

# Dionex\_COMPASS\_September2022

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2023-01-23

## Daily Set up

Read in and Format the raw data - change wd & file names

```
# SULFATE DATA:

## Read in raw data file from Dionex - copied and saved as a txt
Sdat <- read.table("Raw Data/COMPASS_Synoptic_CB_MonMon_202209_S04.txt",sep='\t' , header=T, skip=3)
head(Sdat)

##      X          X.1          X.2 IC.S04 IC.S04.1 IC.S04.2 IC.S04.3 IC.S04.4
## 1 1 Lab Blank Unknown 4.413 0.5525    6.21 0.4551    2.87
## 2 2 Lab Blank Unknown 4.417 0.5520    6.21 0.4546    2.87
## 3 3 Lab Blank Unknown n.a.     n.a.     n.a.     n.a.     n.a.
## 4 4 Lab Blank Unknown n.a.     n.a.     n.a.     n.a.     n.a.
## 5 5 Lab Blank Unknown 4.423 0.5525    6.20 0.4550    2.87
## 6 6 Standard 1 Calibration Standard 4.427 1.1273    6.27 0.9285    5.72

## Only keep the columns that we need
Sdat <- Sdat[,c(2,5)] # dont need this here
head(Sdat)

##          X.1 IC.S04.1
## 1 Lab Blank 0.5525
## 2 Lab Blank 0.5520
## 3 Lab Blank   n.a.
## 4 Lab Blank   n.a.
## 5 Lab Blank 0.5525
## 6 Standard 1 1.1273

## Name the columns correctly
colnames(Sdat) <- c( "Sample_ID", "S04_ppm")
Sdat$Sample_ID <- as.factor(Sdat$Sample_ID)
Sdat$S04_ppm <- as.numeric(Sdat$S04_ppm)

## Warning: NAs introduced by coercion
```

```

Sdat <- as.data.frame(Sdat)
head(Sdat)

##      Sample_ID S04_ppm
## 1 Lab Blank  0.5525
## 2 Lab Blank  0.5520
## 3 Lab Blank     NA
## 4 Lab Blank     NA
## 5 Lab Blank  0.5525
## 6 Standard 1  1.1273

#Chloride data
## Read in raw data file from Dionex - copied and saved as a txt
Cldat <- read.table("Raw Data/COMPASS_Synoptic_CB_MonMon_202209_Cl.txt",sep='\t' , header=T, skip=3)
head(Cldat)

##      X          X.1          X.2 IC.Cl IC.Cl.1 IC.Cl.2 IC.Cl.3 IC.Cl.4
## 1 1 Lab Blank Unknown 3.293   6.0694  93.08  6.8202  47.25
## 2 2 Lab Blank Unknown 3.297   6.0646  93.06  6.8149  47.26
## 3 3 Lab Blank    n.a.    n.a.    n.a.    n.a.    n.a.
## 4 4 Lab Blank Unknown 3.290   0.0090 100.00  0.0101  0.07
## 5 5 Lab Blank Unknown 3.300   6.0747  93.07  6.8262  47.33
## 6 6 Standard 1 Calibration Standard 3.300  12.2365  92.93 13.7503  94.39

## Only keep the columns that we need
Cldat <- Cldat[,c(2,5)]
head(Cldat)

##          X.1 IC.Cl.1
## 1 Lab Blank 6.0694
## 2 Lab Blank 6.0646
## 3 Lab Blank    n.a.
## 4 Lab Blank 0.0090
## 5 Lab Blank 6.0747
## 6 Standard 1 12.2365

## Name the columns correctly
colnames(Cldat) <- c( "Sample_ID", "Cl_ppm")
Cldat$Sample_ID <- as.factor(Cldat$Sample_ID)
Cldat$Cl_ppm <- as.numeric(Cldat$Cl_ppm)

## Warning: NAs introduced by coercion

Cldat <- as.data.frame(Cldat)
head(Cldat)

##      Sample_ID Cl_ppm
## 1 Lab Blank 6.0694
## 2 Lab Blank 6.0646
## 3 Lab Blank     NA
## 4 Lab Blank 0.0090
## 5 Lab Blank 6.0747
## 6 Standard 1 12.2365

```

```

## Bring the data back together:
all_dat <- merge(Sdat, Cldat, by="Sample_ID")
head(all_dat)

##   Sample_ID   S04_ppm   Cl_ppm
## 1           2568.7298 19169.228
## 2           2568.7298  4914.293
## 3           2568.7298     0.009
## 4           2568.7298  4649.136
## 5           2568.7298      NA
## 6           607.8371 19169.228

## Remove empty lines
all_dat <- all_dat[!(is.na(all_dat$Sample_ID) | all_dat$Sample_ID=="") , ]
head(all_dat)

##                               Sample_ID   S04_ppm   Cl_ppm
## 26      1_202209_MSM_UP_LysA_20cm 185.4786 2155.484
## 27      10_202209_MSM_TR_LysA_20cm 505.3590 3550.881
## 28 10_202209_MSM_TR_LysA_20cm_spk 627.6741 3531.850
## 29      100_202209_MSM_WC_RHZ_6  794.4041 8421.266
## 30     100_202209_MSM_WC_RHZ_6_spk 1072.7880 8331.226
## 31      101_202209_MSM_WC_RHZ_7  815.7621 8675.486

all_dat <- droplevels(all_dat[!all_dat$Sample_ID == 'Lab Blank',])
head(all_dat)

##                               Sample_ID   S04_ppm   Cl_ppm
## 26      1_202209_MSM_UP_LysA_20cm 185.4786 2155.484
## 27      10_202209_MSM_TR_LysA_20cm 505.3590 3550.881
## 28 10_202209_MSM_TR_LysA_20cm_spk 627.6741 3531.850
## 29      100_202209_MSM_WC_RHZ_6  794.4041 8421.266
## 30     100_202209_MSM_WC_RHZ_6_spk 1072.7880 8331.226
## 31      101_202209_MSM_WC_RHZ_7  815.7621 8675.486

```

Pull out standards - could do some checks here if we want

```

stds <- all_dat[grep("Standard", all_dat$Sample_ID),]
head(stds)

##   Sample_ID S04_ppm   Cl_ppm
## 1128 Standard 1  1.1273 12.2365
## 1129 Standard 1  1.1273 12.3306
## 1130 Standard 1  1.1273 12.2419
## 1131 Standard 1  1.1273  6.0938
## 1132 Standard 1  1.1501 12.2365
## 1133 Standard 1  1.1501 12.3306

