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A global analysis of root distributions for terrestrial biomes

Received: 11 January / Accepted: 8 July 1996

Abstract Understanding and predicting ecosystem functioning (e.g., carbon and water fluxes) and the role of soils in carbon storage requires an accurate assessment of plant rooting distributions. Here, in a comprehensive literature synthesis, we analyze rooting patterns for terrestrial biomes and compare distributions for various plant functional groups. We compiled a database of 250 root studies, subdividing suitable results into 11 biomes, and fitted the depth coefficient β to the data for each biome (Gale and Grigal 1987). β is a simple numerical index of rooting distribution based on the asymptotic equation $Y = 1 - \beta^d$, where d = depth and Y = the proportion of roots from the surface to depth d . High values of β correspond to a greater proportion of roots with depth. Tundra, boreal forest, and temperate grasslands showed the shallowest rooting profiles ($\beta = 0.913$, 0.943 , and 0.943 , respectively), with 80–90% of roots in the top 30 cm of soil; deserts and temperate coniferous forests showed the deepest profiles ($\beta = 0.975$ and 0.976 , respectively) and had only 50% of their roots in the upper 30 cm. Standing root biomass varied by over an order of magnitude across biomes, from approximately 0.2 to

5 kg m⁻². Tropical evergreen forests had the highest root biomass (5 kg m⁻²), but other forest biomes and sclerophyllous shrublands were of similar magnitude. Root biomass for croplands, deserts, tundra and grasslands was below 1.5 kg m⁻². Root/shoot (R/S) ratios were highest for tundra, grasslands, and cold deserts (ranging from 4 to 7); forest ecosystems and croplands had the lowest R/S ratios (approximately 0.1 to 0.5). Comparing data across biomes for plant functional groups, grasses had 44% of their roots in the top 10 cm of soil ($\beta = 0.952$), while shrubs had only 21% in the same depth increment ($\beta = 0.978$). The rooting distribution of all temperate and tropical trees was $\beta = 0.970$ with 26% of roots in the top 10 cm and 60% in the top 30 cm. Overall, the globally averaged root distribution for all ecosystems was $\beta = 0.966$ ($r^2 = 0.89$) with approximately 30%, 50%, and 75% of roots in the top 10 cm, 20 cm, and 40 cm, respectively. We discuss the merits and possible shortcomings of our analysis in the context of root biomass and root functioning.

Key words Terrestrial biomes · Cumulative root fraction · Root biomass · Rooting density · Soil depth

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Introduction

The formal study of root distributions is over 250 years old, with its origins in studies of crop species (Hales 1727). Historical improvements in techniques of root excavation and *in situ* root studies included using a hose to wash out crop roots in a profile wall (Schubart 1857), observing roots growing against a glass panel (Sachs 1873), and the formalization of root excavations (Weaver 1926). Beginning in the 1950s, tracer techniques provided a powerful tool for assessing functional rooting zones, including radioisotopes, stable isotopes, and stable tracers (e.g., Hall et al. 1953; Dansgaard 1964). More recently, dramatic improvements in video recording and image processing have led to the widespread use of minirhizotrons for *in situ* studies of root growth and demography

(e.g., Taylor 1987). These improvements notwithstanding, the most commonly used technique for biomass assessment remains the coring or excavation of soil and subsequent separation of roots. Böhm (1979) provides an excellent historical overview of methods for root studies.

In spite of this long history of study, our understanding of root distributions, and belowground processes in general, remains inadequate. Gaps in our knowledge include root attributes (e.g., distribution, production, demography), the scaling of soil processes, and the diversity of soil organisms and their role in ecosystem processes (e.g., Burke et al. 1991; Jackson and Caldwell 1993; Hawksworth and Ritchie 1993; Pregitzer et al. 1993; Freckman 1995). Together with litterfall, root production provides the primary input of organic carbon to soils (Raich and Nadelhoffer 1989) and is of obvious importance, since belowground carbon storage is more than twice aboveground storage (Schlesinger 1991). In many non-forest ecosystems, the proportion of plant biomass found in the soil is greater than 80% of total plant biomass (Caldwell and Richards 1986). Even when forests are included, belowground primary production is often 60–80% of total net primary production (Reichle et al. 1973; Coleman 1976; Ågren et al. 1980). Fine roots frequently contribute the majority of belowground production and their life expectancy ranges from weeks to years, depending on the species and environmental conditions (Shaver and Billings 1975; Vogt and Bloomfield 1991; Hendrick and Pregitzer 1993). Coarse, woody roots can be much longer-lived, in some cases effectively as old as the plant itself (Vogt and Bloomfield 1991).

In this review we (1) synthesize data on root distributions, densities, and biomass for major terrestrial biomes, (2) compare root data across biomes for various plant functional groups (grasses, shrubs, and trees), and (3) compute a globally averaged rooting distribution for all biomes. The compiled distributions are based on a comprehensive literature synthesis. Examples of processes where root distributions are important include water fluxes to the atmosphere and groundwater, soil litter decomposition, carbon sequestration, and nutrient cycling. We highlight a number of directions for future research, including incorporating more realistic root distributions into global models for predicting the consequences of global environmental change.

Methods

The database

We first compiled a database of approximately 250 references that were useful for the project (listed and numbered in Appendix 1). These references were found in journals, book chapters, reports, and unpublished manuscripts and include data from all continents except Antarctica. The oldest references date from early this century and several recent publications provided numerous references (e.g., Richards 1986; Rundel and Nobel 1991; Stone and Kalisz 1991). A reference was included in the analysis of root depth dis-

tributions if root samples were taken to at least 50 cm in at least three soil increments. Approximately 80 references met these criteria (Appendix 2), and many included multiple sites per study. Additional studies in the database were used for biomass estimates and root/shoot ratios (see below). In some cases a given study supplied data for several species at a given location and these data were combined into one ecosystem estimate. For each study we also noted the location, latitude and longitude, annual precipitation, soil type or texture, type of roots measured (e.g., fine or total, live or dead), sampling method, and depth of sampling (see Appendix 2). Where possible, the data were analyzed as cumulative root biomass (kg m^{-2} , soil surface-area basis), root density (kg m^{-3}), and cumulative root fraction (the proportion of roots from the soil surface to a given depth in the soil). Where root biomass data were not available (e.g., data presented as root length or number of intersections), a study was included only in the analysis of cumulative root distributions. The data from each reference were separated into 11 biomes: boreal forest, crops, deserts, sclerophyllous shrubland/forest, temperate coniferous forest, temperate deciduous forest, temperate grassland, tropical deciduous forest, tropical evergreen forest, tropical grassland/savanna, and tundra. We have attempted a complete review of the literature for root distributions based on the above criteria, with the exception of crop systems where we merely provide some comparative examples (O'Toole and Bland 1987). In addition to root distributions with depth, we also calculated the average root biomass and root/shoot ratios (R/S) for each biome, based on values in our database and in reviews by Caldwell and Richards (1986); Hilbert and Canadell (1996); Kummerow (1981); O'Toole and Bland (1987); Risser et al. (1981); Rodin and Bazilevich (1967); Rundel and Nobel (1991); Santantonio et al. (1977); Viereck et al. (1986); Vogt et al. (1996). Since R/S ratios sometimes change for systems over time (e.g., decreasing with canopy closure in forests), we emphasized data for mature vegetation.

The model

Gale and Grigal (1987) presented a model of vertical root distribution based on the following asymptotic equation:

$$Y = 1 - \beta^d$$

where Y is the cumulative root fraction (a proportion between 0 and 1) from the soil surface to depth d (cm), and β is the fitted "extinction coefficient". β is the only parameter estimated in the model and provides a simple numerical index of rooting distribution. High β values (e.g., 0.98) correspond to a greater proportion of roots at depth and low β values (e.g., 0.92) imply a greater proportion of roots near the soil surface (Fig. 1). β values were fitted to the data for each biome for those studies that sampled to a minimum soil depth of 1 m. Approximately 50 studies met these criteria, though coverage for some biomes was relatively weak (e.g., boreal forest with three such studies, temperate coniferous forest with four, and tropical deciduous forest with only one).

In addition to biome analyses, we examined the data by plant functional groups using only studies where roots were sampled to depths of 1 m or more. In comparing grass and shrub life forms, we examined data from temperate grasslands, tropical grasslands and deserts (i.e., systems in which the two growth forms co-occur). Many studies in those biomes compared root biomass near shrubs with similar data near grasses, while in other studies nearby shrub and grass sites were compared. To assess trees as a functional group, we combined data for all temperate and tropical forests. We also calculated a globally averaged rooting distribution by pooling all data from systems sampled to at least 1 m depth in the soil.

To create a global map of root distributions, we calculated the percentage of root biomass found in the upper 30 cm of soil for each biome, based on their respective β values. These data were then plotted on a $1^\circ \times 1^\circ$ grid scale for the land-cover classifications of Wilson and Henderson-Sellers (1985). Those classifica-

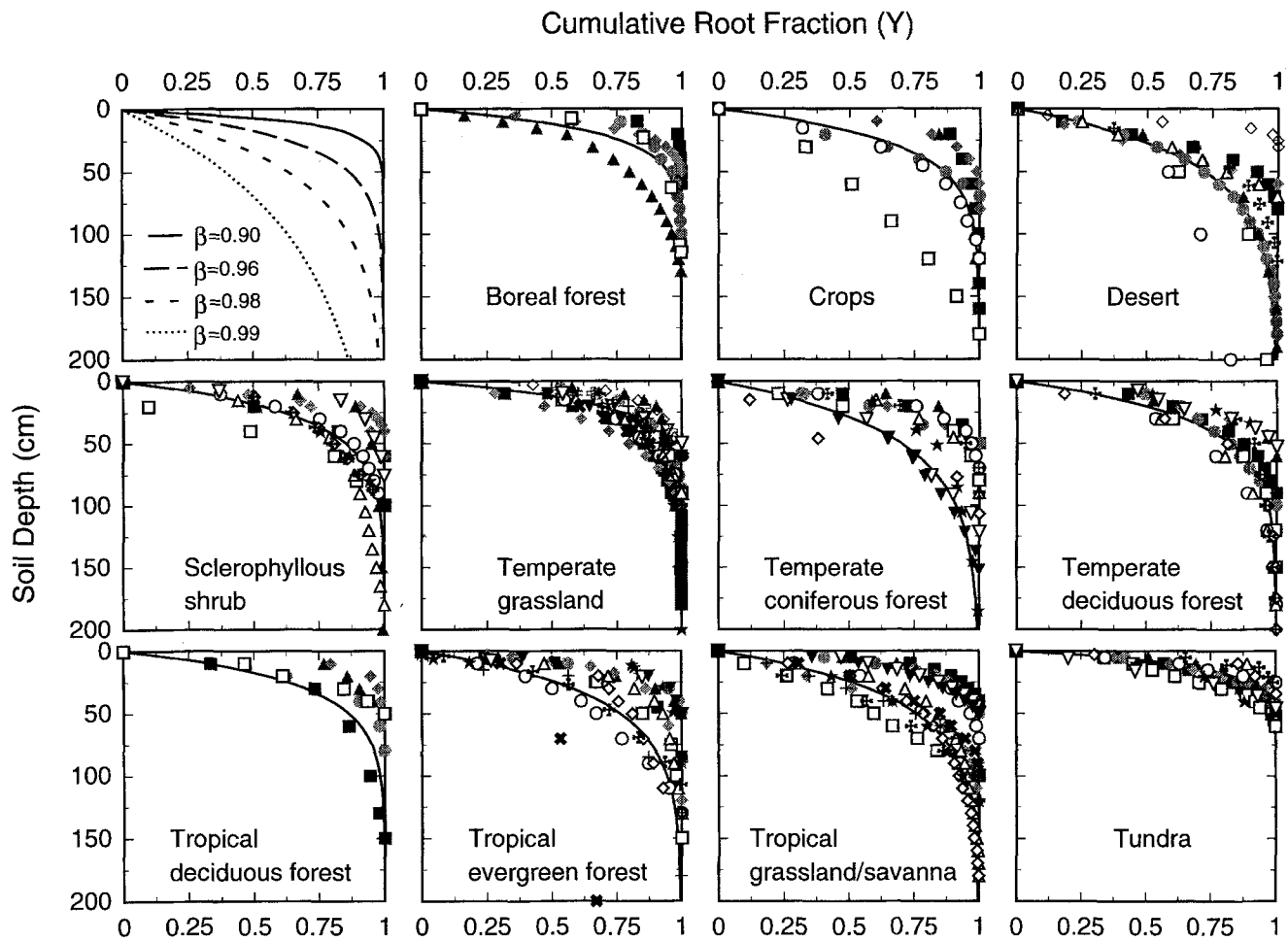


Fig. 1 Cumulative root distribution (cumulative proportion) as a function of soil depth for eleven terrestrial biomes and for the theoretical model of Gale and Grigal (1987). The curve in each biome panel is the least squares fit of β for all studies with data to at least 1 m depth in the soil. The specific β values and the associated r^2

values can be found in Table 1 and the key to the symbols in each panel is in Table 2. Gale and Grigal's equation is of the form $y=1-\beta^d$, where Y is the cumulative root fraction with depth (a proportion between 0 and 1), d is soil depth (in cm), and β is the fitted parameter. Larger values of β imply deeper rooting profiles

Table 1 Values of β (and associated r^2 values) for our data and the model of Gale and Grigal (1987), the percentage of roots in the upper 30 cm of soil, average standing root biomass ($\text{kg} \cdot \text{m}^{-2}$), and root:shoot ratios for each biome. The β values are represented graphically in the panels of Fig. 1. See Methods and Fig. 1 for a description of Gale and Grigal's model; larger values of β imply deeper rooting profiles. The values for root biomass and root:shoot ratios summarize data from our database and the following re-

views: Caldwell and Richards (1986), Hilbert and Canadell (1996), Kummerow (1981), O'Toole and Bland (1987), Risser et al. (1981), Rodin and Bazilevich (1967), Rundel and Nobel (1991), Santantonio et al. (1977), Viereck et al. (1986), and Vogt et al. (1996) (listed in Appendix 1). The dual values for desert root biomass and root:shoot ratios are for cold and warm deserts, respectively

Biome	β	r^2	% Root biomass in upper 30 cm	Root biomass ($\text{kg} \cdot \text{m}^{-2}$)	Root/shoot ratio
Boreal forest	0.943	0.89	83	2.9	0.32
Crops	0.961	0.82	70	0.15	0.10
Desert	0.975	0.95	53	1.2, 0.4	4.5, 0.7
Sclerophyllous shrubs	0.964	0.89	67	4.8	1.2
Temperate coniferous forest	0.976	0.93	52	4.4	0.18
Temperate deciduous forest	0.966	0.97	65	4.2	0.23
Temperate grassland	0.943	0.88	83	1.4	3.7
Tropical deciduous forest	0.961	0.99	70	4.1	0.34
Tropical evergreen forest	0.962	0.89	69	4.9	0.19
Tropical grassland savanna	0.972	0.95	57	1.4	0.7
Tundra	0.914	0.91	93	1.2	6.6

Fig. 2 A global map of the percentage of root biomass found in the upper 30 cm of soil plotted on a $1^\circ \times 1^\circ$ grid scale for the land-cover classifications of Wilson and Henderson-Sellers (1985). *White areas* indicate a lack of information; see Table 1 and Methods for additional information

