   3.8964 2538.4834 57.1846 7.3100312
## 5      130_202210_MSM_PPR_TR_11 439.7246   7.3107 3454.7595 77.8256 13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420   3.5063 5626.6706 63.3699 13.1578915
##          Cl_mM    salinity S04_mM_spk S04_spk_Conc Dilution SampleVol
## 1     1.169275  0.0749276  5.167954 7.797879e-05       50  0.001501
## 2     18.950513  1.2139608  6.346871 7.797879e-05       50  0.001501
## 3     31.166155  1.9964722  6.451229 7.797879e-05        1  0.000001
## 4     71.607430  4.5870655 11.666176 7.797879e-05        1  0.000001
## 5     97.454429  6.2427764 18.027071 7.797879e-05        1  0.000001
## 6    158.721315 10.1674198 21.781335 7.797879e-05     100  0.001475
##          S04_Total_unspkd S04_Total_spkd S04_expctd_spkd spk_recovery S04_spks_flag
## 1     2.676499e-05  1.561756e-04  1.047438e-04    149.10247    NO, rerun
## 2     2.937653e-05  1.918025e-04  1.073553e-04    178.66135    NO, rerun
## 3     2.188391e-06  7.096352e-05  8.016718e-05    88.51941      YES
## 4     7.310031e-06  1.283279e-04  8.528882e-05   150.46278    NO, rerun
## 5     1.371568e-05  1.982978e-04  9.169447e-05   216.25927    NO, rerun
## 6     1.940789e-04  3.234528e-04  2.720577e-04   118.89126      YES