```

```

stds_chk_S <- stds %>%
  group_by(Sample_ID) %>%
  summarise(mean = mean(SO4_ppm), sd = sd(SO4_ppm))
stds_chk_S$cv <- (stds_chk_S$sd/stds_chk_S$mean)*100
stds_chk_S$flag <- ifelse(stds_chk_S$cv <2, 'YES', 'NO, rerun')
head(stds_chk_S)

```

```

## # A tibble: 5 x 5
##   Sample_ID     mean     sd     cv flag
##   <fct>      <dbl>   <dbl>   <dbl> <chr>
## 1 Standard 1  0.991  0.258  26.1 NO, rerun
## 2 Standard 2  1.89   0.562  29.8 NO, rerun
## 3 Standard 3  7.70   4.08   53.0 NO, rerun
## 4 Standard 4 16.8    4.79   28.6 NO, rerun
## 5 Standard 5 NA     NA     NA     <NA>

```

```

stds_chk_Cl <- stds %>%
  group_by(Sample_ID) %>%
  summarise(mean = mean(Cl_ppm), sd = sd(Cl_ppm))
stds_chk_Cl$cv <- (stds_chk_Cl$sd/stds_chk_Cl$mean)*100
stds_chk_Cl$flag <- ifelse(stds_chk_Cl$cv <2, 'YES', 'NO, rerun')
head(stds_chk_Cl)

```

```

## # A tibble: 5 x 5
##   Sample_ID     mean     sd     cv flag
##   <fct>      <dbl>   <dbl>   <dbl> <chr>
## 1 Standard 1  10.7   2.76  25.8 NO, rerun
## 2 Standard 2  19.8   5.62  28.4 NO, rerun
## 3 Standard 3  77.8  40.7  52.3 NO, rerun
## 4 Standard 4 168.   47.4  28.3 NO, rerun
## 5 Standard 5 NA     NA     NA     <NA>

```

## Calculate mmol/L concentrations

```

#remove standards from sample dataframe
sampledat <- all_dat[!grepl("Standard", all_dat$Sample_ID),]
head(sampledat)

```

```

##                               Sample_ID     SO4_ppm     Cl_ppm
## 26      1_202209_MSM_UP_LysA_20cm 185.4786 2155.484
## 27      10_202209_MSM_TR_LysA_20cm 505.3590 3550.881
## 28 10_202209_MSM_TR_LysA_20cm_spk 627.6741 3531.850
## 29      100_202209_MSM_WC_RHZ_6  794.4041 8421.266
## 30      100_202209_MSM_WC_RHZ_6_spk 1072.7880 8331.226
## 31      101_202209_MSM_WC_RHZ_7  815.7621 8675.486

```

```

# Constants needed for calculations:
clmw <- 35.45      #molecular weight of Chloride: 35.45
smw <- 32.06       #molecular weight of sulfur: 32.06

```

```

# Convert ppm to mmol/L
sampledat$SO4_mM <- (sampledat$SO4_ppm / smw)
sampledat$Cl_mM <- (sampledat$Cl_ppm / clmw)

# Calculate Salinity
# calculated using the Knudsen equation
# Salinity = 0.03 + 1.8050 * Chlorinity
# Ref: A Practical Handbook of Seawater Analysis by Strickland & Parsons (P. 11)
# =((1.807*Cl_ppm)+0.026)/1000
sampledat$salinity <- ((1.807 * sampledat$Cl_ppm) + 0.026) / 1000

head(sampledat)

##                               Sample_ID   SO4_ppm   Cl_ppm   SO4_mM   Cl_mM
## 26      1_202209_MSM_UP_LysA_20cm 185.4786 2155.484  5.785359 60.80349
## 27      10_202209_MSM_TR_LysA_20cm 505.3590 3550.881 15.762913 100.16590
## 28     10_202209_MSM_TR_LysA_20cm_spk 627.6741 3531.850 19.578107 99.62906
## 29      100_202209_MSM_WC_RHZ_6    794.4041 8421.266 24.778668 237.55333
## 30     100_202209_MSM_WC_RHZ_6_spk 1072.7880 8331.226 33.461884 235.01341
## 31      101_202209_MSM_WC_RHZ_7    815.7621 8675.486 25.444857 244.72456
##   salinity
## 26  3.894985
## 27  6.416469
## 28  6.382079
## 29 15.217253
## 30 15.054550
## 31 15.676629

```

## Pull out dups and check with percent difference

```

#Show me the data that we have from the calculations
head(sampledat)

```

```

##                               Sample_ID   SO4_ppm   Cl_ppm   SO4_mM   Cl_mM
## 26      1_202209_MSM_UP_LysA_20cm 185.4786 2155.484  5.785359 60.80349
## 27      10_202209_MSM_TR_LysA_20cm 505.3590 3550.881 15.762913 100.16590
## 28     10_202209_MSM_TR_LysA_20cm_spk 627.6741 3531.850 19.578107 99.62906
## 29      100_202209_MSM_WC_RHZ_6    794.4041 8421.266 24.778668 237.55333
## 30     100_202209_MSM_WC_RHZ_6_spk 1072.7880 8331.226 33.461884 235.01341
## 31      101_202209_MSM_WC_RHZ_7    815.7621 8675.486 25.444857 244.72456
##   salinity
## 26  3.894985
## 27  6.416469
## 28  6.382079
## 29 15.217253
## 30 15.054550
## 31 15.676629

