

# Phylogenetic Inference using RevBayes

## *Total-evidence Dating under the FBD Model*

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## 1 Introduction

Ronquist et al. (2012)

Exercise is in Section 3.

### 1.1 Models

#### 1.1.1 Sequence Evolution

In this tutorial, our molecular data are best analyzed with a General Time-Reversible model of sequence evolution and gamma-distributed rate heterogeneity. This model of sequence evolution is covered thoroughly in the RB\_CTMC\_Tutorial.

#### 1.1.2 Morphological Character Change

The incorporation of morphological data from fossils into phylogenetic analyses allows us to directly observe organisms that existed in the past. In 2001, Paul Lewis described a model, the Mk model, to allow for the incorporation of morphology into likelihood and Bayesian analyses. The Mk model uses a generalized Jukes-Cantor matrix. This introduces a number of assumptions. One key assumption is that characters exhibit symmetrical change - that a given character is as likely to transition from a one state to another as it is to reverse. For example, this assumption applied to a binary character would mean that a change from a 0 state to a 1 state is as likely as a change from a 1 state to a 0 state.

In the tutorial, we will relax this assumption through the use of a prior on stationary state frequencies. This prior is visualized in Fig. 1. All of the morphological data we are working with in this tutorial are binary. Therefore, we can draw the stationary state frequencies of our 0 and 1 characters from a Beta distribution. Using the stationary state frequencies drawn from the Beta distribution, we can then create new Q-matrices that allow for asymmetrical change in characters states (i.e., for changing from state 0 to state 1 to be more likely than changing from state 1 to state 0, or vice versa). We can do multiple draws from the Beta distribution, creating multiple different Q-matrices per dataset. This allows for heterogeneity in character change asymmetry across the dataset. Likewise, we will use the Gamma distributed rate heterogeneity described in (other tutorial) to allow for different characters to evolve at different rates.

Because of the way morphological data are collected, we need to exercise caution in how we model the data. Traditionally, phylogenetic trees were built from morphological data using parsimony. Therefore, characters were collected to be parsimony informative - that is, to be useful for discriminating between phylogenetic hypotheses under the maximum parsimony criterion. This means that many morphological datasets do not contain invariant characters or autapomorphies, as these cannot be used to discriminate among hypotheses

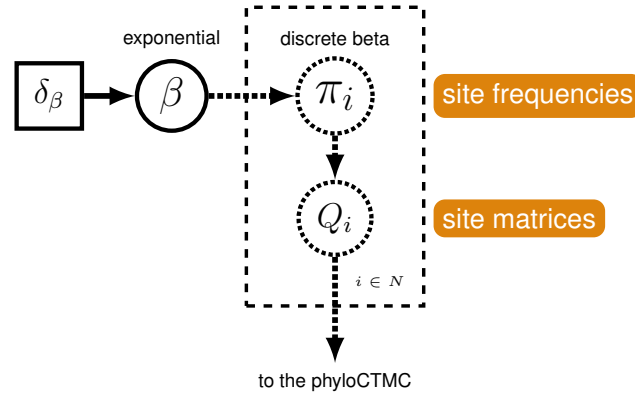


Figure 1: A graphical model of the Beta-distributed frequencies across sites model. In this model, stationary state frequencies are drawn from a discrete Beta distribution and used to generate Q-matrices that allow character change asymmetry.

under parsimony. By excluding these low rate of evolution characters, the estimated branch lengths can be inflated. Therefore, it is important to use models that can condition on the coding bias. RevBayes has two ways of doing this: one is used for datasets in which only parsimony informative characters are observed; the other is for datasets in which parsimony informative characters and non-parsimony informative variable characters (such as autapomorphies) are observed.

### 1.1.3 Lineage-Specific Substitution Rates

Clocks ([Zuckerkandl and Pauling 1962](#)) and relaxing them

### 1.1.4 Lineage Diversification and Sampling

Birth-death processes and FBD

## 2 Prerequisites

What do you need to know before doing this?

### 2.1 Requirements

We assume that you have read and hopefully completed the following tutorials:

- `RB_Getting_Started`
- `RB_Basics_Tutorial`

Note that the `RB_Basics_Tutorial` introduces the basic syntax of `Rev` but does not cover any phylogenetic models. You may skip the `RB_Basics_Tutorial` if you have some familiarity with `R`. We tried to keep this tutorial very basic and introduce all the language concepts on the way. You may only need the `RB_Basics_Tutorial` for a more in-depth discussion of concepts in `Rev`.

### 3 Exercise: Estimating the Phylogeny and Divergence Times of Fossil and Extant Bears

Information about the exercise, citations for the data, questions.

#### 3.1 Data files

We provide the data files which we will use in this tutorial. You may want to use your own data instead. In the **data** folder, you will find the following files

- **bears\_taxa.tsv**: a tab-separated table listing every bear species (both fossil and extant) and their occurrence dates. For extant taxa, the occurrence date is **0.0** (*i.e.*, the present) and for fossil species, the occurrence date is equal to the mean of the age range (the ranges are defined in a separate file).
- **bears\_cytb.nex**: an alignment in NEXUS format of 1,000 bp of cytochrome-b sequences for 10 bear species. This alignment includes 8 living bears and 2 extinct sub-fossil bears.
- **bears\_morphology.nex**: a matrix of 62 discrete, binary (coded **0** or **1**) morphological characters for 18 species of fossil and extant bears.
- **bears\_fossil\_intervals.tsv**: a tab-separated table containing the age ranges (minimum and maximum in millions of years) for 14 fossil bears.

