

# **Fishy Tracking user manual**

**Benjamin Gallois**

---

# Fishy Tracking user manual

Benjamin Gallois

This file is part of Fishy Tracking. Fishy Tracking is free software: you can redistribute it and/or modify it under the terms of the GNU General Public License as published by the Free Software Foundation, either version 3 of the License, or (at your option) any later version. Fishy Tracking is distributed in the hope that it will be useful, but WITHOUT ANY WARRANTY; without even the implied warranty of MERCHANTABILITY or FITNESS FOR A PARTICULAR PURPOSE. See the GNU General Public License for more details. You should have received a copy of the GNU General Public License along with Foobar. If not, see <https://www.gnu.org/licenses/>.

---

# Table of Contents

General information .....	1
Getting started .....	2
Download .....	2
Installation .....	2
Software overview .....	3
Files panel .....	3
Display panel .....	3
Parameters panel .....	3
Help parameter panel .....	4
Replay panel .....	6
Open a tracking analysis .....	6
Correct errors .....	6
Save tracking movie .....	7
Correct rapidly a tracked image sequence .....	7
Input files .....	9
Output files .....	10
For developers .....	11
Performance .....	12

---

## List of Figures

1. The files panel .....	3
2. Open an image .....	5
3. Select an object .....	5
4. Binarize and compute the optimal parameters .....	6
5. The replay panel .....	7
6. Keyboard shortcuts .....	8

---

# General information

Fishy Tracking is an application that allows the user to track multiple animals in video recording. The application takes a sequence of images and outputs a folder with the tracking data, the parameter used to produce the data and a log file. Fishy Tracking can operate on Linux, Mac, and Windows systems.

The software provides to the user two main features:

- An automatic tracking algorithm that can track multiple objects and deals in the most cases with occlusion.
- An manual review of the tracking where errors can be corrected on-the-fly.

---

# Getting started

## Download

Stable versions of Fishy Tracking are released for Linux (AppImage file), MacOS (dmg file) and Windows (installer). Development version is accessible from source and can be compiled for every system.

1. Stable versions:
  - Download the right installer for your system at <http://fishytracking.benjamin-gallois.fr/download.html>.
2. Development versions:
  - Install OpenCV 4 on your system by following instructions at [https://docs.opencv.org/4.0.0/d65/tutorial\\_table\\_of\\_content\\_introduction.html](https://docs.opencv.org/4.0.0/d65/tutorial_table_of_content_introduction.html).
  - Install Qt5 on your system by following instructions at <http://doc.qt.io/qt-5/gettingstarted.html#offline-installation>.
  - Download Fishy Tracking code source at <https://git.benjamin-gallois.fr/bgallois/FishyTracking>.
  - Referred to the developer user guide to compile the project if necessary.

## Installation

1. For Windows:
  - Executes the installer and follows instructions.
2. With Linux AppImage:

Allows FishyTracking.appimage to be executed:

  - Right click on the AppImage file.
  - Click on properties.
  - Click on permissions.
  - Check “allow executing the file as a program”.
3. With MacOS dmg file:
  - Drag the application from the dmg file window into the Applications folder.