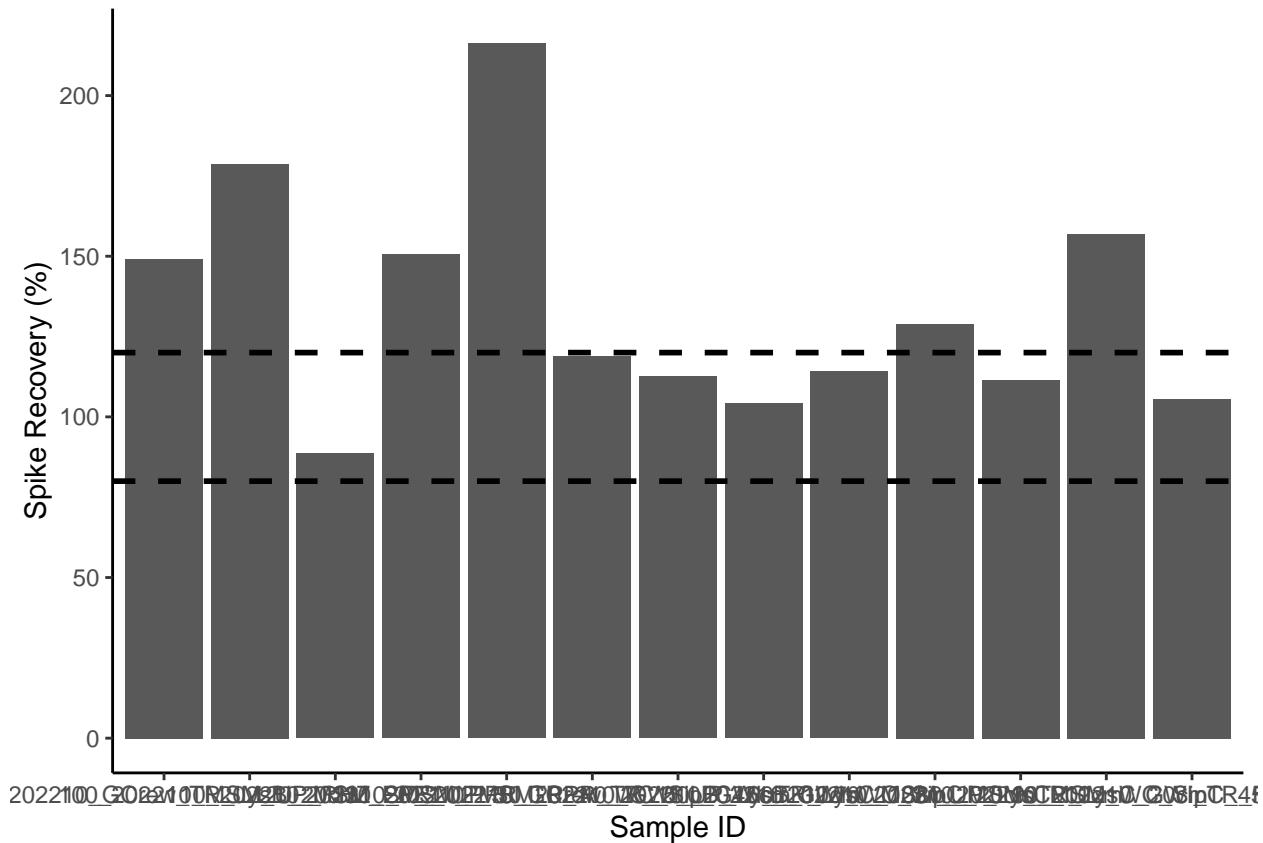
```

```

#plot spk recoveries output as a bar graph to easily check - want any over 10% to be red need to work on this
spksbar <- ggplot(data = QAspks, aes(x = Sample_ID, y = spk_recovery)) +
  geom_bar(stat = 'identity') +
  scale_fill_manual(values=c("lightblue4")) +
  theme_classic() + labs(x= "Sample ID", y="Spike Recovery (%)") +
  theme(legend.position="none") +
  geom_hline(yintercept=80, linetype="dashed", color = "black", size=1) +
  geom_hline(yintercept=120, linetype="dashed", color = "black", size=1)

```

spksbar



```
#check for percent of no, reruns to see if it would warrant reruns
Perc_spks <- QAspks %>%
  group_by(S04_spks_flag) %>%
  summarise(no_rows = length(S04_spks_flag))
Perc_spks$Total <- length(QAspks$S04_spks_flag)
Perc_spks$Percent <- (Perc_spks$no_rows / Perc_spks$Total)*100
head(Perc_spks)
```

```
## # A tibble: 2 x 4
##   S04_spks_flag no_rows Total Percent
##   <chr>          <int> <int>   <dbl>
## 1 NO, rerun      6     13    46.2
## 2 YES            7     13    53.8
```

Pull out spikes and check with area calc

```
#Show me the data that we have from the calculations  
head(sampleddat)
```

```

##                                     Sample_ID  SO4_ppm SO4_Area      Cl_ppm Cl_Area
## 26      1_202210_Gcrew_UP_LysA_20cm  4.1491  0.0690  14.4387  0.3253
## 27      10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338
## 28 10_202210_GCrew_TR_LysB_20cm_spk 165.6846  2.7546  38.8350  0.8748

```

```

## 29      100_202210_MSM_UP_RHZ_SF_1 31.3728  0.5216  671.7957 15.1336
## 30      100_202210_MSM_UP_RHZ_SF_1_spk 203.4807 3.3830 1340.3550 30.1943
## 31      101_202210_MSM_UP_RHZ_SF_2 65.5675  1.0901  933.5416 21.0300
##      SO4_mM     Cl_mM   salinity
## 26 0.1294167 0.4072976 0.02611673
## 27 0.8915721 1.1692750 0.07492760
## 28 5.1679538 1.0954866 0.07020084
## 29 0.9785652 18.9505134 1.21396083
## 30 6.3468715 37.8097320 2.42204748
## 31 2.0451497 26.3340367 1.68693567

#now use area of all samples and calculate undilution corrected concentrations in ug/mL
sampledat$SO4_ugmL <- ((sampledat$SO4_Area-SO4_Int)/SO4_Slope)
sampledat$Cl_ugmL <- (sampledat$Cl_Area-Cl_Int)/Cl_Slope
head(sampledat)