```

```
#pull out any rows that have "dup" in the SampleID column
dups <- sampledat %>%
  filter(str_detect(Sample_ID, "dup"))      #have to change this to match data
head(dups)
```

```
##                               Sample_ID   SO4_ppm    Cl_ppm    SO4_mM    Cl_mM
## 1  109_202209_GWI_TR_RHZ_SF_7_dup  933.3051 6544.394 29.1112009 184.6091312
## 2  19_202209_MSM_WC_SipA_45cm_dup  51.6200 8507.394 1.6101061 239.9828999
## 3  29_202209_GWI_UP_LysA_20cm_dup  772.0785 6490.409 24.0822988 183.0862934
## 4  39_202209_GWI_TR_LysA_45cm_dup  701.8766 8857.495 21.8925951 249.8588124
## 5  49_202209_GWI_WC_SipB_10cm_dup 1907.2701 13781.568 59.4906457 388.7607419
## 6  59_202209_GCrew_TR_LysA_20cm_dup  10.1312    3.942  0.3160075  0.1111989
##           salinity
## 1 11.825745416
## 2 15.372886597
## 3 11.728195244
## 4 16.005519284
## 5 24.903319918
## 6 0.007149194
```

```
#remove these from sample dataframe in a new dataframe
sampledat2 <- sampledat %>%
  filter(!str_detect(Sample_ID, "dup")) %>%
  filter(!str_detect(Sample_ID, "spk"))
head(sampledat2)
```

```
##                               Sample_ID   SO4_ppm    Cl_ppm    SO4_mM    Cl_mM  salinity
## 1  1_202209_MSM_UP_LysA_20cm 185.4786 2155.484  5.785359 60.80349 3.894985
## 2  10_202209_MSM_TR_LysA_20cm 505.3590 3550.881 15.762913 100.16590 6.416469
## 3   100_202209_MSM_WC_RHZ_6 794.4041 8421.266 24.778668 237.55333 15.217253
## 4   101_202209_MSM_WC_RHZ_7 815.7621 8675.486 25.444857 244.72456 15.676629
## 5   102_202209_MSM_WC_RHZ_8 872.0692 8465.177 27.201160 238.79201 15.296600
## 6 103_202209_GWI_TR_RHZ_SF_1 951.8538 6184.023 29.689763 174.44353 11.174556
```

```
#remove the dup from these IDs so we will have duplicate sample names
dups$Sample_ID<-gsub("_dup","",as.character(dups$Sample_ID))
dups <- dups[, -c(2,3)]
colnames(dups) <- c('Sample_ID', 'SO4_mM_dup', "Cl_mM_dup", "salinity_dup")
head(dups)
```

```
##                               Sample_ID SO4_mM_dup    Cl_mM_dup salinity_dup
## 1  109_202209_GWI_TR_RHZ_SF_7 29.1112009 184.6091312 11.825745416
## 2  19_202209_MSM_WC_SipA_45cm 1.6101061 239.9828999 15.372886597
## 3  29_202209_GWI_UP_LysA_20cm 24.0822988 183.0862934 11.728195244
## 4  39_202209_GWI_TR_LysA_45cm 21.8925951 249.8588124 16.005519284
## 5  49_202209_GWI_WC_SipB_10cm 59.4906457 388.7607419 24.903319918
## 6  59_202209_GCrew_TR_LysA_20cm 0.3160075  0.1111989  0.007149194
```

```
#put it back together with the old data set and look for duplicates
QAdups <- merge(sampledat2, dups)
head(QAdups)
```

```

##                               Sample_ID   SO4_ppm     Cl_ppm     SO4_mM     Cl_mM
## 1  109_202209_GWI_TR_RHZ_SF_7  927.5268  6420.4224 28.9309669 181.1120564
## 2  19_202209_MSM_WC_SipA_45cm  52.8767  8548.8314  1.6493044 241.1518025
## 3  29_202209_GWI_UP_LysA_20cm  768.0581  6435.7454 23.9568964 181.5442990
## 4  39_202209_GWI_TR_LysA_45cm  689.8972  8687.1963 21.5189395 245.0549027
## 5  49_202209_GWI_WC_SipB_10cm 1925.9192 13905.9219 60.0723394 392.2686008
## 6 59_202209_GCrew_TR_LysA_20cm  10.4618    4.9391  0.3263194  0.1393258
##           salinity SO4_mM_dup   Cl_mM_dup salinity_dup
## 1 11.601729277 29.1112009 184.6091312 11.825745416
## 2 15.447764340  1.6101061 239.9828999 15.372886597
## 3 11.629417938 24.0822988 183.0862934 11.728195244
## 4 15.697789714 21.8925951 249.8588124 16.005519284
## 5 25.128026873 59.4906457 388.7607419 24.903319918
## 6 0.008950954  0.3160075  0.1111989  0.007149194

QAdups$SO4_dups_chk <- ((abs(QAdups$SO4_mM-QAdups$SO4_mM_dup))/((QAdups$SO4_mM+QAdups$SO4_mM_dup)/2))*100
QAdups$SO4_dups_flag <- ifelse(QAdups$SO4_dups_chk <10, 'YES', 'NO, rerun')

QAdups$Cl_dups_chk <- ((abs(QAdups$Cl_mM-QAdups$Cl_mM_dup))/((QAdups$Cl_mM+QAdups$Cl_mM_dup)/2))*100
QAdups$Cl_dups_flag <- ifelse(QAdups$Cl_dups_chk <10, 'YES', 'NO, rerun')

head(QAdups)

##                               Sample_ID   SO4_ppm     Cl_ppm     SO4_mM     Cl_mM
## 1  109_202209_GWI_TR_RHZ_SF_7  927.5268  6420.4224 28.9309669 181.1120564
## 2  19_202209_MSM_WC_SipA_45cm  52.8767  8548.8314  1.6493044 241.1518025
## 3  29_202209_GWI_UP_LysA_20cm  768.0581  6435.7454 23.9568964 181.5442990
## 4  39_202209_GWI_TR_LysA_45cm  689.8972  8687.1963 21.5189395 245.0549027
## 5  49_202209_GWI_WC_SipB_10cm 1925.9192 13905.9219 60.0723394 392.2686008
## 6 59_202209_GCrew_TR_LysA_20cm  10.4618    4.9391  0.3263194  0.1393258
##           salinity SO4_mM_dup   Cl_mM_dup salinity_dup SO4_dups_chk SO4_dups_flag
## 1 11.601729277 29.1112009 184.6091312 11.825745416      0.6210448      YES
## 2 15.447764340  1.6101061 239.9828999 15.372886597      2.4052434      YES
## 3 11.629417938 24.0822988 183.0862934 11.728195244      0.5220836      YES
## 4 15.697789714 21.8925951 249.8588124 16.005519284      1.7214579      YES
## 5 25.128026873 59.4906457 388.7607419 24.903319918      0.9730331      YES
## 6 0.008950954  0.3160075  0.1111989  0.007149194      3.2107998      YES
##           Cl_dups_chk Cl_dups_flag
## 1      1.9124267      YES
## 2      0.4858941      YES
## 3      0.8457844      YES
## 4      1.9413120      YES
## 5      0.8982656      YES
## 6     22.4544257      NO, rerun

#plot dups output as a bar graph to easily check - want any over 10% to be red need to work on this
Sdupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = SO4_dups_chk, fill=SO4_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="Difference Between SO4 Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
                                             color = "black", size=1)

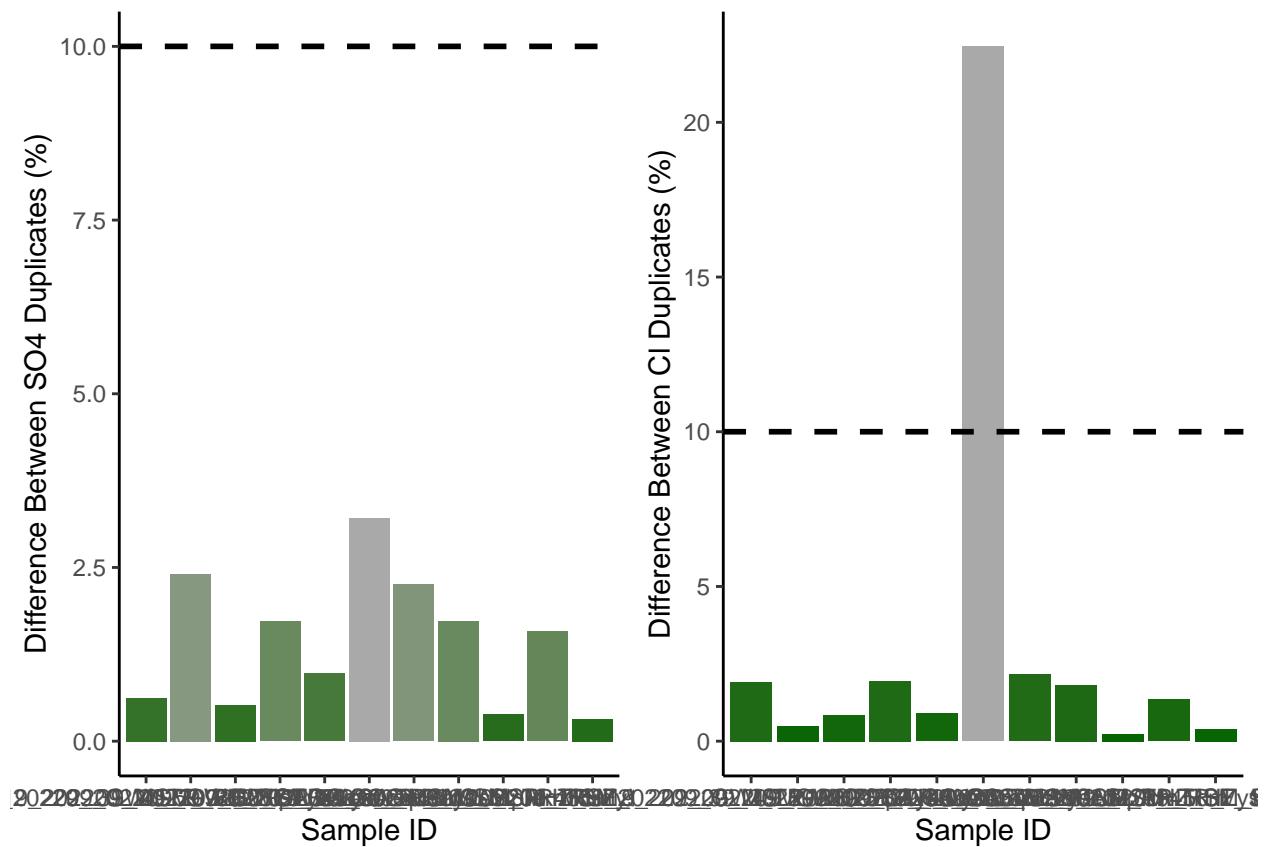
## Warning: Using 'size' aesthetic for lines was deprecated in ggplot2 3.4.0.

