Phylogenetic Inference using `RevBayes`
Branch-Specific Diversification Rate Estimation

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1 Overview: Diversification Rate Estimation

Models of speciation and extinction are fundamental to any phylogenetic analysis of macroevolutionary processes (*e.g.,* divergence time estimation, diversification rate estimation, continuous and discrete trait evolution, and historical biogeography). First, a prior model describing the distribution of speciation events over time is critical to estimating phylogenies with branch lengths proportional to time. Second, stochastic branching models allow for inference of speciation and extinction rates. These inferences allow us to investigate key questions in evolutionary biology.

Diversification-rate parameters may be included as nuisance parameters of other phylogenetic models—*i.e.,* where these diversification-rate parameters are not of direct interest. For example, many methods for estimating species divergence times—such as `BEAST` (Drummond et al. 2012), `MrBayes` (Ronquist et al. 2012), and `RevBayes` (Höhna et al. 2016)—implement 'relaxed-clock models' that include a constant-rate birth-death branching process as a prior model on the distribution of tree topologies and node ages. Although the parameters of these 'tree priors' are not typically of direct interest, they are nevertheless estimated as part of the joint posterior probability distribution of the relaxed-clock model, and so can be estimated simply by querying the corresponding marginal posterior probability densities. In fact, this may provide more robust estimates of the diversification-rate parameters, as they accommodate uncertainty in the other phylogenetic-model parameters (including the tree topology, divergence-time estimates, and the other relaxed-clock model parameters). More recent work, *e.g.,* Heath et al. (2014), uses macroevolutionary models (the fossilized birth-death process) to calibrate phylogenies and thus to infer dated trees.

In these tutorials we focus on the different types of macroevolutionary models to study diversification processes and thus the diversification-rate parameters themselves. Nevertheless, these macroevolutionary models should be used for other evolutionary questions, when an appropriate prior distribution on the tree and divergence times is needed.

1.1 Types of Hypotheses for Estimating Diversification Rates

Many evolutionary phenomena entail differential rates of diversification (speciation – extinction); *e.g.,* adaptive radiation, diversity-dependent diversification, key innovations, and mass extinction. The specific study questions regarding lineage diversification may be classified within three fundamental categories of inference problems. Admittedly, this classification scheme is somewhat arbitrary, but it is nevertheless useful, as it allows users to navigate the ever-increasing number of available phylogenetic methods. Below, we describe each of the fundamental questions regarding diversification rates.

(1) Diversification-rate through time estimation *What is the (constant) rate of diversification in my study group?* The most basic models estimate parameters of the stochastic-branching process (*i.e.,* rates of speciation and extinction, or composite parameters such as net-diversification and relative-extinction

rates) under the assumption that rates have remained constant across lineages and through time; *i.e.,* under a constant-rate birth-death stochastic-branching process model (Nee et al. 1994). Extensions to the (basic) constant-rate models include diversification-rate variation through time (Stadler 2011; Höhna 2015). First, we might ask whether there is evidence of an episodic, tree-wide increase in diversification rates (associated with a sudden increase in speciation rate and/or decrease in extinction rate), as might occur during an episode of adaptive radiation. A second question asks whether there is evidence of a continuous/gradual decrease in diversification rates through time (associated with decreasing speciation rates and/or increasing extinction rates), as might occur because of diversity-dependent diversification (*i.e.,* where competitive ecological interactions among the species of a growing tree decrease the opportunities for speciation and/or increase the probability of extinction, *e.g.,* Höhna (2014)). Third, we can ask whether changes in diversification rates are correlated with environmental factors, such as environmental CO_2 or temperature (Condamine et al. 2013). A final question in this category asks whether our study tree was impacted by a mass-extinction event (where a large fraction of the standing species diversity is suddenly lost, *e.g.,* May et al. (2016)). The common theme of these studies is that the diversification process is tree-wide, that is, all lineages of the study group have the exact same rates at a given time.

(2) Diversification-rate variation across branches estimation   *Is there evidence that diversification rates have varied significantly across the branches of my study group?* Models have been developed to detect departures from rate constancy across lineages; these tests are analogous to methods that test for departures from a molecular clock—*i.e.,* to assess whether substitution rates vary significantly across lineages (Alfaro et al. 2009; Rabosky 2014). These models are important for assessing whether a given tree violates the assumptions of rate homogeneity among lineages. Furthermore, these models are important to answer questions such as: *What are the branch-specific diversification rates?*; and *Have there been significant diversification-rate shifts along branches in my study group, and if so, how many shifts, what magnitude of rate-shifts and along which branches?*

(3) Character-dependent diversification-rate estimation   *Are diversification rates correlated with some variable in my study group?* Character-dependent diversification-rate models aim to identify overall correlations between diversification rates and organismal features (binary and multi-state discrete morphological traits, continuous morphological traits, geographic range, etc.). For example, one can hypothesize that a binary character, say if an organism is herbivorous/carnivorous or self-compatible/self-incompatible, impact the diversification rates. Then, if the organism is in state 0 (*e.g.,* is herbivorous) it has a lower (or higher) diversification rate than if the organism is in state 1 (*e.g.,* carnivorous) (Maddison et al. 2007).

2   Models

We begin this section with a general introduction to the stochastic birth-death branching process that underlies inference of diversification rates in `RevBayes`. This primer will provide some details on the relevant theory of stochastic-branching process models. We appreciate that some readers may want to skip this somewhat technical primer; however, we believe that a better understanding of the relevant theory provides a foundation for performing better inferences. We then discuss a variety of specific birth-death models, but emphasize that these examples represent only a tiny fraction of the possible diversification-rate models that can be specified in `RevBayes`.