##                               Sample_ID  SO4_ppm SO4_Area     Cl_ppm Cl_Area
## 26      1_202210_Gcrew_UP_LysA_20cm 4.1491  0.0690  14.4387  0.3253
## 27      10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338
## 28 10_202210_GCrew_TR_LysB_20cm_spk 165.6846  2.7546  38.8350  0.8748
## 29      100_202210_MSM_UP_RHZ_SF_1 31.3728  0.5216  671.7957 15.1336
## 30      100_202210_MSM_UP_RHZ_SF_1_spk 203.4807 3.3830 1340.3550 30.1943
## 31      101_202210_MSM_UP_RHZ_SF_2 65.5675  1.0901  933.5416 21.0300
##      SO4_mM     Cl_mM   salinity SO4_ugmL     Cl_ugmL
## 26 0.1294167 0.4072976 0.02611673 0.08300035 0.2888063
## 27 0.8915721 1.1692750 0.07492760 0.57164426 0.8290452
## 28 5.1679538 1.0954866 0.07020084 3.31368003 0.7766638
## 29 0.9785652 18.9505134 1.21396083 0.62746178 13.4359223
## 30 6.3468715 37.8097320 2.42204748 4.06962251 26.8071243
## 31 2.0451497 26.3340367 1.68693567 1.31134672 18.6708686

#pull out any rows that have "spk" in the SampleID column
spks <- sampledat %>%
  filter(str_detect(Sample_ID, "spk"))      #have to change this to match data
head(spks)

##                               Sample_ID  SO4_ppm SO4_Area     Cl_ppm Cl_Area     SO4_mM
## 1 10_202210_GCrew_TR_LysB_20cm_spk 165.6846  2.7546  38.835  0.8748 5.167954
## 2 100_202210_MSM_UP_RHZ_SF_1_spk 203.4807  3.3830 1340.355 30.1943 6.346871
## 3 110_202210_MSM_PPR_UP_3_spk 206.8264  3.4386 1094.155 24.6481 6.451229
## 4 120_202210_MSM_PPR_TR_1_spk 374.0176  6.2183 2572.971 57.9615 11.666176
## 5 130_202210_MSM_PPR_TR_11_spk 577.9479  9.6088 3459.534 77.9332 18.027071
## 6 20_202210_GCrew_WC_SipB_45cm_spk 698.3096  5.8043 5526.222 62.2386 21.781335
##      Cl_mM   salinity SO4_ugmL     Cl_ugmL
## 1 1.095487 0.07020084 3.313680 0.7766638
## 2 37.809732 2.42204748 4.069623 26.8071243
## 3 30.864725 1.97716318 4.136507 21.8830928
## 4 72.580279 4.64938442 7.480386 51.4594206
## 5 97.589109 6.25140376 11.559035 69.1907103
## 6 155.887797 9.98590988 6.982359 55.2567187

```

```

## Only keep the columns that we need
spks <- spks[ ,c(1,9,10)] # dont need this here
head(spks)

##                                     Sample_ID  S04_ugmL    Cl_ugmL
## 1 10_202210_GCrew_TR_LysB_20cm_spk 3.313680  0.7766638
## 2 100_202210_MSM_UP_RHZ_SF_1_spk  4.069623 26.8071243
## 3 110_202210_MSM_PPR_UP_3_spk   4.136507 21.8830928
## 4 120_202210_MSM_PPR_TR_1_spk   7.480386 51.4594206
## 5 130_202210_MSM_PPR_TR_11_spk 11.559035 69.1907103
## 6 20_202210_GCrew_WC_SipB_45cm_spk 6.982359 55.2567187

#remove the spk from these IDs so we will have duplicate sample names
spks$Sample_ID<-gsub("_spk","",as.character(spks$Sample_ID))
spks <- spks[ ,c(1,2)]
colnames(spks) <- c('Sample_ID', 'S04_ugmL_spk')
head(spks)