```

```
## i Please use 'linewidth' instead.
## This warning is displayed once every 8 hours.
## Call 'lifecycle::last_lifecycle_warnings()' to see where this warning was
## generated.

Cldupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = Cl_dups_chk, fill=Cl_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="Difference Between Cl Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
                                             color = "black", size=1)

ggarrange(Sdupsbar, Cldupsbar, ncol=2, nrow=1)
```



```
#check for percent of no, reruns to see if it would warrant reruns
Perc_dups <- QAdups %>%
  group_by(S04_dups_flag) %>%
  summarise(S_no_rows = length(S04_dups_flag))
Perc_dups1 <- QAdups %>%
  group_by(C1_dups_flag) %>%
  summarise(C1_no_rows = length(C1_dups_flag))
colnames(Perc_dups) <- c("Flag", "S_no_rows")
colnames(Perc_dups1) <- c("Flag", "C1_no_rows")
Perc_dups2 <- cbind(Perc_dups, Perc_dups1)
```

```

Perc_dups2$Total <- length(QAdups$S04_dups_flag)
Perc_dups2$S_Percent <- (Perc_dups2$S_no_rows / Perc_dups2$Total)*100
Perc_dups2$Cl_Percent <- (Perc_dups2$Cl_no_rows / Perc_dups2$Total)*100
head(Perc_dups2)

```

```

##   Flag S_no_rows      Flag Cl_no_rows Total S_Percent Cl_Percent
## 1  YES      11 NO, rerun      1     11    100  9.090909
## 2  YES      11      YES      10     11    100  90.909091

```

## Pull out dups and check with cv

```

#the cv
# calculate the sd bewteen the two duplicates then divide by the mean and multiply by 100

df2 <- as.data.frame(QAdups$S04_mM)
df2$dups <- QAdups$S04_mM_dup

df2$sds <- apply(df2,1,sd)

QAdups$S04_dups_cv <- (df2$sds)/((QAdups$S04_mM+QAdups$S04_mM_dup)/2) * 100
QAdups$S04_dups_cv_flag <- ifelse(QAdups$S04_dups_cv <11, 'YES', 'NO, rerun')

head(QAdups)