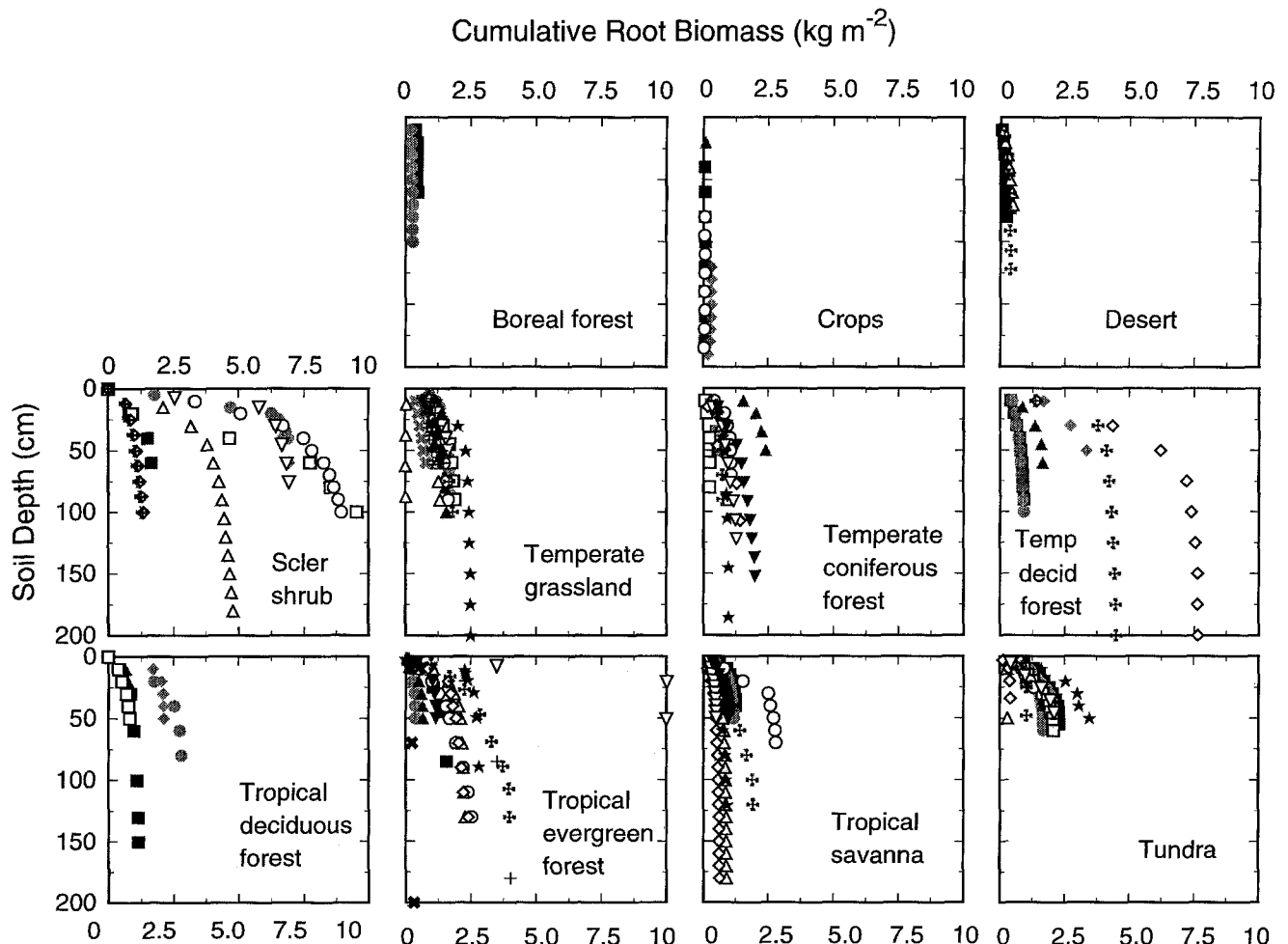
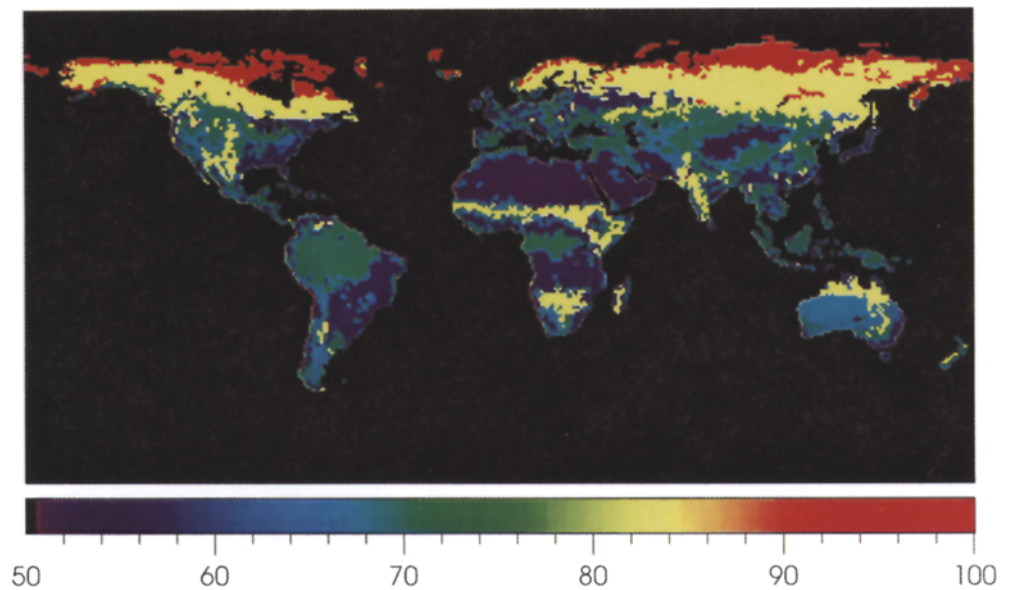


Fig. 3 Cumulative root biomass (kg m^{-2}) for 11 terrestrial biomes. The key to the *symbols* in each panel can be found in Table 2. Actual values for the *two points* shown at the *upper right corner* of tropical evergreen forest are 11.2 and 13.2 kg m^{-2} from Klinge and Herrera (1978)

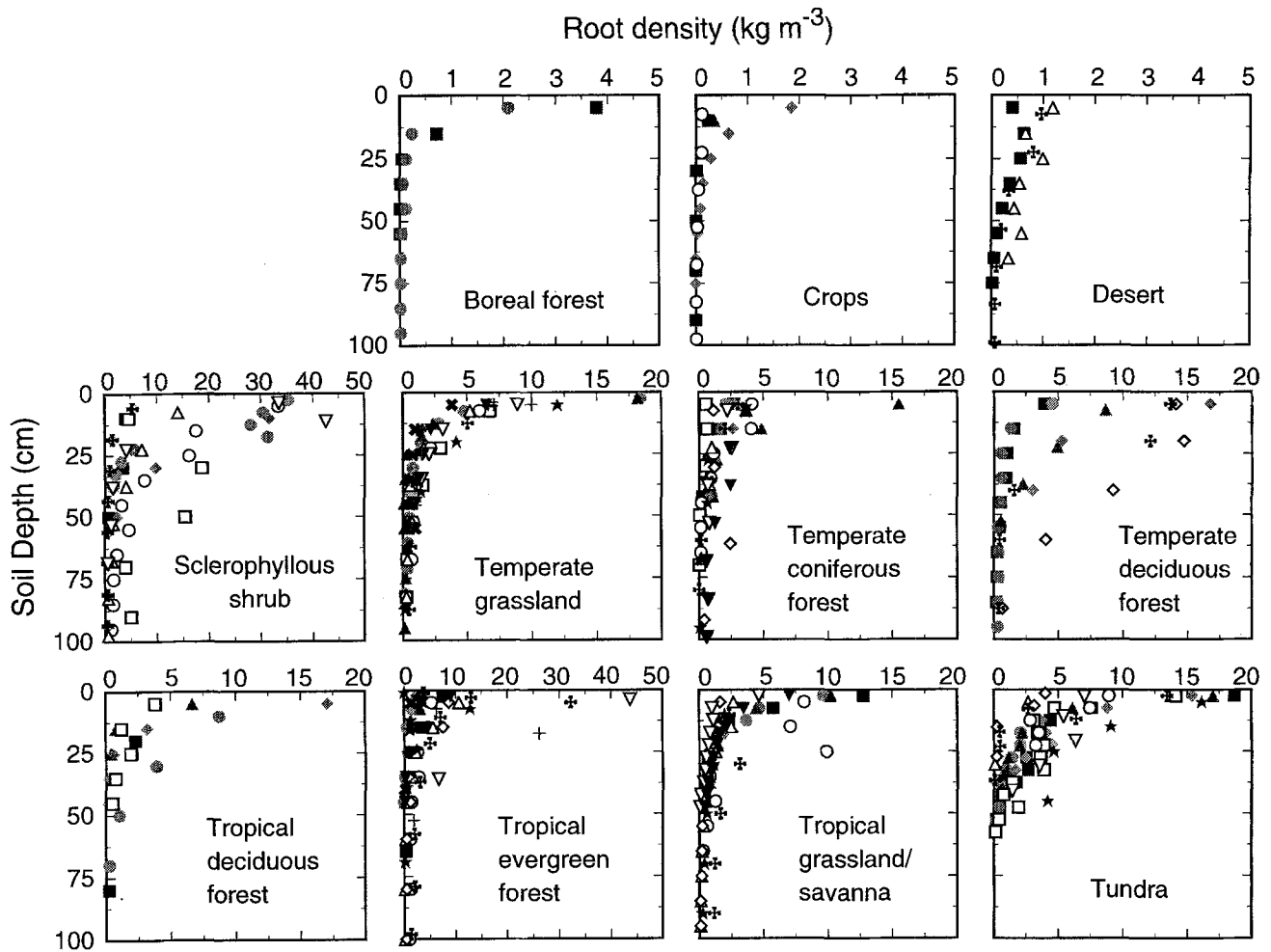


Fig. 4 Root density (kg m^{-3}) for eleven terrestrial biomes. The key to the *symbols* in each panel is in Table 2

tions include tropical broadleaf forest, temperate deciduous forest, mixed coniferous/deciduous forest, boreal coniferous forest, needle-leaf deciduous forest, savanna, temperate grassland, shrubs without ground cover, tundra, desert, and agricultural systems. The data for sclerophyllous shrublands were used for the classification of shrubs without ground cover.

Results

Tundra, boreal forest, and temperate grasslands showed the shallowest rooting profiles ($\beta = 0.913$, 0.943 , and 0.943 , respectively), with 93% of roots occurring in the top 30 cm of soil for tundra and 83% for temperate grasslands and boreal forests (Fig. 1, Table 1). Deserts and temperate coniferous forests showed the deepest rooting profiles ($\beta = 0.975$ and 0.976 , respectively) with only 50% of the roots in the uppermost 30 cm. To further contrast shallow- and deep-rooted systems, tundra typically had 60% of roots in the upper 10 cm of soil while deserts had only 20% of their roots in the same depth increment. Temperate grasslands had a shallower rooting profile than did tropical grasslands/savannas ($\beta = 0.943$

and 0.972 , respectively), though this result was due in large part to the occurrence of woody roots in most tropical grassland/savanna studies. A global map of root distributions by depth (Fig. 2) reveals (1) a predominance of shallowly rooted systems at high latitudes associated with permafrost or waterlogging, (2) shallowly rooted grassland regions, and (3) more deeply rooted woody biomes, particularly deserts, temperate coniferous forests, and tropical savannas.

Average root biomass varied by over an order of magnitude across biomes, to a maximum of 5 kg m^{-2} for forests and sclerophyllous shrublands (Table 1, Fig. 3). Ecosystems with the lowest root biomass were croplands, deserts, tundra, and grasslands, all of which had root biomass $< 1.5 \text{ kg m}^{-2}$. Deserts and croplands were lowest of all, though cold deserts had three times the root biomass of warm deserts. Root biomass in forest ecosystems ranged from approximately 2 to 5 kg m^{-2} (Table 1, Fig. 3). Individual studies finding the greatest root biomass included those in Venezuelan caatinga rainforest (Klinge and Herrera 1978) and the California chaparral (Kummerow et al. 1977; Kummerow and Mangan 1981). Root/shoot ratios for each ecosystem varied from approximately 0.1 to 7 (Table 1). The ecosystem with the smallest R/S ratio was managed croplands ($R/S = 0.1$). For more natural systems, forest ecosystems had the

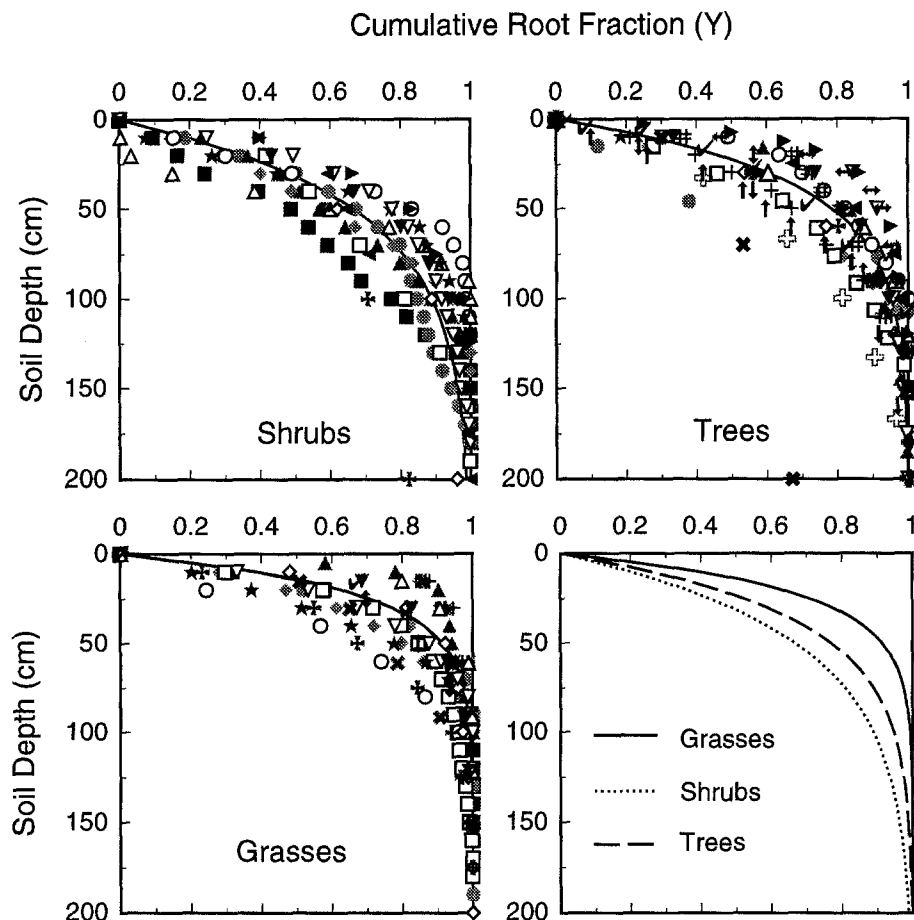


Fig. 5 The distribution of grass, tree, and shrub roots as a function of soil depth across all relevant biomes. The data for trees include temperate deciduous, temperate coniferous, tropical deciduous, tropical evergreen, and tropical savanna trees sampled to at least 1 m depth. The data for grasses and shrubs are from deserts, temperate grasslands, and tropical grasslands sampled to at least 1 m depth where the two life-forms potentially co-occur. The extinction curves derived from these data are $\beta = 0.952$ ($r^2 = 0.88$) for grasses, $\beta = 0.970$ ($r^2 = 0.91$) for trees, and $\beta = 0.978$ ($r^2 = 0.92$) for shrubs (curve fit by least squares minimization; see text for discussion of the model). The key to the grass symbols is as follows (see Appendix 1 for numbered references): ■ *Elymus alinus* (23), ● *Agropyron spicatum* (54), ▲ Belgium grassland (56), ◆ *Bouteloua gracilis* (140), □ Guinea grassland (138), ○ Ghana grassland (175), † Tallgrass prairie (176), ◇ Argentina grassland (203), ✱ *Festuca pallescens* (203), ★ fine-leaved savanna (201), ▽ broad-leaved savanna (201), ▼ *Andropogon furcatus* (243), ► *Andropogon scoparius* (243), ◄ *Bouteloua curtipendula* (243), + *Bouteloua gracilis* (243), × *Agropyron smithii* (243), # *Panicum virgatum* (244), △ *Poa pratensis* (244), ✓ *Buchloe dactyloides* (244). Shrub data: ■ *Chrysothamnus nauseosus* (23), ● *Artemisia tridentata* (23), ▲ *Sarcobatus vermiculatus* (23), ◆ *Atriplex confertifolia* (23), □ *Artemisia tridentata* (54), ○ *Senecio filaginoides* (63), △ *Mulinum spinosum* (63), ◇ *Larrea tridentata* (71), ✱ *Prosopis glandulosa* (71), ★ *Burkea africana* (125), ▽ Guinea shrubs (138), ▼ Ghana shrubs (175), ► *Mulinum spinosum* (203), ◄ *Adesmia campestris* (203). Tree data: ● (189), ▲ (230), ◆ (250) 45 years, □ (250) 80 years, ○ (60), △ (126) Virginia, ◇ (126) Cove, ✱ (126) oak-hickory, ★ (203) *Nothofagus pumila*, ▽ (203) *Nothofagus antarctica*, ▼ (6), ► (82) Kade, ◄ (82) Yangambi, + (105) Banco, × (105) Thalweg, # (105) Yapo, ✓ (123), × (170), † (231), ‡ (94), ↑ (125), ↓ (240)

smallest R/S ratios, reflecting their large aboveground woody biomass. The highest R/S ratios were observed for tundra, grasslands, and the cold-desert component of deserts (R/S ranging from approximately 4 to 6). Average root densities for each biome followed similar relative patterns as root biomass (Fig. 4). Sclerophyllous shrublands and **tropical evergreen forests had the highest root densities, in some cases densities over 40 kg m⁻³ in the shallowest depths.** Deserts and croplands had the lowest densities, with values never more than 5 kg m⁻³ even in the most densely rooted cases.