##                                     Sample_ID S04_ugmL_spk
## 1 10_202210_GCrew_TR_LysB_20cm      3.313680
## 2 100_202210_MSM_UP_RHZ_SF_1       4.069623
## 3 110_202210_MSM_PPR_UP_3         4.136507
## 4 120_202210_MSM_PPR_TR_1        7.480386
## 5 130_202210_MSM_PPR_TR_11       11.559035
## 6 20_202210_GCrew_WC_SipB_45cm    6.982359

#put it back together with the old data set and look for duplicates
QAspks <- merge(sampleddat, spks)
head(QAspks)

##                                     Sample_ID  S04_ppm S04_Area    Cl_ppm Cl_Area     S04_mM
## 1 10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338  0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1   31.3728  0.5216  671.7957 15.1336  0.9785652
## 3 110_202210_MSM_PPR_UP_3     70.1598  1.1665 1104.8402 24.8888  2.1883905
## 4 120_202210_MSM_PPR_TR_1    234.3596  3.8964 2538.4834 57.1846  7.3100312
## 5 130_202210_MSM_PPR_TR_11   439.7246  7.3107 3454.7595 77.8256 13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420  3.5063 5626.6706 63.3699 13.1578915
##                                     Cl_mM salinity S04_ugmL    Cl_ugmL S04_ugmL_spk
## 1    1.169275  0.0749276 0.5716443  0.8290452      3.313680
## 2   18.950513  1.2139608 0.6274618 13.4359223      4.069623
## 3   31.166155  1.9964722 1.4032532 22.0967913      4.136507
## 4   71.607430  4.5870655 4.6872241 50.7696727      7.480386
## 5   97.454429  6.2427764 8.7945036 69.0951807     11.559035
## 6  158.721315 10.1674198 4.2179479 56.2611103      6.982359

#now we need to calculate the spike concentration and calculate the spike recovery
spkconc <- (250)      # in ug
spkvolt <- 10          # in uL
spkvolt <- spkvolt/1000
#spike for these samples was 10uL of the 250mM standard
QAspks$S04_spk_Conc <- (spkconc)*spkvolt           # mmoles of S04
head(QAspks)

```

```

##                               Sample_ID  S04_ppm S04_Area      Cl_ppm Cl_Area      S04_mM
## 1 10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338  0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1   31.3728  0.5216  671.7957 15.1336  0.9785652
## 3 110_202210_MSM_PPR_UP_3     70.1598  1.1665 1104.8402 24.8888  2.1883905
## 4 120_202210_MSM_PPR_TR_1    234.3596  3.8964 2538.4834 57.1846  7.3100312
## 5 130_202210_MSM_PPR_TR_11   439.7246  7.3107 3454.7595 77.8256 13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420  3.5063 5626.6706 63.3699 13.1578915
##                               Cl_mM salinity S04_ugmL Cl_ugmL S04_ugmL_spk S04_spk_Conc
## 1    1.169275  0.0749276 0.5716443  0.8290452  3.313680        2.5
## 2   18.950513  1.2139608 0.6274618 13.4359223  4.069623        2.5
## 3   31.166155  1.9964722 1.4032532 22.0967913  4.136507        2.5
## 4   71.607430  4.5870655 4.6872241 50.7696727  7.480386        2.5
## 5   97.454429  6.2427764 8.7945036 69.0951807 11.559035        2.5
## 6 158.721315 10.1674198 4.2179479 56.2611103  6.982359        2.5

#need to determine dilution factors and initial amount of sample added
#if your samples are all the same dilution just use the first line
#for Steph / COMPASS this depends on the site so...
#QAspks$Dilution <- 1
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 50, QAspks$Dilution)
##$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 50, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 100, QAspks$Dilution)
##$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 50, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 50, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 100, QAspks$Dilution)
##$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 200, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$Dilution)

#Set Sample volumes in uL
QAspks$SampleVol <- 1
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_PPR_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_PPR_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_PPR_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 1462, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$SampleVol)

#change sample volume to L
QAspks$SampleVol <- QAspks$SampleVol/1000
head(QAspks)

```

