# Package 'damr'

September 26, 2020

| Title Interface to Drosophila Activity Monitor System Result Files   |
|--|
| <b>Date</b> 2020-09-14   |
| Version 0.3.7  |
| <b>Description</b> Loads behavioural data from the widely used Drosophila Activity Monitor System (DAMS, TriKinetics <a href="https://trikinetics.com/">https://trikinetics.com/</a> ) into the rethomics framework. |
| <b>Depends</b> R (>= 3.00), behavr, data.table   |
| Imports readr  |
| Suggests testthat, covr, knitr, ggetho, zeitgebr   |
| License GPL-3  |
| Encoding UTF-8   |
| LazyData true  |
| <pre>URL https://github.com/rethomics/damr</pre>   |
| BugReports https://github.com/rethomics/damr/issues RoxygenNote 7.1.1 Roxygen list(markdown = TRUE)  |
| R topics documented:   |
| damr_example   |
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damr\_example

Get path to damr example

#### **Description**

damr comes with a sample DAM2 files in its inst/extdata directory. damr\_example allow make them easy to access.

#### Usage

```
damr_example(path = NULL)
damr_example_dir()
```

#### Arguments

path

Name of file. If NULL, the example files will be listed.

#### Author(s)

Hadley Wickham (modified from readr)

#### **Examples**

```
# list all files
damr_example()
# get path to one file
damr_example("M014.txt")
# get the directory wih all the files
damr_example_dir()
```

link\_dam\_metadata

Link DAM2 or DAM5 metadata to result files

#### **Description**

This function checks and add columns to DAMS metadata. This way, it can subsequently be loaded (via load\_dam).

#### Usage

```
link_dam_metadata(x, result_dir)
```

# Arguments

```
x object such as a data.frame, or the name of a file (see detail) result_dir the root directory where all daily data are saved
```

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#### **Details**

These function will augment metadata from two different types of inputs:

 A data.frame (recommended) In this case, the function will try to match requested data with data available on result\_dir. The provided data.frame or data.table has typically one row per requested individual and the columns (not necessarily in this order):

- file the name of a data file (e.g. "Monitor3.txt"), it has to exists in result\_dir.
- start\_datetime the first day **and time** of the requested experiment (e.g. "2014-12-28 18:00:00").
- stop\_datetime the last day and time of the requested experiment (e.g. "2014-12-30 19:00:00" or simply "2014-12-30").
- region\_id the channel (between 1 and 32) in which the animal was in (e.g. 20). region\_id is optional. If not provided, all 32 channels are loaded *with the same conditions*.
- ??? any number of arbitrary columns to associate conditions/treatments/genotypes/... to the previous columns.
- 2. The name of a CSV file that contains a table as described in 1.

The time in data is expressed relatively to start\_date. In other words, if you do circadian analysis, and your D -> L transitions are at 09:00:00, you want to set start\_datetime = "YYY-MM-DD 09:00". The result\_directory`` is the folder conraining all result (.txt) files (for instance, result\_dir = "C:/where/I/Store/my/txt/files/"')

#### Value

a data.table::data.table with the same rows as x, and extra columns used for further data loading

#### References

- the rethomics workflow on the concept of "linking"
- metadata tutorial how to work with metadata

#### See Also

• load\_dam - to subsequently load the actual data

load\_dam

Load DAMS data from one or several continuous text files

#### **Description**

Uses "linked metadata" to load data from either single beam (DAM2) or multibeam (DAM5) arrays.

#### Usage

```
load_dam(metadata, date_format = "%d %b %y", FUN = NULL, ...)
```

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## **Arguments**

metadata data.table::data.table used to load data (see detail)

date\_format How dates are formatted in the DAM result files (see read\_dam\_file)

FUN function (optional) to transform the data from each animal immediately after is has been loaded.

... extra arguments to be passed to FUN

#### **Details**

The linked metadata should be generated using link\_dam\_metadata.