#### 3.2 Getting Started

On your own computer, create a directory called **RB\_TotalEvidenceDating\_FBD\_Tutorial** (or any name you like).

In this directory download and unzip the archive containing the data files: **data.zip**.

Additionally, create a new directory (in **RB\_TotalEvidenceDating\_FBD\_Tutorial**) called **scripts**

When you execute **RevBayes** in this exercise, you will do so within the main directory you created (**RB\_TotalEvidenceDating\_FBD\_Tutorial**).

#### 3.3 Creating Rev Files

For complex models and analyses, it is best to create **Rev** script files that will contain all of the model parameters, moves, and functions. In this exercise, you will work primarily in your text editor and create a set of modular files that will be easily managed and interchanged. You will write the following files from scratch in a text editor\* and save them in the **scripts** directory:

- **mcmc\_TEFBD.Rev**: the master **Rev** file that loads the data, the separate model files, and specifies the monitors and MCMC sampler.
- **model\_FBDP\_TEFBD.Rev**: specifies the model parameters and moves required for the fossilized birth-death prior on the tree topology, divergence times, fossil occurrence times, and diversification dynamics.
- **model\_UCExp\_TEFBD.Rev**: specifies the components of the uncorrelated exponential model of lineage-specific substitution rate variation.

- **model\_GTRG\_TEFBD.Rev**: specifies the parameters and moves for the general-time reversible model of sequence evolution with gamma-distributed rates across sites (GTR+ $\Gamma$ ).
- **model\_Morph\_TEFBD.Rev**: specifies the model describing discrete morphological character change (binary characters) under a strict morphological clock.

\*Note: if you do not already have a good text editor that you like, we recommend one that has features for syntax coloring, easy navigation between different files, line numbers, etc. A good option is [Sublime Text](#), which is available for Mac OSX, Windows, and Linux.

All of the files that you will create are also provided in the **RevBayes** tutorial repository. Please refer to these files to verify or troubleshoot your own scripts.

### 3.4 Start the Master Rev File and Import Data

Open your text editor and create the master Rev file called **mcmc\_TEFBD.Rev** in the **scripts** directory.

Enter the Rev code provided in this section in the new model file.

The file you will begin in this section will be the one you load into **RevBayes** when you've completed all of the components of the analysis. In this section you will begin the file and write the Rev commands for loading in the taxon list and managing the data matrices. Then, starting in Section 3.5, you will move on to writing modular files for each of the model components. Once the model files are complete, you will return to editing **mcmc\_TEFBD.Rev** and complete the Rev script with the instructions given in Section 3.9.

#### 3.4.1 Load Taxon List

Begin the Rev script by loading in the list of taxon names from the **bears\_taxa.tsv** file using the **readTaxonData()** function.

```
taxa <- readTaxonData("data/bears_taxa.tsv")
```

This function reads a tab-delimited file and creates a variable called **taxa** that is a list of all of the taxon names relevant to this analysis. This list includes all of the fossil and extant bears.

#### 3.4.2 Load Data Matrices

**RevBayes** uses the function **readDiscreteCharacterData()** to load a data matrix to the workspace from a formatted file. This function can be used for both molecular sequences and discrete morphological characters.

Load the cytochrome-b sequences from file and assign the data matrix to a variable called **cytb**.

```
cytb <- readDiscreteCharacterData("data/bears_cytb.nex")
```

Next, import the morphological character matrix and assign it to the variable **morpho**.

```
morpho <- readDiscreteCharacterData("data/bears_morphology.nex")
```

### 3.4.3 Add Missing Taxa

In the descriptions of the files in Section 3.1, we mentioned that the two data matrices have different numbers of taxa. Thus, we must add any taxa that are not found in the molecular (**cytb**) partition (*i.e.*, are only found in the fossil data) to that data matrix as missing data, and do the same with the morphological data partition (**morpho**). In order for all the taxa to appear on the same tree, they all need to be part of the same dataset, as opposed to present in separate datasets. This ensures that there is a unified taxon set that contains all of our tips.

```
cytb.addMissingTaxa( taxa )
morpho.addMissingTaxa( taxa )
```

### 3.4.4 Create Helper Variables

Before we begin writing the Rev scripts for each of the model components, we need to instantiate a couple “helper variables” that will be used by downstream parts of our model specification files. These variables will be used in more than one of the module files so it’s best to initialize them in the master file.

Create a new constant node called **n\_taxa** that is equal to the number of species in our analysis (22).

```
n_taxa <- taxa.size()
```

Next, create a *workspace variable* called **mvi**. This variable is an iterator that will build a vector containing all of the MCMC moves used to propose new states for every stochastic node in the model graph. Each time a new move is added to the vector, **mvi** will be incremented by a value of 1.

```
mvi = 1
```

One important distinction here is that **mvi** is part of the RevBayes workspace and not the hierarchical model. Thus, we use the workspace assignment operator **=** instead of the constant node assignment **<-**.

