Can we detect decline in diversity using molecular phylogenies?

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Introduction

Over the past two decades, it is possible to see the rise and establishment of molecular phylogenies as important information used to estimate rates of diversification (Nee et al. (1994), Ape, Geiger, Diversitree, BAMM). The use of those phylogenies have allowed researchers to evaluate the diversification dynamics of several types of organisms, and especially have allowed the study of fossil-poor clades such as birs for instance. However, some skepticism regarding the extent to which we can rely only on the molecular phylogenies was raised, due to the very small values of extinction rates commonly obtained when using only this type of data (Rabosky (2010), Quental and Marshall (2011)).

- Over the last decade researchers are using molecular phylogenies of extant lineages to estimate rates of diversification (speciation and extinction). Skepticism comes from Rabosky 2010, Quental & Marshall 2011, due to extinction rates being frequently estimated with very small values using molecular phylogenies.
- However, according to the fossil record most lineages are already extinct and many of them are in a declining phase (examples: canids, rhinos, hyena, horses).
- Lineages are characterized by an expansion and a decline phase and it is still an open question whether we can detect it or not using molecular phylogenies (Quental & Marshall 2011 suggest with a simple approach gamma statistics that it is difficult to distinguish between decline and stable diversity; Morlon *et al.* 2011, with more sophisticated methods were able to detect decline; but discuss the limitation of their results).
- However, how general are the results from Morlon *et al.*? Lack of papers that have used these methods, and even greater lack of papers that actually found decline with these methods.
- Curiously BiSSE-like models suggest that some traits might be associated with negative diversification rates (Goldberg *et al.* 2010, Burin *et al.* 2015, among others).
- Additionally, more recently Beaulieau & O'Meara shown by revisiting Rabosky's 2010 that extinction
 rates can be reasonably estimated in some scenarios, although these scenarios are limited by not
 including decline in diversification.
- New methods (Morlon *et al.* (2011) and Rabosky (2014)) allow for extinction to be higher than speciation although none of these two methods were tested thoroughly in their capacity of detecting declines in different points in time. Liow et al. 2010 show that the signature of a given diversity dynamics might change as time goes by.

- Here we present a broad analysis to compare their ability to detect diversity decline in different evolutionary scenarios over different viable parameter combinations within a comprehensive parameter space. Mention that our aim is not only to assess the potential of both methods in estimating the true rates, but also how do both perform when evaluating the diversity trajectory over time.
- In summary, most of the groups analyzed would still be expanding which is in disagreement with the trend suggested by the fossil record.

Goals

Our goal was to explore the performance of two recently proposed methods, Morlon *et al.* (2011) and Rabosky (2014) (BAMM) in detecting diversity declines using molecular phylogenies.

Material and Methods

Parameter space exploration

We divided our simulations into two scenarios: the first scenario had exponential decline on speciation rates and constant extinction rates through time (BVARDCST), whereas the second scenario had constant speciation rates and exponential increase on extinction rates over time (BCSTDVAR). For these two scenarios, four parameters were combined according to each scenario: for BVARDCST, we used two parameters for speciation rates (initial speciation λ_0 and decaying rate α) and one parameter for extinction rates – μ . In the second scenario (BCSTDVAR), we used one parameter for speciation rates (λ) and two for extinction rates (initial extinction μ_0 and decaying rate β). These values were sampled randomly for each of the simulations (simulation process described below) from uniform distributions bounded by the values presented in table 1.

Diversification scenarios and simulations

The simulation process of the 2000 simulated trees for each scenario consisted of 6 steps, as follows. (1) Parameter values $(\lambda, \alpha, \mu \text{ and } \beta)$ were randomly samples from a uniform distribution with the limits indicated in table 1. (2) The expected time of the initiation of the decline phase was calculated (t_{max}) when speciation = extinction). We also calculated the expected peak species diversity at t_{max} . (3) We then estimated the time necessary for losing 80% of the peak diversity. The final species diversity values were forced to lie between 10 and 500 species at the end of the simulation, due to limitations imposed by working with trees too small (low statistical power) or too big (high computational memory/time demand). If the expected final diversity was outside of these limits, the sampled parameter values were stored and discarded, and steps 1-3 were repeated. The time needed for the species diversity to drop to 80% of peak diversity was obtained by numerical approximation, since it is not possible to analytically integrate the speciation and extinction functions. (4) The time calculated in the previous step was then used in the simulation function. The function is available at http://github.com/gburin/labmeme/bamm_rpanda, and

its initial version was kindly provided by Dr. Helene Morlon. The function simulates trees according to time-varying speciation and/or extinction rates, and stores the full resulting phylogeny that contains both extant and extinct species. We modified it to limit the maximum total diversity to 20000 species; if at some point the simulation reached this limit the simulation was interrupted, the parameters were stored and discarded and steps 1-4 were repeated; additionally, some simulated trees went fully extinct before the set time: in this case, the parameter values were also stored and discarded, and the steps 1-4 were repeated. All parameter combinations (valid, extinct or "exploded" were stored in separate files, and used to explore the properties of the parameter space.

After simulating all trees for each scenario, three final steps were performed. (5) We used the estimated diversity estimates for the whole life of each simulated tree to calculate, analogous to step 3, the times needed for the loss of 50% and 20% of peak species diversity. Furthermore, to check for false positives, we also calculated the time needed to reach 20% less of peak diversity but still on the increase phase, were the methods are expected to not detect decline in diversity. (6) All trees had then their extinct species pruned to give us the corresponding molecular phylogenies.

Model fitting and parameter estimation

We used two recently described methods (RPANDA and BAMM) to retrieve information about speciation and extinction rates for the resulting molecular trees. RPANDA estimates the parameters of the functions that describe the variation of speciation and extinction rates through time, whereas BAMM provides average instantaneous rates for each time step within the duration of the tree. Both models were thus fitted to each one of the trees in both scenarios. RPANDA models were fitted within the R environment, whereas BAMM is coded in python. The two models provide distinct types of information: while RPANDA returns parameter estimates for the model(s) one is evaluating, BAMM generates point mean rate estimates for discrete time intervals along a tree history. Thus, in order to compare both models, we analyzes only the initial and final values of the varying rate and of net diversification as well as the constant rate values.

References

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