#### Value

A behavr::behavr table. In addition to the metadata, it contains the data, whith the columns:

- id autogenerated unique identifier, one per animal
- t time
- activity number of beam crosses

#### References

• damr tutorial – how to use this function in practice

#### See Also

- behavr::behavr the class of the resulting object
- read\_dam\_file to load data from a single file (without metadata)

#### **Examples**

```
# This is where our toy data lives
root_dir <- damr_example_dir()</pre>
# Metadata already made for us.
# It defines condition and genotype of each animal
data(single_file_metadata)
print(single_file_metadata)
# Linking:
metadata <- link_dam_metadata(single_file_metadata, root_dir)</pre>
# We find and load the matching data
dt <- load_dam(metadata)</pre>
print(dt)
# An example of the use of FUN,
# we load only the first few reads by run `head()` on each animal,
# on the fly (no pun intended)
dt <- load_dam(metadata, FUN = head)</pre>
print(dt)
```

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| read_dam_file | Reads data from a single DAM2 single beam or a DAM5 multibeam |
|---------------|---|
|               | file  |

# Description

This function retrieves activity data in a DAMS text file. It allows selection of a date range and channels (i.e. regions).

#### Usage

```
read_dam_file(
  path,
  region_id = 1:32,
  start_datetime = -Inf,
  stop_datetime = +Inf,
  tz = "UTC",
  date_format = "%d %b %y"
)
```

#### **Arguments**

```
path location of the file to read (character)
region_id vector of unique regions to read
start_datetime, stop_datetime
the start and the end of an the experiment (see details)
tz the timezone (see OlsonNames for a list)
date_format the format of the dates in the DAM file (see details)
```

#### **Details**

start\_datetime and stop\_datetime are formatted as "YYYY-MM-DD HH:MM:SS". start\_datetime is used as the reference time (ZT0). Therefore, if you are interested in circadian analysis and D -> L transitions are at 10:00:00, you probably want to set start\_datetime = "YYYY-MM-DD 10:00:00".

According to the acquisition system, the date format can be inconsistently formatted between DAM Systems. Specify the format using strptime syntax. For instance:

```
"%d %b %y" – the default, to parse "15 Nov 2019"
"%d-%m-%y" – to parse "15-11-2019"
"%Y-%m-%d" – the default to parse "2019-11-15"
```

#### Value

A behave table. The metadata contains an autogenerated id per animal. The data has the columns:

- id autogenerated unique identifier, one per animal
- t time
- activity number of beam crosses

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#### See Also

load\_dam – to load data from many files and biological conditions using metadata (the recommended alternative)

#### **Examples**

```
path <- damr_example("M064.txt")
dt <- read_dam_file(path, region_id = c(1:3), start_datetime = "2017-06-30 15:00:00")
print(dt)</pre>
```

single\_file\_metadata

A simple toy metadata defining the experimental conditions of 32 animals monitored at the same time in a single DAM2 monitor. Each animal has its own channel (region\_id), as well as a condition and genotype. It serves as an example for link\_dam\_metadata.

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A simple toy metadata defining the experimental conditions of 32 animals monitored at the same time in a single DAM2 monitor. Each animal has its own channel (region\_id), as well as a condition and genotype. It serves as an example for link\_dam\_metadata.

#### Usage

```
single_file_metadata
```

#### **Format**

An object of class data. frame with 32 rows and 6 columns.

#### Author(s)

Quentin Geissmann

two\_files\_metadata

A toy metadata defining the experimental conditions of 64 animals monitored at the same time in two separate DAM2 monitors. Each animal has its own channel (region\_id), as well as a condition and genotype. It serves as an example for link\_dam\_metadata.

#### **Description**

A toy metadata defining the experimental conditions of 64 animals monitored at the same time in two separate DAM2 monitors. Each animal has its own channel (region\_id), as well as a condition and genotype. It serves as an example for link\_dam\_metadata.

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# Usage

two\_files\_metadata

# **Format**

An object of class  $\mbox{\tt data.frame}$  with  $64\ rows$  and  $6\ columns.$ 

# Author(s)

Quentin Geissmann

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