Save your current working version of **model\_FBDP\_TEFBD.Rev** in the **scripts** directory.

We will now move on to the next Rev file and will complete **model\_FBDP\_TEFBD.Rev** in Section 3.9.

### 3.5 The Fossilized Birth-Death Process

Open your text editor and create the fossilized birth-death model file called `model_FBDP_TEFBD.Rev` in the `scripts` directory.

Enter the Rev code provided in this section in the new model file.

Two key parameters of the FBD process are the speciation rate (the rate at which lineages are added to the tree) and the extinction rate (the rate at which lineages are removed from the tree). We'll place exponential priors on both of these values. An exponential prior with a  $\lambda = 10$  places a higher probability on values closer to zero than one for these parameters.

```
speciation_rate ~ dnExponential(10)
extinction_rate ~ dnExponential(10)
```

Now that the priors have been specified, we give RevBayes some information on how to sample values for our parameters. We'll use a scaling move, which proposes multiplicative changes to a parameter. We will use three different tuning parameters, which govern the size of the move. Including multiple tuning parameters improves mixing.

```
moves[mvi++] = mvScale(speciation_rate, lambda=0.01, weight=1)
moves[mvi++] = mvScale(speciation_rate, lambda=0.1, weight=1)
moves[mvi++] = mvScale(speciation_rate, lambda=1.0, weight=1)

moves[mvi++] = mvScale(extinction_rate, lambda=0.01, weight=1)
moves[mvi++] = mvScale(extinction_rate, lambda=0.1, weight=1)
moves[mvi++] = mvScale(extinction_rate, lambda=1, weight=1)
```

In addition to the speciation and extinction rates, we may also be interested in inferring diversification (speciation minus extinction) and turnover (extinction divided by speciation). Since these parameters can be expressed as a deterministic transformation of the speciation and extinction rates, we can monitor their values by creating two deterministic nodes:

```
diversification := speciation_rate - extinction_rate
turnover := extinction_rate/speciation_rate
```

All extant bears are represented in this dataset. Therefore, we can fix the sampling probability of extant lineages to 1.

```
rho <- 1.0
```

The fossil sampling rate ( $\psi$ ), on the other hand is not known. We will use an exponential prior on this parameter as well, and use a scale move to sample values from the posterior distribution.

```
psi ~ dnExponential(10)
moves[mvi++] = mvScale(psi, lambda=0.01, weight=1)
moves[mvi++] = mvScale(psi, lambda=0.1, weight=1)
moves[mvi++] = mvScale(psi, lambda=1, weight=1)
```

Under the FBD model, the process is can be conditioned on the age of the origin, or the age of the root. In our case, we will condition on the origin time of bears, and we will specify a uniform distribution on the origin age. For this parameter, we will use a sliding window move. A sliding window samples a parameter uniformly within an interval (defined by the half-width **delta**). Sliding window moves can be tricky for small values, as the window may overlap zero. However, for parameters such as the origin age, there is little risk of this being an issue.

```
origin_time ~ dnUnif(37.0, 55.0)
moves[mvi++] = mvSlide(origin_time, delta=0.01, weight=5.0)
moves[mvi++] = mvSlide(origin_time, delta=0.1, weight=5.0)
moves[mvi++] = mvSlide(origin_time, delta=1, weight=5.0)
```

All the parameters of the FBD process have now been specified. The next step is to use these parameters to define the FBD tree prior distribution.

```
fbd_dist = dnFBDP(origin=origin_time, lambda=speciation_rate, mu=extinction_rate, psi=
  psi, rho=rho, taxa=taxa)
```

Note that we created the distribution as a workspace variable using the workspace assignment operator `=`. This is because we still need to include a topology constraint in our final specification of the tree prior. Specifically, we do not have any morphological or molecular data for the fossil species *Ursus abstrusus*. Therefore, in order to use the age of this fossil as a calibration, we need to specify to which clade it belongs. In this case, *Ursus abstrusus* belongs to the subfamily Ursinae, so we define a clade for Ursinae including *Ursus abstrusus*.

```
clade_ursinae = clade("Melursus_ursinus", "Ursus_arctos", "Ursus_maritimus",
  "Helarctos_malayanus", "Ursus_americanus", "Ursus_thibetanus",
  "Ursus_abstrusus", "Ursus_spelaeus")
```

Then we can specify the final constrained tree prior distribution by creating a vector of constraints, and providing it along with the workspace FBD distribution to the constrained topology distribution. Here we use the stochastic assignment operator `~` to create a stochastic node for our tree variable.

```
constraints = v(clade_ursinae)
fbd_tree ~ dnConstrainedTopology(fbd_dist, constraints=constraints)
```

Next, in order to sample from the posterior distribution of trees, we need to specify moves that propose changes to the topology and node times. We also include a proposal that toggles whether a fossil is a ‘sampled ancestor’ - a fossil species that has sampled descendants present on the tree. When conditioning on the origin time, we also need to explicitly sample the root age.

```
moves[mvi++] = mvFNPR(fbd_tree, weight=15.0)
moves[mvi++] = mvCollapseExpandFossilBranch(fbd_tree, origin_time, weight=6.0)
moves[mvi++] = mvNodeTimeSlideUniform(fbd_tree, weight=40.0)
moves[mvi++] = mvRootTimeSlideUniform(fbd_tree, origin_time, weight=5.0)
```