To obtain a globally averaged rooting distribution, we combined all studies in which roots were sampled to at least 1 m depth (which included data from every biome except tundra). The global average for all ecosystems was $\beta = 0.966$ ($r^2 = 0.89$; data not shown). Consequently, in the average global root profile approximately 30% of roots were in the top 10 cm, 50% in the top 20 cm, and 75% in the top 40 cm. In addition, we also compared rooting patterns for various plant functional groups across biomes, including grasses, shrubs, and trees. While grasses had 44% of their root biomass on average in the top 10 cm of soil, shrubs had only 21% of their roots in the same depth increment (Fig. 5). Grasses had 75% of their root biomass in the top 30 cm, compared to 47% for shrubs. The respective extinction coefficients were $\beta = 0.952$ ($r^2 = 0.88$) for grasses and $\beta = 0.978$ ($r^2 = 0.92$) for shrubs (Fig. 5). The average

Table 2 Key to the symbols for Figs. 1, 3, and 4. Each *number* in the table identifies a reference in Appendix 1. Each *column* contains all of the references for a given biome in alphabetical order

Symbol	Boreal forest	Crops	Desert	Sclerophyllous shrubs	Temperate coniferous forest	Temperate deciduous forest	Temperate grassland	Tropical deciduous forest	Tropical evergreen forest	Tropical grassland/savanna	Tundra
■	184	3	9	33	2	60	23	6	15	65	50
●	186	76	23	39	2	60	45	37	81	65	50
▲	216	102	54	98	2	89	56	137	81	65	50
◆	216	206	62	128	2	118	140	137	82	125	50
□	218	222	71	131	77	126	141	137	82	125	50
○		249	71	133	89	126	141		105	136	52
△			166	142	151	126	141		105	138	99
◇			172	149	189	203	152		105	138	106
⊗			220	150	228	203	176		123	175	106
★				160	230	204	203		123	175	121
▽				212	250	252	207		124	178	121
▼					250		207		155	178	
+							207		231	201	
×							207		170	201	
#							207				
†							209				
✓							244				

rooting distribution for all temperate and tropical trees was $\beta = 0.970$ ($r^2 = 0.91$), with 26% of roots in the top 10 cm, 60% in the top 30 cm, and 78% in the top 50 cm (Fig. 5). Boreal forest trees were considerably more shallowly rooted ($\beta = 0.943$, see above). Combining data from 25 studies of all woody plants (trees and shrubs), the average rooting distribution was $\beta = 0.975$ ($r^2 = 0.90$), with 40% of roots in the top 20 cm (data not shown).

Discussion

One goal of our root analysis was to provide a database for use in assessing soil C distributions and in examining the effect of roots on C, H₂O, and nutrient fluxes between soil, plants, and the atmosphere. One of the only approaches for addressing such questions at regional and global scales, and for predicting the consequences of global change, is modeling. Currently, the most explicit root descriptions in well accepted biome or global models are simple two- or three-layer representations that separate shallow and deep water at arbitrary depth (e.g., Potter et al. 1993; Neilson 1995). For example, MAPSS (Neilson 1995) is an ecosystem-biogeographic model that links vegetation with water balance processes. It has three soil layers (L1 from 0–50 cm, L2 from 50–150 cm, and L3 below 150 cm), with grasses extracting water only from L1, shrubs from L1 and L2, and L3 containing no roots (but consisting of a pool of H₂O for gravity-driven drainage to streams). CASA (Potter et al. 1993) is a process model of terrestrial ecosystem production that uses two sets of rooting depths. For water uptake the soil rooting depth is 1.0 m for grasslands, tundra, and croplands and 2.0 m for forests; the scalar used to estimate C turnover and N mineralization includes a depth of only 0.3 m (Potter et al. 1993). Other models, including TEM (Raich et al. 1991; Melillo et al. 1993), BIOME2 (Pren-

tice et al. 1992) and BIOME-BGC (e.g., Running and Hunt 1993), either do not specifically include soil depth and root distributions, or use only a single biome-specific soil depth parameter. CENTURY (Parton et al. 1988, 1992), an ecosystem model used to simulate patterns of plant primary production, soil organic matter dynamics, and nutrient cycling, is a notable exception with five soil depths: 0–15 cm, 15–30 cm, 30–45 cm, 45–60 cm, and 60–90 cm.

Mechanistic models that examine the feedbacks between vegetation and climate (including atmospheric CO₂) are critical for predicting the consequences of global change and for understanding the cycling of C, H₂O, and nutrients (e.g., Vitousek and Matson 1984; Wullschlegel et al. 1994; Paruelo and Sala 1995; Field et al. 1995). Given these models as examples, how might information on root distributions improve predictions of ecosystem response to global change? One promising approach would be to incorporate the observed root distributions into biome or global models. One or more of the models might then be linked to a GCM (general circulation or global change model) to quantify feedbacks between vegetation and climate. Such feedbacks are necessary for dynamic models that allow biomes to fluctuate geographically, both affected by and affecting the earth's climate. More specific questions based on plant functional groups or a subset of biomes might also be addressed by combining our root data with models. For example, if the world's grasslands were converted to shrublands, how would H₂O fluxes and C sequestration be altered? How might the conversion of tropical forests to pasture affect C distributions in the soil, and what would be the consequences for recirculation of H₂O? The observed root distributions provide information to help answer many such questions.

By far the majority of ecosystem root biomass resides in the upper 1 m of soil (Fig. 3; Table 1). Despite

this predominance of biomass in the upper soil layers, our knowledge of the importance of the deep soil to nutrient and water balances could be much improved, particularly considering how few studies have quantitatively sampled roots below 2 m. We found only nine studies that measured root distributions to at least 2 m depth in the soil. Those studies included one each in cold and warm deserts (Dobrowolski et al. 1990; Freckman and Virginia 1989), one chaparral dataset (the mountain fynbos of Higgins et al. 1987), three forest studies (the pine plantation of Van Rees and Comerford 1986; Kochenderfer 1973 in temperate hardwood forest; and Nepstad et al. 1994 for the Amazon), two savanna studies (*Prosopis glandulosa* data in Heitschmidt et al. 1988; Watts 1993) and data for five sites in Patagonia, Argentina (a transect from *Nothofagus* forest through grassland to desert, Schulze et al. 1996). From a practical perspective it is interesting to ask how much information was gained in these studies by sampling below 1 m depth. In five of the nine studies, 93%–100% of the roots observed in the profile occurred in the uppermost 1 m. Two minor exceptions were Heitschmidt et al. (1988), who found 90% of roots at 133 cm, and Watts (1993), who found 92% of root biomass at 120 cm. The two notable exceptions were Freckman and Virginia (1989) and Nepstad et al. (1994). The Jornada desert data in Freckman and Virginia (1989) included two community types, one dominated by the phreatophyte *Prosopis glandulosa* and one dominated by *Larrea tridentata*. *P. glandulosa*, one of the most deeply rooted species in the world (Canadell et al. 1996), had 30% of its roots below 1 m, while *L. tridentata*, with a shallower rooting profile, had only 11% below 1 m. Nepstad et al. (1994) measured fine-root biomass (< 1 mm) to a depth of approximately 6.5 m in eastern Amazonia, Brazil. Their data show small but consistent fine-root biomass between 1 m and 6.5 m, enough to contribute substantially to total fine-root biomass; 50% of fine roots in that system occurred in the upper 70 cm of soil, but nearly one-third were below 2 m. Based on Table 1 and the data in Nepstad et al. (1994), we estimate fine root biomass in that system to be approximately 10% of total root biomass (assuming 5 kg m⁻² for the latter). Deep roots are likely to be important for C and H₂O dynamics in a number of ecosystems that experience periodic drought. An examination of deep-rooted species, including *Acacia*, *Prosopis*, and *Eucalyptus* spp., shows that they are most often found in water-limited systems (see recent reviews by Stone and Kalisz 1991; Canadell et al. 1996).

The root distributions presented here (Fig. 1) are based primarily on root biomass in the upper 1–2 m of soil (Appendix 2). What additional factors may be important for belowground resource capture and ecosystem attributes? In addition to biomass, root surface area is important for resource uptake, with important contributions from the relative activity of roots (Newman 1974; Fitter 1982; Jackson et al. 1990) and root symbioses (e.g., Vincent 1974; Allen 1991). Although the uptake

of nutrients may be limited primarily to upper soil layers, a relatively small proportion of roots deep in the soil can be quite important for water uptake. To assess functional rooting zones, tracer techniques and other approaches are an important supplement to direct excavation. Relevant tracers include radioisotopes, stable isotopes, and stable tracers (e.g., Fox and Lipps 1964; McKane et al. 1990). Both short term and seasonal fluctuations in deep soil water can indicate root activity (Holmes and Colville 1970). Seasonality of CO₂ concentrations in soil air with depth can also indicate activity of roots. Richter and Markewitz (1995) show substantial soil acidity (pH ≤ 4.2) to at least 6 m depth in the soil, considerably more acidity than for the underlying parent material (pH 7.9).

The data for certain systems were quite variable. Sclerophyllous shrublands include such diverse systems as the shallowly rooted mountain fynbos of South Africa, dominated by *Protea* spp. (Higgins et al. 1987), to the potentially deep-rooted chaparral of southern California (Kummerow and Mangan 1981, though we were unable to identify any southern California study that quantitatively sampled roots to > 1 m depth). R/S ratios in sclerophyllous shrublands ranged from approximately 0.3 to 5, while R/S ratios in forest systems were much more consistent. In general, variation in root distributions requires more detailed spatial and temporal integration in some systems if accurate root assessments are to be made. Deserts are comprised of shallow-rooted ephemerals, shallow-rooted perennials, and deep-rooted perennials (Rundel and Nobel 1991). Where desert root distributions have been examined, studies have typically focused on individual species. Shallow-rooted ephemerals typically avoid drought, with root depths less than 20 cm (Evenari et al. 1971; Golluscio and Sala 1993). Shallow-rooted perennials include cacti, which rarely grow roots below 50 cm (Cannon 1911; Nobel 1989). In contrast, roots of phreatophytes such as *Prosopis glandulosa* or *Zizyphus lotus* can reach depths of 50 m or more in the soil (Phillips 1963; Zohary 1961; Canadell et al. 1996). Accurately assessing desert rooting patterns requires spatial integration (either by examining a number of species or by random sampling) and temporal integration (to capture the changing phenologies of root abundance). Forest studies face similar difficulties, and should address sampling distance from the tree and tree density. A few examples of attempts to integrate such spatial or temporal dynamics in woody systems are Reich et al. (1980), Farrish (1991), and Le Roux et al. (1995).

Relative root distributions among and within biomes differ in part because of physical barriers to growth. For example, permafrost restricts rooting depth in tundra and in some boreal forests (e.g., Bonan 1992), though less commonly in the boreal forests of North America (Solomon 1992). In addition, waterlogging can also inhibit root growth (Kane et al. 1992). These and other factors make tundra ecosystems the most shallowly rooted of all biomes examined, and lead to a shallower rooting profile

for boreal forests than for other forest types (Fig. 1, Table 1). Poor soil aeration from waterlogging can decrease rooting depth in all ecosystems (Klinge and Herrera 1978; Drew 1990; Rundel and Nobel 1991). Strong mechanical resistance to root penetration can be found in arid and semi-arid ecosystems with a substantial caliche layer (Gile et al. 1966), or in tropical savannas and tropical forests with a prevalent ironpan (Richards 1986). Not surprisingly, shallow bedrock also inhibits root growth, but channels and cracks can sometimes increase functional rooting depth. Though all of these factors can limit rooting depth, high temperatures can result in decreased root abundance near the soil surface. In unshaded desert soils the surface temperature can reach 70°C (Buxton 1925), reducing or eliminating roots in the upper soil layer (e.g., Nobel 1988).

The ideal root study provides data to compare not just total root biomass, but fine roots alone, coarse roots, the distribution of root length and surface area with depth, the proportion of live and dead roots, and root distributions for ecosystems and individual species. Not surprisingly, few studies include all of this information. Although it is unrealistic to expect every study to do so, there are simple improvements that could be made to increase the benefit of many future studies. One such improvement would be to document the sampling methods more clearly. Spelling out the core diameter or area over which sampling occurred, and accompanying depth increments, enables data to be converted easily between a soil-density and soil surface-area basis (e.g., kg m⁻³ or kg m⁻²). This conversion is important when, for example, the same data are used to compare total ecosystem biomass (where a soil surface-area basis is appropriate) and soil organic matter concentrations (where density is appropriate). Studies should be specific about whether root mass included dead roots, a subset of root size classes, or total root biomass. One chronic problem is the underestimation of fine root biomass. Grier et al. (1981) estimated that two-thirds of net primary production in a 180-year-old stand of *Abies amabilis* went to fine root production, but such roots are often overlooked in biomass estimates for woody vegetation. In some forests, the majority of fine roots are < 1.0 mm in diameter and may be difficult to recover from the soil (Hendrick and Pregitzer 1993). Certain techniques, such as separating roots from soil with pressurized water, almost certainly underestimate fine roots.

This review provides a current synthesis of the literature, to be improved with the addition of new studies and the inclusion of older studies that were unintentionally omitted. There are a number of important questions on the controls of plant rooting distributions that we hope to use the database to address, with an emphasis on global environmental change. Global change may induce strong feedbacks between plant rooting distributions and climate. The relatively large global warming predicted for polar regions could have a profound effect on permafrost depth and, consequently, tundra rooting patterns and net C efflux (Chapin et al. 1992; Oechel et

al. 1994). Deforestation in the Amazon and other regions could alter recirculation of water between terrestrial ecosystems and the atmosphere, regional hydrology in general, and C storage (e.g., Dickinson and Henderson-Sellers 1988; Lean and Warrilow 1989; Nepstad et al. 1994), though net C loss can be mitigated to some extent by intercropping and by selecting relatively deep-rooted pasture species (Fisher et al. 1994). Increased atmospheric CO₂ and land-use change may alter the proportion of shrubs and grasses across the globe (e.g., Archer 1995; Polley et al. 1996), changing C distributions in the soil and the recirculation of water. We plan to incorporate root distributions into existing biome and global models for more realistic representations of belowground processes. With model developers, we could then address the effects of changes in land use or climate for the cycling of C, H₂O, and nutrients. Our long-term goal is to link one or more of these global terrestrial models with a GCM to examine the feedbacks between vegetation and climate. Such global models, together with paleo-analyses, provide the only integrative method for predicting the potential consequences of global environmental change.

Acknowledgements We thank Jessica Pitelka, who began the tedious process of compiling references, and the many researchers who provided them. J. Randerson assisted with the global map of root distributions. L.J. Anderson, M.M. Caldwell, M.R. Gale, S.G. Jackson, and K.S. Pregitzer provided helpful comments on the manuscript. We also thank the Max Planck Institut, NASA-EOS (NAS 5-31726), and NIGEC/DOE (TUL-038-95/96) for support of this study.