```

```

##                   Sample_ID    S04_ppm      Cl_ppm      S04_mM      Cl_mM
## 1 109_202209_GWI_TR_RHZ_SF_7 927.5268 6420.4224 28.9309669 181.1120564
## 2 19_202209_MSM_WC_SipA_45cm 52.8767 8548.8314 1.6493044 241.1518025
## 3 29_202209_GWI_UP_LysA_20cm 768.0581 6435.7454 23.9568964 181.5442990
## 4 39_202209_GWI_TR_LysA_45cm 689.8972 8687.1963 21.5189395 245.0549027
## 5 49_202209_GWI_WC_SipB_10cm 1925.9192 13905.9219 60.0723394 392.2686008
## 6 59_202209_GCrew_TR_LysA_20cm 10.4618 4.9391 0.3263194 0.1393258
##           salinity S04_mM_dup      Cl_mM_dup salinity_dup S04_dups_chk S04_dups_flag
## 1 11.601729277 29.1112009 184.6091312 11.825745416 0.6210448 YES
## 2 15.447764340 1.6101061 239.9828999 15.372886597 2.4052434 YES
## 3 11.629417938 24.0822988 183.0862934 11.728195244 0.5220836 YES
## 4 15.697789714 21.8925951 249.8588124 16.005519284 1.7214579 YES
## 5 25.128026873 59.4906457 388.7607419 24.903319918 0.9730331 YES
## 6 0.008950954 0.3160075 0.1111989 0.007149194 3.2107998 YES
##      Cl_dups_chk Cl_dups_flag S04_dups_cv S04_dups_cv_flag
## 1 1.9124267      YES 0.4391450      YES
## 2 0.4858941      YES 1.7007639      YES
## 3 0.8457844      YES 0.3691688      YES
## 4 1.9413120      YES 1.2172546      YES
## 5 0.8982656      YES 0.6880383      YES
## 6 22.4544257      NO, rerun 2.2703783      YES

```

```

#plot dups output as a bar graph to easily check - want any over 10% to be red need to work on this
Sdupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = S04_dups_cv, fill=S04_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +

```

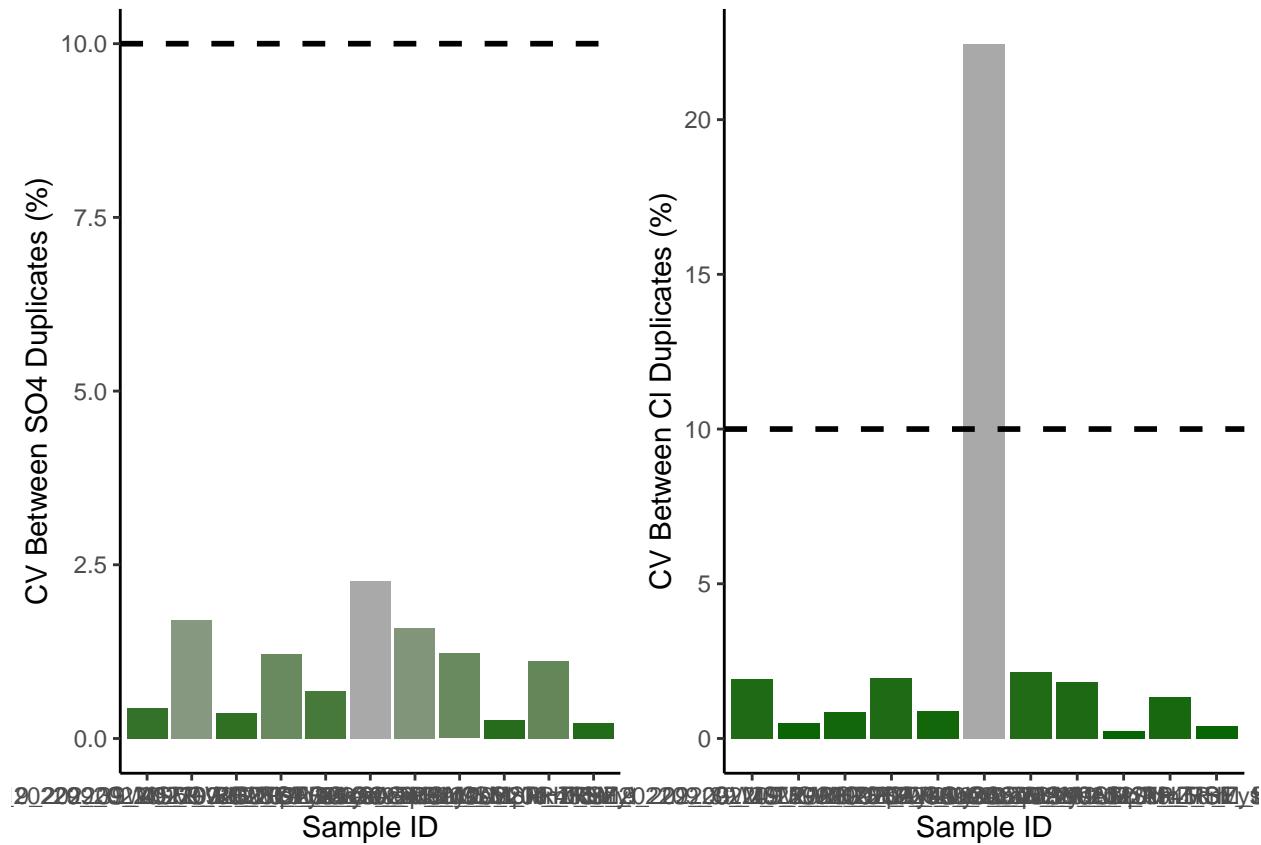
```

theme_classic() + labs(x = "Sample ID", y="CV Between SO4 Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
                                             color = "black", size=1)

Cldupsbar <- ggplot(data = QAdups, aes(x = Sample_ID, y = Cl_dups_chk, fill=Cl_dups_chk)) +
  geom_bar(stat = 'identity') +
  scale_fill_gradient2(low='darkgreen', mid='darkgreen', high='darkgrey', space='Lab') +
  theme_classic() + labs(x= "Sample ID", y="CV Between Cl Duplicates (%)") +
  theme(legend.position="none") + geom_hline(yintercept=10, linetype="dashed",
                                             color = "black", size=1)

ggarrange(Sdupsbar, Cldupsbar, ncol=2, nrow=1)