---

# Software overview

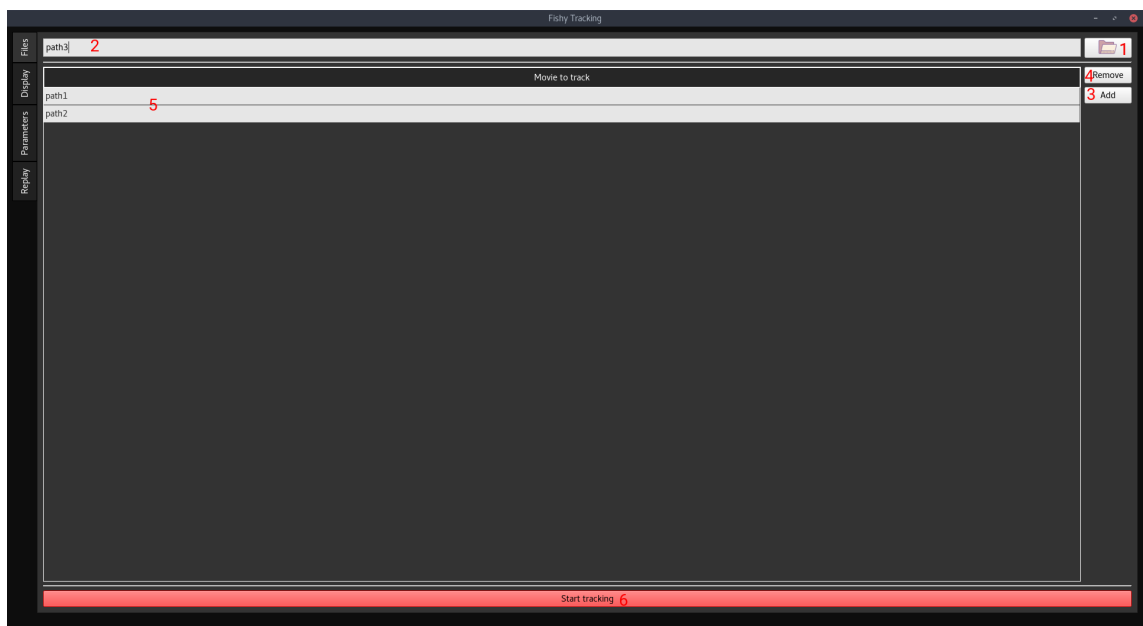
## Files panel

The files panel allows the user to add paths to folders where the image sequence to analyze are stored. Multiple paths can be added to analyze data in a bunch.

How to add image sequences to analyze?

1. Add the path to the folder where the image sequence is stored by clicking on the (1) open folder button to select a folder or copy the folder path directly into (2) the path bar.
2. Click on the (3) add path button to add the path to the list of movie to analyze , visible in the (5) paths table.
3. The added path appears now in the (5) table.
4. If necessary you can remove a path from the (5) paths table by clicking on it, it will become high-lighted in blue. Finally click on the (4) remove path button to remove the path from the (5) paths table.
5. Start the analysis by clicking on the (6) button start. The path will be deleted from the (5) paths table when the tracking analysis is finished.

**Figure 1. The files panel**



1: Open folder button. 2: path bar. 3: Add path button. 4 Remove path button. 5: Paths table. 6: Start tracking button.

## Display panel

The display panel shows the current analysis processes by displaying the binary image (1) and the tracking image (2). The tracking can be extremely fast, to review the tracking please referred to the replay panel.

## Parameters panel

The parameters panel provides a mean to change the tracking parameters before or during analysis. We recommend finding a set of parameters by changing parameters one by one and controlling effects

in the display panel. When an optimal set of parameters is found, restart the analysis. An interactive help to determine some parameters is available by clicking on the help button (1). All parameters are saved and reloaded at each closing and opening of the program.

Parameter	Range	Description
Dilatation	0 to inf	Dilates detected objects. Sets to 0 for no dilatation.
Registration	yes, no	Registers images.
Light background	yes, no	yes: dark objects on light background. no: light object on dark background
ROI bottom y	0 to image height	0: no ROI.y-coordinate of the bottom right corner of the rectangle ROI.
ROI bottom x	0 to image width	0: no ROI.x-coordinate of the bottom right corner of the rectangle ROI.
ROI top y	0 to ROI bottom y	0: no ROI.y-coordinate of the top left corner of the rectangle ROI.
ROI top x	0 to ROI bottom x	0: no ROI.x-coordinate of the top left corner of the rectangle ROI.
Number of images background	1 to number of images in the image sequences	Number of images to average to compute the background. The images are taken at equal interval in the image sequence.
Arrow size	0 to inf	Size of the arrow displayed on the object in the tracking image in the display panel.
Maximal occlusion	0 to inf	Maximal distance in pixels that an object can travel when occluded.
Weight	0 to 1	Weight between difference in displacement or difference in direction to compute the cost. Closer to 1 the matching is done with the difference in direction. Closer to 0 the matching is done only the angular difference.
Maximal angle	0 to 360	Angle in degree that the object direction can change between two frames.
Maximal length	0 to inf	Length that an object can travel between two frames
Spot to track	0, 1, 2	0: head, 1: tail, 2: body. Body part parameters (position and angle) used for the matching process.
Binary threshold	0 to 255	Threshold value that separates background and foreground.
Minimal size	0 to maximal size	Minimal size in pixels of the tiniest object to track in the image sequence.
Maximal size	minimal size to inf	Maximal size in pixels of the biggest object to track in the image sequence.
Object number	1 to inf	Number of object to track.