Next we need to account for uncertainty in the age estimates of our fossils. We do this by assuming each fossil can occur with uniform probability anywhere within its observed stratigraphic range. Stratigraphic range data for each fossil species is provided in the file `bears_fossil_intervals.tsv`. First, we read this file into a matrix, then we loop over this matrix and for each fossil create a uniform random variable. Note that this is somewhat different from the typical approach to node calibration. Here, instead of treating the calibration density as an additional prior distribution on the tree, we treat it as the *likelihood* of our fossil data given the tree parameter. Specifically, we assume the likelihood of a particular fossil observation is equal to one if the fossil’s inferred occurrence time falls within its observed stratigraphic range. This is equivalent to saying the likelihood of the observed median age is uniform within the stratigraphic interval surrounding the inferred fossil occurrence time.

```
fossil_intervals = readDataDelimitedFile(file="data/bears_fossil_intervals.tsv",
    header=true)

for(i in 1:fossil_intervals.size())
{
  taxon = fossil_intervals[i][1]
  obs_min = fossil_intervals[i][2]
  obs_max = fossil_intervals[i][3]

  t[i] := tmrca(fbd_tree, clade(taxon))

  half = (obs_max - obs_min)/2
  umin[i] := max(v(t[i] - half, 0.0))
  umax[i] := t[i] + half

  fossil[i] ~ dnUniform(umin[i], umax[i])
}
```



```
fossil[i].clamp( obs_min + half )
}
```

Finally, we can also create deterministic nodes for other quantities we might be interested in monitoring. Below, we will define a monitor that prints the number of fossils in the tree that are sampled ancestors. We will also define a deterministic node for the age of the crown group of extant bears. In addition, we can monitor the sampled age of a particular fossil.

```
num_samp_anc := fbd_tree.numSampledAncestors()

clade_extant = clade("Ailuropoda_melanoleuca","Tremarctos_ornatus","Melursus_ursinus",
                    "Ursus_arctos","Ursus_maritimus","Helarctos_malayanus",
                    "Ursus_americanus","Ursus_thibetanus")
age_extant := tmrca(fbd_tree, clade_extant)

age_Kretzoiarctos_beatrix := tmrca(fbd_tree, clade("Kretzoiarctos_beatrix"))
```

### 3.6 The Uncorrelated Exponential Relaxed-Clock Model

Open your text editor and create the lineage-specific branch-rate model file called **model\_UCExp\_TEFBD.Rev** in the **scripts** directory.

Enter the Rev code provided in this section in the new model file.

```
branch_rates_mean ~ dnExponential(10.0)

moves[mvi++] = mvScale(branch_rates_mean, lambda=0.01, weight=1.0)
moves[mvi++] = mvScale(branch_rates_mean, lambda=0.1, weight=1.0)
moves[mvi++] = mvScale(branch_rates_mean, lambda=1.0, weight=1.0)
```

Before creating a rate parameter for each branch, we need to get the number of branches in the tree. For rooted trees with  $n$  taxa, the number of branches is  $2n - 2$ .

```
n_branches <- 2 * n_taxa - 2
```

Then, use a for loop to define a rate for each branch. The branch rates are independently and identically exponentially distributed with mean equal to the mean branch rate parameter we specified above. For each rate parameter we also create scale proposal moves.

```
for(i in 1:n_branches){
  branch_rates[i] ~ dnExp(1/branch_rates_mean)
  moves[mvi++] = mvScale(branch_rates[i], lambda=1.0, weight=1.0)
  moves[mvi++] = mvScale(branch_rates[i], lambda=0.1, weight=1.0)
  moves[mvi++] = mvScale(branch_rates[i], lambda=0.01, weight=1.0)
}
```

Lastly, we use a vector scale move to propose changes to all branch rates simultaneously. This way we can sample the total branch rate independently of each individual rate, which can improve mixing.

```
moves[mvi++] = mvVectorScale(branch_rates, lambda=0.01, weight=4.0)
moves[mvi++] = mvVectorScale(branch_rates, lambda=0.1, weight=4.0)
moves[mvi++] = mvVectorScale(branch_rates, lambda=1.0, weight=4.0)
```

### 3.7 The General Time Reversible + Gamma Model of Nucleotide Sequence Evolution

Open your text editor and create the molecular substitution model file called **model\_GTRG\_TEFBD.Rev** in the **scripts** directory.

Enter the Rev code provided in this section in the new model file.

```
sf_hp <- v(1,1,1,1)
sf ~ dnDirichlet(sf_hp)

er_hp <- v(1,1,1,1,1,1)
er ~ dnDirichlet(er_hp)
```

Special moves are required to propose changes to a Dirichlet random variable, also known as a simplex (a vector constrained sum to one). Here, we use a **SimplexElementScale** move, which scales a single element of a simplex and then renormalizes it to sum to one. The tuning parameter **alpha** specifies how conservative the proposal should be, with larger values of **alpha** leading to proposals closer to the current value.

```
moves[mvi++] = mvSimplexElementScale(er, alpha=10.0, weight=5.0)
moves[mvi++] = mvSimplexElementScale(sf, alpha=10.0, weight=5.0)
```