2.1   The birth-death branching process

Our approach is based on the *reconstructed evolutionary process* described by Nee et al. (1994); a birth-death process in which only sampled, extant lineages are observed. Let $N(t)$ denote the number of species at time

t. Assume the process starts at time t_1 (the 'crown' age of the most recent common ancestor of the study group, t_{MRCA}) when there are two species. Thus, the process is initiated with two species, $N(t_1) = 2$. We condition the process on sampling at least one descendant from each of these initial two lineages; otherwise t_1 would not correspond to the t_{MRCA} of our study group. Each lineage evolves independently of all other lineages, giving rise to exactly one new lineage with rate $b(t)$ and losing one existing lineage with rate $d(t)$ (Figure 1 and Figure 2). Note that although each lineage evolves independently, all lineages share both a common (tree-wide) speciation rate $b(t)$ and a common extinction rate $d(t)$ (Nee et al. 1994; Höhna 2015). Additionally, at certain times, $t_{\mathbb{M}}$, a mass-extinction event occurs and each species existing at that time has the same probability, ρ, of survival. Finally, all extinct lineages are pruned and only the reconstructed tree remains (Figure 1).
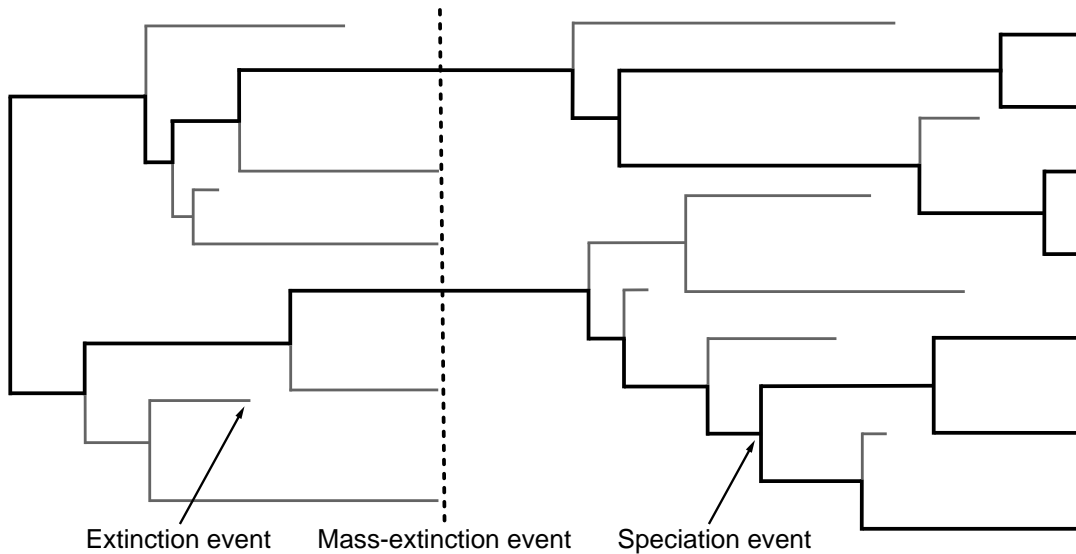

Figure 1: A realization of the birth-death process with mass extinction. Lineages that have no extant or sampled descendant are shown in gray and surviving lineages are shown in a thicker black line.
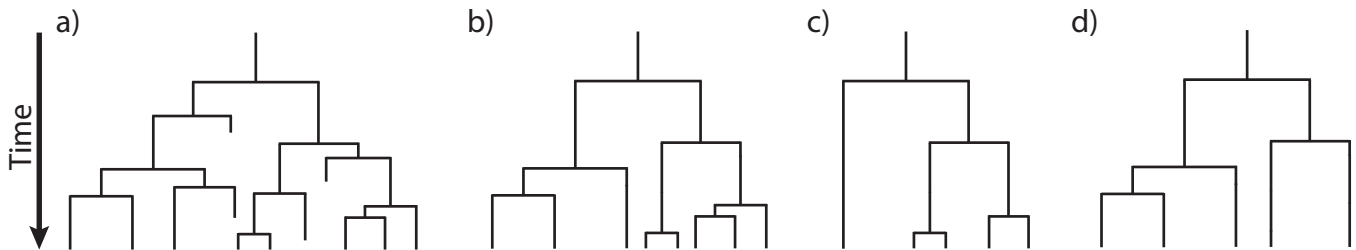

Figure 2: **Examples of trees produced under a birth-death process.** The process is initiated at the first speciation event (the 'crown-age' of the MRCA) when there are two initial lineages. At each speciation event the ancestral lineage is replaced by two descendant lineages. At an extinction event one lineage simply terminates. (A) A complete tree including extinct lineages. (B) The reconstructed tree of tree from A with extinct lineages pruned away. (C) A *uniform* subsample of the tree from B, where each species was sampled with equal probability, ρ. (D) A *diversified* subsample of the tree from B, where the species were selected so as to maximize diversity.

To condition the probability of observing the branching times on the survival of both lineages that descend from the root, we divide by $P(N(T) > 0|N(0) = 1)^2$. Then, the probability density of the branching times,

$\mathbb{T}$, becomes

$$P(\mathbb{T}) = \underbrace{\frac{\overbrace{P(N(T)=1 \mid N(0)=1)^2}^{\text{both initial lineages have one descendant}}}{P(N(T)>0 \mid N(0)=1)^2}}_{\text{both initial lineages survive}} \times \prod_{i=2}^{n-1} \overbrace{i \times b(t_i)}^{\text{speciation rate}} \times \overbrace{P(N(T)=1 \mid N(t_i)=1)}^{\text{lineage has one descendant}},$$

and the probability density of the reconstructed tree (topology and branching times) is then

$$P(\Psi) = \frac{2^{n-1}}{n!(n-1)!} \times \left(\frac{P(N(T)=1 \mid N(0)=1)}{P(N(T)>0 \mid N(0)=1)} \right)^2$$
$$\times \prod_{i=2}^{n-1} i \times b(t_i) \times P(N(T)=1 \mid N(t_i)=1) \tag{1}$$

