

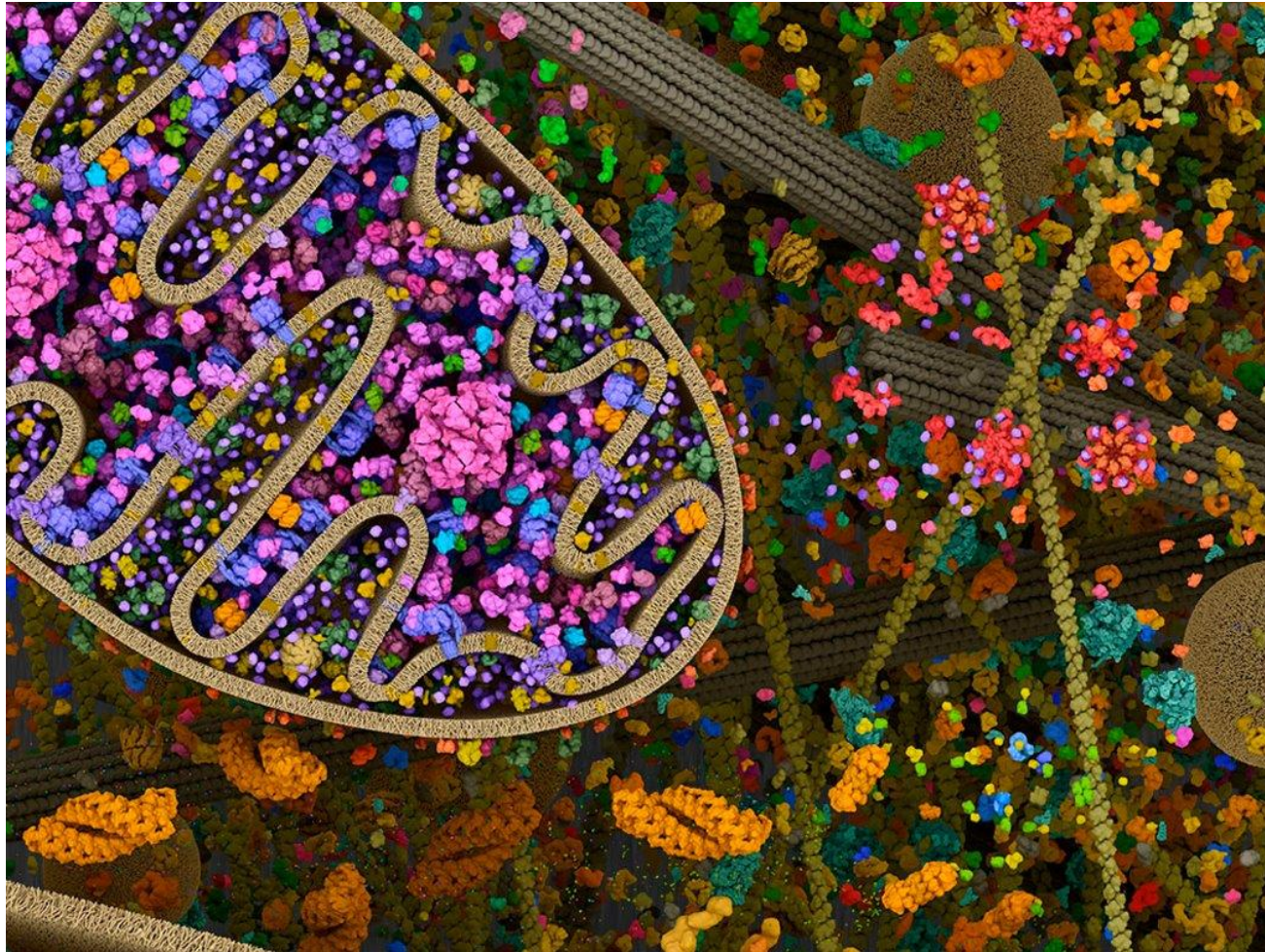
Computational Models of Self Organization of Neuronal Cytoskeleton

Christopher Manry, Calvin Sprouse, Dr. Craig

CWU Computational Biophysics Lab

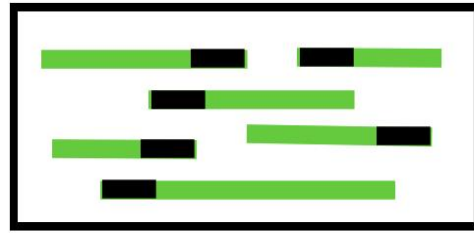
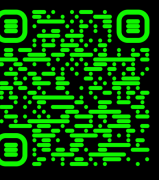