Then we can define a deterministic node for our GTR substitution matrix using the special GTR matrix function.

```
Q_cytb := fnGTR(er,sf)
```

Next, in order to model gamma-distributed rates across, we create an exponential parameter  $\alpha$  for the shape of the gamma distribution, along with scale proposals.

```
alpha_cytb ~ dnExponential( 1.0 )

moves[mvi++] = mvScale(alpha_cytb, lambda=0.01, weight=1.0)
moves[mvi++] = mvScale(alpha_cytb, lambda=0.1, weight=1.0)
moves[mvi++] = mvScale(alpha_cytb, lambda=1, weight=1.0)
```

Then we create a  $\text{Gamma}(\alpha, \alpha)$  distribution, discretized into 4 rate categories using the **DiscretizeGamma** function. Here, **rates\_cytb** is a deterministic vector of rates computed as the mean of each category.

```
rates_cytb := fnDiscretizeGamma( alpha_cytb, alpha_cytb, 4 )
```

Finally, we can create the **PhyloCTMC** distribution for our sequence data, including the gamma-distributed site rate categories, as well as the branch rates defined as part of our exponential relaxed clock. We set the value of this distribution equal to our observed data and identify it as a static part of the likelihood using the **clamp** method.

```
phySeq ~ dnPhyloCTMC(tree=fbd_tree, Q=Q_cytb, siteRates=rates_cytb, branchRates=
  branch_rates, type="DNA")
phySeq.clamp(cytb)
```

### 3.8 Modeling the Evolution of Binary Morphological Characters

Open your text editor and create the morphological character model file called **model\_Morph\_TEFBD.Rev** in the **scripts** directory.

Enter the Rev code provided in this section in the new model file.

As stated in the introduction, we will use a prior on state frequencies to relax the key assumption of the Mk model. Because we are working with binary data, we can use a discrete Beta distribution to describe the variation in stationary state frequencies across characters. The Beta distribution has two parameters,  $\alpha$  and  $\beta$  which describe its shape. For simplicity, we will assume that  $\alpha = \beta$ . The below code draws a value for  $\beta$  from an exponential distribution and places a move on it.

```
beta_hp ~ dnExponential( 1.0 )

moves[mvi++] = mvScale(beta_hp, lambda=1, weight=1.0 )
moves[mvi++] = mvScale(beta_hp, lambda=0.1, weight=1.0 )
moves[mvi++] = mvScale(beta_hp, lambda=0.01, weight=1.0 )
```

Next, we'll create a vector containing four different site stationary state frequencies. This is similar to allowing gamma-distributed rate variation across sites. We will then use these stationary frequencies to generate a set of new Q-matrices which do not enforce the assumptions of the same transition rates at every site and equal forward and backwards transition rates.

```
beta_cats := fnDiscretizeBeta( beta_hp, beta_hp, 4)
for(i in 1:beta_cats.size())
{
  Q_morpho[i] := fnFreeBinary(v(1-beta_cats[i], beta_cats[i]))
}
```

As in the molecular data partition, we will allow gamma-distributed rate heterogeneity among sites.

```
alpha_morpho ~ dnExponential( 1.0 )
rates_morpho := fnDiscretizeGamma( alpha_morpho, alpha_morpho, 4, false )

moves[mvi++] = mvScale(alpha_morpho, lambda=0.01, weight=1.0)
moves[mvi++] = mvScale(alpha_morpho, lambda=0.1, weight=1.0)
moves[mvi++] = mvScale(alpha_morpho, lambda=1, weight=1.0)
```

Each data partition has to have a clock rate. For simplicity, we will assume a strict clock rate drawn from an exponential distribution.

```
clock_morpho ~ dnExponential(1.0)

moves[mvi++] = mvScale(clock_morpho, lambda=0.01, weight=4.0)
moves[mvi++] = mvScale(clock_morpho, lambda=0.1, weight=4.0)
moves[mvi++] = mvScale(clock_morpho, lambda=1, weight=4.0)
```

As in our molecular data partition, we now combine our data and our model. There are some unique aspects to doing this for morphology.

You will notice that we have a variable called ‘coding’. This variable allows us to condition on biases in the way the morphological data were collected. This specifies the ascertainment bias correction discussed in ??.

We use the flag ‘siteMatrices=true’ to indicate that we are providing multiple Q matrices generated as a function of our state frequency variation model.

```
phyMorpho ~ dnPhyloCTMC(tree=fbd_tree, siteRates=rates_morpho, branchRates=
  clock_morpho, Q=Q_morpho, type="Standard", coding="variable", siteMatrices=true)
phyMorpho.clamp(morpho)
```

### 3.9 Complete MCMC File

Return to the master Rev file you created in Section 3.4 called **mcmc\_TEFBD.Rev** in the **scripts** directory.

Enter the Rev code provided in this section in this file.