```



```

#check for percent of no, reruns to see if it would warrant reruns
Perc_dups <- QAdups %>%
  group_by(SO4_dups_flag) %>%
  summarise(S_no_rows = length(SO4_dups_flag))
Perc_dups1 <- QAdups %>%
  group_by(Cl_dups_flag) %>%
  summarise(Cl_no_rows = length(Cl_dups_flag))
colnames(Perc_dups) <- c("Flag", "S_no_rows")
colnames(Perc_dups1) <- c("Flag", "Cl_no_rows")
Perc_dups2 <- cbind(Perc_dups, Perc_dups1)

```

```

Perc_dups2$Total <- length(QAdups$S04_dups_flag)
Perc_dups2$S_Percent <- (Perc_dups2$S_no_rows / Perc_dups2$Total)*100
Perc_dups2$Cl_Percent <- (Perc_dups2$Cl_no_rows / Perc_dups2$Total)*100
head(Perc_dups2)

##   Flag S_no_rows      Flag Cl_no_rows Total S_Percent Cl_Percent
## 1  YES       11 NO, rerun      1     11    100  9.090909
## 2  YES       11      YES      10     11    100  90.909091

```

## Pull out spikes and check

```

#Show me the data that we have from the calculations
head(sampledat)

##                               Sample_ID   S04_ppm   Cl_ppm   S04_mM   Cl_mM
## 26      1_202209_MSM_UP_LysA_20cm 185.4786 2155.484  5.785359 60.80349
## 27      10_202209_MSM_TR_LysA_20cm 505.3590 3550.881 15.762913 100.16590
## 28 10_202209_MSM_TR_LysA_20cm_spk 627.6741 3531.850 19.578107 99.62906
## 29      100_202209_MSM_WC_RHZ_6 794.4041 8421.266 24.778668 237.55333
## 30     100_202209_MSM_WC_RHZ_6_spk 1072.7880 8331.226 33.461884 235.01341
## 31      101_202209_MSM_WC_RHZ_7 815.7621 8675.486 25.444857 244.72456
##   salinity
## 26 3.894985
## 27 6.416469
## 28 6.382079
## 29 15.217253
## 30 15.054550
## 31 15.676629

#pull out any rows that have "spk" in the SampleID column
spks <- sampledat %>%
  filter(str_detect(Sample_ID, "spk"))      #have to change this to match data
head(spks)

##                               Sample_ID   S04_ppm   Cl_ppm   S04_mM   Cl_mM
## 1 10_202209_MSM_TR_LysA_20cm_spk 627.6741 3531.850 19.57811 99.62906
## 2 100_202209_MSM_WC_RHZ_6_spk 1072.7880 8331.226 33.46188 235.01341
## 3 110_202209_GWI_TR_RHZ_SF_8_spk 793.6349 4702.940 24.75468 132.66403
## 4 20_202209_MSM_WC_SipB_10cm_spk 1122.1770 9663.288 35.00240 272.58922
## 5 30_202209_GWI_UP_LysA_20cm_spk 945.0486 5694.335 29.47750 160.63003
## 6 40_202209_GWI_TR_LysB_10cm_spk 1990.9739 12742.654 62.10149 359.45428
##   salinity
## 1 6.382079
## 2 15.054550
## 3 8.498238
## 4 17.461587
## 5 10.289689
## 6 23.026002

```

```

#remove the dup from these IDs so we will have duplicate sample names
spks$Sample_ID<-gsub("_spk","",as.character(spks$Sample_ID))
spks <- spks[ ,-c(2,3, 5,6)]
colnames(spks) <- c('Sample_ID', 'SO4_mM_spk')
head(spks)

##                               Sample_ID SO4_mM_spk
## 1 10_202209_MSM_TR_LysA_20cm   19.57811
## 2 100_202209_MSM_WC_RHZ_6    33.46188
## 3 110_202209_GWI_TR_RHZ_SF_8 24.75468
## 4 20_202209_MSM_WC_SipB_10cm 35.00240
## 5 30_202209_GWI_UP_LysA_20cm 29.47750
## 6 40_202209_GWI_TR_LysB_10cm 62.10149

#put it back together with the old data set and look for duplicates
QAspks <- merge(sampledat, spks)
head(QAspks)

##                               Sample_ID SO4_ppm      Cl_ppm      SO4_mM      Cl_mM salinity
## 1 10_202209_MSM_TR_LysA_20cm 505.3590 3550.881 15.76291 100.1659 6.416469
## 2 100_202209_MSM_WC_RHZ_6   794.4041 8421.266 24.77867 237.5533 15.217253
## 3 110_202209_GWI_TR_RHZ_SF_8 962.0158 6500.256 30.00673 183.3641 11.745989
## 4 20_202209_MSM_WC_SipB_10cm 860.1581 9655.185 26.82964 272.3606 17.446945
## 5 30_202209_GWI_UP_LysA_20cm 658.0800 5681.553 20.52651 160.2695 10.266592
## 6 40_202209_GWI_TR_LysB_10cm 1698.7295 12719.267 52.98595 358.7946 22.983742
##      SO4_mM_spk
## 1     19.57811
## 2     33.46188
## 3     24.75468
## 4     35.00240
## 5     29.47750
## 6     62.10149

#now we need to calculate the spike concentration and calculate the spike recovery
spkconc <- (250/smw)      # in mM
spkvol <- 10                # in uL
spkvol <- spkvol/1000000
#spike for these samples was 10uL of the 250mM standard
QAspks$SO4_spk_Conc <- (spkconc)*spkvol          # mmoles of SO4
head(QAspks)

##                               Sample_ID SO4_ppm      Cl_ppm      SO4_mM      Cl_mM salinity
## 1 10_202209_MSM_TR_LysA_20cm 505.3590 3550.881 15.76291 100.1659 6.416469
## 2 100_202209_MSM_WC_RHZ_6   794.4041 8421.266 24.77867 237.5533 15.217253
## 3 110_202209_GWI_TR_RHZ_SF_8 962.0158 6500.256 30.00673 183.3641 11.745989
## 4 20_202209_MSM_WC_SipB_10cm 860.1581 9655.185 26.82964 272.3606 17.446945
## 5 30_202209_GWI_UP_LysA_20cm 658.0800 5681.553 20.52651 160.2695 10.266592
## 6 40_202209_GWI_TR_LysB_10cm 1698.7295 12719.267 52.98595 358.7946 22.983742
##      SO4_mM_spk SO4_spk_Conc
## 1     19.57811 7.797879e-05
## 2     33.46188 7.797879e-05
## 3     24.75468 7.797879e-05