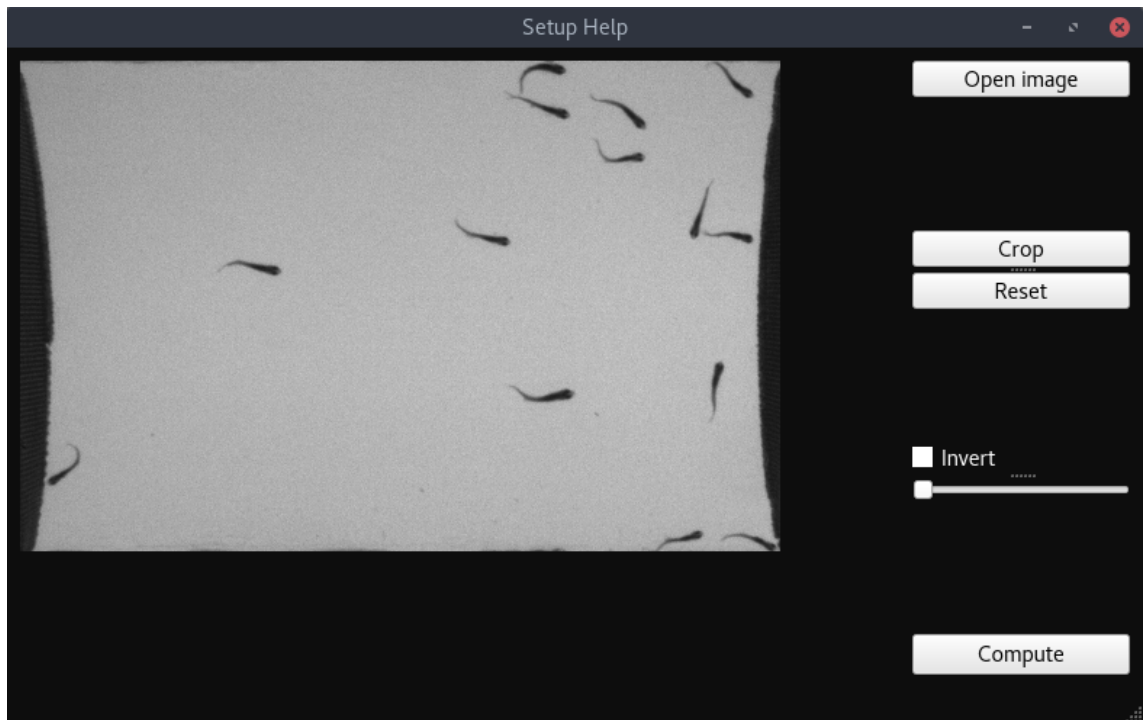
## Help parameter panel

An semi-automatic help is provided to help the user to setup the size of tracked objects. Open the Setup Help window by clicking the (1) help button.



- Open an image by clicking on the Open image button.

**Figure 2. Open an image**



- Select a typical object by clicking and dragging the mouse to enclose the object in the green rectangle.
- Click on the Crop button to crop the image.