Appendix 1

References to works included in the database. *Numbers* are referred to in Table 2

1. Abbott ML, Fraley L, Reynolds Jr, TD (1991) Root profiles of selected cold desert shrubs and grasses in disturbed and undisturbed soils. *Environ Exp Bot* 31:165-178
2. Ares A, Peinemann N (1992) Fine-root distribution of coniferous plantations in relation to site in southern Buenos Aires, Argentina. *Can J For Res* 22:1575-1582
3. Armstrong EL, Pate JS, Tennant D (1994) The field pea crop in South Western Australia-patterns of water use and root growth in genotypes of contrasting morphology and growth habit. *Aust J Plant Physiol* 21:517-532
4. Atkinson D (ed) (1991) Plant root growth: an ecological perspective, vol. 10. (Special publication of the British Ecological Society). Blackwell, Oxford
5. Backeus I (1990) Production and depth distribution of fine roots in a boreal open bog. *Ann Bot Fenn* 27:261-265
6. Bang-xing Wu (1991) Studies on the vertical structure of seasonal rain-forest in Xishuangbanna of Yunnan. *Act Bot Sin* 33:232-239
7. Bannan MW (1940) The root systems of northern Ontario conifers growing in sand. *Am J Bot* 27:108-114
8. Barbour MG (1973) Desert dogma reexamined: root/shoot productivity and plant spacing. *Am Midl Nat* 89:41-57
9. Barbour MG, MacMahon JA, Bamberg SA, Ludwig JA (1977) The structure and distribution of *Larrea* communities. In: Mabry TJ, Hunziker JH, Difeo DR Jr. (eds) *Creosote bush: biology and chemistry of Larrea in New World deserts*. Dowden, Hutchinson and Ross, Stroudsburg, pp 227-251

10. Bathke GR, Cassel DK, Hargrove WL, Porter PM (1992) Modification of soil physical properties and root growth response. *Soil Sci* 154:316–329
11. Beese F (1986) Parameter des Stickstoffumsatzes in Ökosystemen mit Böden unterschiedlicher Acidität. *Göttinger Bodenk Ber* 90:1–344
12. Bell KL, Bliss LC (1978) Root growth in a polar semidesert environment *Can J Bot* 56:2470–2490
13. Belsky AJ, Amundson RG, Duxbury JM, Riha SJ, Ali AR, Mwonga SM (1989) The effects of trees on their physical, chemical, and biological environments in a semi-arid savanna in Kenya. *J Appl Ecol* 26:1005–1024
14. Bennett OL, Doss BD (1960) Effect of soil moisture level on root distribution of cool-season forage species. *Agron J* 52: 204–207
15. Berish CW (1982) Root biomass and surface area in three successional tropical forests. *Can J For Res* 12:699–704
16. Berish CW, Ewel JJ (1988) Root development in simple and complex tropical successional ecosystems. *Plant Soil* 106: 73–84
17. Bernard JM, Fiala K (1986) Distribution and standing crop of living and dead roots in three wetland *Carex* species. *Bull Torrey Bot Club* 113:1–5
18. Bishop DM (1962) Lodgepole pine rooting habits in the Blue Mountains of northeastern Oregon. *Ecology* 43:140–142
19. Biswell HH (1935) Effects of environment upon the root habits of certain deciduous forest trees. *Bot Gaz* 96:676–708
20. Blaschke H (1991) Distribution, mycorrhizal infection, and structure of roots of calcicole floral elements at treeline, Bavarian Alps, Germany. *Arct Alp Res* 23:444–450
21. Böhm W (1979) Methods of studying root systems. Springer Berlin Heidelberg New York
22. Braecke FH (1992) Root biomass changes after drainage and fertilization of a low-shrub pine bog. *Plant Soil* 143:33–43
23. Branson FA, Miller RF, McQueen IS (1976) Moisture relationships in twelve northern desert shrub communities near Grand Junction, Colorado. *Ecology* 57:1104–1124
24. Braun EL (1936) Notes on root behavior of certain trees and shrubs of the Illinoisian till plain of southwestern Ohio. *Ohio J Sci* 36:141–146
25. Bray JR (1963) Root production and the estimation of net productivity. *Can J Bot* 41:65–72
26. Bray JR, Dudkiewicz LA (1963) The composition, biomass, and productivity of two *Populus* forests. *Bull Torrey Bot Club* 90:298–308
27. Brown JH Jr, Woods FW (1968) Root extension of trees in surface soils of the North Carolina Piedmont. *Bot Gaz* 129: 126–132
28. Brown JR, Archer S (1990) Water relations of a perennial grass and seedling vs. adult woody plants in a subtropical savanna, Texas. *Oikos* 57:366–374
29. Brundrett MC, Kendrick B (1988) The mycorrhizal status, root anatomy, and phenology of plants in a sugar maple forest. *Can J Bot* 66:1153–1173
30. Burton GW, DeVane EH, Carter RL (1954) Root penetration, distribution, and activity in southern grasses measured by yields, drought symptoms, and P₃₂ uptake. *Agron J* 46:229–238
31. Caldwell MM, Camp LB (1974) Belowground productivity of two cool desert communities. *Oecologia* 17:123–130
32. Caldwell MM, Richards JH (1986) Competing root systems: morphology and models of absorption. In: Givnish TJ (ed) *On the economy of plant form and function*. Cambridge University Press, Cambridge, pp 251–273
33. Canadell J, Roda F (1991) Root biomass of *Quercus ilex* in a montane Mediterranean forest. *Can J For Res* 21:1771–1778
34. Cannon WA (1911) The root habits of desert plants (Publication 131). Carnegie Institution, Washington
35. Cannon WA (1949) A tentative classification of root systems. *Ecology* 30:542–548
36. Carbon BA, Bartle GA, Murray AM, MacPherson DK (1980) The distribution of root length, and the limits to flow of soil water to roots in a dry sclerophyll forest. *For Sci* 26:656–664
37. Castellanos J, Maass, Kummerow J (1991) Root biomass of a dry deciduous tropical forest in Mexico. *Plant Soil* 131: 225–228
38. Cavelier J (1992) Fine-root biomass and soil properties in a semideciduous and a lower montane rain forest in Panama. *Plant Soil* 142:187–201
39. Chapman SB (1970) The nutrient content of the soil and root system of a dry heath ecosystem. *J Ecol* 58:445–452
40. Cholick FA, Welsh JR, Cole CV (1977) Rooting patterns of semi-dwarf and tall winter wheat cultivars under dryland field conditions. *Crop Sci* 17:637–639
41. Clements FE, Weaver JE, Hanson HC (1929) Plant competition: an analysis of community function (Publication 398). Carnegie Institution, Washington
42. Comstock JP, Ehleringer JR (1992) Plant adaptation in the Great Basin and Colorado Plateau. *Great Basin Nat* 52:195–215
43. Coutts MP, Lewis GJ (1983) When is the structural root system determined in Sitka spruce? *Plant Soil* 71:155–160
44. Cuevas E, Brown S, Lugo AE (1991) Aboveground and belowground organic matter storage and production in a tropical pine plantation and a paired broadleaf secondary forest. *Plant Soil* 135:257–268
45. Dahlman RC, Kucera CL (1965) Root productivity and turnover in native prairie. *Ecology* 46:84–89
46. Davis SD, Mooney HA (1986) Water use patterns of four co-occurring chaparral shrubs. *Oecologia* 70:172–177
47. Day MW (1944) The root system of aspen. *Am Midl Nat* 32:502–509
48. Dell B, Bartle JR, Tacey WH (1983) Root occupation and root channels of Jarrah forest subsoils. *Aust J Bot* 31:615–627
49. Denmead OT, Shaw RH (1962) Availability of soil water to plants as affected by soil moisture and meteorological conditions. *Agron J* 54:385–390
50. Dennis JG, Johnson PL (1970) Shoot and rhizome-root standing crops of tundra vegetation at Barrow, Alaska. *Arct Alp Res* 2:253–266
51. Dennis JG (1977) Distribution patterns of belowground standing crop in arctic tundra at Barrow, Alaska. *Arct Alp Res* 9:111–125
52. Dennis JG, Tieszen LL, Vetter MA (1978) Seasonal dynamics of above- and belowground production of vascular plants at Barrow, Alaska. In: Tieszen LL (ed) *Vegetation and production ecology of an Alaskan Arctic tundra*, Springer Berlin Heidelberg New York, pp 113–140
53. Dittmer HJ (1959) A study of the root systems of certain sand dune plants in New Mexico. *Ecology* 40:265–273
54. Dobrowolski JP, Caldwell MM, Richards JH, (1990) Basin hydrology and plant root systems. In: Osmond CB, Pitelka LF, Hidy GM (eds) *Plant biology of the Basin and Range*. Springer Berlin Heidelberg New York, pp 243–292
55. Drexhage M (1994) Die Wurzelentwicklung 40-jähriger Fichten (*Picea abies* [L.] Karst.) in der Langen Bramke (Harz).
56. Dumortier M (1991) Below-ground dynamics in a wet grassland ecosystem. In: Atkinson D (ed) *Plant root growth: an ecological perspective*. Blackwell, Oxford, pp 301–309
57. Edwards PJ, Grubb PJ (1977) Studies of mineral cycling in a montane rain forest in New Guinea. I. The distribution of organic matter in the vegetation and soil. *J Ecol* 65:943–969
58. Ehrenfeld JG, Kaldor E, Parmelee RW (1992) Vertical distribution of roots along a soil toposequence in the New Jersey Pinelands. *Can J For Res* 22:1929–1936
59. Escamilla JA, Comerford NB, Neary DG (1991) Spatial pattern of slash pine roots and its effect on nutrient uptake. *Soil Sci Soc Am J* 55:1752–1757
60. Farrish KW (1991) Spatial and temporal fine-root distribution in three Louisiana forest soils. *Soil Sci Soc Am J* 55: 1752–1757
61. Feller MC (1980) Biomass and nutrient distribution in two eucalypt forest ecosystems. *Aust J Ecol* 5:309–333
62. Fernandez OA, Caldwell MM (1975) Phenology and dynamics of root growth of three cool semi-desert shrubs under field conditions. *J Ecol* 63:703–714

63. Fernandez RJ, Paruelo JM (1988) Root systems of two Patagonian shrubs: a quantitative description using a geometrical method. *J Range Manage* 41:220–223
64. Fiala K (1979) Estimation of annual increment of underground plant biomass in a grassland community. *Fol Geobot Phytotax* 14:1–10
65. Fiala K, Herrera R (1988) Living and dead belowground biomass and its distribution in some savanna communities in Cuba. *Fol Geobot* 23:225–237
66. Fitter AH (1986) Spatial and temporal patterns of root activity in a species-rich alluvial grassland. *Oecologia* 69:594–599
67. Fitter AH (1987) An architectural approach to the comparative ecology of plant root systems. *New Phytol* 106:61–77
68. Fogel R (1983) Root turnover and productivity of coniferous forests. *Plant Soil* 71:75–85
69. Franco AC, Nobel PS (1990) Influences of root distribution and growth on predicted water uptake and interspecific competition. *Oecologia* 82:151–157
70. Fraser DA, McGuire D (1969) Total growth of a black spruce (*Picea mariana*) tree at Chalk River, Ontario, Canada. *Can J Bot* 47:73–84
71. Freckman DW, Virginia RA (1989) Plant-feeding nematodes in deep-rooting desert ecosystems. *Ecology* 70:1665–1678
72. Gale MR (1987) A forest productivity index model based on soil and root-distributional characteristics. Ph-D dissertation, Univ of Minnesota, St. Paul
73. Gale MR, Grigal DK (1987) Vertical root distributions of northern tree species in relation to successional status. *Can J For Res* 17:829–834
74. Gallagher JL, Plumley FG (1979) Underground biomass profiles and productivity in Atlantic coastal marshes. *Am J Bot* 66:156–161
75. Gardner WR (1964) Relation of root distribution to water uptake and availability. *Agron J* 56:41–45
76. Gäth S, Meuser H, Abitz C-A, Wessolek G, Renger M (1989) Determination of potassium delivery to the roots of cereal plants. *Z Pflanzenernähr Bodenk* 152:143–149
77. Gehrman J, Gerriets M, Puhe J, Ulrich B (1984) Untersuchungen an Boden, Wurzeln, Nadeln und erste Ergebnisse von Depositionsmessungen im hils. In: *Berichte des Forschungszentrums Waldökosysteme/Waldsterben*, vol 2.
78. Gifford GF (1966) Aspen root studies on three sites in northern Utah. *Am Midl Nat* 75:132–141
79. Glinski DS, Karnok KJ, Carrow RN (1993) Comparison of reporting methods for root growth data from transparent-interface measurements. *Crop Sci* 33:310–314
80. Golluscio RA, Sala OE (1993) Plant functional types and ecological strategies in Patagonian forbs. *J Veg Sci* 4:839–846
81. Gower ST (1987) Relations between mineral nutrient availability and fine root biomass in two Costa Rican tropical wet forests: a hypothesis. *Biotropica* 19:171–175
82. Greenland DJ, Kowal JML (1960) Nutrient content of the moist tropical forest of Ghana. *Plant Soil* 12:154–174
83. Gregory PJ, Lake JV, Rose DA, (eds) (1987) Root development and function. Cambridge University Press, Cambridge
84. Grier CC, Logan RS (1977) Old-growth *Pseudotsuga menziesii* communities of a western Oregon watershed: biomass distribution and production budgets. *Ecol Monogr* 47:373–400
85. Grier CC, Vogt KA, Keyes MR, Edmonds RL (1981) Biomass distribution and above- and below-ground production in young and mature *Abies amabilis* zone ecosystems of the Washington Cascades. *Can J For Res* 11:155–167
86. Groenendijk AM, Vink-Lievaert MA (1987) Primary production and biomass on a Dutch salt marsh: emphasis on the below-ground component. *Vegetatio* 70:21–27
87. Habib R, Chadoeuf J (1989) Errors in estimating total root length by sub-samples method. *Plant Soil* 115:129–134
88. Hamblin A, Tennant D (1987) Root length density and water uptake in cereals and grain legumes: how well are they correlated? *Aust J Agric Res* 38:513–527
89. Harris WF, Kinerson RA Jr, Edwards NT (1977) Comparison of belowground biomass of natural deciduous forests and loblolly pine plantations. In: Marshall JK (ed) *The belowground ecosystem: a synthesis of plant-associated processes*. Colorado State University Press, Fort Collins, pp 29–37
90. Harris WF, Kinerson RS Jr, Edwards NT (1977) Comparison of belowground biomass of natural deciduous forest and loblolly pine plantations. *Pedobiologia* 17:369–381
91. Håland B, Braekke FH (1989) Distribution of root biomass in a low-shrub pine bog. *Scand J For Res* 4:307–316
92. Heeraman DA, Juma NG (1993) A comparison of minirhizotron, core and monolith methods for quantifying barley (*Hordeum vulgare* L.) and fababean (*Vicia faba* L.) root distribution. *Plant Soil* 148:29–41
93. Hellmers H, Horton JS, Juhren G, O'Keefe J (1955) Root systems of some chaparral plants in southern California. *Ecology* 36:667–678
94. Heitschmidt RK, Ansley RJ, Dowhower SL, Jacoby PW, Price DL (1988) Some observations from the excavation of honey mesquite root systems. *J Range Manage* 41:227–230
95. Hendrick RL, Pregitzer KS (1993) The dynamics of fine root length, biomass and nitrogen content in two northern hardwood ecosystems. *Can J For Res* 12:2507–2520
96. Hendrick RL, Pregitzer KS (1996) Temporal and depth-related patterns of fine root dynamics in northern hardwood forests. *J Ecol* 84:167–176
97. Hermann RK, Petersen RG (1969) Root development and height increment of Ponderosa pine in pumice soils of central Oregon. *For Sci* 15:226–237
98. Higgins KB, Lamb AJ, Wilgen BW van (1987) Root systems of selected plant species in mesic fynbos in the Jonkershoek Valley, south-western Cape Province. *S Afr J Bot* 53:249–257
99. Hobbie SE (1995) The effects of increased temperature on tundra plant community composition and the consequences for ecosystem processes. Ph D Dissertation, University of California, Berkeley
100. Hoffmann A, Kummerow J (1978) Root studies in the Chilean matorral. *Oecologia* 32:57–69
101. Holch AE, Hertel EW, Oakes WO, Whitwell HH (1941) Root habits of certain plants of the foothill and alpine belts of Rocky Mountain National Park. *Ecol Monogr* 11:327–345
102. Huck MG, Peterson CM, Hoogenboom G, Busch CD (1986) Distribution of dry matter between shoots and roots of irrigated and nonirrigated determinate soybeans. *Agron J* 78:807–813
103. Hunt ER Jr, Nobel PS (1987) A two-dimensional model for water uptake by desert succulents: implications of root distribution. *Ann Bot* 59:559–569
104. Hunt ER Jr, Nobel PS (1987) Allometric root/shoot relationships and predicted water uptake for desert succulents. *Ann Bot* 59:571–577
105. Huttel C (1975) Root distribution and biomass in three Ivory Coast rain forest plots. In: Golley FB, Medina E (eds) *Tropical ecological systems*. Springer Berlin Heidelberg New York, pp 123–130
106. Ignatenko IV, Khamizyanova FI (1971) Soils and total phytomass reserves in dwarf birch-white dryas and willow tundras of the east European northlands. *Ekologiya* 4:17–24
107. Jackson RB, Caldwell MM (1993) Geostatistical patterns of soil heterogeneity around individual perennial plants. *J Ecol* 81:683–692
108. Jeník J (1969) Root structure and underground biomass in equatorial forests. In: *Productivity of forest ecosystems*. Proc Brussels Symp 4:323–331
109. Jenkins MB, Virginia RA, Jarrell WM (1988) Depth distribution and seasonal population fluctuations of mesquite-nodulating rhizobia in warm desert ecosystems. *Soil Sci Soc Am J* 52:1644–1650
110. Jonasson S, Callaghan TV (1992) Root mechanical properties related to disturbed and stressed habitats in the Arctic. *New Phytol* 122:179–186
111. Jordan CF, Escalante G (1980) Root productivity in an Amazonian rain forest. *Ecology* 61:14–18
112. Jordan PW, Nobel PS (1984) Thermal and water relations of roots of desert succulents. *Ann Bot* 54:705–717