Physics of Cell Biology



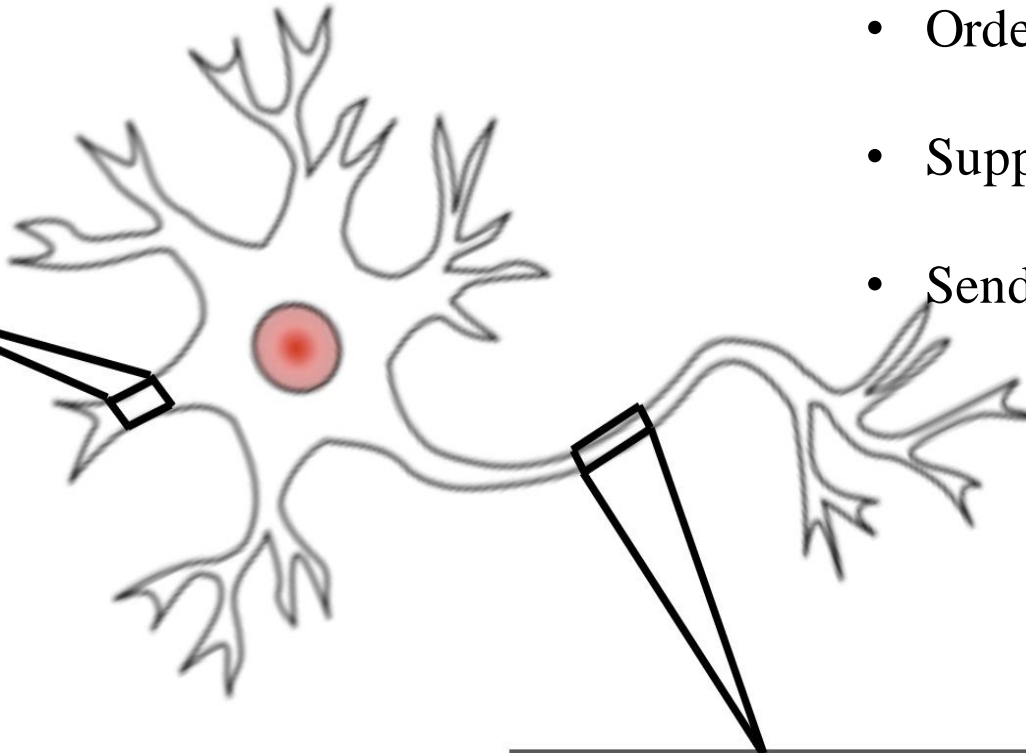
- Crowded
- Highly viscous
- Intricately-timed dynamic processes
- Physical principles determine biological function