#### 3.9.1 Source Model Scripts

This step will load in the model scripts we have written in the text editor.

```
source("scripts/model_FBDP_TEFBD.Rev")
source("scripts/model_UCExp_TEFBD.Rev")
source("scripts/model_GTRG_TEFBD.Rev")
source("scripts/model_Morph_TEFBD.Rev")
```

#### 3.9.2 Create Model Object

```
mymodel = model(sf)
```

#### 3.9.3 Specify Monitors and Output Filenames

```

mni = 1
monitors[mni++] = mnModel(filename="output/bears.log", printgen=10)
monitors[mni++] = mnFile(filename="output/bears.trees", printgen=10, fbd_tree)
monitors[mni++] = mnScreen(printgen=10, age_extant, num_samp_anc, origin_time)

```

### 3.9.4 Set up the MCMC

```

mymcmc = mcmc(mymodel, monitors, moves)

mymcmc.run(generations=10000)

```

Save and close all files.

## 3.10 Run it

```
./rb
```

Execute the MCMC analysis:

```
source("scripts/mcmc_TEFBD.Rev")
```

```

Processing file "scripts/mcmc_TEFBD.Rev"
Successfully read one character matrix from file 'data/bears_cytb.nex'
Successfully read one character matrix from file 'data/bears_morphology.nex'
Processing file "scripts/model_FBDP_TEFBD.Rev"
Processing of file "scripts/model_FBDP_TEFBD.Rev" completed
Processing file "scripts/model_UCExp_TEFBD.Rev"
Processing of file "scripts/model_UCExp_TEFBD.Rev" completed
Processing file "scripts/model_GTRG_TEFBD.Rev"
Processing of file "scripts/model_GTRG_TEFBD.Rev" completed
Processing file "scripts/model_Morph_TEFBD.Rev"
Processing of file "scripts/model_Morph_TEFBD.Rev" completed

```

```

Running MCMC simulation
This simulation runs 1 independent replicate.
The simulator uses 163 different moves in a random move schedule with 267 moves per iteration

```

Iter	Posterior	Likelihood	Prior	age_extant	num_samp_anc	origin_time	elapsed	ETA
0	-8174.01	-8053.8	-120.209	34.8641	0	44.4332	00:00:00	--:--:--
10	-4654.95	-4611.2	-43.7495	4.32618	7	45.4494	00:00:01	--:--:--
20	-4294.05	-4266.91	-27.1443	4.58804	7	46.5636	00:00:01	00:08:19
30	-4267.35	-4233.41	-33.94	6.8467	6	45.9177	00:00:02	00:11:04
40	-4226.63	-4188.32	-38.3037	6.40484	8	44.3696	00:00:02	00:08:18
...								

## 3.11 Summarize Your Results

### 3.11.1 Evaluate MCMC

For this part of the tutorial, we will use a software called Tracer. Tracer allows researchers to visualize the MCMC sample for any parameters in the log file. Start Tracer by double-clicking on the Tracer executable. Click the plus sign on the left-hand side of the screen to add your log file (see Fig. 3). Navigate to your log file, and click ‘OK’.