113. Joslin D, Henderson GS (1987) Organic matter and nutrients associated with fine root turnover in a white oak stand. *For Sci* 33:330–346
114. Joslin JD, Wolfe MH (1992) Red spruce soil solution chemistry and root distribution across a cloud water deposition gradient. *Can J For Res* 22:893–904
115. Kaspar TC, Bland WL (1992) Soil temperature and root growth. *Soil Sci* 154:290–299
116. Kaufman CM (1945) Root growth of jack pine on several sites in the Cloquet Forest, Minnesota. *Ecology* 26:10–23
117. Kelly JM (1975) Dynamics of root biomass in two eastern Tennessee old-field communities. *Am Midl Nat* 94:54–61
118. Kelly JM, Joslin JD (1989) Mass and chemical composition of roots in two second-growth oak forests in eastern Tennessee. *For Ecol Manage* 27:87–92
119. Kelly JM, Mays PA (1989) Root zone physical and chemical characteristics in southeastern spruce-fir stands. *Soil Sci Soc Am J* 53:1248–1255
120. Keyes MR, Grier CC (1981) Above- and below-ground net production in 40-year-old Douglas fir stands on low and high productivity sites. *Can J For Res* 11:599–605
121. Khodachek EA (1969) Vegetal matter of tundra phytocoenoses in the western part of Taimyr Peninsula. *J Bot* 54:1059–1073
122. Kittredge J (1955) Litter and forest floor of the chaparral in parts of the San Dimas Experimental Forest, California. *Hilgardia* 23:563–596
123. Klinge H (1973) Root mass estimation in lowland tropical rain forests of central Amazonia, Brazil. I. Fine root masses of a pale yellow latosol and a giant humus podzol. *Trop Ecol* 14:29–38
124. Klinge H, Herrera R (1978) Root biomass studies in Amazon caatinga forest in southern Venezuela. I. Standing crop of composite root mass in selected stands. *Trop Ecol* 19:93–110
125. Knoop WT, Walker BH (1985) Interactions of woody and herbaceous vegetation in a southern African savanna. *J Ecol* 73:235–253
126. Kochenderfer JN (1973) Root distribution under some forest types native to West Virginia. *Ecology* 54:445–448
127. Körner Ch, Renhardt U (1987) Dry matter partitioning and root length/leaf area ratios in herbaceous perennial plants with diverse altitudinal distribution. *Oecologia* 74:411–418
128. Kummerow J, Krause D, Jow W (1977) Root systems of chaparral shrubs. *Oecologia* 29:163–177
129. Kummerow J, Krause D, Jow W (1978) Seasonal changes of fine root density in the southern California chaparral. *Oecologia* 37:201–212
130. Kummerow J (1981) Structure of roots and root systems. In: Castri F di, Goodall DW, Specht RL (eds) *Mediterranean-Type Shrublands*, Elsevier, New York, pp 269–288
131. Kummerow J, Mangan R (1981) Root systems in *Quercus dumosa* Nutt. dominated chaparral in southern California. *Acta Oecol* 2:177–188
132. Kummerow J, Castellanos J, Maas M, Larigauderie A (1990) Production of fine roots and the seasonality of their growth in a Mexican deciduous dry forest. *Vegetatio* 90:75–80
133. Kummerow J, Kummerow M, Traub L (1990) Root biomass, root distribution and the fine-root dynamics of *Quercus coccifera* L. in the garrigue of southern France. *Vegetatio* 87:37–44
134. Kutschera L (1960) *Wurzelatlas Mitteleuropäischer Ackerunkräuter und Kulturpflanzen*. DLG, Frankfurt
135. Laitakari E (1929) The root system of pine (*Pinus silvestris*). A morphological investigation. *Acta For Fenn* 33:1–380
136. Lawson GW, Jenik J, Armstrong-Mensah KO (1968) A study of a vegetation catena in guinea savanna at Mole Game Reserve (Ghana). *J Ecol* 56:505–522
137. Lawson GW, Armstrong-Mensah KO, Hall JB (1970) A catena in tropical moist semi-deciduous forest near Kade, Ghana. *J Ecol* 58:371–398
138. Le Roux X, Bariac T, Mariotti A (1995) Spatial partitioning of the soil water resource between grass and shrub components in a West African humid savanna. *Oecologia* 104:147–155
139. Leaf AL, Leonard RE, Berglund JV (1971) Root distribution of a plantation-grown red pine in an outwash soil. *Ecology* 52:153–158
140. Lee CA, Lauenroth WK (1994) Spatial distributions of grass and shrub root systems in the shortgrass steppe. *Am Midl Nat* 132:117–123
141. Liang YM, Hazlett DL, Lauenroth WK (1989) Biomass dynamics and water use efficiencies of five plant communities in the shortgrass steppe. *Oecologia* 80:148–153
142. Low AB, Lamont BB (1990) Aerial and below-ground phytomass of *Banksia* scrub-heath at Eneabba, south-western Australia. *Aust J Bot* 38:351–359
143. Lugo AE (1992) Comparison of tropical tree plantations with secondary forests of similar age. *Ecol Monogr* 62:1–41
144. Lyr H, Hoffmann G (1967) Growth rates and growth periodicity of tree roots. *Int Rev For Res* 1:181–236
145. Mackie-Dawson LA, Atkinson D (1991) Methodology for the study of roots in field experiments and the interpretation of results. In: Atkinson D (ed) *Plant root growth: an ecological perspective*, Blackwell, Oxford, pp 25–47
146. Manning SJ, Barbour MG (1988) Root systems, spatial patterns, and competition for soil moisture between two desert subshrubs. *Am J Bot* 75:885–893
147. Markle MS (1981) Biomass and production of an aspen-mixed hardwood-spodosol ecosystem in northern Wisconsin. *Can J For Res* 11:132–138
148. Marshall JK (ed) (1977) *The belowground ecosystem: a synthesis of plant-associated processes*. Range Science Department science series (vol 26). Colorado State University, Fort Collins
149. Martínez F, Merino J, Martín Vicente A, unpublished. Biomass and root structure in a mediterranean shrub community
150. Martínez García F, Rodríguez JM (1988) Distribución vertical de las raíces del matorral de Doñana. *Lagascalia* 15:549–557
151. McClaugherty CA, Aber JD, Melillo JM (1982) The role of fine roots in the organic matter and nitrogen budgets of two forested ecosystems. *Ecology* 63:1481–1490
152. McKell CM, Jones MB, Perrier ER (1962) Root production and accumulation of root material on fertilized annual range. *Agron J* 54:459–462
153. McMinn RG (1963) Characteristics of Douglas fir root systems. *Can J Bot* 41:105–122
154. Mengel DB, Barber SA (1974) Development and distribution of the corn root system under field conditions. *Agron J* 66:341–344
155. Mensah KOA, Jeník J (1968) Root system of tropical trees. 2. Features of the root system of iroko. *Preslia* 40:21–27
156. Milchunas DG, Lauenroth WK (1989) Three-dimensional distribution of plant biomass in relation to grazing and topography in the shortgrass steppe. *Oikos* 55:82–86
157. Milchunas DG, Lauenroth WK, Chapman PL, Kazempour MK (1989) Effects of grazing, topography, and precipitation on the structure of a semiarid grassland. *Vegetatio* 80:11–23
158. Milchunas DG, Lee CA, Lauenroth WK, Coffin DP (1992) A comparison of C14, Rb86, and total excavation for determination of root distributions of individual plants. *Plant Soil* 144:125–132
159. Milchunas DG, Lauenroth WK (1993) Quantitative effects of grazing on vegetation and soils over a global range of environments. *Ecol Monogr* 63:327–366
160. Miller PC, Ng E (1977) Root:shoot biomass ratios in shrubs in southern California and Central Chile. *Madroño* 24:215–223
161. Miller PC, Mangan R, Kummerow J (1982) Vertical distribution of organic matter in eight vegetation types near Eagle Summit, Alaska. *Hol Ecol* 5:117–124
162. Moir WH, Bachelard EP (1969) Distribution of fine roots in three *Pinus radiata* plantations near Canberra, Australia. *Ecology* 50:658–662
163. Molz FJ (1971) Interaction of water uptake and root distribution. *Agron J* 63:608–610
164. Montaña C, Cavagnaro B, Briones O (1995) Soil water use by co-existing shrubs and grasses in the southern Chihuahuan Desert, Mexico. *J Arid Environ* 31:1–13

165. Moore RT, West NE (1973) Distribution of *Galleta* roots and rhizomes at two Utah sites. *J Range Manage* 26:34–36
166. Moorhead DL, Reynolds JF, Fonteyn PJ (1989) Patterns of stratified soil water loss in a Chihuahuan desert community. *Soil Sci* 148:244–249
167. Mortimer SR (1992) Root length/leaf area ratios of chalk grassland perennials and their importance for competitive interactions. *J Veg Sci* 3:665–672
168. Murphy PG, Lugo AE (1986) Structure and biomass of a subtropical dry forest in Puerto Rico. *Biotropica* 18:89–96
169. Murphy PG, Lugo AE, Murphy AJ, Nepstad DC (1995) The dry forests of Puerto Rico's south coast. In: Lugo AE, Lowe C (eds) *Tropical forests: management and ecology*. Springer, New York, pp 178–209.
170. Nepstad DC, Carvalho CR de, Davidson EA, Jipp PH, Lefebvre PA, Negreiros GH, Silva ED da, Stone TA, Trumbore SE, Vieira S (1994) The role of deep roots in the hydrological and carbon cycles of Amazonian forests and pastures. *Nature* 372:666–669
171. Newman EI (1966) A method for estimating the total length of root in a sample. *J Appl Ecol* 3:139–145
172. Nobel PS (1989) Temperature, water availability, and nutrient levels at various soil depths – consequences for shallow-rooted desert succulents, including nurse plant effects. *Am J Bot* 76:1486–1492
173. Nobel PS (1991) Ecophysiology of roots of desert plants, with special emphasis on agaves and cacti. In: Waisel AEY, Kafkafi U (eds) *Plant roots: the hidden half*. Marcel Dekker, New York, pp 839–866
174. Nobel PS, Huang B, Garcia-Moya E (1993) Root distribution, growth, respiration, and hydraulic conductivity for two highly productive agaves. *J Exp Bot* 44:747–754
175. Okali DUU, Hall JB, Lawson GW (1973) Root distribution under a thicket clump on the Accra Plains, Ghana: its relevance to clump localization and water relations. *J Ecol* 61:439–454
176. Old SM (1969) Microclimate, fire, and plant production in an Illinois prairie. *Ecol Monogr* 39:355–384
177. Ovington JD, Heitkamp D, Lawrence DB (1963) Plant biomass and productivity of prairie, savanna, oakwood, and maize field ecosystems in central Minnesota. *Ecology* 44:52–63
178. Pandey CB, Singh JS (1992) Influence of rainfall and grazing on belowground biomass dynamics in a dry tropical savanna. *Can J Bot* 70:1885–1890
179. Pastor J, Bockheim JG (1981) Biomass and production of an aspen-mixed hardwood-spodosol ecosystem in northern Wisconsin. *Can J For Res* 11:132–138
180. Pavlychenko TK (1937) Quantitative study of the entire root systems of weed and crop plants under field conditions. *Ecology* 18:62–69
181. Persson H (1975) Deciduous woodland at Andersby, eastern Sweden: field-layer and below-ground production. *Act Phytogeogr Suec* 62:1–72
182. Persson H (1979) Fine-root production, decomposition, and mortality in forest ecosystems. *Vegetatio* 41:101–109
183. Persson H (1980) Spatial distribution of fine-root growth, mortality and decomposition in a young Scots pine stand in Central Sweden. *Oikos* 34:77–87
184. Persson H (1982) Changes in the tree and dwarf shrub fine-roots after clear-cutting in a mature scots pine stand. Swedish Coniferous Forest Project. Report Number 31
185. Persson H (1983) The distribution and productivity of fine roots in boreal forests. *Plant Soil* 71:87–101
186. Persson H, Fircks Y von, Majdi H, Nilsson LO (1995) Root distribution in a Norway spruce (*Picea abies* (L.) Karst.) stand subjected to drought and ammonium-sulphate application. *Plant Soil* 168–169:161–165
187. Price SR (1911) The roots of some North African desert grasses. *New Phytol* 10:328–340
188. Reinhardt DR, Miller RM (1990) Size classes for root diameter and mycorrhizal fungal colonization in two temperate grassland communities. *New Phytol* 116:129–136
189. Reynolds ERC (1970) Root distribution and the cause of its spatial variability in *Pseudotsuga taxifolia* (Poir.) Britt. *Plant Soil* 32:501–517
190. Richards JH (1986) Root form and depth distribution in several biomes. In: Carlisle D, Berry WL, Kaplan IR, Wattersson JR (eds) *Mineral exploration: biological systems and organic matter*. Prentice-Hall, Englewood Cliffs, pp 82–97
191. Risser PG, Birney EC, Blocker HD, May SW, Parton WJ, Wiens JA (eds) (1981) *The true prairie ecosystem*. (US/IBP synthesis series, vol 16). Hutchinson Ross, Stroudsburg
192. Rodin LE, Basilevich NI (1966) The biological productivity of the main vegetation types in northern hemisphere of the Old World. *For Abstr* 27:369–372
193. Rodin LE, Basilevich NI (1967) Production and mineral cycling in terrestrial vegetation. Oliver and Boyd, Edinburgh
194. Rowe JS, Action DF (1985) Taproots of jack pine and soil tongues in Saskatchewan. *Can J For Res* 15:646–650
195. Ruark GA, Bockheim JG (1988) Biomass, net primary production, and nutrient distribution for an age sequence of *Populus tremuloides* ecosystems. *Can J For Res* 18:435–443
196. Rundel PW, Nobel PS (1991) Structure and function in desert root systems. In: Atkinson D, Plant root growth: an ecological perspective. Blackwell, Oxford, pp 349–378
197. Safford LO (1974) Effect of fertilization on biomass and nutrient content of fine roots in a beech-birch-maple stand. *Plant Soil* 40:349–363
198. Sainju UM, Good RE (1993) Vertical root distribution in relation to soil properties in New Jersey Pinelands forest. *Plant Soil* 150:87–97
199. Sanford RL Jr (1985) Root ecology and successional Amazon forests. Ph D, University of California, Berkeley
200. Schneider BU, Meyer J, Schulze E-D, Zech W (1989) Root and mycorrhizal development in healthy and declining Norway spruce stands. In: Schulze E-D, Oren R (eds) *Forest decline and air pollution*. Springer Berlin Heidelberg New York, pp 370–391
201. Scholes RJ, Walker BH (eds) (1993) *An African savanna*. Cambridge University Press, Cambridge
202. Schulze E-D, Hantschel R, Werk KS, Horn R (1989) Water relations of two Norway spruce stands at different stages of decline. In: Schulze E-D, Oren R. *Forest decline and air pollution*. Springer Berlin Heidelberg New York, pp 341–351
203. Schulze E-D, Bauer G, Buchmann N, Canadell J, Ehleringer JR, Jackson RB, Jobbagy E, Loreti J, Mooney HA, Oesterheld M, Sala OE (1996) Water availability, rooting depth, and vegetation zones along an aridity gradient in Patagonia. *Oecologia*, in press
204. Scully NJ (1942) Root distribution and environment in a maple-oak forest. *Bot Gaz* 103:492–517
205. Shaver GR, Billings WD (1975) Root production and root turnover in a wet tundra ecosystem, Barrow, Alaska. *Ecology* 56:401–409
206. Siddique KHM, Belford RK, Tennant D (1990) Root:shoot ratios of old and modern, tall and semi-dwarf wheats in a mediterranean environment. *Plant Soil* 121:89–98
207. Sims PL, Singh JS (1978) The structure and function of ten western North American grasslands. II. Intra-seasonal dynamics in primary producer compartments. *J Ecol* 66:547–572
208. Singh JS, Coleman DC (1973) A technique for evaluating functional root biomass in grassland ecosystems. *Can J Bot* 51:1867–1870
209. Singh JS, Coleman DC (1977) Evaluation of functional root biomass and translocation of photoassimilated C14 in a short-grass prairie ecosystem. In: Marshall JK (ed) *The below-ground ecosystem: a synthesis of plant-associated processes*. Colorado State University Press, Fort Collins, pp 29–37
210. Smith P, Every L (1980) Rooting habits of selected commercial tree species of the eastern United States (Bibliographies and literature 10). USDA Forest Service, Washington
211. Soriano A, Golluscio RA, Satorre E (1987) Spatial heterogeneity of the root system of grasses in the Patagonian arid steppe. *Bull Torrey Bot Club* 114:103–108