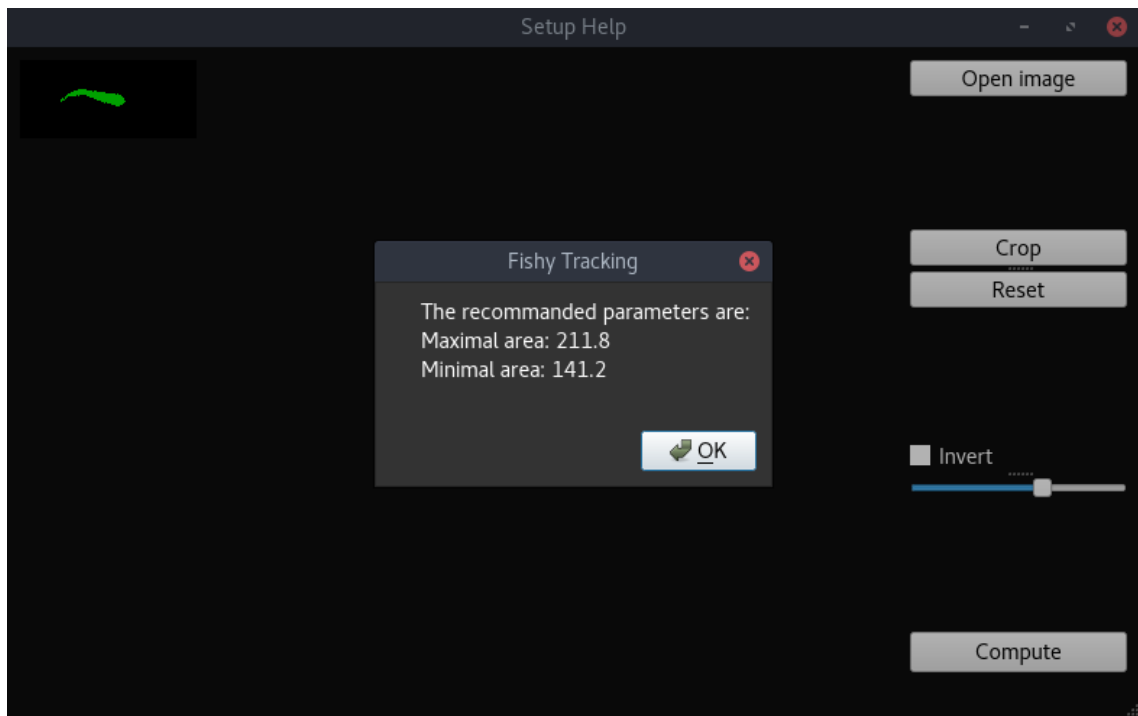
**Figure 3. Select an object**



- Threshold the object by displacing the slider.

- Tick/untick the Invert tick-box to have a white object on black background.
- Click on the Compute button to display the recommended maximal and minimal area parameters.

**Figure 4. Binarize and compute the optimal parameters**



## Replay panel

The replay panel allows the user to open and see the result of a tracking analysis and to correct the tracking if errors are detected.

## Open a tracking analysis

1. Clicks on the (1) open folder button to open a folder where an image sequence previously analyzed is stored (if no tracking data are detected, the software will display only the image sequence). Be careful to open the image sequence folder and not the tracking\_result folder.
2. The image sequence is displayed in the (6) display screen and can be played by clicking on the (5) play button or by moving the (7) play cursor. The frame rate can be chosen with the (7) fps box.
3. You can change the display options with (2) options boxes.

## Correct errors

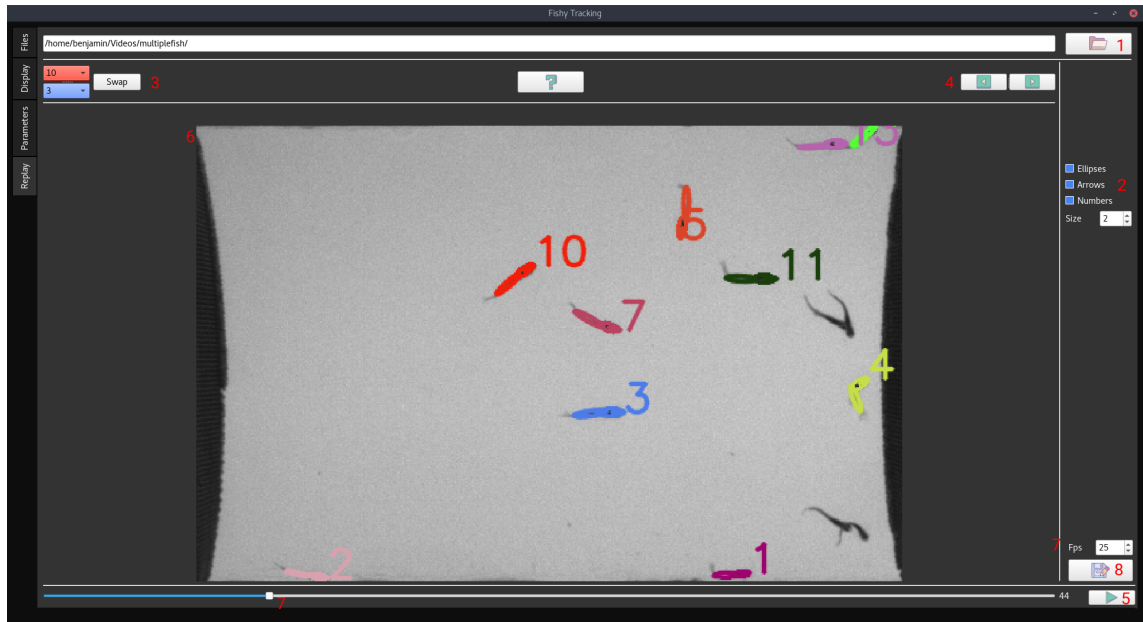
To correct a tracking error (mismatching between two objects):

1. Get to the frame where the error occurs:
  - By playing the image sequence and visually identify errors.
  - By clicking on (4) occlusion buttons to go to previous and next occlusion events where mismatching are the most probable to occurs.
2. Left click on the first object or select the number of the first object in the (3) swap boxes. The box displays the number and the color of the selected object.
3. Left click on the second object or select the number of the second object in the (3) swaps boxes. The box displays the number and the color of the selected object.
4. Right click on the (6) display screen or click on the (3) swap button to swap the two objects from the image displayed to the end of the image sequence.

