Date: 2022-08-17

Tags: 1\_Wetlab 3\_Expression Created by: Stefanie Brands

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(Written by Stefanie Brands\_)

(\_Last update: 2022.08.18\_)

After cloning or receiving a new strain, flask expression is conducted with the aim to generate an active enzyme or folded protein.

{29.09.2017|Date of experiment}

Overnight precultures of {P. putida KT2440 pVLT33::pigC|expression strain} and the {empty vector strain (:as:)|:negative control:} are grown from a {single colony|inoculum} in {5|mL|:LB Kan:|growth media} at {30|°C|temperature} and {250|rpm|shaking} {overnight|time}.

{10.10.2017|Date of experiment}

Two {100|mL|:LB Kan:|expression media} cultures in {unbaffled Erlenmeyer|:flasks:} per strain were inoculated to an OD600 of {0.05|inoculation OD} with the overnight cultures.

Strain	OD600 preculture	V [mL] inoculation @OD600 = 0.05
P. putida KT2440 pVLT33::pigC	4.57	0.05 / 4.57 * 100 mL = 1.090 mL
P. putida KT2440 pVLT33 (EV)	7.92	0.05 / 7.92 * 100 mL = 0.631 mL

The cultures were incubated at  $\{30|^{\circ}C|\text{temperature}\}\$  and  $\{250|\text{rpm}|\text{shaking}\}\$  to an OD600 of  $\{0.4\text{-}0.6|\text{induction OD}\}\$  and induced by addition of  $\{0.5|\text{mM}|\text{:isopropyI-$\square$-$D$-thiogalactopyranoside}\$  (IPTG):|inducer molecule}.  $\{4|\text{h}|\text{:after induction:}|\text{time of substrate supplementation}\}\$ , the cultures were supplemented with  $\{0.5|\text{mM}|\text{:MAP}\ \text{and MBC:}|\text{substrates}\}\$  in order to confirm the presence of active PigC enzyme.

During and after induction, samples of the corresponding volume of  $\{1|mL|:OD = 1:|sample volume\}$  were taken at  $\{1|h|:intervals:\}$  for SDS-PAGE analysis. Samples were pelleted for  $\{1|min|centrifugation time\}$  at  $\{11,000|xg|centrifugation speed\}$  and stored at  $\{-20|^{\circ}C|storage temperature\}$ .

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**Table 1** - Growth of expression cultures.

Time [h]		P. putida KT2440 pVLT33::pigC		P. putida KT2440 pVLT33 (EV)		
Date	Time	t [h]	OD <sub>600</sub> (+ MAP/MBC)	OD <sub>600</sub> (- MAP/MBC)	OD <sub>600</sub> (+ MAP/MBC)	OD <sub>600</sub> (- MAP/MBC)
10.10.2017	08:35	0	0.037	0.040	0.033	0.030
	10:35	2	0.082	0.077	0.123	0.117
	11:35	3	0.121	0.132	0.228	0.206
	12:35	4	0.203	0.184	0.447 (t0) +IPTG	0.407 (t0) +IPTG
	13:35	5	0.360	0.359	0.93	0.93
	14:35	6	0.635 (t0) +IPTG	0.612 (t0) +IPTG	1.64 (t2)	1.70 (t2)
	15:35	7	1.13	1.12	2.10	2.00
	16:35	8	1.59 (t2)	1.63 (t2)	2.05 (t4) +MAP/MBC	2.14 (t4) +MAP/MBC
	17:35	9	2.19	2.44	2.83	2.90
	18:35	10	2.69 (t4) +MAP/MBC	2.79 (t4) +MAP/MBC	3.07	3.31
11.10.2017	10:35	26	5.45 (t16)	5.48	5.60	7.00
	12:35	28	5.40	5.67	5.60 (t20)	7.05
	14:35	30	5.62 (t20)	5.71	5.87	6.96

