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Demo 1 – Learning a localization model

This demo is intended to get you started using Deep-STORM3D. Given a new optical setup, new phase mask, fluorescent dye, objective lens, etc. you need to train a new neural network for localization. In order to do so, you need to create a training set that can be fed into the training process. The involved steps are listed below with accompanying snapshots to ease the process.

Recovering the experimental phase mask

To calibrate the experimental phase mask, you will need to acquire a z-stack of a bead on the coverslip. This z-stack should cover the entire z-range that the user intends to recover in the actual experiment. An example such z-stack with the Tetrapod PSF is given in **Fig. 1**.

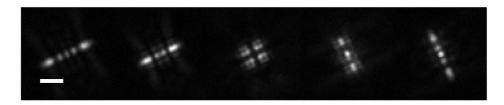


Figure. 1 - 5 slices taken from a z-stack of a fluorescent microsphere that was used to calibrate the PSF in the STORM experiment (main text Fig. 3). Scale bar is 2 microns.

Next, to retrieve the phase using the measurements in **Fig. 1**, the user should use the VIPR [1] phase retrieval method. The code of VIPR is written in MATLAB 2019a, and is publicly available¹. For more details on using this software given the acquired z-stack (**Fig. 1**) please refer to its own documentation. The expected output from VIPR is a phase mask modelling the experimental system, as shown in **Fig. 2**.

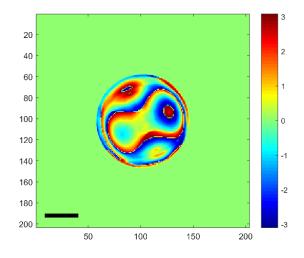


Figure. 2 - phase mask retrieved by VIPR[1]. SLM pixel size is 30 microns. Scale bar is 1 mm.

¹ https://github.com/Borisfer/VIPR---Vectorial-Phase-Retrieval-for-microscopy

Experimental stack preprocessing and SNR estimation

Usually in STORM experiments the acquired frames have a non-uniform background component stemming from autofluorescence. The simplest way to partially eliminate this non-uniform background is to subtract the minimal value per pixel from the entire stack (**Fig. 3**).

To train a net to localize the experimental data we need to match the signal to noise ratio in simulations. To get a rough estimate of the remaining baseline and the background standard deviation we can examine emitter-empty regions in the experimental frames (**Fig. 3b** red square). Similarly, the signal intensity can be estimated from subtracting the mean background counts in emitter-occupied regions (**Fig. 3b** green square).

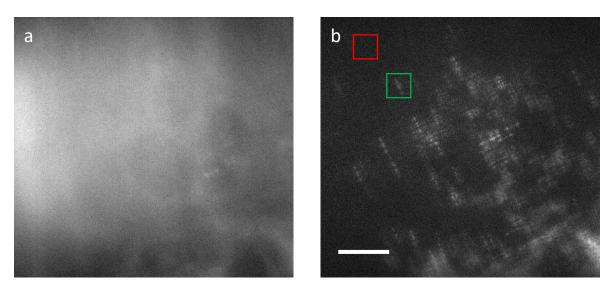


Figure. 3 - background subtraction in Fiji [2,3]. a) minimum frame of the experimental stack. b) Experimental frames after minimum subtraction. Scale bar is 10 microns.

Setting the simulation parameters

Now that we have characterized the experimental conditions, we can train a localization CNN. The simulation parameters needed to simulate the training data and learn the localization CNN are encapsulated in the script "Demos/parameter_setting_demo1.py". Next, let us go over the list of parameters and discuss their role:

- First section is used to set the mode of training: either learning a phase mask or learning a localization CNN. In case we are learning a localization CNN, the Boolean flag "learn_mask" (Fig. 4 blue rectangle) is set to False, and the "initial" mask is set to the mask recovered by VIPR (Fig. 4 green rectangle).
- 2. Second section is used to specify the optical system and the imaging assumptions (**Fig. 4** red rectangle). These include: the fluorophore mean emission wavelength (lamda) in microns, the objective numerical aperture (NA), the immersion oil refractive index (noil), the imaging medium refractive index (nwater), the pixel size of the sensor in microns (pixel_size_CCD), the pixel size

of the spatial light modulator in microns (pixel_size_SLM), the optical magnification (M), and the 4f lenses focal length in microns (f_4f).