```

##                               Sample_ID  SO4_ppm SO4_Area      Cl_ppm Cl_Area      SO4_mM
## 1 10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338  0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1   31.3728  0.5216  671.7957 15.1336  0.9785652
## 3 110_202210_MSM_PPR_UP_3     70.1598  1.1665 1104.8402 24.8888  2.1883905
## 4 120_202210_MSM_PPR_TR_1    234.3596  3.8964 2538.4834 57.1846  7.3100312
## 5 130_202210_MSM_PPR_TR_11   439.7246  7.3107 3454.7595 77.8256  13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420  3.5063 5626.6706 63.3699  13.1578915
##                               Cl_mM  salinity SO4_ugmL Cl_ugmL SO4_ugmL_spk SO4_spk_Conc
## 1    1.169275  0.0749276 0.5716443  0.8290452  3.313680          2.5
## 2   18.950513  1.2139608 0.6274618 13.4359223  4.069623          2.5
## 3   31.166155  1.9964722 1.4032532 22.0967913  4.136507          2.5
## 4   71.607430  4.5870655 4.6872241 50.7696727  7.480386          2.5
## 5   97.454429  6.2427764 8.7945036 69.0951807 11.559035          2.5
## 6  158.721315 10.1674198 4.2179479 56.2611103  6.982359          2.5
##   SampleVol
## 1    1.501
## 2    1.501
## 3    1.501
## 4    1.501
## 5    1.501
## 6    1.475

#gives us the total SO4 in the sample in mmoles
QAspks$SO4_Total_unspkd <- QAspks$SO4_ugmL*QAspks$SampleVol

##total SO4 in spiked sample in mmoles
QAspks$SO4_Total_spkd <- (QAspks$SO4_ugmL_spk)*(QAspks$SampleVol+spkvol)

QAspks$SO4_expctd_spkd <- (QAspks$SO4_Total_unspkd + QAspks$SO4_spk_Conc)
QAspks$spk_recovery <- (QAspks$SO4_Total_spkd/QAspks$SO4_expctd_spkd)*100
QAspks$SO4_spks_flag <- ifelse(QAspks$spk_recovery <=120 & QAspks$spk_recovery >=80 , 'YES' , 'NO', rerun)

head(QAspks)

##                               Sample_ID  SO4_ppm SO4_Area      Cl_ppm Cl_Area      SO4_mM
## 1 10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338  0.8915721
## 2 100_202210_MSM_UP_RHZ_SF_1   31.3728  0.5216  671.7957 15.1336  0.9785652
## 3 110_202210_MSM_PPR_UP_3     70.1598  1.1665 1104.8402 24.8888  2.1883905
## 4 120_202210_MSM_PPR_TR_1    234.3596  3.8964 2538.4834 57.1846  7.3100312
## 5 130_202210_MSM_PPR_TR_11   439.7246  7.3107 3454.7595 77.8256  13.7156769
## 6 20_202210_GCrew_WC_SipB_45cm 421.8420  3.5063 5626.6706 63.3699  13.1578915
##                               Cl_mM  salinity SO4_ugmL Cl_ugmL SO4_ugmL_spk SO4_spk_Conc
## 1    1.169275  0.0749276 0.5716443  0.8290452  3.313680          2.5
## 2   18.950513  1.2139608 0.6274618 13.4359223  4.069623          2.5
## 3   31.166155  1.9964722 1.4032532 22.0967913  4.136507          2.5
## 4   71.607430  4.5870655 4.6872241 50.7696727  7.480386          2.5
## 5   97.454429  6.2427764 8.7945036 69.0951807 11.559035          2.5
## 6  158.721315 10.1674198 4.2179479 56.2611103  6.982359          2.5
##   SampleVol SO4_Total_unspkd SO4_Total_spkd SO4_expctd_spkd spk_recovery
## 1    1.501        0.8580380      5.006971       3.358038  149.1040
## 2    1.501        0.9418201      6.149200       3.441820  178.6613
## 3    1.501        2.1062830      6.250263       4.606283  135.6899
## 4    1.501        7.0355234     11.302863       9.535523  118.5343
## 5    1.501       13.2005499     17.465702      15.700550  111.2426

```