Microtubule Organization in the Neuron

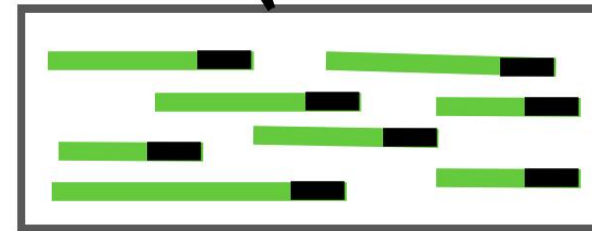


Dendrite

- Unordered
- Supports multi-directional transport
- Receives incoming signals

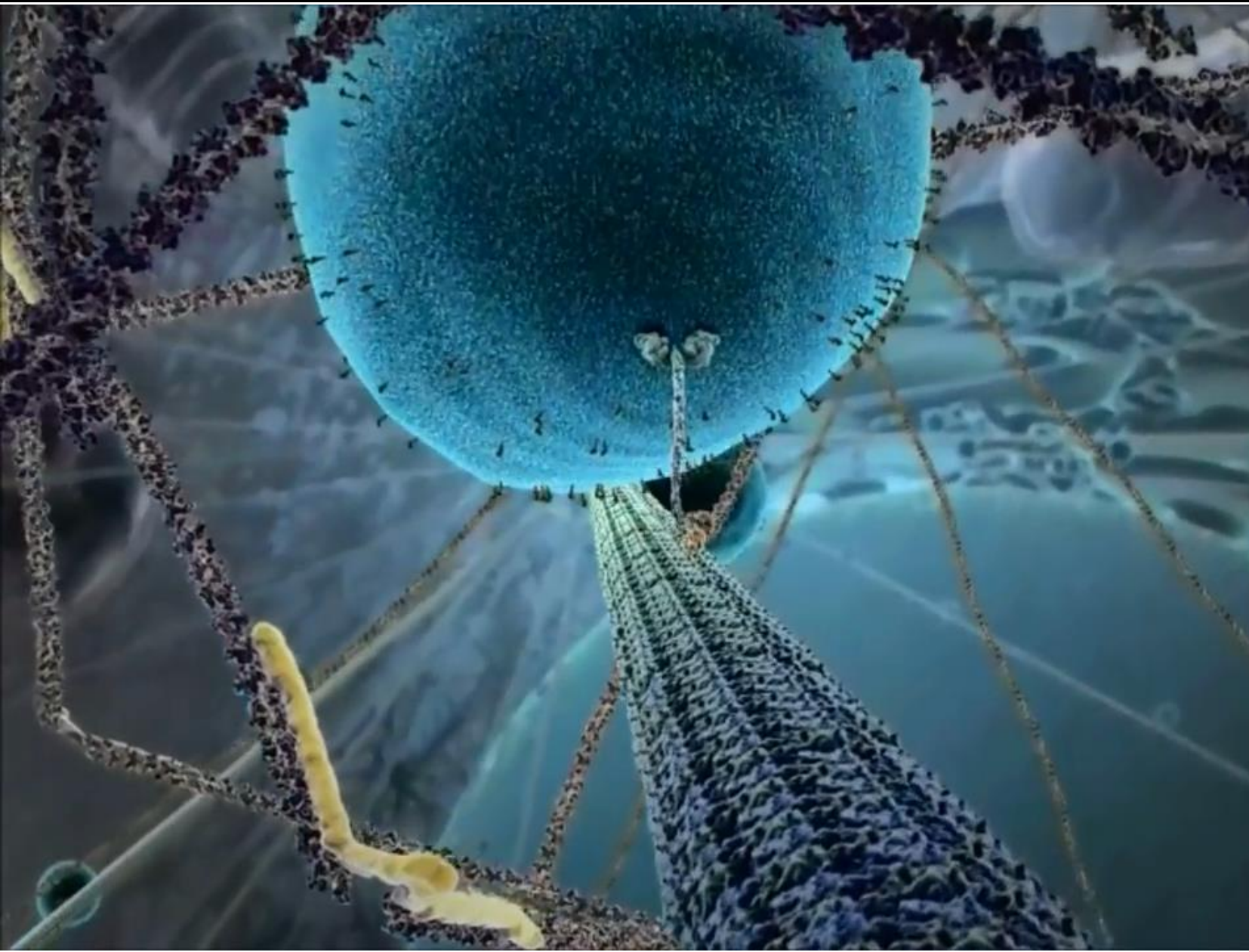


- Ordered in healthy neurons
- Supports one-way transport
- Sends the outgoing signals