## Save tracking movie

The result of the tracking can be saved in a video file. Select display options and the frame rate wanted and click on the (8) save button. The movie will be saved in .avi container with the MJPEG codec. In case of error, check that you have the MJPEG codec available on your system.

**Figure 5. The replay panel**



1: Open folder button. 2: Option boxes. 3: Swap boxes and swap button. 4: Occlusion buttons. 5: Play button. 6: Display screen. 7: Fps box. 8: Save button. 9: Play button. 10: Keyboard shortcuts help.

## Correct rapidly a tracked image sequence

Mouse and keyboards shortcuts are provided to rapidly correct a tracked image sequence. The user can move inside the image sequence and review all occlusion events with the keyboard shortcuts and correct the tracking by swapping objects with the mouse.

For AZERTY keyboard:

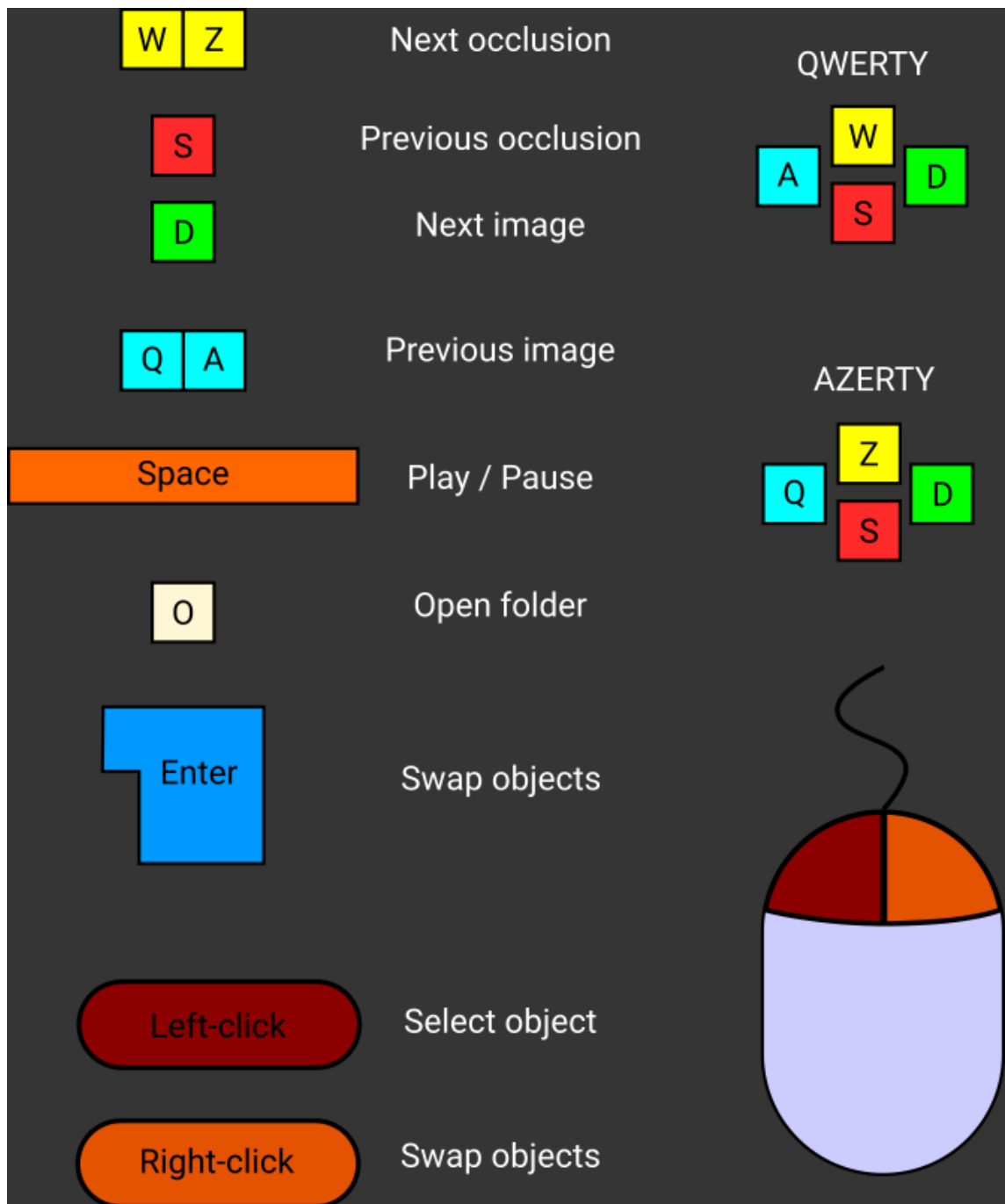
- Next image: Z
- Previous image: S
- Next occlusion: Q
- Previous occlusion: D
- Play: Space