We can expand Equation (1) by substituting $P(N(T)>0 \mid N(t)=1)^2 \exp(r(t,T))$ for $P(N(T)=1 \mid N(t)=1)$, where $r(u,v) = \int_u^v d(t) - b(t)dt$; the above equation becomes

$$P(\Psi) = \frac{2^{n-1}}{n!(n-1)!} \times \left(\frac{P(N(T)>0 \mid N(0)=1)^2 \exp(r(0,T))}{P(N(T)>0 \mid N(0)=1)} \right)^2$$
$$\times \prod_{i=2}^{n-1} i \times b(t_i) \times P(N(T)>0 \mid N(t_i)=1)^2 \exp(r(t_i,T))$$
$$= \frac{2^{n-1}}{n!} \times \left(P(N(T)>0 \mid N(0)=1) \exp(r(0,T)) \right)^2$$
$$\times \prod_{i=2}^{n-1} b(t_i) \times P(N(T)>0 \mid N(t_i)=1)^2 \exp(r(t_i,T)). \tag{2}$$

For a detailed description of this substitution, see Höhna (2015). Additional information regarding the underlying birth-death process can be found in (Thompson 1975; Equation 3.4.6) and Nee et al. (1994) for constant rates and Höhna (2013; 2014; 2015) for arbitrary rate functions.

To compute the equation above we need to know the rate function, $r(t,s) = \int_t^s d(x) - b(x)dx$, and the probability of survival, $P(N(T)>0|N(t)=1)$. Yule (1925) and later Kendall (1948) derived the probability that a process survives ($N(T)>0$) and the probability of obtaining exactly n species at time T ($N(T)=n$) when the process started at time t with one species. Kendall's results were summarized in Equation (3) and Equation (24) in Nee et al. (1994)

$$P(N(T)>0|N(t)=1) = \left(1 + \int_t^T \left(\mu(s) \exp(r(t,s)) \right) ds \right)^{-1} \tag{3}$$

$$P(N(T)=n|N(t)=1) = (1 - P(N(T)>0|N(t)=1) \exp(r(t,T)))^{n-1}$$
$$\times P(N(T)>0|N(t)=1)^2 \exp(r(t,T)) \tag{4}$$

An overview for different diversification models is given in Höhna (2015).

> **Sidebar: Phylogenetic trees as observations**
>
> The branching processes used here describe probability distributions on phylogenetic trees. This probability distribution can be used to infer diversification rates given an "observed" phylogenetic tree. In reality we never observe a phylogenetic tree itself. Instead, phylogenetic trees themselves are estimated from actual observations, such as DNA sequences. These phylogenetic tree estimates, especially the divergence times, can have considerable uncertainty associated with them. Thus, the correct approach for estimating diversification rates is to include the uncertainty in the phylogeny by, for example, jointly estimating the phylogeny and diversification rates. For the simplicity of the following tutorials, we take a shortcut and assume that we know the phylogeny without error. For publication quality analysis you should always estimate the diversification rates jointly with the phylogeny and divergence times.

3 Estimating Branch-Specific Speciation & Extinction Rates

3.1 Outline

This tutorial describes how to specify a branch-specific branching-process models in `RevBayes`; a birth-death process where diversification rates vary among branches, similar to Rabosky (2014). The probabilistic graphical model is given for each component of this tutorial. The goal is to obtain estimate of branch-specific diversification rates using Markov chain Monte Carlo (MCMC).

3.2 Requirements

We assume that you have read and hopefully completed the following tutorials:

- RB_Getting_Started

- RB_Basics_Tutorial

- RB_BayesFactor_Tutorial

- RB_BasicDiversificationRate_Tutorial

Note that the RB_Basics_Tutorial introduces the basic syntax of `Rev` but does not cover any phylogenetic models. You may skip the RB_Basics_Tutorial if you have some familiarity with `R`. The RB_BayesFactor_Tutorial introduces Bayesian model selection by means of Bayes factors, which can be skipped by readers familiar with Bayesian model selection. We tried to keep this tutorial very basic and introduce all the language concepts and theory on the way. You may only need the RB_Basics_Tutorial for a more in-depth discussion of concepts in `Rev`.

4 Data and files

We provide the data file(s) which we will use in this tutorial. You may want to use your own data instead. In the **data** folder, you will find the following files

- **primates_tree.nex**: Dated primates phylogeny including 233 out of 367 species from Magnuson-Ford and Otto (2012).

→   Open the tree `data/primates_tree.nex` in FigTree.
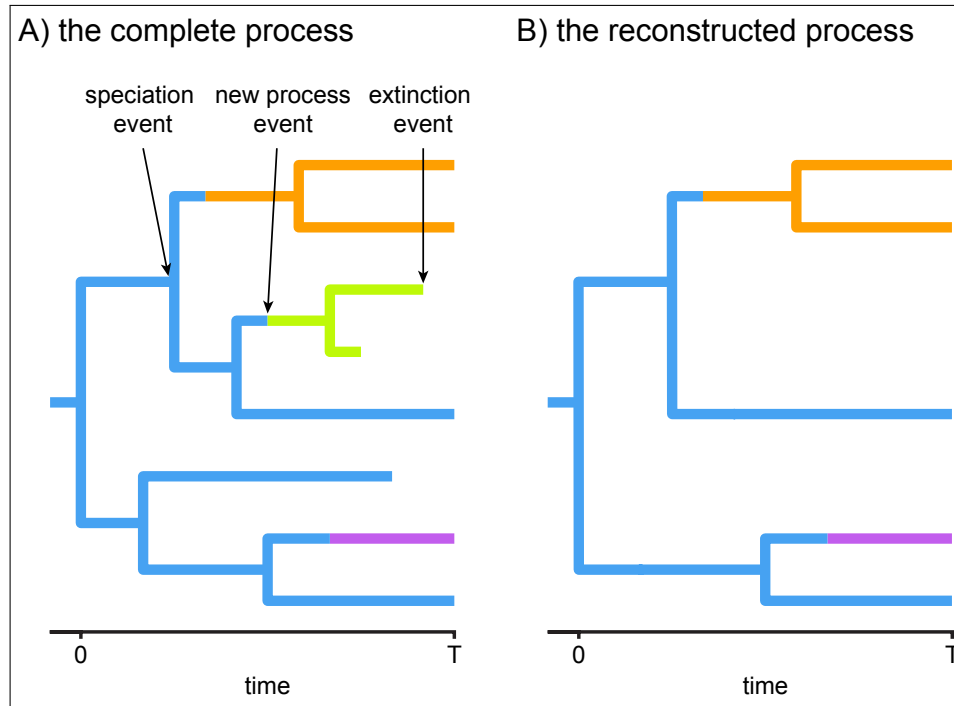
5   Branch-Specific Birth-Death Model

Figure 3: Cartoon of a branch-specific birth-death process. On the left we see the full process. On the right we only see the branches of the reconstructed tree, thus missing one rate-shift event.