```

## 6      1.475      6.2214732     10.368803     8.721473     118.8882
## S04_spks_flag
## 1      NO, rerun
## 2      NO, rerun
## 3      NO, rerun
## 4      YES
## 5      YES
## 6      YES

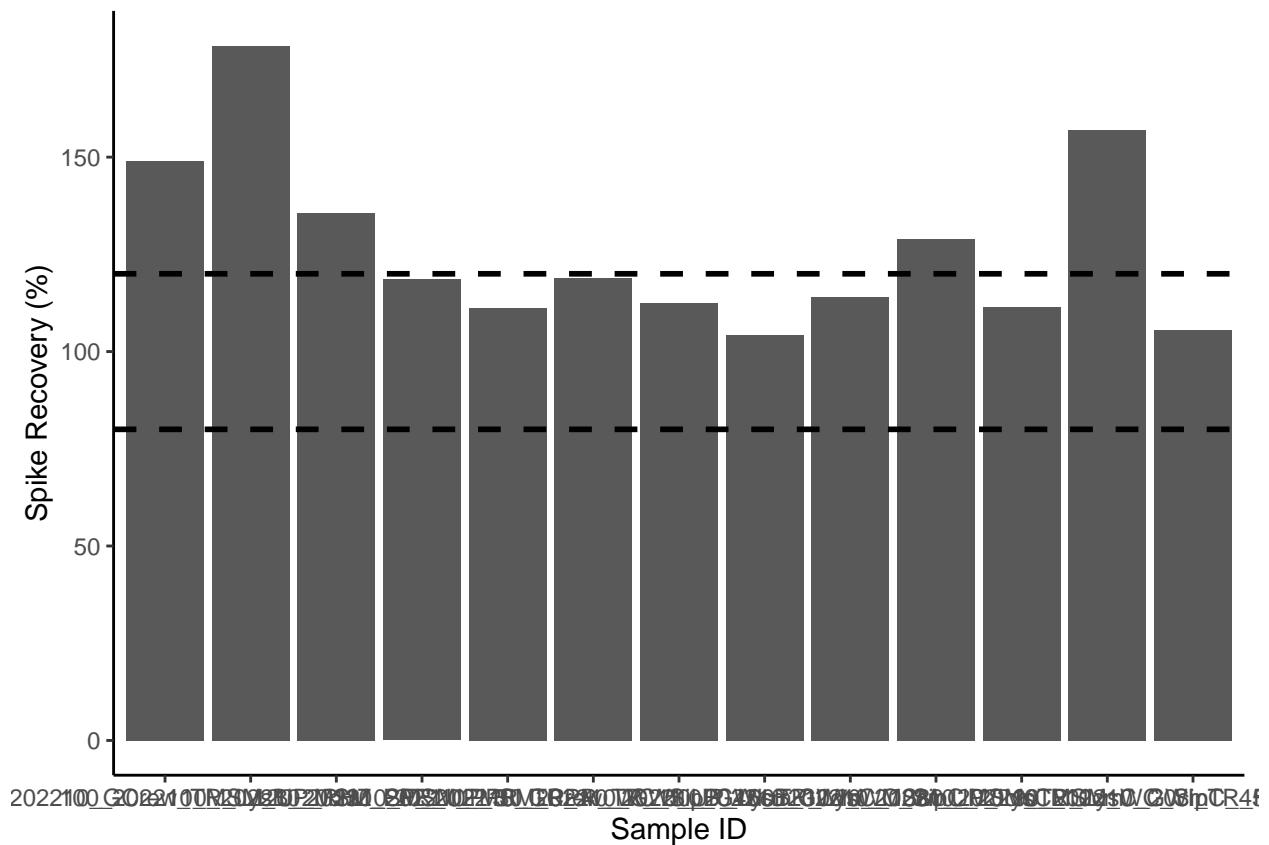
```

```

#plot spk recoveries output as a bar graph to easily check - want any over 10% to be red need to work on
spksbar <- ggplot(data = QAspks, aes(x = Sample_ID, y = spk_recovery)) +
  geom_bar(stat = 'identity') +
  scale_fill_manual(values=c("lightblue4")) +
  theme_classic() + labs(x= "Sample ID", y="Spike Recovery (%)") +
  theme(legend.position="none") +
  geom_hline(yintercept=80, linetype="dashed", color = "black", size=1) +
  geom_hline(yintercept=120, linetype="dashed", color = "black", size=1)

```

```
spksbar
```



```
Perc_spks$Percent <- (Perc_spks$no_rows / Perc_spks$Total)*100
head(Perc_spks)
```

```
## # A tibble: 2 x 4
##   S04_spks_flag no_rows Total Percent
##   <chr>          <int>  <dbl>
## 1 NO, rerun      5     13    38.5
## 2 YES            8     13    61.5
```

Make final dataframe with IDs

```
#for steph <- pull out identifiers of the sample names
#pull the sample ID and separate it by the underscores
IDs <- data.frame(do.call('rbind', strsplit(as.character(sampledat2$Sample_ID), '_', fixed=TRUE)))

## Warning in rbind(c("1", "202210", "Gcrew", "UP", "LysA", "20cm"), c("10", :
## number of columns of result is not a multiple of vector length (arg 1)

colnames(IDs) <- c("Analysis_No" , "Date", "Site", "Zone", "Replicate", "Depth", "Tree")
head(IDs)
```

```