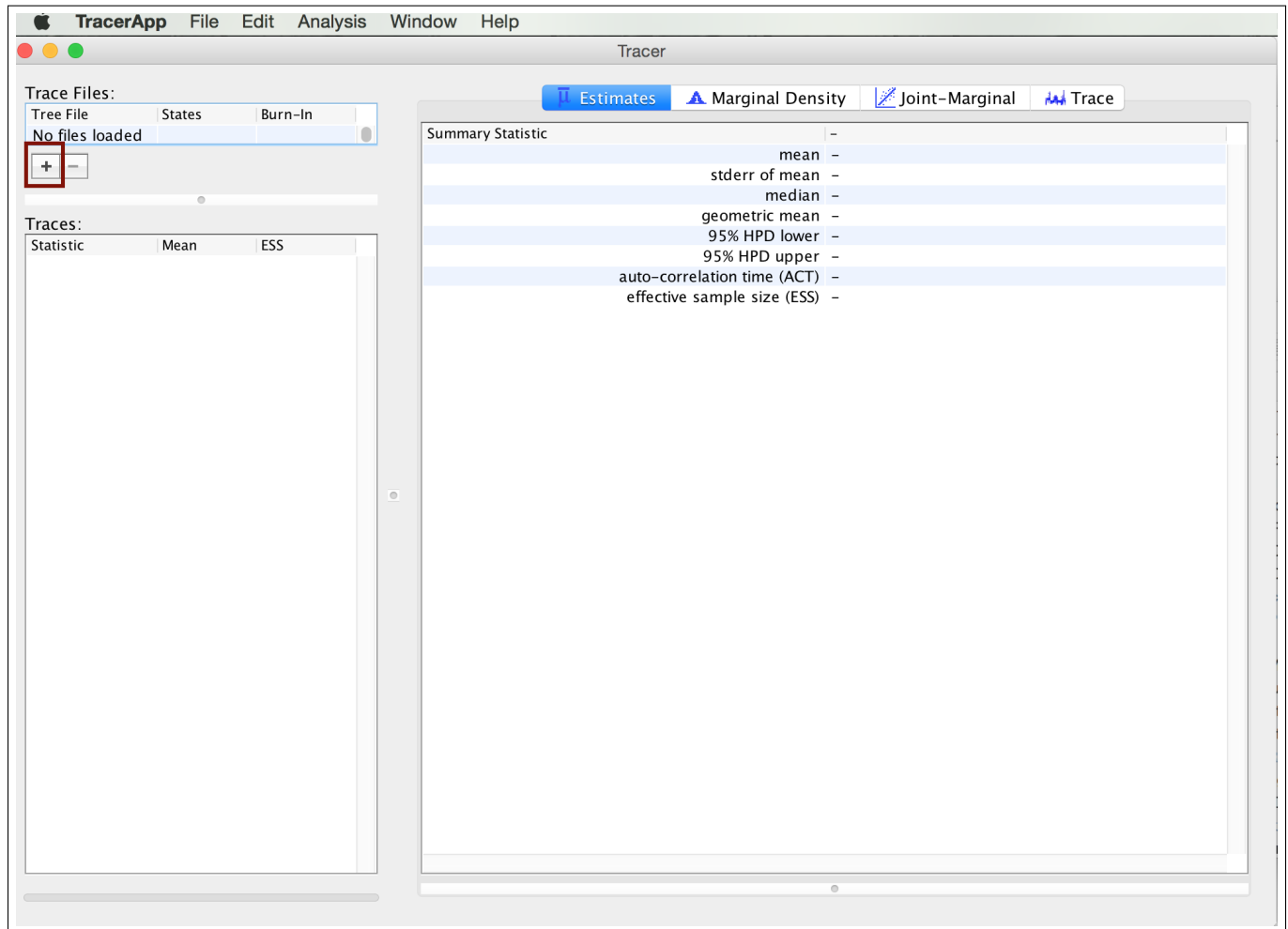


Figure 2: The Tracer window. To add data, click on the ‘+’ sign, highlighted in red above.

Immediately upon loading, you will view the ‘Estimates’ window, which shows the actual values sampled in the MCMC. In this window, the statistics on the left-hand side are the mean of the distribution and the Effective Sample Size (ESS) for that parameter. Parameters in a phylogenetic model may be correlated. The ESS is meant to provide a representation of the number of independent draws represented by the MCMC sample. This quantity will typically be much smaller than the number of generations of the chain. Poor values for the ESS will be colored red. You will likely see a lot of red values in the log file you made today because we did not run the chain very long. The data used to make the screenshots can be found in the bears\_long.zip file in the output directory.

Take a look at some of the parameters. Are there any that have really low ESS? Why do you think that might be?