The basic idea behind the model is that speciation and extinction rates are allowed to vary across branches of the tree (see Figure 3). Unfortunately, it is not possible to model rates drawn from a continuous distribution directly, as done for example in BAMM, because in that case one needs to integrate over any number of possible rate shifts, any time of these shifts and most importantly over all possible new rates. This is unfeasible to do and failure to do so has been shown to make parameter estimates unreliable (Moore et al. 2016).

Here we adopt an approach using (few) discrete rate categories instead. This allows us to numerically integrate over all possible rate categories using a system of differential equation originally described by Maddison et al. (2007) (see also FitzJohn et al. (2009) and FitzJohn (2010)). The numerical procedure beaks time into very small time intervals and sums over all possible events occurring in that interval (see Figure 4).

→   You don't need to worry about any of the technical details. It is important for you to realize that this model assumes that new rates at a rate-shift event are drawn from a given (discrete) set of rates.

In RevBayes we have two implementations (*i.e.,* distributions) for modeling a branch-specific birth-death process. The first distribution is the **dnBirthDeathMultiRate** (or its alias **dnMRBDP**) and the second is the **dnHeterogeneousBirthDeath** (or its alias **dnHBDP**). We have designed this tutorial so that each section can be read independently although we recommend that you work through both of them. You will find that some parts are redundant, which is intentional to emphasize the similarities between the analysis but
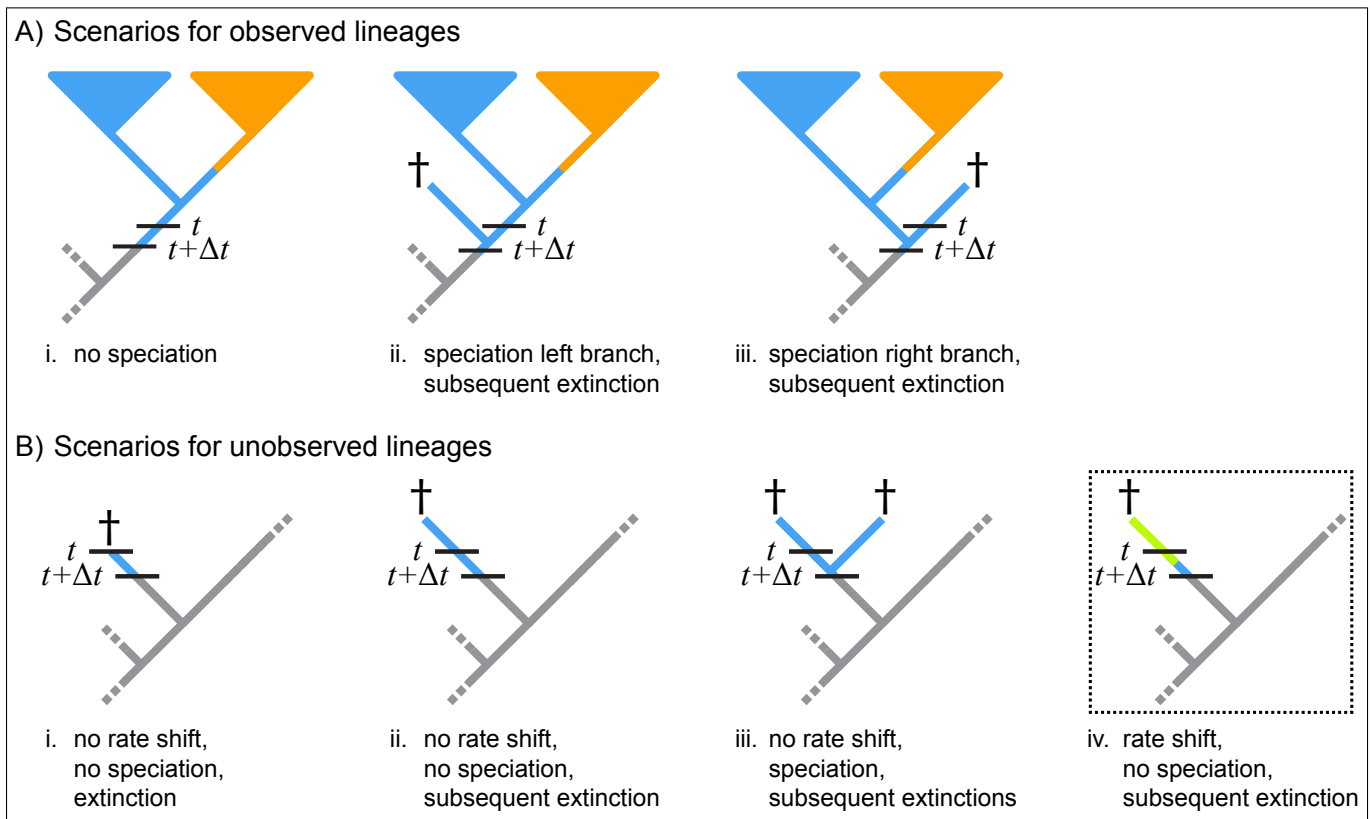
Figure 4: Cartoon of the likelihood computation using numerical integration.

also to make the sections independent.

6 Testing for Branch-Specific-Diversification Rates

In this first exercise we are interested in knowing if there is diversification-rate variation among branches for our study tree. That is, we want to see if we can reject a constant rate birth-death process. Therefore, we don't focus on branch-specific parameter estimates but instead on the marginal likelihood estimation for model testing.

