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4 CONTENTS

Prerequisites

This is a sample book written in Markdown. You can use anything that Pandoc's Markdown supports, e.g., a math equation $a^2 + b^2 = c^2$.

The **bookdown** package can be installed from CRAN or Github:

```
install.packages("bookdown")
# or the development version
# devtools::install_github("rstudio/bookdown")
```

Remember each Rmd file contains one and only one chapter, and a chapter is defined by the first-level heading #.

To compile this example to PDF, you need XeLaTeX. You are recommended to install TinyTeX (which includes XeLaTeX): https://yihui.name/tinytex/.

Introduction

This section will become the section for the administrative updates/organization once we have figured out how to use all of the bookdown features for our purposes.

You can label chapter and section titles using {#label} after them, e.g., we can reference Chapter 2. If you do not manually label them, there will be automatic labels anyway, e.g., Chapter 5.

Figures and tables with captions will be placed in figure and table environments, respectively.

```
par(mar = c(4, 4, .1, .1))
plot(pressure, type = 'b', pch = 19)
```

Reference a figure by its code chunk label with the fig: prefix, e.g., see Figure 2.1. Similarly, you can reference tables generated from knitr::kable(), e.g., see Table 2.1.

```
knitr::kable(
  head(iris, 20), caption = 'Here is a nice table!',
  booktabs = TRUE
)
```

You can write citations, too. For example, we are using the **bookdown** package (Xie, 2019) in this sample book, which was built on top of R Markdown and **knitr** (Xie, 2015).



Figure 2.1: Here is a nice figure!

Table 2.1: Here is a nice table!

Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
5.1	3.5	1.4	0.2	setosa
4.9	3.0	1.4	0.2	setosa
4.7	3.2	1.3	0.2	setosa
4.6	3.1	1.5	0.2	setosa
5.0	3.6	1.4	0.2	setosa
5.4	3.9	1.7	0.4	setosa
4.6	3.4	1.4	0.3	setosa
5.0	3.4	1.5	0.2	setosa
4.4	2.9	1.4	0.2	setosa
4.9	3.1	1.5	0.1	setosa
5.4	3.7	1.5	0.2	setosa
4.8	3.4	1.6	0.2	setosa
4.8	3.0	1.4	0.1	setosa
4.3	3.0	1.1	0.1	setosa
5.8	4.0	1.2	0.2	setosa
5.7	4.4	1.5	0.4	setosa
5.4	3.9	1.3	0.4	setosa
5.1	3.5	1.4	0.3	setosa
5.7	3.8	1.7	0.3	setosa
5.1	3.8	1.5	0.3	setosa

Project CC: Bullets and Cartridge Cases

For both bullets and cartridge cases we are dealing with several inter-related aspects, that we want to address independently.

Those are:

- 1. data collection
- 2. computational tools
- 3. similarity scores
 - 1. for bullet lands:
 - a. crosscut identification
 - b. groove location
 - c. curvature removal
 - d. alignment of signatures
 - e. feature extraction
 - f. matching with trained Random Forest
 - 2. for breech faces
- 4. analysis of results
- 5. communication of results and methods

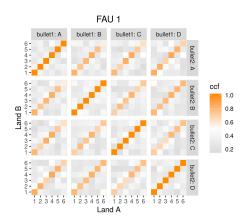


Figure 3.1: Results from assessing scans of barrel FAU 1 similarity.

3.1 Data Collection

3.1.1 LAPD

All bullets are collected by Srinivasan Rathinam, LAPD.

3.1.1.1 Main study

4 bullets per barrel for 626 Beretta $92~{\rm F/FS}$ firearms , ammunition used are 9 mm Luger Winchester 115 grain with a Copper surface.

scans are on Raven.

evaluation: Yawei is going to work through all 626 barrels of knowns to assess similarity scores

3.1.1.2 follow-up study

4 bullets per barrel for 96 of the original 626 Beretta firearms using different ammunition

bullets are being scanned

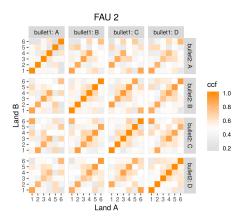


Figure 3.2: Results from assessing scans of barrel FAU 2 similarity.