212. Specht RL, Rayson P (1957) Dark Island heath (Ninety-mile Plain, South Australia). III. The root systems. *Aust J Bot* 5:103–114
213. Sperry TM (1935) Root systems in Illinois prairie. *Ecology* 16:178–202
214. Stark N, Spratt M (1977) Root biomass and nutrient storage in rain forest oxisols near San Carlos de Rio Negro. *Trop Ecol* 18:1–9
215. Stone EL, Kalisz PJ (1991) On the maximum extent of tree roots. *For Ecol Manage* 46:59–102
216. Strong WL, La Roi GH (1983) Rooting depths and successional development of selected boreal forest communities. *Can J For Res* 13:577–588
217. Strong WL, La Roi GH (1983) Root-system morphology of common boreal forest trees in Alberta, Canada. *Can J For Res* 13:1164–1173
218. Strong WL, La Roi GH (1985) Root density-soil relationships in selected boreal forests of central Alberta, Canada. *For Ecol Manage* 12:233–251
219. Sturges DL (1977) Soil water withdrawal and root characteristics of big sagebrush. *Am Midl Nat* 98:257–274
220. Sturges DL (1980) Soil water withdrawal and root distribution under grubbed, sprayed, and undisturbed big sagebrush vegetation. *Great Basin Nat* 40:157–164
221. Tabler RD (1964) The root system of *Artemisia tridentata* at 9,500 feet in Wyoming. *Ecology* 45:633–636
222. Taylor HM, Klepper B (1973) Rooting density and water extraction patterns for corn (*Zea mays* L.). *Agron J* 65:965–968
223. Tesarova M, Fiala K, Studeny V (1982) Live and dead roots – their mass ratio in several grassland stands. *Fol Geobot Phytotax* 17:427–430
224. Thomas CM, Davis SD (1989) Recovery patterns of three chaparral shrub species after wildfire. *Oecologia* 80:309–320
225. Titlyanova A, Rusch G, Van Der Maarel E (1988) Biomass structure of limestone grasslands on dland in relation to grazing intensity. *Act Phytogeogr Suec* 76:125–134
226. Turner LM (1936) A comparison of roots of southern short-leaf pine in three soils. *Ecology* 17:649–658
227. Ukpong IE (1992) The structure and soil relations of *Avicennia* mangrove swamps in southeastern Nigeria. *Trop Ecol* 33:1–16
228. Ulrich B (1986) Berichte des Forschungszentrum Waldökosysteme/Waldsterben, Reihe B, Band 2. In: Ulrich B (ed) *Raten der Deposition, Akkumulation und des Austrags toxischer Luftverunreinigungen als Maß der Belastung und Belastbarkeit von Waldökosystemen*.
229. USDA (1981) Root characteristics of some important trees of eastern forests: a summary of the literature. USDA Forest Service, Washington
230. Van Rees KCJ, Comerford NB (1986) Vertical root distribution and strontium uptake of a slash pine stand on a Florida spodosol. *Soil Sci Soc Am J* 50:1042–1046
231. Vance ED, Nadkarni NM (1992) Root biomass distribution in a moist tropical montane forest. *Plant Soil* 142:31–39
232. Veresoglou DS, Fitter AH (1984) Spatial and temporal patterns of growth and nutrient uptake of five coexisting grasses. *J Ecol* 72:259–272
233. Virginia RA, Jarrell WM, Whitford WG, Freckman DW (1992) Soil biota and soil properties associated with the surface rooting zone of mesquite (*Prosopis glandulosa*) in historical and recently desertified habitats. *Biol Fert Soils* 14:90–98
234. Vogt KA, Moore EE, Vogt KA, Redlin MR, Edmonds RL (1983) Conifer fine root and mycorrhizal root biomass within the forest floors of Douglas fir stands of different ages and site productivities. *Can J For Res* 13:429–437
235. Vogt KA, Grier CC, Vogt DJ (1986) Production, turnover, and nutrient dynamics of above- and below-ground detritus of world ecosystems. *Adv Ecol Res* 15:303–377
236. Vogt KA, Vogt DJ, Moore EE, Fatuga BA, Redlin MR, Edmonds RL (1987) Conifer and angiosperm fine-root biomass in relation to stand age and site productivity in Douglas Fir forests. *J Ecol* 75:857–870
237. Waisel Y, Eshel A, Kafakafi U, (ed) (1991) *Plant roots: the hidden half*. Marcel Dekker, New York
238. Wallace A, Bamberg SA, Cha JW (1974) Quantitative studies of roots of perennial plants in the Mojave Desert. *Ecology* 55:1160–1162
239. Wallace A, Romney EM, Cha JW (1980) Depth distribution of roots of some perennial plants in the Nevada test site area of the northern Mojave Desert. *Great Basin Nat Mem* 4:201–207
240. Watts SE (1993) Rooting patterns of co-occurring woody plants on contrasting soils in a subtropical savanna. MS dissertation, Texas A&M University, College Station
241. Weaver JE (1919) The ecological relations of roots (publication 286). Carnegie Institution, Washington
242. Weaver JE, Kramer J (1932) Root system of *Quercus macrocarpa* in relation to the invasion of prairie. *Bot Gaz* 94:51–85
243. Weaver JE, Darland RW (1949) Soil-root relationships of certain native grasses in various soil types. *Ecol Monogr* 19:303–338
244. Weaver JE (1954) North American prairie. Donnelley, Chicago
245. Weaver T (1977) Root distribution and soil water regimes in nine habitat types of the northern Rocky Mountains. In: Marshall JK (ed) *The belowground ecosystem: a synthesis of plant-associated processes*. Colorado State University Press, Fort Collins, pp 239–244
246. Webber PJ, May DE (1977) The magnitude and distribution of belowground plant structures in the alpine tundra of Niwot Ridge, Colorado. *Arct Alp Res* 9:157–174
247. Westman WE, Rogers RW (1977) Biomass and structure of a subtropical eucalypt forest, North Stradbroke Island. *Aust J Bot* 25:171–191
248. Whittaker RH, Bormann FH, Likens GE, Siccama TG (1974) The Hubbard Brook ecosystem study: forest biomass and production. *Ecol Monogr* 44:233–254
249. Wilhelm WW, Mielke LN, Fenster CR (1982) Root development of winter wheat as related to tillage practice in western Nebraska. *Agron J* 74:85–88
250. Wright TW (1955) Profile development in the sand dunes of Culbin Forest, Morayshire. *J Soil Sci* 6:270–283
251. Yamaguchi J, Tanaka A (1989) Root profiles of some native and exotic plant species in southeastern Idaho. *Environ Exp Bot* 29:241–248
252. Yin X, Perry JA, Dixon RK (1989) Fine-root dynamics and biomass distribution in a *Quercus* ecosystem following harvesting. *For Ecol Manage* 27:159–177
253. Zohary M (1961) On the hydro-ecological relations of the near east desert vegetation. *UNESCO Arid Zone Res* 16:198–212

Appendix 2

References meeting criteria for inclusion in analysis of root depth distributions (listed in Appendix 1)

Vegetation type	Reference	Specifics	Location	Coordinates	Annual precip.	Soil type texture	Root type	Method	Measurement	Other
Boreal coniferous forest	Persson 1982	Table 2	Central Sweden	60:49 N 16:30 E	607 mm		Fine and coarse	Monolith	$\text{g} \cdot \text{m}^{-2}$ to 60 cm	<i>Pinus sylvestris</i> stand
	Persson et al. 1995	Table 1	SW-Sweden	56:33 N 13:13 E		Haplic podzol	Live and dead fine roots	Monolith	$\text{g} \cdot \text{m}^{-2}$ to 100 cm	<i>Picea abies</i> stand
	Strong & La Roi 1983 & 3	Figs. 2 & 3	Alberta, Canada		475 mm	Sandy and Eutric Brunisols	Total	Excavated soil pits	# roots dm^{-2} to 140 cm	Four boreal forest stands (<i>Larix</i> , <i>Picea</i> , <i>Pinus</i> , and <i>Populus</i>)
	Strong & La Roi 1985	Table 1	Alberta, Canada			Eutric Brunisolic or Gray Luvisolic	Total (5 diam classes)	Profile face	# roots to 115 cm	<i>Populus</i> , <i>Pinus</i> , and <i>Picea</i>
Crops	Armstrong et al. 1994	Fig. 5	Wongan Hills, Western Australia	30:51 S 116:43 E			Nodulated roots	10 cm soil cores	$\text{g} \cdot \text{m}^{-2}$ to 160 cm	6 field pea genotypes <i>Pisum sativum</i>
	Gäth et al. 1989	Fig. 1	25 sites in Germany			Silty-loam and sandy	Total	Soil core and profile methods	Length density (cm cm^{-3}) to 70 cm	Cereal crops
	Huck et al. 1986	Table 1	Alabama, USA			Marvyn loamy sand	Total	Excavation	$\text{g} \cdot \text{m}^{-2}$ to 180 cm	Soybeans (<i>Glycine max</i>)
	Siddique et al. 1990	Fig. 2	Merredin, W-Australia	31:29 S 118 E		Duplex profile of grey sand over sandy clay	Total	10 cm soil cores	$\text{g} \cdot \text{m}^{-3}$ and $\text{cm} \cdot \text{cm}^{-3}$ to 80 cm	Wheat (<i>Hordeum vulgare</i>)
	Taylor & Klepper 1973	Table 2	Alabama, USA			Cahaba loamy fine sand	Total	Glass wall	Root density to 180 cm	Corn (<i>Zea mays</i>)
	Wilhelm et al. 1982	Fig. 1	Nebraska, USA			Alliance silt loam	Total	Hydraulic probe 7.6-cm-diam. core	$\text{mg} \cdot \text{dm}^{-3}$ to 120 cm	Wheat (<i>Hordeum vulgare</i>)
Desert	Barbour et al. 1977	Fig. 9-6	Arizona, USA				Total	Unknown	kg/ha to 100 cm	Data for three shrub species and in the open
	Branson et al. 1976	Fig. 19	Colorado, USA		<230 cm	Shallow weathered mantle over bedrock	Total	50 cm^2 soil samples	$\text{g} \cdot \text{dm}^{-3}$ to 180 cm	Data for 12 communities
	Dobrowolski et al. 1990	Fig. 7.8	Utah, USA	41:45 N 111:48 W	468 mm	Rocky Mollisols formed on alluvial fan material	Total	Profile wall mapping	Intersections m^{-2} to 2.5 m	Data for big sagebrush and crested wheatgrass
	Fernandez & Caldwell 1975	Table 1	Utah, USA	41:05 N 113:05 W	230 mm	Lacustrine, silty loams with high salinity	Total	Root observation chambers	Intersections m^{-2} to 60 cm	Three shrub species in two size classes

Appendix 2 (continued)

Vegetation type	Reference	Specifics	Location	Coordinates	Annual precip.	Soil type texture	Root type	Method	Measurement	Other
Desert	Freckman & Virginia 1989	Fig. 1	New Mexico, USA	32:30 N 106:45 W	211 mm	Haplargid, Torrifluent, or Torripsamment	Total	Drilling system, 6.5 cm core	Root fresh mass, $\text{mg} \cdot \text{kg}^{-1}$	Five Jornada sites, <i>Larrea</i> and <i>Prosopis</i>
	Jordan & Nobel 1984	Fig. 1	California, USA	33:38 N 116:24 W			Total	Monoliths	Length per soil volume ($\text{micro-m} \cdot \text{mm}^{-3}$) to 150 cm	Data for 2 succulent species
	Montana et al. 1995	Fig. 2	Durango, Mexico	26:40 N 103:40 W	264 mm	Haplic Yermosol	Total	Trench wall	# of roots to 70 cm	Three shrubs, one grass
	Moorhead et al. 1989	Fig. 1	New Mexico, USA			Calciorthid and Typic Haplargids	Fine	Soil pit	$\text{g} \cdot \text{m}^{-2}$ to 70 cm	Creosote community
	Nobel 1989	Fig. 1	California, USA	33:38 N 116:24 W			Total	Unknown	Length (% of total) to 30 cm	Data for 3 succulent species
Miscellaneous	Rundel & Nobel 1991	Fig. 13	New Mexico, USA				Total	Unknown to 12 m	$\text{mg fw} \cdot \text{kg}^{-1}$ to 30 cm	<i>Prosopis glandulosa</i>
	Sturges 1980	Fig. 4	Wyoming, USA		500 mm	Developed from sandstone, Argic Cryoboroll subgroup	Total	7.6 cm soil cores	Water depletion and root weight to 122 cm	<i>Artemisia tridentata</i>
	Beese 1986	Table 2.32	Germany	57:13 N 5:65 E	600 mm	Parabrownearth		Harvest		<i>Avena sativa</i>
	Bernard & Fiala 1986	Table 1	New York, USA			Mineral and peat soils	Total, live/dead	Monoliths	$\text{g} \cdot \text{m}^{-2}$ to 20 cm	3 <i>Carex</i> species, wet meadow
	Håland & Brække 1989	Table 2	Ekebergmosen, Trøgstad, Norway	59:38 N 11:14 E		Peat layer over sandy marine shore deposits	Fine/small (<10 mm)	56 mm soil cores	$\text{g} \cdot \text{m}^{-2}$ to 40 cm	pine bog
	Richards 1986	Fig. 5-3	Various	Varied	Varied	Varied	Total	Varied	$\text{g} \cdot \text{m}^{-2}$ to various depths	All but Wallace et al. 1980 recorded elsewhere
	Weaver 1977	Table 4	Montana, USA	Varied (all within 30 km of Bozeman)	338-909 mm	Varied	Live feeder root (<5 mm)	2 cm soil cores to 70 cm	$\text{g} \cdot \text{m}^{-2}$	Various grass and shrub spp.
	Webber & May 1977	Fig. 3	Colorado, USA	40:03 N 105:36 W		Coarse with thin organic-rich surface horizons, often with loess fraction	Live/dead	5 x 5 cm soil monoliths	$\text{g} \cdot \text{m}^{-2}$ to 100 cm	Alpine tundra

Appendix 2 (continued)