For QWERTY keyboard:

- Next image: W
- Previous image: S
- Next occlusion: A
- Previous occlusion: D
- Play: Space

Mouse shortcuts:

- Left click to select objects.
- Right click to swap objects.

**Figure 6. Keyboard shortcuts**

---

# Input files

Fishy Tracking analyzes image sequence. “pgm”, “png”, “jpeg”, “jpg”, “tiff”, “tif”, “bmp”, “dib”, “jpe”, “jp2”, “webp”, “pbm”, “ppm”, “sr”, “ras”, “tif” image formats are supported.

If you have recording in a video format, you need to convert it into image sequence.

1. For Linux, Windows, and MacOS in command lines:
  - Install ffmpeg.
  - In a terminal type: `ffmpeg -i video.webm image-%03d.png`
2. For Linux, Windows, and MacOS in graphical:
  - Use Handbrake: <https://handbrake.fr/> or ImageJ.

# Output files

When an analysis is started, Fishy Tracking creates a folder named Tracking\_Result containing:

- tracking.txt containing the tracking data (in the frame of reference of the full image with the origin in the top left corner) as following:

xHead	yHead	Angle	xTail	yTail	Angle	xBody	yBody	Angle	Curvature	ImageLength	headMajorAxisLength	tailMajorAxisLength	tailMinorAxisLength
x coordinate of the head. Object 1	y coordinate of the head. Object 1	Angle of the head. Object 1	x coordinate of the tail. Object 1	y coordinate of the tail. Object 1	Angle of the tail. Object 1	x coordinate of the body. Object 1	y coordinate of the body. Object 1	Angle of the body. Object 1	Curvature of the body. Object 1	ImageLength of the head. Object 1	Length of the minor axis of the head. Object 1.	Length of the major axis of the tail. Object 1.	Length of the minor axis of the tail. Object 1.
x coordinate of the head. Object 2	y coordinate of the head. Object 2	Angle of the head. Object 2	x coordinate of the tail. Object 2	y coordinate of the tail. Object 2	Angle of the tail. Object 2	x coordinate of the body. Object 2	y coordinate of the body. Object 2	Angle of the body. Object 2	Curvature of the body. Object 2	ImageLength of the head. Object 2.	Length of the minor axis of the head. Object 2.	Length of the major axis of the tail. Object 2.	Length of the minor axis of the tail. Object 2.
x coordinate of the head. Object 1	y coordinate of the head. Object 1	Angle of the head. Object 1	x coordinate of the tail. Object 1	y coordinate of the tail. Object 1	Angle of the tail. Object 1	x coordinate of the body. Object 1	y coordinate of the body. Object 1	Angle of the body. Object 1	Curvature of the body. Object 1	ImageLength of the head. Object 1.	Length of the minor axis of the head. Object 1.	Length of the major axis of the tail. Object 1.	Length of the minor axis of the tail. Object 1.
...	...	...	...	...	...	...	...	...	...	...			

Note: if you want to extract the data for a particular object:  $data_i = (objectNumber) * imageNumber + objectNumber$  with objectNumber the object number {1, 2, 3, ...}, imageNumber the image number {0, 1, 2, ...}.

- parameters.txt containing the set of parameters at the beginning of the analysis, changes during the analysis are not saved.

---

# For developers

A complete API and developer documentation is available. It can be generated for the latest version of Fishy Tracking by executing `./generateDocumentation.sh`. A folder `Documentation` is created with the developer documentation.

---

# Performance

The performance can vary with the selected parameters. Registration and dilatation can cause a loss of 50% in performance. The number of objects influences the performance as the cube of the object number.

Revised: 2019/01/03