3.1.2 Hamby Sets

Scans for Hamby Sets 10, 36, 44, and 224 Scans for 3 replicates of clones for Hamby 224

3.1.3 Houston Tests

contact: Melissa Nally, Houston FSI

3.1.3.1 Pre-study

3 kits with 23 bullets each evaluation included in submission to JFI

3.1.3.2 Study

4 kits with 20 bullets each scans done, evaluation finished, some scans of doubtful quality

Houston set 3

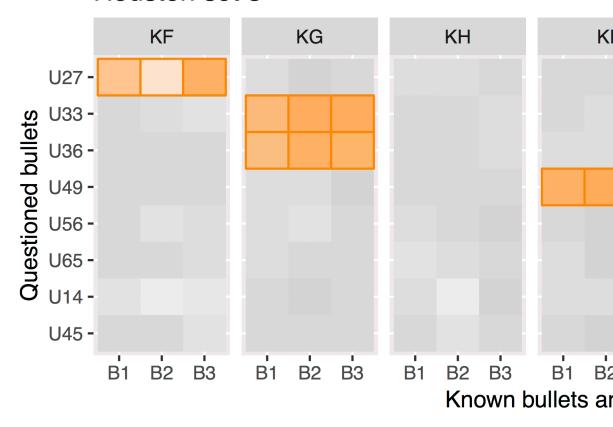


Figure 3.3: Bullet-to-bullet similarity scores for questioned bullets (y-axis) compared to all other bullets of the test set (x-axis).

3.1.4 Houston Persistence

contact: Melissa Nally, Houston FSI 8 barrels with 40 fired bullets each

3.1.5 St Louis persistence

contact: Steve Kramer, St Louis PD

2 barrels with 192 fired bullets each (2 bullets collected every 25 shots)

3.1.6 DFSC Cartridge cases

Breech face data for knowns are scanned and available on a private github repository

evaluation

3.2 Computational Tools

3.2.1 x3ptools

x3ptools is an R package for working with files in x3p format. x3p is an ISO standard for describing 3d topographic surface measurements. x3ptools is available on CRAN, i.e. can be installed with the command install.packages("x3ptools"). The development version is available from github. Installation instructions and basic usage can be found at https://heike.github.io/x3ptools/

3.2.2 bulletxtrctr

bulletxtrctr is a developmental R package available from github (see https://heike.github.io/bulletxtrctr/) that allows an assessment of similarity scores using the data extraction pipeline described in Hare et al. (2016).

3.2.3 grooveFinder

grooveFinder is a developmental R package providing different methods for identifying the location of grooves in scans of bullets. Installation instructions and some basic usage can be found at https://heike.github.io/grooveFinder/

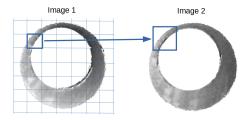


Figure 3.4: Comparing a cell in image 1 to a larger region in image 2. We wish to find the translations of the image 1 cell that yield the highest correlation within the image 2 region.

3.3 Similarity Scores

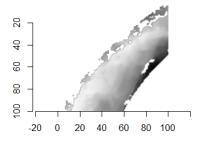
- 3.3.1 **Bullet Lands**
- 3.3.1.1 Approaches to identify groove locations
- 3.3.2 Cartridge Cases
- Analysis of Results 3.4

3.5 Communication of Results and Methods

Congruent Matching Cells (CMC) algorithm for 3.5.1comparing cartridge case breech faces

Joe 9/5/19 Update: Dealing with missing values in the x3p scans continues to be an issue. The Fast Fourier Transform method for calculating cross-correlation can't handle missing data in an image, so we've attempted a few "fixes" that haven't necessarily turned out as well as expected. One idea we had was to replace the NA values in a cell with the average pixel value. However, this is artificially introducing a signal where before there was none. This can (and demonstrably has) led to inflated/incorrect correlations between cells that shouldn't have much at all in common. Unfortunately, this may be the only solution if we still wish to adhere to the CMC algorithm as described in Song et al. (2015). One improvement that I've implemented is to "crop out" the rows and columns of an image that only contain NAs. This at least means that we've weakened the strength of the artificial signal relative to the breechface's signal.