Vegetation type	Reference	Specifics	Location	Coordinates	Annual precip.	Soil type texture	Root type	Method	Measurement	Other
Sclero-phyllous shrubland	Canadell & Roda 1991	Table 5	NE-Spain		870 mm	Dystic Xerochrepts sandy-loams	Fine	4 cm diam. cores	tons/ha to 60 cm	<i>Quercus ilex</i>
	Chapman 1970	Fig. 1	Dorset, England			Well-developed humus iron podzols	Total	9 cm soil cores	kg/ha to 40 cm	Dry heath
	Higgins et al. 1987	Table 5	Cape Province, South Africa	33:57 S 18:55 S	1700 mm	See Table 1 of article	Total	Water jets	% root mass by depth	Fynbos
	Kummerow et al. 1977	Table 3	California, USA	32:54 N 116:39 W	550 mm	Sandy loam, clay, and decomposing granite	Total	Plant excavations	$g \cdot 70 m^{-2}$ to 60 cm	Data for 5 species
	Kummerow & Mangen 1981	Table IV	California, USA		460 mm	Sandy and clay loam	Total, fine for 1 species	Plant excavations and soil cores	$g \cdot m^{-2}$ to 80 cm; fine roots ($g \cdot dm^{-3}$) to 40 cm	Data for 5 species
	Kummerow et al. 1990	Fig. 2	Montpelier, France		900 mm	Rich, loamy soil 30–50 cm deep, underlain by cracks with sandy loam	Total	Trenches	% roots to 1 m	<i>Quercus coccifera</i>
	Low & Lamont 1990	Table 3	Enaebba, SW Australia	29:52 S 115:15 E	530 mm	Podsolized Sand, acidic	Total	Excavation	$g \cdot m^{-2}$ to 180 cm	<i>Banksia</i> scrub heath
	Martinez et al., unpublished work	Table 1	SW Spain		620 mm	Dystic Quaeztipsammit	Total	20-cm-diam. cores	$g \cdot m^{-2}$ to 100 cm	Mediterranean shrub
	Martinez Garcia & Rodriguez 1988	Table 1	SW Spain		620 mm	Dystic Quaeztipsammit	Total	20-cm-diam. cores	$g \cdot m^{-2}$ to 100 cm	Matorral
	Miller & Ng 1977	Table 3	California, USA	32:54 N 116:39 W	550 mm	Sandy loam, underlain by decomposed granite at CA site	Total	Plant excavations	$g \cdot m^{-3}$ to 1 m	Chaparral shrubs
Temperate conifer	Specht & Rayson 1957	Fig. 10	Ninety-Mile Plain, South Australia		457 mm	Deep, acid sand	Total	Excavations	1000 kg/3 in. depth/acre to 6 feet	25-year-old heath stands
	Ares & Peinemann 1992	Table 7	Buenos Aires, Argentina			Primarily Mollisols	Fine (<2 mm)	7 cm soil cores and monoliths	kg/ha to 50 cm	Plantations (<i>Pinus</i> , <i>Cedrus</i> & <i>Cupressus</i>)
	Gehrman et al. 1984	Fig. 6	Germany	57:52 N 5:50 E		Podsol	Fine (<2 mm)	Root cores		<i>Picea abies</i> plantation
	Harris et al. 1977	Table 3	Tennessee, USA		1390 mm	Typic Paleudults	Total	Excavation and soil cores	kg/ha to 60 and 70 cm	<i>Pinus taeda</i>
			North Carolina, USA		1160 mm	Typic Hapludults				

Appendix 2 (continued)

Vegetation type	Reference	Specifics	Location	Coordinates	Annual precip.	Soil type texture	Root type	Method	Measurement	Other
Temperate deciduous forest	McClaugherty et al. 1982	Table 1	Harvard Forest, USA			Entic Haplorthods (Spodosol), very stony	Fine: live/dead	19 mm and 50 mm	Mg/ha to depth of soil cores rooting zone (0.6–1.2 m)	<i>Pinus resinosa</i> (53-yr old)
	Reynolds 1970	Table 4	Oxford, England			coarse sand or sandy loam	Total	6 cm diam cores	kg · m ⁻² to 107 cm	36-yr old Douglas Fir <i>Pseudotsuga taxifolia</i>
	Ulrich 1986	Abb. 23	Germany	57:52 N 5:50 E		Podsol	Fine (<2 mm)	Root cores		<i>Picea abies</i>
	Van Rees & Comerford 1986	Table 2	Florida, USA		1330 cm	Sandy, Ultic Haplaquads	Total	10 cm soil cores	g · m ⁻² for all species	<i>Pinus elliotii</i> to 245 cm
	Wright 1955	Fig. 1	Morayshire, Scotland		607 mm	Coarse and fine sand	Total	6-inch cubes (216 in ³)	g in ⁻³ to approx. 5 feet	Dune, Corsican pine, Scots pine, and birch
	Farrish 1991	Tables 2, 4	Louisiana, USA	32 N 92 W		Upland: Fine loamy, siliceous, thermic Typic Paleudults; Bottomland: fine-silty, thermic Typic Glossaqualfs	Live, fine	8 cm soil cores	Mass (mg · cm ⁻³) and surface area (cm ² · cm ⁻³) to 90 cm (upland) and 100 cm (bottomland)	Bottomland hardwood forest
	Harris et al. 1977	Table 3	Tennessee, USA		1390 mm	Fullerton and Bodine	Total	Excavation and soil cores	kg/ha to 60 and 70 cm	Mixed deciduous forest
	Kelly & Joslin 1989	Table 2	Tennessee, USA		1160 mm	(typic paleudults); Granville series (typic hapludults)	Total	10 cm soil cores	ton/ha to 50 cm	<i>Quercus coccinea</i>
						Hapludults (derived from weathered sandstone and siltstone)				
	Kochenderfer 1973	Table 1	West Virginia, USA		1300 mm	Various silt loams	Total	Strip-mine high walls and road cuts	% total root endings to 2.1 m	3 forest types: northern hardwood, cove hardwood, and oak-hickory
	McClaugherty et al. 1982	Table 1	Harvard Forest, USA			Entic Haplorthods (Spodosol), very stony	Fine: live/dead	19 mm and 50 mm	Mg/ha to depth of soil cores rooting zone (0.6–1.2 m)	Mixed hardwood stand
	Schulze et al. 1996		Patagonia, Argentina		770 mm		Total	Monoliths	g · m ⁻² to 200 cm	<i>Nothofagus pumila</i>
					522 mm		Total	Monoliths	g · m ⁻² to 225 cm	<i>Nothofagus antarctica</i>
	Scully 1942	Table 2	Wisconsin, USA		800 mm	Bellefontaine silt loam	Total	Trenches	# of roots ft ⁻² ; % root area ft ⁻² to 3 ft. (1 ft. increments)	Maple-Oak forest

Appendix 2 (continued)

Vegetation type	Reference	Specifics	Location	Coordinates	Annual precip.	Soil type texture	Root type	Method	Measurement	Other
Temperate grassland	Yin et al. 1989	Fig. 1	Wisconsin, USA	44:06 N 91:12 W	792 mm	Typic Hapludalf, loam and silt loam	Fine	10 cm soil cores	% biomass to 60 cm	<i>Quercus</i> ecosystem
	Dahlman & Kucera 1965	Table 1	Missouri, USA		1016	Fine loess with claypan subsoil	Total	1.65 inch soil cores	$g \cdot m^{-2}$ to 34 inches	Central Missouri Prairie
	Dumortier 1991	Fig. 1	Bourgoyen Ossemeersen, Belgium	51:06 N 3:40 E		Humuficuous upper layer and clay	Total	8.2 cm soil cores	$g \cdot m^{-2}$ to 100 cm for two plots	Two hayfields
	Fernández & Paruelo 1988	Fig. 5	Chubut, Argentina	45:25 S 70:20 W	142 mm	Calcicorthid with high gravel content	Total	Excavation	Root length (cm per plant) to 120 cm	<i>Mulinum</i> and <i>Senecio</i> Two shrub species
	Lee & Lauenroth 1994	Fig. 2	Colorado, USA	40:49 N 104:47 W	321 mm	Sandy clay loam	Total	Monolith	to 110 cm	Shortgrass steppe
	Liang et al. 1989	Fig. 2	Colorado, USA	40:49 N 104:46 W	311 mm	Sandy loam or clay loam	Fine	5 cm soil cores	$g \cdot m^{-2}$ to 90 cm	Shortgrass steppe
	McKell et al. 1962	Fig. 2	California, USA		889 mm	Sutherland fine gravelly clay loam	Macro organic matter	2.37 inch soil cores	$g \cdot ft^{-2}$ to 24 inches	Unimproved annual grassland
	Old 1969	Table 8	Illinois, USA		910 mm	Mollisol or Alfisol	Total	8 cm soil cores	$g \cdot m^{-2}$ to 100 cm	Tall grass prairie, <i>Andropogon</i> spp.
	Schulze et al. 1996		Patagonia, Argentina		290 mm		Total	Monolith	$g \cdot m^{-2}$ to 200 cm	Patagonia grassland
	Sims & Singh 1978	Table 2	Bridger (Montana)	45:57 N 110:47 W	900 mm	Silt loam, stony	Total	See Sims et al. 1978	$g \cdot m^{-2}$ to 60 cm	Montana grassland
			Cottonwood (South Dakota)	43:57 N 101:52 W	400 mm	Silty clay loam				South Dakota grassland
			Dickinson (North Dakota)	46:54 N 102:49 W	400 mm	Loamy fine sand				North Dakota grassland
			Hays (Kansas)	38:52 N 99:23 W	600 mm	Loam, shallow bedrock				Kansas grassland
			Jornada (New Mexico)	32:36 N 106:51 W	250 mm	Loamy fine sand, caliche				New Mexico grassland
			Osage (Oklahoma)	36:57 N 96:33 W	900 mm	Silty clay				Oklahoma grassland
			Pantex (Texas)	35:18 N 101:32 N	500 mm	Silty clay loam				Texas grassland
			Pawnee (Colorado)	40:49 N 104:46 W	300 mm	Fine sandy loam				Colorado grassland
	Singh & Coleman 1977	Table 2	Colorado, USA	40:49 N 104:46 W	300 mm	Fine sandy loam	Live/dead	4.5 cm soil cores	$g \cdot m^{-2}$ to 60 cm	Shortgrass prairie
	Weaver 1954	p. 163	Nebraska, USA		580–840 mm	Silty clay-loam and silt-loam	Total	Soil monoliths	% biomass to 5 feet (see Weaver and Darland 1949)	<i>Andropogon</i> , <i>Bouteloua</i> , and <i>Smittii</i>

Appendix 2 (continued)

Vegetation type	Reference	Specifics	Location	Coordinates	Annual precip.	Soil type texture	Root type	Method	Measurement	Other
Tropical deciduous	Bang-xing 1991	Table 4	Yunnan, China	21:44 N 100:40 E	1515–1606 mm		Fine	Unknown	$\text{g} \cdot \text{cm}^{-2}$ to 150 cm	Seasonal rainforest
	Castellanos et al. 1991	Fig. 1	Chamela, Mexico	19:30 N, 2 km east of Pacific coast	707 mm	Deep sandy loam	Total/coarse	0.5 m \times 2 m excavated trenches	$\text{kg} \cdot \text{m}^{-2}$ to 80 cm	Chamela deciduous forest
	Lawson et al. 1970	Fig. 14	Kade, Ghana	06:09 N 0:55 W	1650 mm	Reddish yellow latosols consisting of silty clay over sandy clay	Total (by size), fine	25 \times 25 \times 10 cm soil monoliths	$\text{g} \cdot 10000 \text{ cm}^{-3}$	<i>Celtis</i> , <i>Triplochiton</i>
Tropical evergreen	Berish 1982	Table 1	Florencia Norte Forest, Costa Rica	9:53 N 83:40 W	2700 mm	Typic Dystrandept	Total (minus large dead roots >2 mm)	4.2 cm soil cores, 25 \times 25 cm soil blocks	$\text{g} \cdot \text{m}^{-2}$ to 85 cm, fine root surface area to 85 cm	Successional forest
	Gower 1987	Table 1	La Selva, Costa Rica	10:26 N 83:59 W	3800 mm	Fluvaquentic Hapludoll (River site) and Oxic Dystrandept (Arboleada site)	Fine: live/total (up to 5 mm)	7 cm soil cores	$\text{g} \cdot \text{m}^{-2}$ to 50 cm	La Selva forest
	Greenland & Kowal 1960	Table 8	Ghana		1650 mm	Oxysols or ochrosols	Total	4-cm-diam. cores	to 150 cm	<i>Diospyros</i> , <i>Strombosia</i>
	Huttl 1975	Fig. 10-3	Ivory Coast		Banco: 2100 mm Yapo: 1800 mm	Sandy with high clay and silt content	Total	Soil cores, unearthing roots	$\text{g} \cdot \text{dm}^{-3}$ to 130 cm	<i>Diospyros</i> , <i>Mapania</i>
	Klinge 1973	Tables 1, 4	Central Amazonia, Brazil			Pale yellow latosol (loamy), humus podzol (sandy)	Fine	1 m soil pits	kg/ha and length to 18 and 40 cm	Lowland forest
	Klinge & Herrera 1978	Table 3	Southern Venezuela			Spodosols		Excavation	kg/ha to approximately 60 cm	Amazon Caatinga, <i>Micrandra</i>
	Mensah & Jenik 1968	Figs. 4, 5, 6	Kade, Ghana	06:0:20 N 0:45 W			Total, fine	Soil monoliths	$\text{g} \cdot 6250 \text{ cm}^{-3}$	<i>Chlorophora excelsa</i>
	Nepstad et al. 1994	Fig. 2	Para, Brazil	06:09 N 0:55 W	1750 mm	deeply weathered clay soils	Fine	Auger borings	$\text{mg} \cdot \text{cm}^{-3}$ to 6 m	forest and adjacent pasture
	Vance & Nadkarni 1992	Table 3	Monteverde, Costa Rica	10:18 NN 84:48 W	2000 mm	Typic Dystrandept	Live: total/fine	10 cm soil cores, 1 m 2 excavated pits	$\text{g} \cdot \text{m}^{-2}$ to 180 cm	Monteverde cloud forest

Appendix 2 (continued)

Vegetation type	Reference	Specifics	Location	Coordinates	Annual precip.	Soil type texture	Root type	Method	Measurement	Other
Tropical Grassland/Savanna	Fiala & Herrera 1988	Tables 1, 4	Cuba	22:15 N 80:41 W 21:38 N 82:59 W 22:53 N 82:53 W 22:59 N 82:23 W	1000–1500 mm 1165–1795 mm 2013 mm 1600–1800 mm	fine deep siliceous gleyed coarse sands fine sandy loam ferrallitic red clay	Total, Live/dead	10×10 cm soil monoliths	% biomass to 50 cm	<i>Byrsonima</i> <i>Andropogonetum</i> <i>Phyllantho-</i> <i>Aristidetum</i> <i>Axonopus compressus</i> <i>Panicum maximum</i>
	Heitschmidt et al. 1988	Fig. 5	Texas, USA		650 mm	Typic Paleustoll	Total	4 m-wide by 2 m-deep profile face	# roots	Texas savanna <i>Prosopis glandulosa</i>
	Knoop & Walker 1985	Fig. 1	South Africa	25 S 29 E	630 mm	Sandy	Woody/ Herbaceous	Trenches	Density of 5 mm root lengths m ⁻²	Burkea site: broad-leaf Acacia site: fine-leaf
	Lawson et al. 1968	Fig. 17	Mole Game Reserve, Ghana			Colluvial, with deep sandy loam	Total	26×25×70 cm soil monolith	g · 10000 cm ⁻³ to 70 cm	Guinea savanna
	Le Roux et al. 1995	Fig. 1	Cote D'Ivoire, Africa	6:13 N 5:02 W	1210 mm		Fine	4.4-cm diam. cores	g · m ⁻³ to 180 cm	Humid savanna
Tundra	Okali et al. 1973	Fig. 5	Accra Plains, Ghana	5:42 N 0:07 W	750 mm	Black loamy soil surrounded by pale sand over mottled sandy clay	Total	25×25 cm soil monoliths	g/monolith for 3 samples and 3 sizes to 120 cm	Grassland and thicket clump
	Pandey & Singh 1992	Fig. 5, Table 1	Vindhyan plateau, India	24:19 N 82:78 E	926–1145 mm	Residual ultisols with sandy loam texture	Total	15×15 cm soil monoliths	% biomass to 50 cm	Northern India plateau
	Scholes & Walker 1993	Fig. 14.3	South Africa	25 S 29 E	630 mm		Fine; woody/grass	0.5 m ² soil profiles	Length density (m · m ⁻³) to 1 m	<i>Eragrostis</i> , <i>Burkea</i> , <i>Terminalia</i>
	Watts 1993	Fig. 2	Texas, USA	27:39 N 98:13 W	716 mm	Sandy loam	Total, live	20×20 cm soil monoliths	g · m ⁻² to 200 cm	<i>Prosopis glandulosa</i>
	Dennis & Johnson 1970	Fig. 2	Alaska, USA	71:20 N 156:39 W	104 mm	Marine and lacustrine sediments; loamy texture	Total, live	Soil cores	g · m ⁻² to 30–60 cm	5 sites with data from 5 sampling dates, many spp.
Tundra	Dennis et al. 1978	Table 5	Alaska, USA	71:20 N 156:39 W	104 mm	Marine and lacustrine sediments; loamy texture	Live/dead	Soil cores	g · m ⁻² to 25 cm	Barrow tundra, many spp.
	Hobbie 1995	Appendix	Alaska, USA	68:38 N 149:34 W	400 mm	Histosols	Live	Soil monolith	g · m ⁻² to 25 cm	Toolik Lake tundra
	Ignatenko & Khakimzyanova 1971	Table 3	Pribaidaratskii region		340 mm		Total	Unknown	g · m ⁻² to 48 cm	Permafrost at 50 cm Dwarf Birch, Dryas, Willow
	Khodachek 1969	Tabelle III	Taimyr Peninsula				Total	Monolith	g · m ⁻² to 50 cm	<i>Dryas</i> , <i>Carex</i>