##   Analysis_No Date Site Zone Replicate Depth Tree
## 1           1 202210 Gcrew  UP     LysA 20cm   1
## 2          10 202210 GCrew  TR     LysB 20cm  10
## 3         100 202210   MSM  UP     RHZ   SF   1
## 4         101 202210   MSM  UP     RHZ   SF   2
## 5         102 202210   MSM  UP     RHZ   SF   3
## 6         103 202210   MSM  UP     RHZ   SF   4
```

```
#rejoin them to the dataframe
alldat <- cbind(IDs, sampledat2)
head(alldat)
```

```
##   Analysis_No Date Site Zone Replicate Depth Tree
## 1           1 202210 Gcrew  UP     LysA 20cm   1
## 2          10 202210 GCrew  TR     LysB 20cm  10
## 3         100 202210   MSM  UP     RHZ   SF   1
## 4         101 202210   MSM  UP     RHZ   SF   2
## 5         102 202210   MSM  UP     RHZ   SF   3
## 6         103 202210   MSM  UP     RHZ   SF   4
##                               Sample_ID S04_ppm S04_Area   Cl_ppm Cl_Area   S04_mM
## 1 1_202210_Gcrew_UP_LysA_20cm  4.1491  0.0690  14.4387  0.3253 0.1294167
## 2 10_202210_GCrew_TR_LysB_20cm 28.5838  0.4752  41.4508  0.9338 0.8915721
## 3 100_202210_MSM_UP_RHZ_SF_1  31.3728  0.5216  671.7957 15.1336 0.9785652
## 4 101_202210_MSM_UP_RHZ_SF_2  65.5675  1.0901  933.5416 21.0300 2.0451497
## 5 102_202210_MSM_UP_RHZ_SF_3 105.7117  1.7575 1230.2357 27.7136 3.2973082
## 6 103_202210_MSM_UP_RHZ_SF_4  54.7246  0.9098 1060.9960 23.9011 1.7069432
##                               Cl_mM salinity
## 1  0.4072976 0.02611673
```

```
## 2 1.1692750 0.07492760
## 3 18.9505134 1.21396083
## 4 26.3340367 1.68693567
## 5 34.7034048 2.22306191
## 6 29.9293653 1.91724577
```

Make final dataframe with IDs

```
write.csv(alldat, file="Processed Data/COMPASS_SynopticCB_PW_Processed_C1_S04_202210.csv") #Change file
```

END