Figure. 4 –Training mode, initial mask, and optical setup parameters.

- 3. Third section is used to control the dimensions of the Fourier space (**Fig. 5** blue rectangle), the image space (**Fig. 5** green rectangle), and the discretization in the z axis (**Fig. 5** red rectangle). The z-values are set like expected in the experimental sample. If the imaged emitters are directly on the coverslip, then "zmin" = 0. Otherwise, the distance of the lowest emitter from the coverslip needs to be calibrated for accurate depth recovery. The nominal focus position "NFP" should be set such that the PSF approximately spans the entire axial range. The number of voxels in z ("D") is what sets the axial discretization. Smaller voxels require more training time but will lead to more accurate depth recovery. Normally, for an axial range of 4 microns this is set to be within the range [81,121]. The resulting voxel size in z is $\Delta_z = \frac{z_{max} z_{min}}{D}$ [μm].
- 4. Fourth section is used to control the training density. The parameter "num_particles_range" defines the upper and lower limit on the number of particles that will be simulated within the field of view (FOV) (Fig. 5 yellow rectangle). Normally, a large range during training makes the model more robust to density variations in the experimental data.

Figure. 5 – Dimensions, discretization in z, and emitter density.

- 5. Fifth section of the parameters is used to control the distribution of the signal counts and threshold on the minimal detectable signal for the given phase mask (**Fig. 6** blue rectangle). The Boolean flag "nsig_unif" specifies whether the counts distribution for different emitters is uniform or gamma distributed. In case it is True, then the user needs to define the upper and lower limit of the uniform distribution in the parameter "nsig_unif_range". Otherwise, the counts distribution will be Gamma with a shape and scale parameters as defined in "nsig_gamma_params". Finally, for gamma distributed counts, the user can set a threshold on the detectable number of counts given the SNR conditions. This is controlled by the parameter "nsig_thresh".
- 6. Sixth section of the parameters control the range of the standard deviation of the Gaussian blur used to smooth the simulated PSFs (**Fig. 6** green rectangle). For true point sources, the parameter "blur_std_range" can be set to the estimated parameter in VIPR. Otherwise, for finite size emitters (like in the telomere sample), this parameter is set to the range [0.75, 1.25] to account for variable emitter size.

Figure 6 – Signal distribution and blur standard deviation.