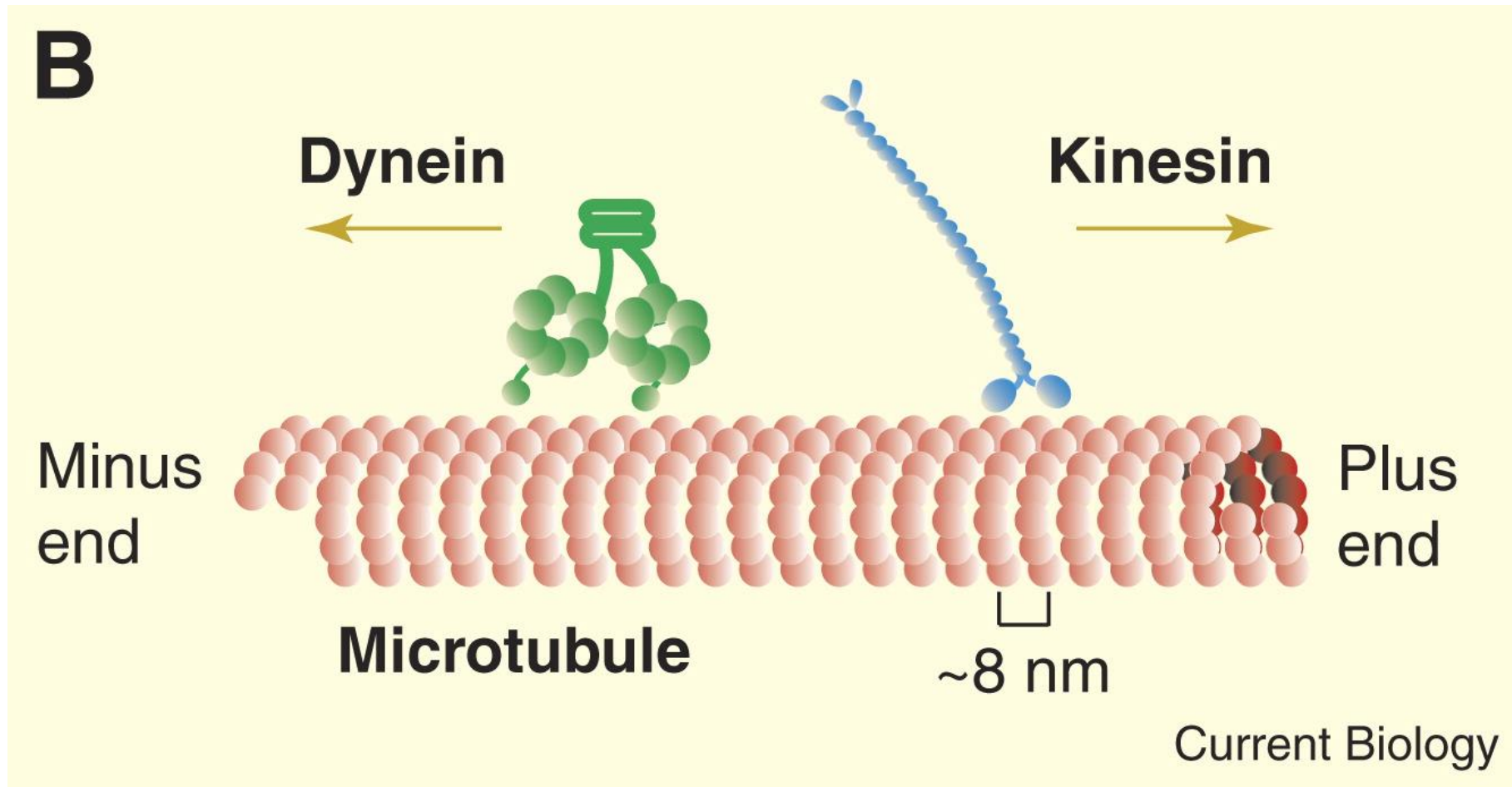
Axon

The Role of Motor Proteins



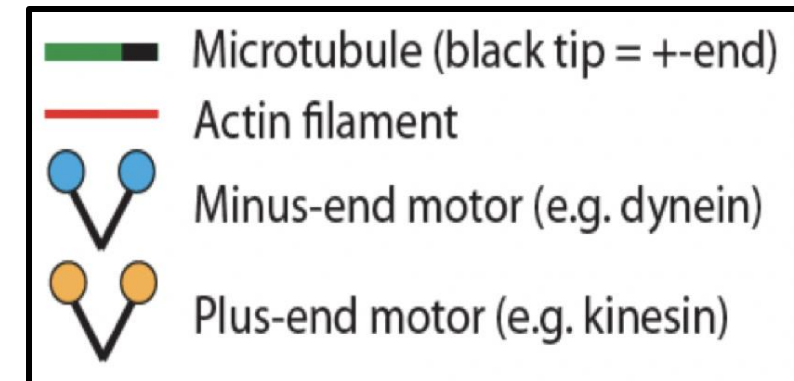
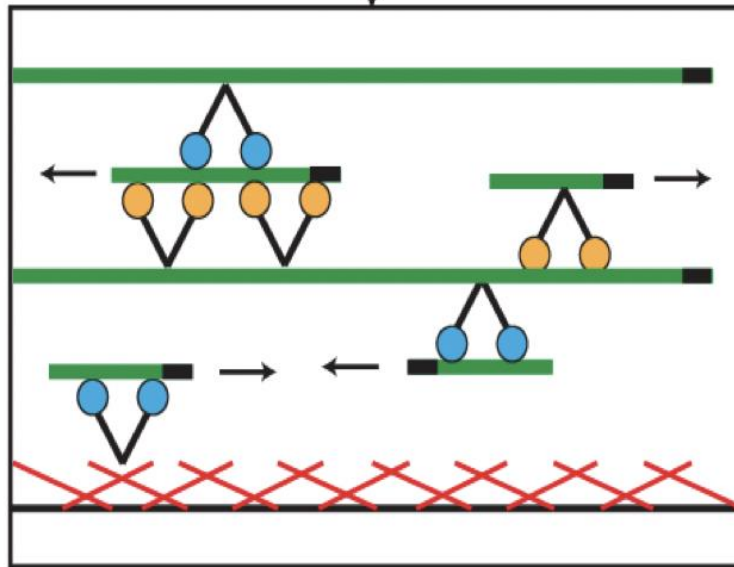
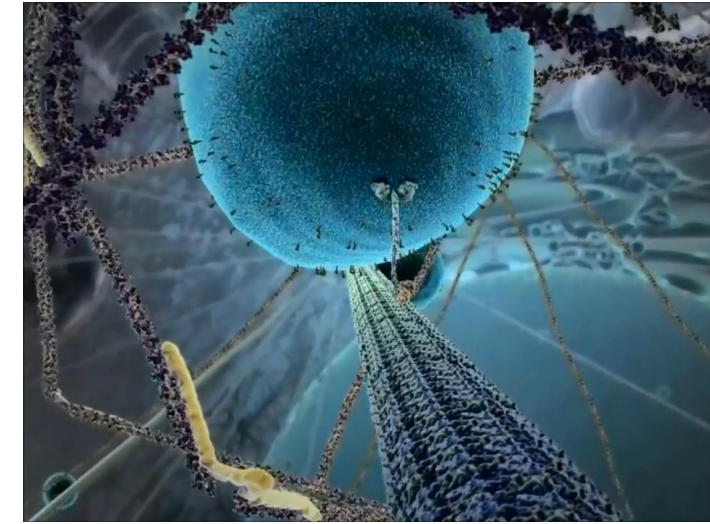
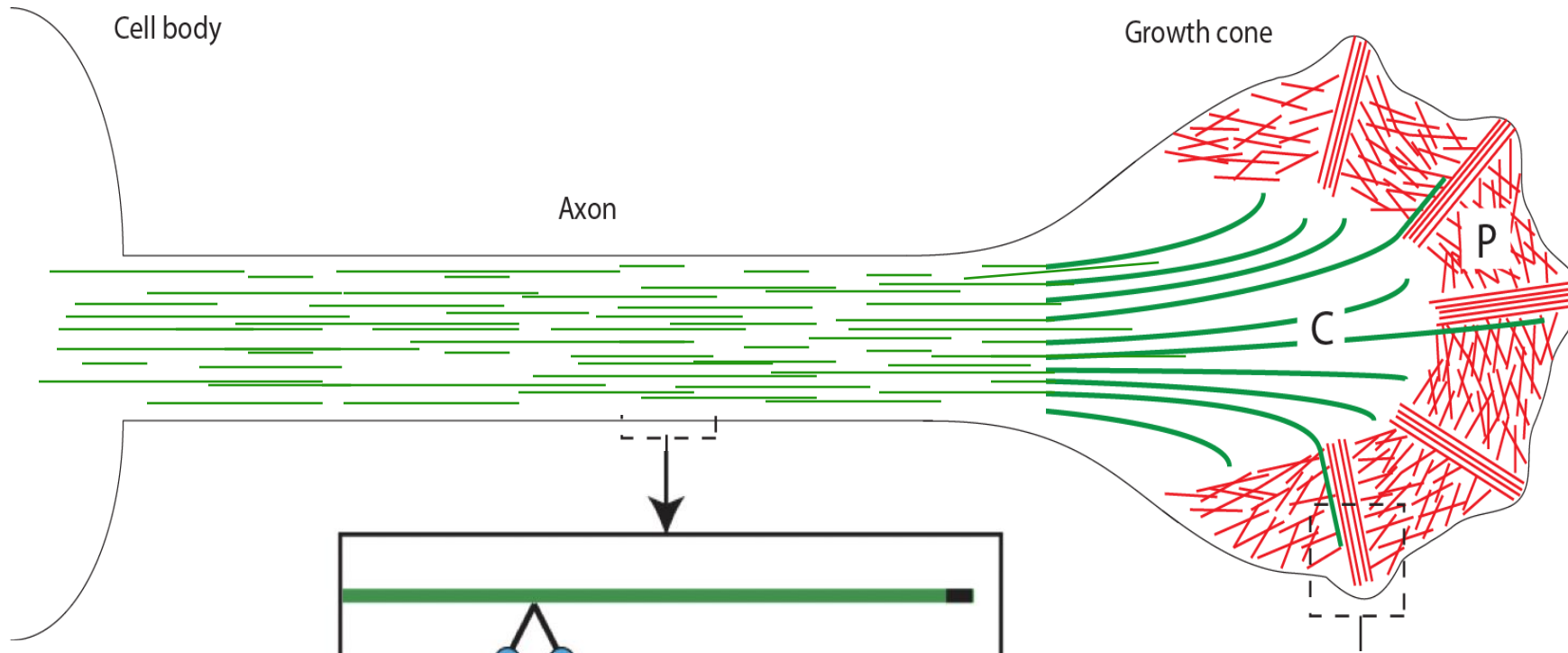
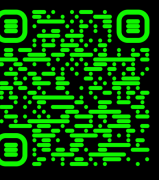
- A motor protein carries cargo along a microtubule
- The motion of the “feet” has been smoothed; on the cellular level it is random with a tendency towards forward motion