Below is a series of images that illustrate how we might compare a cell in one image to a region of another image.



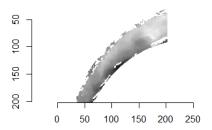
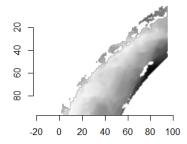


Figure 3.5: (Left) A cell from image 1. (Right) A region from image 2 centered in the same location as the image 1 cell, yet quadruple the area.

For the sake of an example, let's focus on the blue outlined cell in image 1. Our goal is to use the image 1 cell to "search" a corresponding larger region in image 2 for the horizontal/vertical translations needed to produce the highest correlation. Below is a zoomed-in version of the blue outlined image 1 cell on the left and the larger image 2 region (approximately: I made the gridded image above by-hand outside of R while the images below are from R). The image 1 cell may look larger than the image 2 region, but we can see from the axes that the image 2 region is indeed larger. Any white pixels in the two images are NA values that need to be dealt with in some way before we can use FFTs to calculate the cross-correlation.

As already discussed above, one "solution" is to replace the NA values with the average pixel value of each image. However, to avoid creating a stronger artificial signal than necessary, we can crop-out the NA rows and columns from the two images above. Below is the cropped version of the two images. The cropping



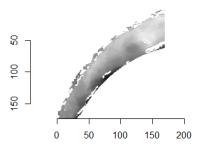


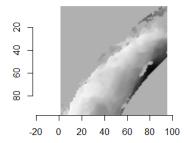
Figure 3.6: The same images as above after cropping NA rows/columns.

doesn't produce signficantly different images in this case, but you could imagine other examples in which a cell has captured only small amount of breechface in the corner. Such examples are fairly common and cropping signficantly changes the resulting correlation values.

The last step before calculating correlation for these cells is to replace the remaining NAs with the average pixel value. This is shown below.

The cross-correlation is then calculated between these two images via a standard fast fourier transform process (see Cross-Correlation Theorem). The benefit of using such a process is that (as the name suggests) it's faster than calculating the raw correlation between the two images. Also, the translations that produce the highest correlation between the image 1 cell and the image 2 region fall out of the calculation for free.

This pre-processing/cross-correlation calculation procedure is repeated for every cell in image 1 that contains breech face. Because it is not valid to assume



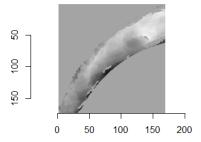


Figure 3.7: The NA-cropped images with remaining NAs replaced with the image's average pixel values.

that the two images are rotationally aligned by default, we perform the same procedure repeatedly while rotating image 2. Currently, we perform a "rough" grid search of $\theta \in [-177.5, 180]$ by increments of 2.5°. Theoretically, the final results tell us how we need to horizontally/vertically translate and rotate the two images to be correctly aligned.

3.5.1.1 Congruent Matching Tori: a promising solution to the missing value problem

As discussed above, dealing with missing values is provign to be a pain. The good news is that the currently-implemented CMC as described above yields results very similar to those published in Song et al. (2015) that originally describes that CMC algorithm. While our results seem to agree with currently published results, it would be nice if we could avoid needing to artifically replace missing values. We can do so if, rather than breaking up the circular breech face scans into disjoint squares, we break up the breech face into donut-shaped regions containing only breech face. Below is an example of such a toroidal region.

By comparing such regions instead of the square cells, we would presumably only need to fill in a few missing value "holes" in the breech face scan rather than completely replacing a non-existent signal with an artificial one. In the near-future, I hope to finish up the pre-processing needed for this Congruent Matching Tori method by performing a polar transformation on these images to make them into strips that can easily be compared via an FFT.