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12.10.2017	10:35	50	5.32	5.35	5.23	6.16
	12:35	52	nd	nd	nd (t44)	nd
	14:35	54	nd (t44)	nd	nd	nd

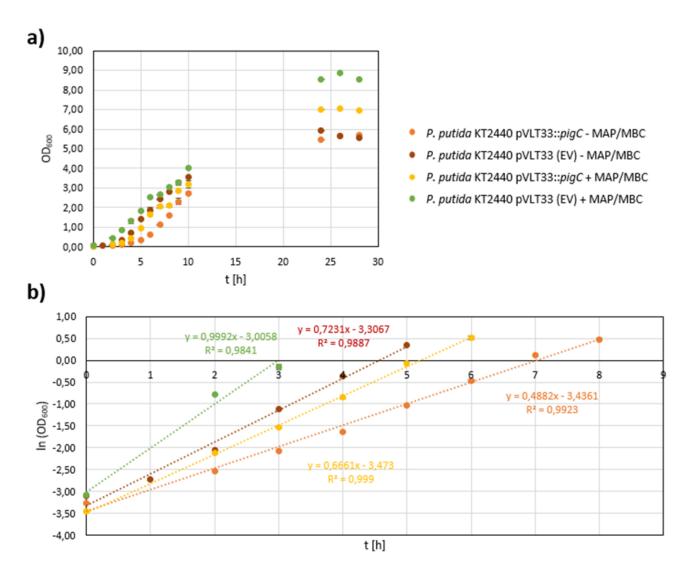
blue - sample for SDS-PAGE (1 mL OD1)

orange - sample for prodiginine analysis (1 mL)

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**Fig. 1: a.** Growth curves of P. putida KT2440 pVLT33::pigC and P. putida KT2440 pVLT33 (EV) with and without substrates (MAP/MBC). **b.** Growth rates of the cultures.

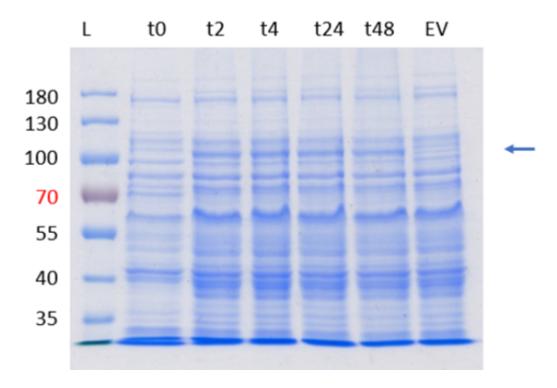
#### {25.10.2017|Date of experiment}

Samples from the expression cultures were thawn on ice and resuspended in  $\{30|\mu L|:PBS\}$  buffer:|resuspension volume}. After  $\{1|\min|:sonification:\}$  in an ultrasonic bath, resuspended samples were mixed with  $\{10|\mu L|:4x\ SDS\ loading\ dye:\}$ , incubated for  $\{5\ min\ at\ 95^{\circ}C|incubation\}$  and  $\{vortex(:ed:)\ thoroughly|process\}$  in order to shear genomic DNA. The boiled samples were spinned down for  $\{1\ min\ at\ maximal\ speed|centrifugation\}$  and  $\{5|\mu L|(:of:)\ :supernatant:\}$  was loaded on a  $\{10|\%|:SDS\ -PAGE\ separating\ gel:|acrylamide\ concentration\}$ .

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**Fig 2:** SDS-PAGE of pigC expression with P. putida KT2440 pVLT33::pigC. EV = empty vector control after 48 h. The arrow marks the PigC size of ~ 100 kDa.

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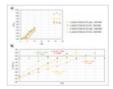
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### **Attached files**

### fig1.png

sha256: f2f568ee870efe0cabe8f66374ee50d82166a506ad26262519fefd772e5a893a



#### fig2.png

sha256: aaba59bc60e0a24c052fa549279e3c6c717cfea7d65af79fcb0ecfcdb1bfd4a4