```

```

## 4 35.00240 7.797879e-05
## 5 29.47750 7.797879e-05
## 6 62.10149 7.797879e-05

#need to determine dilution factors and initial amount of sample added
#if your samples are all the same dilution just use the first line
#for Steph / COMPASS this depends on the site so...
QAspks$Dilution <- 1
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 50, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 100, QAspks$Dilution)
QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 200, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$Dilution)
#QAspks$Dilution <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$Dilution)

#Set Sample volumes in uL
QAspks$SampleVol <- 1
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "MSM_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_UP"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_TR"), 1501, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GCrew_WC"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_UP"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_TR"), 1475, QAspks$SampleVol)
QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "GWI_WC"), 1462, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_UP"), 100, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_TR"), 100, QAspks$SampleVol)
#QAspks$SampleVol <- ifelse(str_detect(QAspks$Sample_ID, "SWH_WC"), 100, QAspks$SampleVol)

#change sample volume to L
QAspks$SampleVol <- QAspks$SampleVol/1000000
head(QAspks)

##                               Sample_ID    SO4_ppm     Cl_ppm    SO4_mM     Cl_mM salinity
## 1 10_202209_MSM_TR_LysA_20cm 505.3590 3550.881 15.76291 100.1659 6.416469
## 2 100_202209_MSM_WC_RHZ_6   794.4041 8421.266 24.77867 237.5533 15.217253
## 3 110_202209_GWI_TR_RHZ_SF_8 962.0158 6500.256 30.00673 183.3641 11.745989
## 4 20_202209_MSM_WC_SipB_10cm 860.1581 9655.185 26.82964 272.3606 17.446945
## 5 30_202209_GWI_UP_LysA_20cm 658.0800 5681.553 20.52651 160.2695 10.266592
## 6 40_202209_GWI_TR_LysB_10cm 1698.7295 12719.267 52.98595 358.7946 22.983742
##      SO4_mM_spk SO4_spk_Conc Dilution SampleVol
## 1 19.57811 7.797879e-05       50 0.001501
## 2 33.46188 7.797879e-05      100 0.001475
## 3 24.75468 7.797879e-05      100 0.001475
## 4 35.00240 7.797879e-05      100 0.001475
## 5 29.47750 7.797879e-05      100 0.001475
## 6 62.10149 7.797879e-05      100 0.001475

```

```

#gives us the total SO4 in the sample in mmoles
QAspks$SO4_Total_unspkd <- (QAspks$SO4_mM/QAspks$Dilution)*(QAspks$SampleVol)

##total SO4 in spiked sample in mmoles
QAspks$SO4_Total_spkd <- (QAspks$SO4_mM_spk/QAspks$Dilution)*(QAspks$SampleVol+spkvol)

QAspks$SO4_expctd_spkd <- (QAspks$SO4_Total_unspkd + QAspks$SO4_spk_Conc)
QAspks$spk_recovery <- (QAspks$SO4_Total_spkd/QAspks$SO4_expctd_spkd)*100
QAspks$SO4_spks_flag <- ifelse(QAspks$spk_recovery <=120 & QAspks$spk_recovery >=80 , 'YES' , 'NO', rerun)

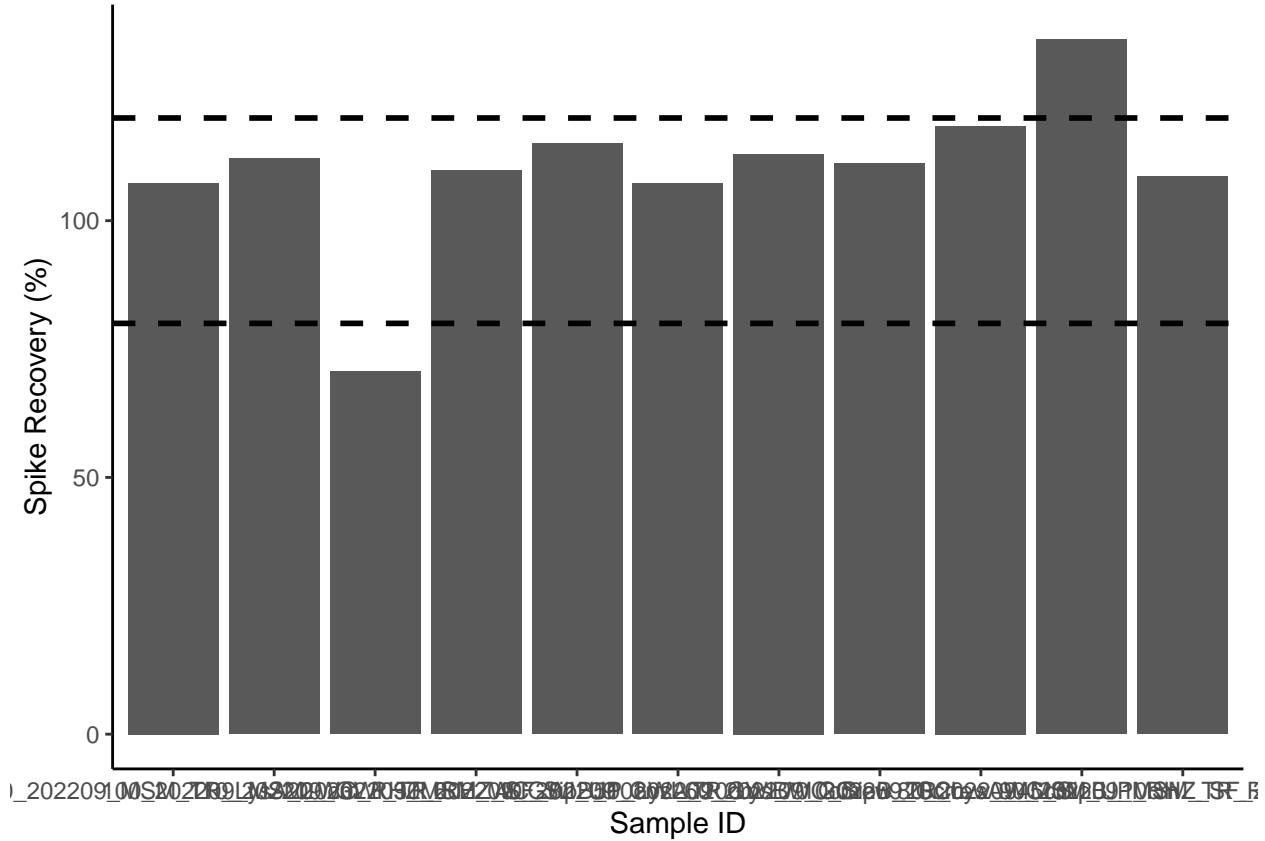
head(QAspks)

##          Sample_ID    SO4_ppm     Cl_ppm    SO4_mM     Cl_mM salinity
## 1 10_202209_MSM_TR_LysA_20cm 505.3590 3550.881 15.76291 100.1659 6.416469
## 2 100_202209_MSM_WC_RHZ_6   794.4041 8421.266 24.77867 237.5533 15.217253
## 3 110_202209_GWI_TR_RHZ_SF_8 962.0158 6500.256 30.00673 183.3641 11.745989
## 4 20_202209_MSM_WC_SipB_10cm 860.1581 9655.185 26.82964 272.3606 17.446945
## 5 30_202209_GWI_UP_LysA_20cm 658.0800 5681.553 20.52651 160.2695 10.266592
## 6 40_202209_GWI_TR_LysB_10cm 1698.7295 12719.267 52.98595 358.7946 22.983742
##   SO4_mM_spk SO4_spk_Conc Dilution SampleVol SO4_Total_unspkd SO4_Total_spkd
## 1 19.57811 7.797879e-05      50 0.001501 0.0004732027 0.0005916504
## 2 33.46188 7.797879e-05     100 0.001475 0.0003654854 0.0004969090
## 3 24.75468 7.797879e-05     100 0.001475 0.0004425993 0.0003676069
## 4 35.00240 7.797879e-05     100 0.001475 0.0003957371 0.0005197857
## 5 29.47750 7.797879e-05     100 0.001475 0.0003027661 0.0004377409
## 6 62.10149 7.797879e-05     100 0.001475 0.0007815427 0.0009222072
##   SO4_expctd_spkd spk_recovery SO4_spks_flag
## 1 0.0005511814    107.34222      YES
## 2 0.0004434641    112.05167      YES
## 3 0.0005205781    70.61514      NO, rerun
## 4 0.0004737159    109.72519      YES
## 5 0.0003807449    114.96960      YES
## 6 0.0008595215    107.29309      YES

#plot spk recoveries output as a bar graph to easily check - want any over 10% to be red need to work on this
spksbar <- ggplot(data = QAspks, aes(x = Sample_ID, y = spk_recovery)) +
  geom_bar(stat = 'identity') +
  scale_fill_manual(values=c("lightblue4")) +
  theme_classic() + labs(x= "Sample ID", y="Spike Recovery (%)") +
  theme(legend.position="none") +
  geom_hline(yintercept=80, linetype="dashed", color = "black", size=1) +
  geom_hline(yintercept=120, linetype="dashed", color = "black", size=1)

spksbar