→ We assume that you have completed the RB_BasicDiversificationRate_Tutorial to estimate the marginal likelihood under a constant-rate birth-death process. If you haven't done so, then you should go back and do this now!

6.1 Read the tree

Begin by reading in the observed tree.

```
observed_phylogeny <- readTrees("data/primates_tree.nex")[1]
```

From this tree, we can get some helpful variables:

```
taxa <- observed_phylogeny.taxa()
root <- observed_phylogeny.rootAge()
tree_length <- observed_phylogeny.treeLength()
```

Additionally, we can initialize an iterator variable for our vector of moves and monitors:

```
mvi = 0
mni = 0
```

Finally, we create a helper variable that specifies the number of discrete rate categories, another helper variable for the expected number of rate-shift events, the total number of species, and the variation in rates.

```
NUM_RATE_CATEGORIES = 4
EXPECTED_NUM_EVENTS = 2
NUM_TOTAL_SPECIES = 367
H = 0.587405
```

Using these variables we can easily change our script, for example, to use more or fewer categories and test the impact. For example, setting `NUM_RATE_CATEGORIES = 1` gives the constant rate birth-death process.

6.2 Specifying the model
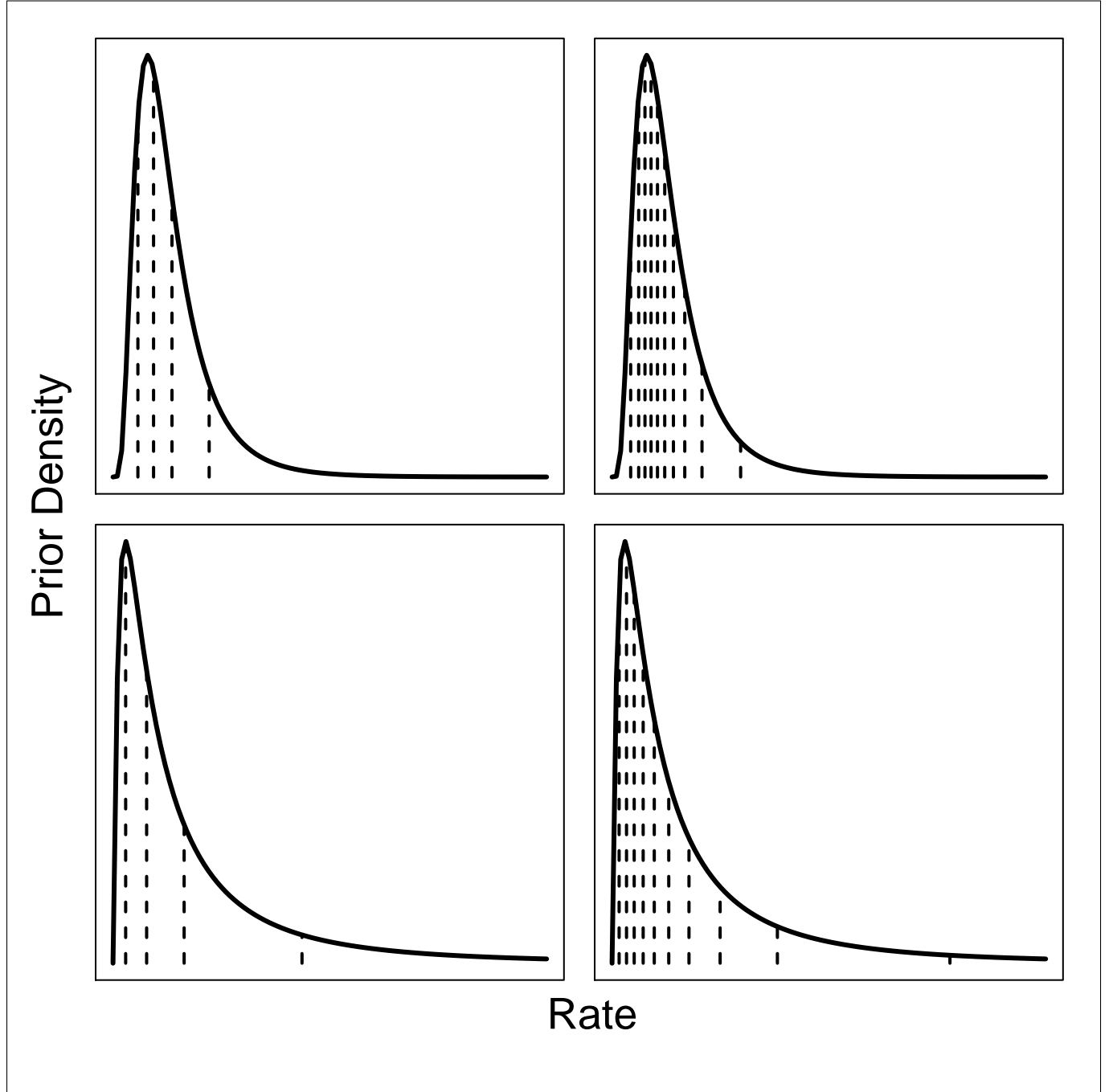
6.2.1 Priors on rates

Figure 5: Discretization of a lognormal distribution. The two left figures have 4 rate categories and the two right plots have 10 rate categories. The top plots have the 95% probability interval spanning one order of magnitude (**sd** = 0.587405) and the bottom plots have the 95% probability interval spanning two orders of magnitude (**sd** = 2 ∗ 0.587405)