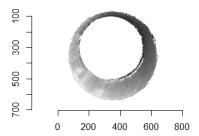
3.6 People involved

3.6.1 Faculty

- Heike Hofmann
- Susan VanderPlas

3.6.2 Graduate Students

- Ganesh Krishnan
- Kiegan Rice
- Nate Garton
- Charlotte Roigers
- Joe Zemmels
- Yawei Ge



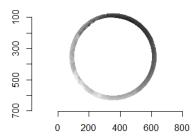


Figure 3.8: (Left) The original breech face scan image. (Right) A donut-shaped region cut out of the original image.

${\bf 3.6.3}\quad {\bf Undergraduates}$

- Talen Fisher (fix3p)
- Andrew Maloney
- Mya Fisher, Allison Mark, Connor Hergenreter, Carley McConnell, Anyesha Ray (scanner)

Handwriting

We describe our methods for going about the handwriting project here.

Glass

Shoes

6.1 Longitudinal Shoe Study

Github repository

6.1.1 Original Study Description

6.1.2 Database Paper

Paper subdirectory of Github repository

6.1.2.1 Methods and Data Description

Methods and data description handed off to Alicia for editing

6.1.2.2 Data Analysis Tools

• Working with the EBImage package - very fast processing of images

6.1.2.2.1 Film and Powder Images

Analysis Steps:

- 1. Create threshold mask
 - a. Invert the image

- b. Blur image (circular/gaussian blur, diameter 5)
- c. Threshold image (adaptive threshold, 10×10 region, keep anything with an average higher than .90 from the mean)
- d. Expand mask (default parameters rad1 = 5, rad2 = 91, proportion, expand_rad = 50)
 - 1. erode mask image (circle, diameter rad1)
 - 2. dilate mask image (circle, diameter rad2)
 - 3. label disjoint regions of the image
 - 4. prune small image regions (area < proportion parameter)
 - 5. set background color
 - 6. create dataframe of useful (non-background) pixels
 - 7. fill in holes and concave regions in mask, then expand by expand_rad vertically and horizontally (similar to "convex hull", but faster and with additional expansion)
- 2. Mask image to remove extra variability unrelated to the shoe
- 3. Threshold masked image?

6.1.2.2.2 Wear Characterization

Ideas: - average intensity of cleaned image - length of border/edges detected

- 6.2 Passive Shoe Recognition
- 6.3 Maximum Clique Matching
- 6.4 Cocoa Powder Citizen Science

Theoretical foundations

7.1 Nate's Updates 9/5/2019

- Currently in Virginia
- RA for this semester (year?) is under Danica
- Central Goals:
 - continue work started by Danica and Peter Vergeer on the analysis of likelihood ratios
 - study the differences between specific source (SS) and common source (CS) likelihood ratios (LRs) in an information theoretic way
 - does the CS or SS LR have more "information"?
 - can be the CS or SS hypotheses (prosecution or defense) be formally compared in terms of being easier to "prove" or "disprove"?

• Basic Setup

- H_p , H_d are CS prosecution and defense hypotheses upon which we will place priors
- -A and B are discrete r.v.'s representing two "sources" of evidence
 - * distributions for A and B defined conditionally based on the hypothesis
 - * SS hypothesis is represented by the conditional random variable $H_p|A$
- -X is data coming from A, Y is data coming from B
- compare information contained in (X,Y) about H_p and $H_p|A$
 - * this is what I'm working on now
 - * important quantities:
 - · Kullback-Leibler divergences, entropy, mutual information

Outreach activities

Bibliography

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- Xie, Y. (2015). Dynamic Documents with R and knitr. Chapman and Hall/CRC, Boca Raton, Florida, 2nd edition. ISBN 978-1498716963.
- Xie, Y. (2019). bookdown: Authoring Books and Technical Documents with R Markdown. R package version 0.13.