- 7. Seventh section of the parameters control the Poisson background settings, namely, uniform/non-uniform (Fig. 7 blue rectangle). The parameter "unif_bg" specifies the uniform background component in counts. The Boolean flag "nonunif_bg_flag" specifies whether to include the non-uniform background modelled by a super-Gaussian. The supper-Gaussian center is randomly shifted within the range specified by "nonunif_bg_offset", and the peak and valey of the super-Gaussian are given in "non_unif_minvals". These values are perturbed by 50% randomly during training data generation. Finally, the rotation angle of the super-Gaussian is randomly chosen within the range given in "nonunif_bg_theta_range". Note that even if the flag "nonunif_bg_flag" is set to False, the remaining values are required as these are also potentially used in defining the read noise spatial distribution discussed next.
- 8. Eight section of the parameters control the read noise settings (Fig. 7 green rectangle). The Boolean flag "read_noise_flag" is used to decide whether to add read noise. The Boolean flag "read_noise_nonunif" is used to decide whether the spatial distribution of the read noise is non-uniform across the FOV (higher in the middle). If set to True, the standard deviation of the read-noise across the FOV is assumed to be distributed using the super-Gaussian from section 7. The upper and lower limits for the read noise standard deviation are given in the parameter "read_noise_std_range". Finally, after subtracting the minimal frame from the experimental data, normally we are left with a varying "baseline" across the FOV. To account for this, during data generation we add a random baseline in the range "read_noise_baseline_range".

```
# uniform/ono-unifora background settings

# uniform/ono-unifora background settings

# uniform background value per pixel
unif bg = 0 # in (counts)

# boolean flag whether or not to include a non-uniform background in pixels
nonunif_bg_flag = False

# maximal offset for the center of the non-uniform background in pixels
nonunif_bg_nter=[0, 10] # in (counts)

# peak and valley minimal values for the super-gaussian; randomized with addition of up to 50%
nonunif_bg_ntrvals = [20.0, 100.0] # in (counts)

# minimal and maximal angle of the super-gaussian for augmentation
nonunif_bg_theta_range = [-pi/4, pi/4] # in [radians]

# read_noise_flag = True

# flag whether or not to include read noise
read_noise_flag = True

# flag whether of not the read noise standard deviation is not uniform across the FOV
read_noise_nonuinf = True

# range of baseline_range = [20.0, 40.0] # in [counts]

# read_noise_baseline_range = [20.0, 40.0] # in [counts]

# read_noise_dictionary
read_noise_std_range = [8.0, 12.0] # in [counts]

# read_noise_dictionary
read_noise_dictionary
read_noise_dictionary
read_noise_dictionary
read_noise_std_range = range_rand_noise_baseline_range;
read_noise_dictionary
read_noise_std_range': read_noise_baseline_range;
read_noise_dictionary
```

Figure 7 – Non-uniform background and read noise.

- 9. Nineth section of the parameters control the image normalization for training (**Fig. 8** blue rectangle). The Boolean flag "project_01" specifies whether each training image should be normalized to the range [0,1] using the transformation: $I_{[0,1]} = \frac{I I_{min}}{I_{max} I_{mn}}$. This is helpful especially when the SNR is changing from frame to frame (e.g. Fig. S32 in supplementary information, or Fig. 6 in the main text). If "project_01" is set to False, the parameter "global_factors" is used to normalize the training images via the transformation: $I_{norm} = \frac{I global_factors[1]}{global_factors[2]}.$ This is normally useful when the SNR doesn't change much in between frames.
- 10. Tenth section of the parameters control the size of the training and validation sets (**Fig. 8** green rectangle). "ntrain" defines the number of examples for training, and "nvalid" defines the number of examples for validation. The parameter "training_data_path" defines the path where the generated training examples will be saved. The Boolean flag "visualize" controls whether the images are shown during data generation.

```
# image normalization factors for STORM (subtract the first and divide by the second)

# image normalization factors for STORM (subtract the first and divide by the second)

# image normalization factors for STORM (subtract the first and divide by the second)

# image normalization dictionary

# training data settings

# training data settings

# training data settings

# number of training and validation examples

# number of training examples: images + locations for localization net or locations + photons for PSF learning

# path for saving training examples: images + locations for localization net or locations + photons for PSF learning

# boolean flag whether to visualize examples while created

visualize = True

# training data dictionary

# training data dictionary
```

Figure. 8 – Image normalization and training/validation size.

- 11. Eleventh section of the parameters control the learning settings (**Fig. 9** blue rectangle). The parameter "results_path" defines the path were the learning results (including the CNN weights) will be saved. The Boolean flag "dilation_flag" controls whether the maximal dilation will be $d_{max}=16$ or $d_{max}=4$. This controls the network receptive field as explained in Fig. S2 in the supplementary information. The parameter "batch_size" controls the batch size used for training the CNN. Normally this is set to the biggest number that can still fit the entire model on GPU for training (in this case 4). The parameter "max_epochs" defines the maximal number of epochs for training the CNN, and the parameter "initial_learning_rate" defines the initial learning rate for the ADAM optimizer. Finally, the parameter "scaling_factor" is used to control the scaling factor of the emitters in the loss function. This parameter is used to mitigate the class imbalance between empty voxels and voxels containing emitters in the prediction grid.
- 12. Last section allows the user to resume training a model from a previously saved checkpoint in case of system crash/ Transfer learning (**Fig. 9** green rectangle). This feature is advanced and not recommended for new users.