Instead of using a continuous probability distribution we will use a discrete approximation of the distribu-

tion, as done for modeling rate variation across sites (Yang 1994) and for modeling relaxed molecular clocks (Drummond et al. 2006). That means, we assume that the speciation rates are drawn from one of the N quantiles of the lognormal distribution. For this we will use the function **fnDiscretizeDistribution** which takes in a distribution as its first argument and the number of quantiles as the second argument. The return value is a vector of quantiles. We use it as a deterministic variable and every time the parameters of the base distribution (*i.e.,* the lognormal distribution in our case) change the quantiles will update automatically as well. Thus we only need to specify parameters for our base distribution, the lognormal distribution. We choose a stochastic variable for the mean parameter of the lognormal distribution drawn from yet another lognormal prior distribution. We fix the prior mean on this mean speciation rate on our expected diversification rate, which is $\ln(\ln(\frac{\#Taxa}{2})/age)$. Remember that the median of a lognormal distribution is equal to the exponential of the mean parameter. This is why we used a log-transform of the actual mean. This prior density is analogous to the prior on the speciation-rate parameter in the constant-rate birth-death process.

```
speciation_prior_mean <- ln( ln(NUM_TOTAL_SPECIES/2.0) / root )
speciation_mean ~ dnLognormal(mean=speciation_prior_mean, sd=H)
moves[++mvi] = mvScale(speciation_mean,lambda=1,tune=true,weight=5)
```

Additionally, we choose a fixed standard deviation of $2 * H$ ($0.587405 * 2$) for the speciation rates because it represents two orders of magnitude variance in the rate categories.

```
speciation_sd <- 2*H
speciation_categories := fnDiscretizeDistribution( dnLognormal(ln(speciation_mean),
    speciation_sd), NUM_RATE_CATEGORIES )
```

We also need discretized extinction-rate categories. We are completely free to choose how we construct these rate categories. For example, we could choose a similar discretization of a lognormal distribution using its quantiles to provide different extinction-rate categories. For simplicity, this is how we specify the current model. Alternatively, we could assume that each rate category has the same extinction rate.

```
extinction_prior_mean <- ln( ln(NUM_TOTAL_SPECIES/2.0) / root )
extinction_mean ~ dnLognormal(mean=extinction_prior_mean,sd=2*H)
moves[++mvi] = mvScale(extinction_mean,lambda=1.0,tune=true,weight=3.0)
```

As with the speciation rate, we discretize the lognormal distribution into a finite number of rate categories.

```
extinction_categories := fnDiscretizeDistribution( dnLognormal(ln(extinction_mean), H)
    , NUM_RATE_CATEGORIES )
```

Now, we must create a vector that contains each combination of speciation- and extinction-rates. This allows the rate of speciation to change without changing the rate of extinction and vice versa. The resulting vector should be N^2 elements long. We call these the 'paired' rate categories.

```
k = 1
for(i in 1:NUM_RATE_CATEGORIES) {
    for(j in 1:NUM_RATE_CATEGORIES) {
        speciation[k]  := speciation_categories[i]
        extinction[k++] := extinction_categories[j]
    }
}
```

Next, we need a rate parameter for the rate-shifts events. We do not have much prior information about this rate but we can provide some realistic ranges. For example, we can specify a mean rate so that the resulting number of expected rate-shift events is 2 (as specified in our global variable **EXPECTED_NUM_EVENTS**). Furthermore, we can say that the 95% prior ranges exactly one order of magnitude. We achieve all this by specifying a lognormal prior distribution with mean **ln(EXPECTED_NUM_EVENTS/tree_length)** and standard deviation of **H**. Remember that this is only possible if the tree is known and not estimated simultaneously because only if the tree is do we also know the tree length. As usual for rate parameter, we apply a scaling move to the **event_rate** variable.

```
event_rate ~ dnLognormal( ln( EXPECTED_NUM_EVENTS/tree_length ), H)
moves[++mvi] = mvScale(event_rate,lambda=1,tune=true,weight=5)
```

Additionally, we need a rate-matrix parameter providing the relative rates between paired rate categories. In this case we simply use equal rates between each rate category; and thus use the Jukes-Cantor rate matrix. You could, for example, also use an ordered rate matrix where the process needs to go through rate 2 before going to rate 3 when starting in rate 1.

```
rate_matrix <- fnJC( NUM_RATE_CATEGORIES * NUM_RATE_CATEGORIES )
```

Furthermore, we need prior probabilities for the process being in either paired rate category at the root. Given our lack of prior knowledge we create a flat prior distribution giving each rate category equal weight. We do this by create a constant variable using the simplex function.

```
rate_category_prior <- simplex( rep(1, NUM_RATE_CATEGORIES * NUM_RATE_CATEGORIES) )
```

6.2.2   Incomplete Taxon Sampling

We know that we have sampled 233 out of 367 living primate species. To account for this we can set the sampling parameter as a constant node with a value of 233 / 367.

```
rho <- observed_phylogeny.ntips() / NUM_TOTAL_SPECIES
```

6.2.3 Root age

The birth-death process requires a parameter for the root age. In this exercise we use a fix tree and thus we know the age of the tree. Hence, we can get the value for the root from the Magnuson-Ford and Otto (2012) tree. This is done using our global variable **root** defined above and nothing else has to be done here.

6.2.4 The time tree

Now we have all of the parameters we need to specify the full episodic birth-death model. We initialize the stochastic node representing the time tree.

```
timetree ~ dnMRBDP(lambda=speciation, mu=extinction, Q=rate_matrix, rootAge=root, rho=
    rho, pi=rate_category_prior, delta=event_rate, taxa=taxa)
```

And then we attach data to it.

```
timetree.clamp(observed_phylogeny)
```

Finally, we create a workspace object of our whole model using the **model()** function.

```
mymodel = model(speciation)
```

The **model()** function traversed all of the connections and found all of the nodes we specified.

6.3 Running a marginal likelihood estimation

6.3.1 Specifying Monitors

For the marginal likelihood analysis we don't necessarily need monitors because we are not going to look into the samples. However, as good practice we still define our two standard monitors: the model monitor and a screen monitor

```
monitors[++mni] = mnModel(filename="output/primates_MRBD.log",printgen=10, separator =
    TAB)
monitors[++mni] = mnScreen(printgen=10, diversification_mean, turnover)
```

6.3.2 Initializing and Running the MCMC Simulation

→   If you don't feel comfortable with Bayesian model selection anymore, then have a look at the RB_BayesFactor_Tutorial again.