```



```
#check for percent of no, reruns to see if it would warrant reruns
Perc_spks <- QAspks %>%
  group_by(S04_spks_flag) %>%
  summarise(no_rows = length(S04_spks_flag))
Perc_spks$Total <- length(QAspks$S04_spks_flag)
Perc_spks$Percent <- (Perc_spks$no_rows / Perc_spks$Total)*100
head(Perc_spks)
```

```
## # A tibble: 2 x 4
##   S04_spks_flag no_rows Total Percent
##   <chr>          <int> <int>   <dbl>
## 1 NO, rerun      2     11    18.2
## 2 YES            9     11    81.8
```

## Make final dataframe with IDs

```
#for steph <- pull out identifiers of the sample names
#pull the sample ID and separate it by the underscores
IDs <- data.frame(do.call('rbind', strsplit(as.character(sampledat2$Sample_ID), '_', fixed=TRUE)))

## Warning in rbind(c("1", "202209", "MSM", "UP", "LysA", "20cm"), c("10", :
## number of columns of result is not a multiple of vector length (arg 1)
```

```
colnames(IDs) <- c("Analysis_No" , "Date", "Site", "Zone", "Replicate", "Depth", "Tree")
head(IDs)
```

```
##   Analysis_No Date Site Zone Replicate Depth Tree
## 1           1 202209  MSM    UP      LysA  20cm    1
## 2          10 202209  MSM    TR      LysA  20cm   10
## 3         100 202209  MSM    WC      RHZ     6 100
## 4         101 202209  MSM    WC      RHZ     7 101
## 5         102 202209  MSM    WC      RHZ     8 102
## 6         103 202209  GWI    TR      RHZ     SF    1
```

```
#rejoin them to the dataframe
alldat <- cbind(IDs, sampledat2)
head(alldat)
```

```
##   Analysis_No Date Site Zone Replicate Depth Tree          Sample_ID
## 1           1 202209  MSM    UP      LysA  20cm    1 1_202209_MSM_UP_LysA_20cm
## 2          10 202209  MSM    TR      LysA  20cm   10 10_202209_MSM_TR_LysA_20cm
## 3         100 202209  MSM    WC      RHZ     6 100 100_202209_MSM_WC_RHZ_6
## 4         101 202209  MSM    WC      RHZ     7 101 101_202209_MSM_WC_RHZ_7
## 5         102 202209  MSM    WC      RHZ     8 102 102_202209_MSM_WC_RHZ_8
## 6         103 202209  GWI    TR      RHZ     SF    1 103_202209_GWI_TR_RHZ_SF_1
##   S04_ppm  Cl_ppm  S04_mM  Cl_mM salinity
## 1 185.4786 2155.484 5.785359 60.80349 3.894985
## 2 505.3590 3550.881 15.762913 100.16590 6.416469
## 3 794.4041 8421.266 24.778668 237.55333 15.217253
## 4 815.7621 8675.486 25.444857 244.72456 15.676629
## 5 872.0692 8465.177 27.201160 238.79201 15.296600
## 6 951.8538 6184.023 29.689763 174.44353 11.174556
```

## Make final dataframe with IDs

```
write.csv(alldat, file="Processed Data/COMPASS_SynopticCB_PW_Processed_Cl_S04_Final_202209.csv") #Change
```

END