References

- Ågren GI, Axelsson B, Flower-Ellis JGK, Linder S, Persson H, Staaf H, Troeng E (1980) Annual carbon budget for a young Scots pine. *Ecol Bull* 32:307–313
- Allen M (1991) The ecology of mycorrhizae. Cambridge University Press, Cambridge
- Archer S (1995) Tree-grass dynamics in a subtropical savanna: reconstructing the past, predicting the future. *Ecoscience* 2:83–99
- Böhm W (1979) Methods of studying root systems. Springer Berlin Heidelberg New York
- Bonan GB (1992) Soil temperature as an ecological factor in boreal forests. In: a systems analysis of the global boreal forest. Shugart HH, Leemans R, Bonan GB (eds) Cambridge University Press, Cambridge, pp 126–143
- Burke IC, Kittel TGF, Lauenroth WK, Snook P, Yonker CM, Parton WJ (1991) Regional analysis of the Central Great Plains. *BioScience* 41:685–692
- Buxton PA (1925) The temperature of the surface of deserts. *J Ecol* 12:127–134
- Caldwell MM, Richards JH (1986) Competing root systems: morphology and models of absorption. In: Givnish TJ (ed) On the economy of plant form and function. Cambridge University Press, Cambridge, pp 251–273
- Canadell J, Jackson RB, Ehleringer JR, Mooney HA, Sala OE, Schulze ED (1996) Maximum rooting depth for vegetation types at the global scale. *Oecologia*, in press
- Cannon WA (1911) The root habits of desert plants (Publication 131). Carnegie Institution, Washington
- Chapin FS III, Jefferies RL, Reynolds JF, Shaver GR, Svoboda J (1992) Arctic ecosystems in a changing climate. Academic Press, San Diego
- Coleman DC (1976) A review of root production processes and their influence on soil biota in terrestrial ecosystems. In: Macfadyen JMA (ed) The role of terrestrial and aquatic organisms in decomposition processes. Blackwell, Oxford
- Dansgaard W (1964) Stable isotopes in precipitation. *Tellus* 16:436–468
- Dickinson RE, Henderson-Sellers A (1988) Modelling tropical deforestation: study of GCM land-surface parameterizations. *Q J Meteorol Soc* 114:439–462
- Dobrowolski JP, Caldwell MM, Richards JH, (1990) Basin hydrology and plant root systems. In: Osmond CB, Pitelka LF, Hidy GM (eds) Plant biology of the Basin and Range. Springer Berlin Heidelberg New York, pp 243–292
- Drew MC (1990) Sensing soil oxygen. *Plant Cell Environ* 13:681–693
- Evenari M, Shanan L, Tadmor N (1971) The Negev: challenge of a desert. Harvard University Press, Cambridge
- Farrish KW (1991) Spatial and temporal fine-root distribution in three Louisiana forest soils. *Soil Sci Soc Am J* 55:1752–1757
- Field CB, Jackson RB, Mooney HA (1995) Stomatal responses to increased CO₂: implications from the plant to the global scale. *Plant Cell Environ* 18: 1214–1225
- Fisher MJ, Rao IM, Ayarza MA, Lascano CE, Sanz JI, Thomas RJ, Vera RR (1994) Carbon storage by introduced deep-rooted grasses in the South American savannas. *Nature* 371:236–238
- Fitter AH (1982) Morphometric analysis of root systems: application of the technique and influence of soil fertility on root system development in two herbaceous species. *Plant Cell Environ* 5:313–322
- Fox RL, Lipps RC (1964) A comparison of stable strontium and ³²P and tracers for estimating alfalfa root activity. *Plant Soil* 20:337–350
- Freckman DW (1995) Life in the soil: soil biodiversity and its importance to ecosystem processes. The Natural History Museum, London
- Freckman DW, Virginia RA (1989) Plant-feeding nematodes in deep-rooting desert ecosystems. *Ecology* 70:1665–1678
- Gale MR, Grigal DF (1987) Vertical root distributions of northern tree species in relation to successional status. *Can J For Res* 17:829–834
- Gile LH, Peterson FF, Grossman RB (1966) Morphological and genetic sequences of carbonate accumulation in desert soils. *Soil Sci* 101:347–360
- Golluscio RA, Sala OE (1993) Plant functional types and ecological strategies in Patagonian forbs. *J Veg Sci* 4:839–846
- Grier CC, Vogt KA, Keyes MR, Edmonds RL (1981) Biomass distribution and above- and below-ground production in young and mature *Abies amabilis* zone ecosystems of the Washington Cascades. *Can J For Res* 11:155–167
- Hales S (1727) Vegetable statics, current edition (1961). London Scientific Book Guild, London
- Hall NS, Chandler WF, Bavel CHM van, Reid PH, Anderson JH (1953) A tracer technique to measure growth and activity of plant root systems. *N C Agric Exp Sta Tech Bull* 101: 1–40
- Hawksworth DL, Ritchie JM (1993) Biodiversity and biosystematic priorities: microorganisms and invertebrates. CAB International, Wallingford
- Heitschmidt RK, Ansley RJ, Dowhower SL, Jacoby PW, Price DL (1988) Some observations from the excavation of honey mesquite root systems. *J Range Manage* 41:227–230
- Hendrick RL, Pregitzer KS (1993) Patterns of fine root mortality in two sugar maple forests. *Nature* 361:59–61
- Higgins KB, Lamb AJ, Wilgen BW van (1987) Root systems of selected plant species in mesic fynbos in the Jonkershoek Valley, south-western Cape Province. *S Afr J Bot* 53:249–257
- Hilbert DW, Canadell J (1996) Biomass partitioning and resource allocation of plants from Mediterranean-type ecosystems: possible responses to elevated atmospheric CO₂. In: Moreno JM, Oechel WC (eds) Global change and mediterranean-type ecosystems. Ecological studies 117, Springer Berlin Heidelberg New York, pp 76–101
- Holmes JW, JS Colville (1970) Forest hydrology in a karstic region of southern Australia. *J Hydrol* 10:59–74
- Jackson RB, Caldwell MM (1993) Geostatistical patterns of soil heterogeneity around individual perennial plants. *J Ecol* 81:683–692
- Jackson RB, Manwaring JH, Caldwell MM (1990) Rapid physiological adjustment of roots to localized soil enrichment. *Nature* 344:58–60
- Kane DL, Hinzman LD, Woo M, Everett KR (1992) Arctic hydrology and climate change. In: III Chapin FS, Jefferies RL, Reynolds JF, Shaver GR, Svoboda J (eds) Arctic ecosystems in a changing climate. Academic Press, San Diego, pp 35–51
- Klinge H (1973) Root mass estimation in lowland tropical rain forests of central Amazonia, Brazil. I. Fine root masses of a pale yellow latosol and a giant humus podzol. *Trop Ecol* 14:29–38
- Klinge H, Herrera R (1978) Root biomass studies in Amazon caatinga forest in southern Venezuela. I. Standing crop of composite root mass in selected stands. *Trop Ecol* 19:93–110
- Kochenderfer JN (1973) Root distribution under some forest types native to West Virginia. *Ecology* 54:445–448
- Kummerow J (1981) Structure of roots and root systems. In: Castri F di, Goodall DW, Specht RL (eds) Mediterranean-Type Shrublands, Elsevier, New York, pp 269–288
- Kummerow J, Mangan R (1981) Root systems in *Quercus dumosa* Nutt. dominated chaparral in southern California. *Acta Oecol* 2:177–188
- Kummerow J, Krause D, Jow W (1977) Root systems of chaparral shrubs. *Oecologia* 29:163–177
- Lean J, Warrilow DA (1989) Simulation of the regional climatic impact of Amazon deforestation. *Nature* 342:411–413
- Le Roux X, Bariac T, Mariotti A (1995) Spatial partitioning of the soil water resource between grass and shrub components in a West African humid savanna. *Oecologia* 104:147–155
- McKane R B, Grigal DF, Russelle MP (1990) Spatial and temporal differences in ¹⁵N uptake and the organization of an old-field plant community. *Ecology* 71:1126–1132
- Melillo JM, McGuire AD, Kicklighter DW, Moore B III, Vorosmarty CJ, Schloss AL (1993) Global climate change and terrestrial net primary production. *Nature* 363:234–240
- Nepstad DC, Carvalho CR de, Davidson EA, Jipp PH, Lefebvre PA, Negreiros GH, Silva ED da, Stone TA, Trumbore SE, Vieira S (1994) The role of deep roots in the hydrological and carbon cycles of Amazonian forests and pastures. *Nature* 372:666–669

- Newman EI (1974) Root and soil water relations. In: Carson EW (ed) The plant root and its environment. University Press of Virginia, Charlottesville, pp 363–440
- Neilson RP (1995) A model for predicting continental-scale vegetation distribution and water balance. *Ecol Appl* 5:362–385
- Nobel PS (1988) Environmental biology of agaves and cacti. Cambridge University Press, New York
- Nobel PS (1989) Temperature, water availability, and nutrient levels at various soil depths – consequences for shallow-rooted desert succulents, including nurse plant effects. *Am J Bot* 76:1486–1492
- Oechel WC, Cowles S, Grulke N, Hastings SJ, Lawrence B, Prudhomme T, Riechers G, Strain B, Tissue D, Vourlitis G (1994) Transient nature of CO₂ fertilization in Arctic tundra. *Nature* 371:500–503
- O'Toole JC, Bland WL (1987) Genotypic variation in crop plant root systems. *Adv Agron* 41:91–145
- Parton WJ, Stewart JWB, Cole CV (1988) Dynamics of C, N, P, and S in grassland soils: a model. *Biogeochemistry* 5:109–131
- Parton WJ, McKeown B, Kirchner V, Ojima D (1992) Century Users Manual. Natural Resource Ecology Laboratory, Colorado State University, Fort Collins
- Paruelo JM, Sala OE (1995) Water losses in the Patagonian steppe: a modelling approach. *Ecology* 76:510–520
- Phillips WS (1963) Depth of roots in soil. *Ecology* 44:424
- Polley HW, Mayeux HS, Johnson JB, Tischler CR (1996) Implications of rising atmospheric CO₂ concentration for soil water availability and shrub/grass ratios on grasslands and savannas. *J Range Manage*, in press
- Potter CS, Randerson JT, Field CB, Matson PA, Vitousek PM, Mooney HA, Klooster SA (1993) Terrestrial ecosystem production: a process model based on global satellite and surface data. *Global Biogeochem Cycles* 7:811–841
- Pregitzer KS, Hendrick RL, Fogel R (1993) The demography of fine roots in response to patches of water and nitrogen. *New Phytol* 125:575–580
- Prentice IC, Cramer W, Harrison SP, Leemans R, Monserud RA, Solomon AM (1992) A global biome model based on plant physiology and dominance, soil properties and climate. *J Biogeogr* 19:117–134
- Raich JW, Nadelhoffer KJ (1989) Belowground carbon allocation in forest ecosystems: global trends. *Ecology* 70:1346–1354
- Raich JW, Rastetter EB, Melillo JM, Kicklighter DW, Steudler PA, Peterson BJ, Grace AL, Moore B III, Vörösmarty CJ (1991) Potential net primary productivity in South America: application of a global model. *Ecol Appl* 1:399–429
- Reich PB, Teskey RO, Johnson PS, Hinckley TM (1980) Periodic root and shoot growth in oak. *For Sci* 26:590–598
- Reichle DE, Dinger BE, Edwards NT, Harris WF, Sollins P (1973) Carbon flow and storage in a forest ecosystem in: Woodwell GM, Pecan EV (eds) Carbon and the biosphere. US Atomic Energy Commission, Brookhaven Symposium in Biology, AEC Conf-720510, pp 345–365
- Richards JH (1986) Root form and depth distribution in several biomes. In: Carlisle D, Berry WL, Kaplan IR, Watterson JR (eds) Mineral exploration: biological systems and organic matter. Prentice-Hall, Englewood Cliffs, pp 82–97
- Richter DD, Markewitz D (1995) How deep is soil? *BioScience* 45:600–609
- Risser PG, Birney EC, Blocker HD, May SW, Parton WJ, Wiens JA (eds) (1981) The true prairie ecosystem. (US/IBP synthesis series, vol 16). Hutchinson Ross, Stroudsburg
- Rodin LE, Basilevich NI (1966) The biological productivity of the main vegetation types in northern hemisphere of the Old World. *For Abstr* 27:369–372
- Rodin LE, Basilevich NI (1967) Production and mineral cycling in terrestrial vegetation. Oliver and Boyd, Edinburgh
- Rundel PW, Nobel PS (1991) Structure and function in desert root systems. In: Atkinson D, Plant root growth: an ecological perspective. Blackwell, Oxford, pp 349–378
- Running SW, Hunt ER Jr (1993) Generalization of a forest ecosystem process model for other biomes, BIOME-BGC, and an application for global-scale models. In: Ehleringer JR, Field CB (eds) Scaling physiological processes: leaf to globe. Academic Press, San Diego, pp 141–158
- Sachs J (1873) Über das Wachstum der Haupt- und Nebenwurzeln. *Arb Bot Inst Würzburg* 3:395–477
- Santantonio D, Hermann RK, Overton WS (1977) Root biomass studies in forest ecosystems. *Pedobiologia* 17:1–31
- Schlesinger WH (1991) *Biogeochemistry: an analysis of global change*. Academic Press, San Diego
- Schubart A (1857) Ueber die Wurzelbildung der Cerealien, beobachtet bei Ausspülungen derselben in ihren verschiedenen Lebensperioden. Hoffmann, Leipzig
- Schulze E-D, Bauer G, Buchmann N, Canadell J, Ehleringer JR, Jackson RB, Jobbagy E, Loreti J, Mooney HA, Oosterheld M, Sala OE (1996) Water availability, rooting depth, and vegetation zones along an aridity gradient in Patagonia. *Oecologia*, in press
- Shaver GR, Billings WD (1975) Root production and root turnover in a wet tundra ecosystem, Barrow, Alaska. *Ecology* 56:401–409
- Solomon AM (1992) The nature and distribution of past, present and future boreal forests: lessons for a research and modeling agenda. In: Shugart HH, Leemans R, Bonan GB (eds) A systems analysis of the global boreal forest. Cambridge University Press, New York, pp 291–301
- Stone EL, Kalisz PJ (1991) On the maximum extent of tree roots. *For Ecol Manage* 46:59–102
- Taylor HM (1987) Minirhizotron observation tubes: methods and applications for measuring rhizosphere dynamics (American Society of Agronomy special publication 50). American Society of Agronomy, Madison
- Van Rees KCJ, Comerford NB (1986) Vertical root distribution and strontium uptake of a slash pine stand on a Florida spodosol. *Soil Sci Soc Am J* 50:1042–1046
- Viereck LA, Van Cleve K, Dyrness CT (1986) Forest ecosystem distribution in the taiga environment. In: Van Cleve K, Chapin FS III, Flanagan PW, Viereck LA, Dyrness CT (eds) Forest ecosystems in the Alaskan taiga. Springer Berlin Heidelberg New York, pp 22–43
- Vincent JM (1974) Root-nodule symbioses with *Rhizobium*. In: Quispel A (ed) The biology of nitrogen fixation. North-Holland Publishing, Amsterdam, pp 266–307
- Vitousek PM, Matson PA (1984) Mechanisms of nitrogen retention in forest ecosystems: a field experiment. *Sci* 225:51–52
- Vogt KA, Bloomfield J (1991) Tree root turnover and senescence. In: Waisel AEY, Kafkafi U (eds) Plant roots: the hidden half. Marcel Dekker, New York, pp 281–306
- Vogt KA, Vogt DJ, Boon P, O'Hara J, Asbjornsen H (1996) Factors controlling the contribution of roots to ecosystem carbon cycles in boreal, temperate and tropical forests. *Plant Soil*, in press
- Watts SE (1993) Rooting patterns of co-occurring woody plants on contrasting soils in a subtropical savanna. MS dissertation, Texas A&M University, College Station
- Weaver JE (1926) Root development of field crops. McGraw-Hill, New York
- Wilson MF, Henderson-Sellers A (1985) A global archive of land cover and soils data for use in general circulation models. *J Climatol* 5:119–143
- Wullschlegel SD, Lynch JP, Berntson GM (1994) Modeling the belowground response of plants and soil biota to edaphic and climatic change: what can we expect to gain? *Plant Soil* 165:149–160
- Zohary M (1961) On the hydro-ecological relations of the near east desert vegetation. *UNESCO Arid Zone Res* 16:198–212