The Type of Motor Proteins



Each motor has a preferred direction to walk on the microtubule. Kinesin walks in the same direction the microtubule points while Dynein walks in the opposite direction.

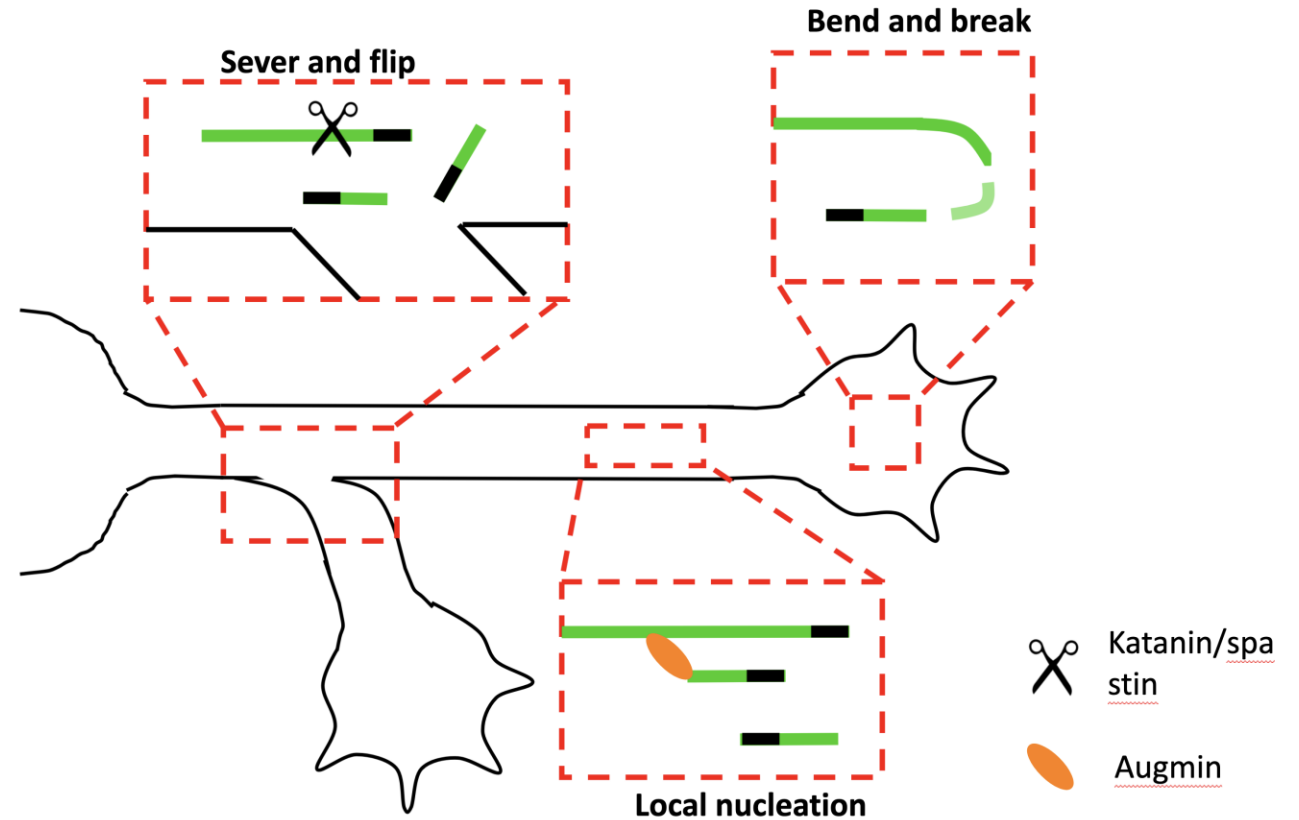
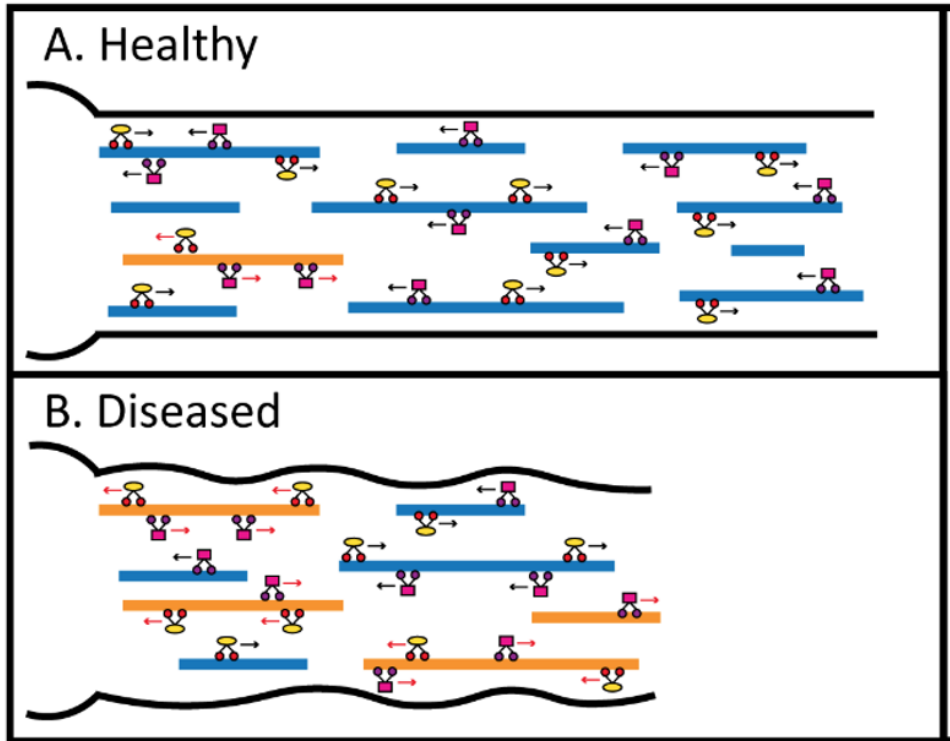
Microtubule Organization in the Axon



Corruption of the Polarity Pattern

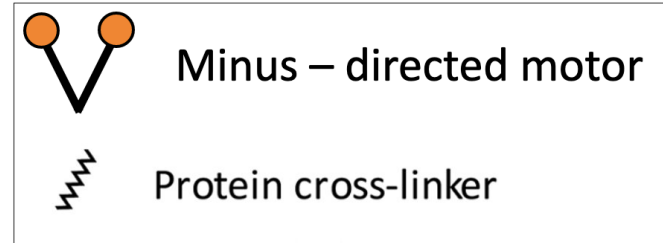
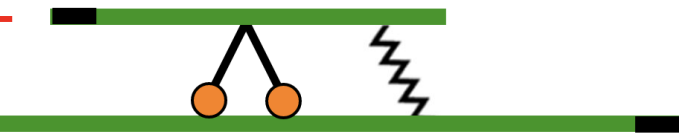


Axon's uniform polarity pattern is continually at risk of corruption



Damage to the polarity pattern may arise from a variety of sources. Significant local damage to the polarity pattern impacts cargo transport and creates traffic jams of molecular motor proteins.