First, we create the variable containing the power posterior. This requires us to provide a model and vector of moves, as well as an output file name. The **cats** argument sets the number of power steps.

```
pow_p = powerPosterior(mymodel, moves, "output/MRBD_powp.out", cats=100)
```

We can start the power posterior by first burning in the chain and and discarding the first 5000 states.

```
pow_p.burnin(generations=5000,tuningInterval=200)
```

Now execute the run with the **.run()** function:

```
pow_p.run(generations=2000)
```

Once the power posteriors have been saved to file, create a stepping-stone sampler. This function can read any file of power posteriors and compute the marginal likelihood using stepping-stone sampling.

```
ss = steppingStoneSampler(file="output/MRBD_powp.out", powerColumnName="power",
    likelihoodColumnName="likelihood")
```

Compute the marginal likelihood under stepping-stone sampling using the member function **marginal()** of the **ss** variable and record the value in Table 1.

```
ss.marginal()
```

Path sampling is an alternative to stepping-stone sampling and also takes the same power posteriors as input.

```
ps = pathSampler(file="output/MRBD_powp.out", powerColumnName="power", likelihoodColumnName="
    likelihood")
```

Compute the marginal likelihood under stepping-stone sampling using the member function **marginal()** of the **ps** variable and record the value in Table 1.

```
ps.marginal()
```

→   The Rev file for performing this analysis: `ml_MRBD.Rev`.

6.4 Exercise 1

- Enter the marginal likelihood estimate from the previous exercise on the constant-rate birth-death process in the table below.

- Compute the marginal likelihood under the 2-rate model, *i.e.,* set the NUM_Rate_CATEGORIES variable to 2.

- Repeat the estimation of the marginal likelihoods with other number of rate categories to fill out the table.

- What is the most supported model? Can we reject the constant-rate birth-death process?

Table 1: Marginal likelihoods and Bayes factors*.

Estimate	*Stepping-stone*	*Path sampling*
Marginal likelihood constant-rate ($N = 1$)		
Marginal likelihood two rate ($N = 2$)		
Marginal likelihood four rate ($N = 4$)		
Marginal likelihood six rate ($N = 6$)		
Marginal likelihood eight rate ($N = 8$)		
Marginal likelihood ten rate ($N = 10$)		
Supported model?		

*you can edit this table

7 Estimating Branch-Specific Diversification Rates

In this second analysis we are interested in estimating the branch-specific diversification rates. We are going to use a very similar model to the one described in the previous section. However, now we are going to use the **dnHBDP** distribution instead which will require some slightly different parameterization and moves. The main difference, as mentioned above, is that the **dnHBDP** uses a data-augmentation scheme to sample the locations and parameters of rate-shift events across branches of the tree.

7.1 Read the tree

Begin by reading in the observed tree.

```
observed_phylogeny <- readTrees("data/primates_tree.nex")[1]
```

From this tree, we can get some helpful variables:

```
taxa <- observed_phylogeny.taxa()
root <- observed_phylogeny.rootAge()
tree_length <- observed_phylogeny.treeLength()
```

Additionally, we can initialize an iterator variable for our vector of moves:

```
mvi = 0
mni = 0
```

Finally, we create a helper variable that specifies the number of discrete rate categories, another helper variable for the expected number of rate-shift events, the total number of species, and the variation in rates.

```
NUM_RATE_CATEGORIES = 4
EXPECTED_NUM_EVENTS = 2
NUM_TOTAL_SPECIES = 367
H = 0.587405
```

Using these variables we can easily change our script, for example, to use more or fewer categories and test the impact.

7.2 Specifying the model

7.2.1 Priors on rates

Similar the previous section, we will set up the rate categories using the exact same model and `Rev` syntax. Thus, we first create our hyper-prior on the mean speciation rate, which is drawn from a lognormal distribution.

```
speciation_prior_mean <- ln( ln(NUM_TOTAL_SPECIES/2.0) / root_age )
speciation_mean ~ dnLognormal(mean=speciation_prior_mean, sd=H)
moves[++mvi] = mvScale(speciation_mean,lambda=1,tune=true,weight=5)
```

Additionally, we choose a fixed standard deviation of $H * 2$ for the speciation rates because it represents two orders of magnitude variance in the rate categories.

```
speciation_sd <- H*2
speciation_categories := fnDiscretizeDistribution( dnLognormal(ln(speciation_mean),
    speciation_sd), NUM_RATE_CATEGORIES )
```

We define the prior on the extinction rate in the same way as we did for the speciation rate, with the only difference that we allow for two orders of magnitude of uncertainty.

```
extinction_prior_mean <- ln( ln(NUM_TOTAL_SPECIES/2.0) / root_age )
extinction_mean ~ dnLognormal(mean=extinction_prior_mean,sd=H*2)
moves[++mvi] = mvScale(extinction_mean,lambda=1.0,tune=true,weight=3.0)
```

As with the speciation rate, we discretize the lognormal distribution into a finite number of rate categories.

```
extinction_categories := fnDiscretizeDistribution( dnLognormal(ln(extinction_mean), H)
    , NUM_RATE_CATEGORIES )
```

Now, we must create a vector that contains each combination of speciation- and extinction-rates. This allows the rate of speciation to change without changing the rate of extinction and vice versa. The resulting vector should be N^2 elements long. We call these the 'paired' rate categories.

```
k = 1
for(i in 1:NUM_RATE_CATEGORIES) {
    for(j in 1:NUM_RATE_CATEGORIES) {
        speciation[k]  := speciation_categories[i]
        extinction[k++] := extinction_categories[j]
    }
}
```

Next, we need a rate parameter for the rate-shifts events. We do not have much prior information about this rate but we can provide some realistic ranges. For example, we can specify a mean rate so that the resulting number of expected rate-shift events is 2 (as specified in our global variable **EXPECTED_NUM_EVENTS**). Furthermore, we can say that the 95% prior ranges exactly one order of magnitude. We achieve all this by specifying a lognormal prior distribution with mean **ln(EXPECTED_NUM_EVENTS/tree_length)** and standard deviation of **H**. Remember that this is only possible if the tree is known and not estimated simultaneously because only if the tree is do we also know the tree length. As usual for rate parameter, we apply a scaling move to the **event_rate** variable.

```
event_rate ~ dnLognormal( ln( EXPECTED_NUM_EVENTS/tree_length ), H)
moves[++mvi] = mvScale(event_rate,lambda=1,tune=true,weight=5)
```