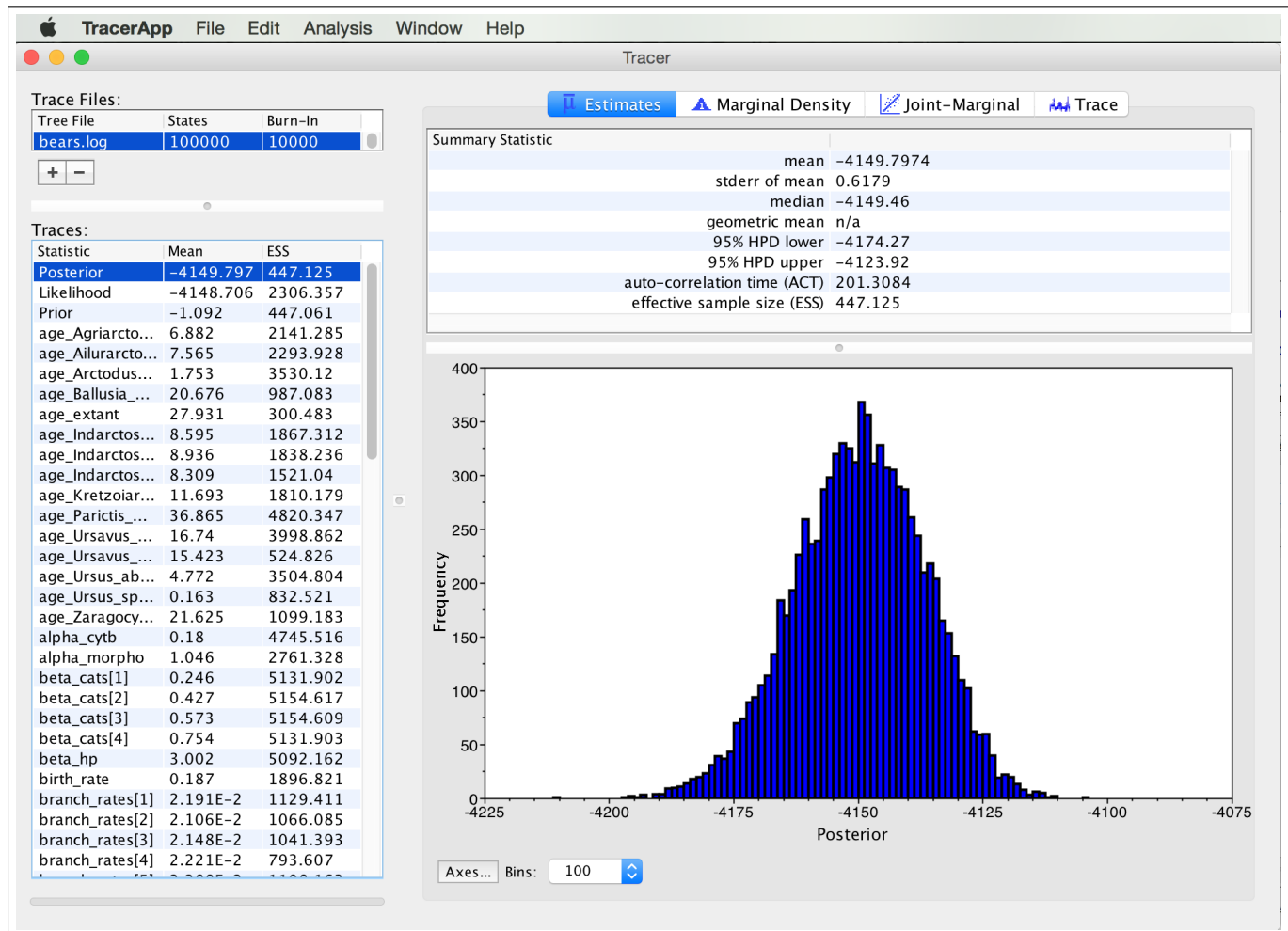


Figure 3: The Estimates window. The left-hand window provides mean and ESS of the chain. The right-hand window visualizes the distribution of samples.

### 3.11.2 Summarize Tree

Start up RevBayes at the command line. You should do this from within the `RB_TotalEvidenceDating_FBD_Tutorial` directory.

```
./rb
```

Read in the MCMC sample of trees from file.

```
trace = readTreeTrace("output/bears.trees")
for(i in 1:trace.size())
```



```
{
  trees[i] = fnPruneTree(trace.getTree(i), pruneTaxa=v(taxa[17],taxa[20]))
}

trace_pruned = treeTrace(trees)
mccTree(trace_pruned, "output/bears.mcc.tre" )
```

See Fig. 4

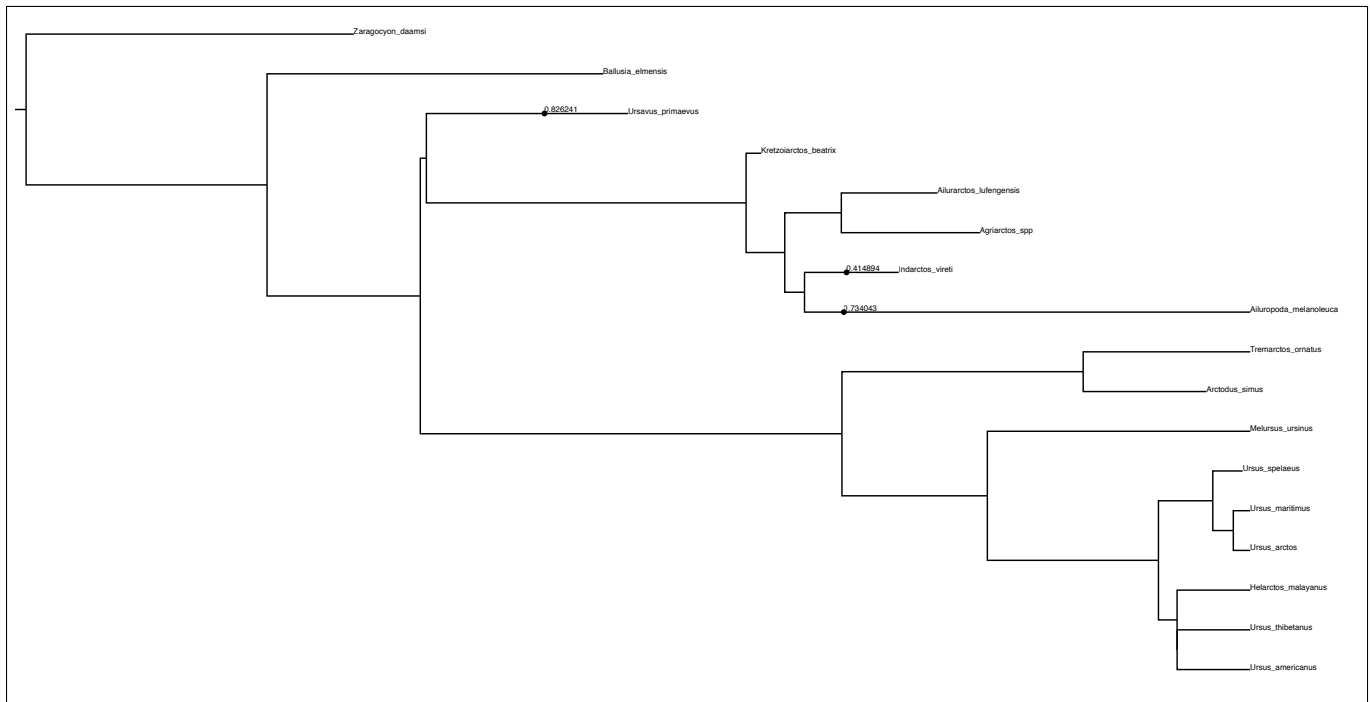


Figure 4: This is a place-holder figure.

## References

- Ronquist, F., S. Klopstein, L. Vilhelmsen, S. Schulmeister, D. L. Murray, and A. P. Rasnitsyn. 2012. A total-evidence approach to dating with fossils, applied to the early radiation of the Hymenoptera. *Systematic Biology* 61:973–999.
- Zuckerkandl, E. and L. Pauling. 1962. Molecular disease, evolution, and genetic heterogeneity. Pages 189–225 *in* Horizons in Biochemistry (M. Kasha and B. Pullman, eds.) Academic Press, New York.

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