Figure. 9 – Learning settings and resuming from checkpoint.

Training a localization CNN

Now, after setting the parameters for data generation and network training the user needs to generate the training data and learn a localization CNN. This is achieved by using the function "gen_data" from the module "GenerateTrainingExamples.py", and the function "learn_localization_cnn" from the module "Training_Localization_Model". Both steps are demonstrated in the script "demo1.py" (**Fig. 10**). Note that it takes approximately 35 hours on a Titan Xp GPU to learn a localization CNN with the specified parameters in "Demos/parameter_setting_demo1.py".

Figure. 10 – demo 1.

Localization using a trained CNN

After learning a localization CNN, the model can be tested using the function "test_model" from the module "Testing_Localization_Model.py" as demonstrated in "demo2.py" (Fig. 11). The function "test_model" accepts 4 input parameters:

- 1. "path results" which is the path to the saved training results.
- 2. "postprocessing_params" which is a dictionary with 2 entries: "thresh" and "radius" specifying the postprocessing threshold (in the range [0, "scaling_factor"]) and the radius for local maxima finding in recovery voxels. These 2 parameters ultimately control the Jaccard Index and the RMSE of the network. Usually "radius"=4 and "thresh" in the range [0.05, 0.2]*"scaling_factor" are a good guess for most scenarios.
- 3. "path_exp_data" which is the path to the experimental data for testing the model. In case the folder only contains a single experimental frame (as in "demo5.py"), the function will plot the recovered 3D positions and the regenerated input image for visual comparison with the experimental image. Otherwise, if the folder contains several frames (such as in STORM experiments) the function will save a csv file in the experimental folder named "localizations.csv". This file will have 5 columns ("frame", "x [nm]", "y [nm]", "z [nm]", and "intensity [au]"), and can be used afterwards for drift correction and visualization in ThunderSTORM [4] or ZOLA-3D [5]. If the number of images in the folder is below 100, the script will also plot the images with localizations on top marked by red crosses. Finally, if this parameter is set to "None" the function will generate a random testing image and show the recovered 3D positions alongside the GT positions and calculate quantitative performance metrics. These include the Jaccard Index, the lateral RMSE and the axial RMSE. In addition, the input image will be compared to the regenerated image visually.
- 4. "seed" is the seed used to generate the testing image in case no experimental data is supplied. You can change with this parameter to randomize the testing image in "demo3.py" (Fig. 12).

Figure. 11 - demo 2.

Figure. 12 – demo 3.

References

- [1] B. Ferdman, E. Nehme, L. E. Weiss, R. Orange, O. Alalouf, and Y. Shechtman, "VIPR: Vectorial Implementation of Phase Retrieval for fast and accurate microscopic pixel-wise pupil estimation," bioRxiv, 2020.
- [2] J. Schindelin, C. T. Rueden, M. C. Hiner, and K. W. Eliceiri, "The ImageJ ecosystem: An open platform for biomedical image analysis," *Mol. Reprod. Dev.*, vol. 82, no. 7–8, pp. 518–529, 2015.
- [3] J. Schindelin *et al.*, "Fiji: An open-source platform for biological-image analysis," *Nat. Methods*, vol. 9, no. 7, pp. 676–682, 2012.
- [4] M. Ovesný, P. Křížek, J. Borkovec, Z. Švindrych, and G. M. Hagen, "ThunderSTORM: A comprehensive ImageJ plug-in for PALM and STORM data analysis and super-resolution imaging," *Bioinformatics*, vol. 30, no. 16, pp. 2389–2390, 2014.
- [5] A. Aristov, B. Lelandais, E. Rensen, and C. Zimmer, "ZOLA-3D allows flexible 3D localization microscopy over an adjustable axial range," *Nat. Commun.*, vol. 9, no. 1, pp. 1–8, 2018.