Additionally, we need a parameter for the category of the process at root. We use a uniform prior distribution on the indices 1 to N^2 since we do not have any prior information in which rate category the process is at the root. The move for this random variable is a random integer walk because the random variable is defined only on the indices (*i.e.,* with real number).

```
root_category ~ dnUniformNatural(1,NUM_RATE_CATEGORIES * NUM_RATE_CATEGORIES)
moves[++mvi] = mvRandomIntegerWalk(root_category,weight=1)
```

7.2.2   Incomplete Taxon Sampling

We know that we have sampled 233 out of 367 living primate species. To account for this we can set the sampling parameter as a constant node with a value of 233 / 367.

```
rho <- observed_phylogeny.ntips() / NUM_TOTAL_SPECIES
```

7.2.3   Root age

The birth-death process requires a parameter for the root age. In this exercise we use a fix tree and thus we know the age of the tree. Hence, we can get the value for the root from the Magnuson-Ford and Otto (2012) tree. This is done using our global variable **root** defined above and nothing else has to be done here.

7.2.4   The time tree

Now we have all of the parameters we need to specify the full branch-specific birth-death model. We initialize the stochastic node representing the time tree.

```
timetree ~ dnHBDP(lambda=speciation, mu=extinction, rootAge=root, rho=rho, rootState=
    root_category, delta=event_rate, taxa=taxa )
```

And then we attach data to it.

```
timetree.clamp(observed_phylogeny)
```

This specific implementation of the branch-specific birth-death process augments the tree with rate-shift events. In order to sample the number, the location, and the types of the rate-shift events, we have to apply special moves to the tree. These moves will not change the tree but only the augmented rate-shift events. We use a **mvBirthDeathEvent** to add and remove events, a **mvEventTimeBeta** move to change the time and location of the events, and a **mvDiscreteEventCategoryRandomWalk** to change the the paired-rate category to which a rate-shift event belongs.

```
moves[++mvi] = mvBirthDeathEvent(timetree,weight=2)
moves[++mvi] = mvEventTimeBeta(timetree,weight=2)
moves[++mvi] = mvDiscreteEventCategoryRandomWalk(timetree,weight=2)
```

In this analysis, we are interested in the branch-specific diversification rates. So far we do not have any variables that directly give us the number of rate-shift events per branch or the rates per branch. Fortunately, we can construct deterministic variables and query these properties from the tree. These function are made available by the branch-specific birth-death process distribution.

```
num_events := timetree.numberEvents()
avg_lambda := timetree.averageSpeciationRate()
avg_mu     := timetree.averageExtinctionRate()
avg_net    := avg_lambda - avg_mu
avg_rel    := avg_mu / avg_lambda

total_num_events := sum( num_events )
```

Finally, we create a workspace object of our whole model using the **model()** function.

```
mymodel = model(speciation)
```

The **model()** function traversed all of the connections and found all of the nodes we specified.

7.3 Running an MCMC analysis

7.3.1 Specifying Monitors

For our MCMC analysis, we need to set up a vector of *monitors* to record the states of our Markov chain. First, we will initialize the model monitor using the **mnModel** function. This creates a new monitor variable that will output the states for all model parameters when passed into a MCMC function.

```
monitors[++mni] = mnModel(filename="output/primates_BSBD.log",printgen=10, separator =
    TAB)
```

Additionally, we create an extended-Newick monitor. The extended-Newick monitor writes the tree to a file and adds parameter values to the branches and/or nodes of the tree. We can thus print the tree with the average speciation and extinction rates, as well as the net diversification (speciation - extinction) and relative extinction (extinction / speciation) rates, for each branch into a file. We will need this file later to estimate and visualize the posterior distribution of the rates at the branches.

```
monitors[++mni] = mnExtNewick(filename="output/primates_BSBD.trees", isNodeParameter=
    FALSE, printgen=10, separator = TAB, tree=timetree, avg_lambda, avg_mu, avg_net,
    avg_rel)
```

Finally, create a screen monitor that will report the states of specified variables to the screen with **mnScreen**:

```
monitors[++mni] = mnScreen(printgen=10, event_rate, mean_speciation, root_category,
    total_num_events)
```

7.3.2 Initializing and Running the MCMC Simulation

With a fully specified model, a set of monitors, and a set of moves, we can now set up the MCMC algorithm that will sample parameter values in proportion to their posterior probability. The `mcmc()` function will create our MCMC object:

```
mymcmc = mcmc(mymodel, monitors, moves)
```

First, we will run a pre-burnin to tune the moves and to obtain starting values from the posterior distribution.

```
mymcmc.burnin(generations=1000,tuningInterval=200)
```

Now, run the MCMC:

```
mymcmc.run(generations=5000)
```

When the analysis is complete, you will have the monitored files in your output directory. You can then visualize the branch-specific rates by attaching them to the tree. This is actually done automatically in our `mapTree` function.

```
treetrace = readTreeTrace("output/primates_BSBD.trees", treetype="clock")
map_tree = mapTree(treetrace,"output/primates_BSBD_MAP.tree")
```

Now you can open the tree in `FigTree`.

→ The `Rev` file for performing this analysis: `mcmc_BSBD.Rev`.

7.4 Exercise

- Run an MCMC simulation to estimate the posterior distribution of the speciation rate and extinction rate.

- Visualize the branch-specific rates in `FigTree`.

- Do you see evidence for rate decreases or increases? What is the general trend?

- Run the analysis using a different number of categories, *e.g.,* 2 or 6. How do the rates change?

- Modify the model by specifying a prior on the log-diversification and log-turnover rate and then estimate the diversification rates through time. Do you see any differences in the estimates?

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