Computational Model for Forces on Microtubules



Number of attached cytoplasmic dynein motors, N_d :

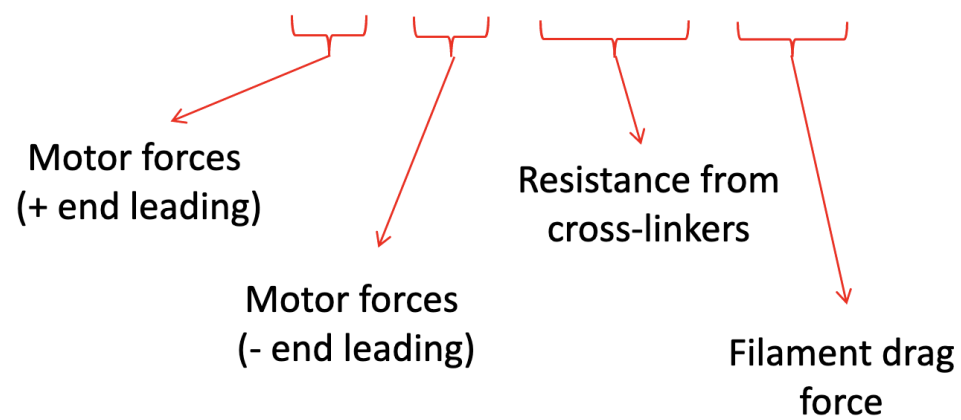
$$\frac{dN_d}{dt} = r_{d,on} - r_{d,off}N_d$$

Attachment
depends on # of
available sites)

Detachment
(Force-dependent)

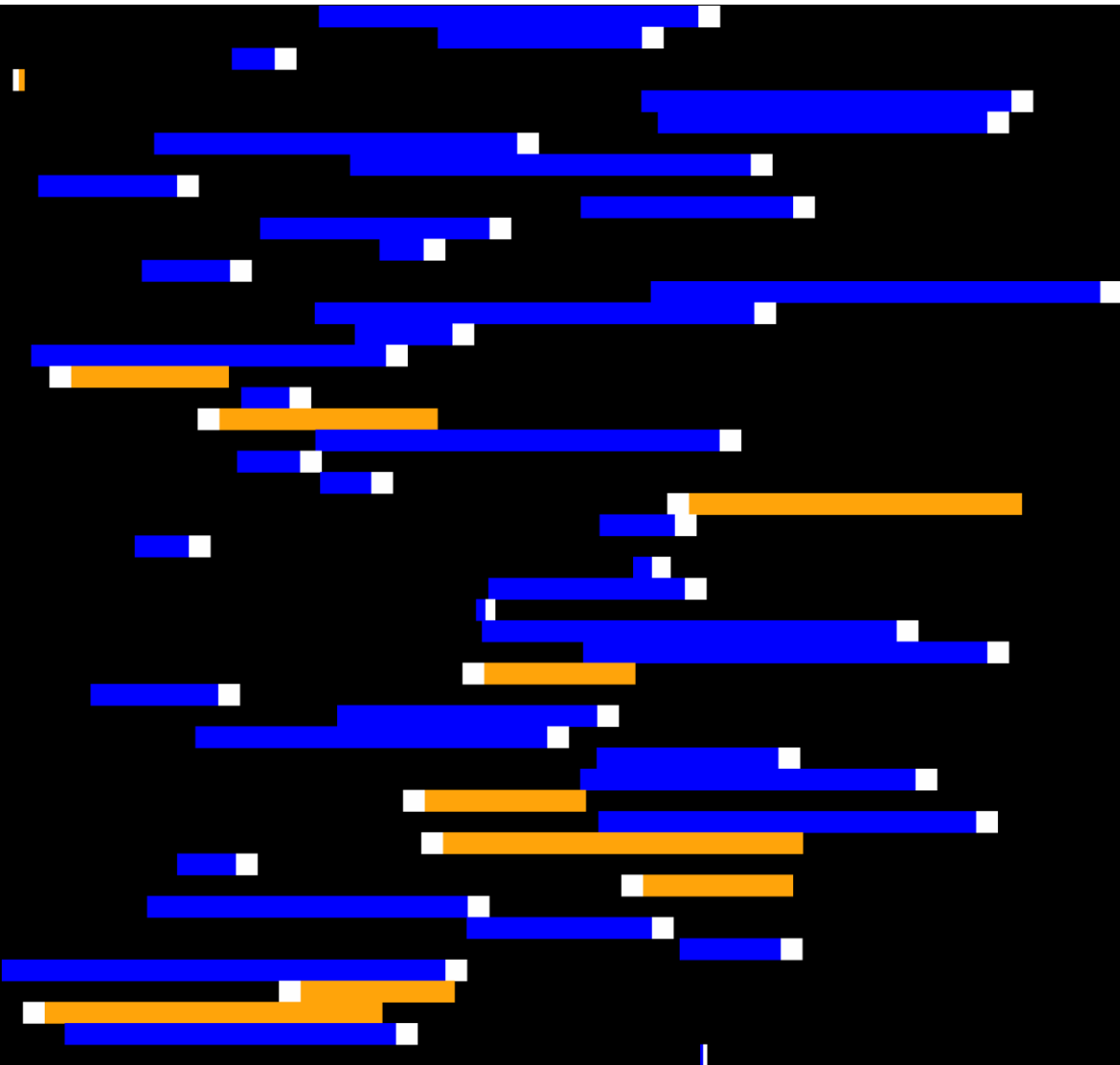
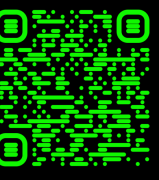
Balance of forces:

$$F_{d+} = F_{d-} + N_x \gamma v + \xi L v$$



- Net 0 force assumption due to high fluid viscosity
- Essentially objects do not move except when under active forces

Computational Model for Forces on MTs



Frame: 0 Time: 0.0

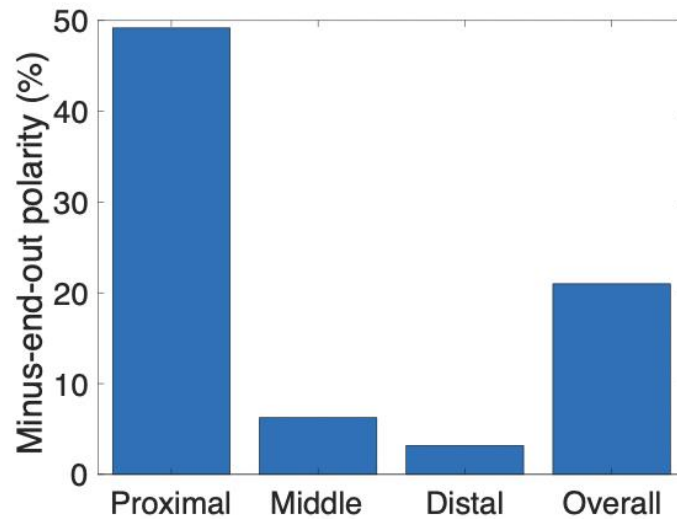
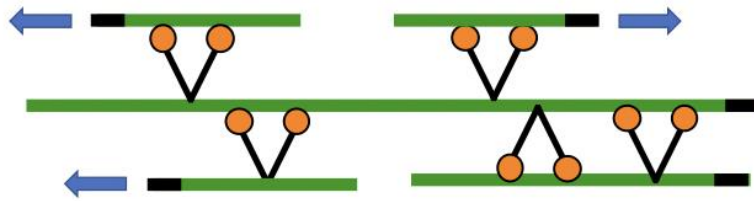
```
191
192
193
194
195 % Update position at time i+1 based on position and velocity at time i
196
197 A(i+1,2,j)=A(i,2,j)+A(i,3,j)*dt; % Translocate MT based on vel calculated for this time step
198 A(i+1,11,j)=A(i,11,j); % Default is for orientation to stay the same. Several possibilities later for this to flip.
199 % Apply conditions for if MT reaches a boundary
200 if (A(i+1,2,j)<0) % MT has been cleared from axon into cell body
201     'MT cleared';
202     % Replaced cleared MT with a new short growing MT, random
203     % location and orientation
204     A(i+1,2,j)=Laxon*rand([1,1,1]); % random location
205     A(i+1,3,j)=0.0; % Initial velocity (v)
206     A(i+1,4,j)=0.0; % Initial force (Fdp)
207     A(i+1,5,j)=0.0; % Initial force (Fdm)
208     A(i+1,6,j)= 0.0; % Initial dynein attachment number, forward pulling (Ndp)
209     A(i+1,7,j)=0.0; % Initial dynein attachment number, backward pulling (Ndm)
210     A(i+1,8,j)=0.0; % Initial cross-linker attachment number, parallel MTs (Nxpar)
211     A(i+1,9,j)=0.0; % Initial cross-linker attachment number, anti-parallel MTs (Nxanti)
212     A(i+1,10,j)=0.1; % Short initial length, 0.1micron for newly nucleated MT
213
214 % Fraction of plus-out MTs at new MT's location
215 loc=round(A(i+1,2,j));
216 Fmin=Polarity(loc+1,3);
217 if(rand<Fmin) % Newly nucleated MT has random orientation
218     A(i+1,11,j)=1; % minus-end-out
219     'minus out new MT';
220 else
221     A(i+1,11,j)=0; % plus-end-out
222     'plus out new MT';
223 end
224
225 A(i+1,12,j)=1; % dynamic and growing
226 elseif(A(i+1,2,j)>Laxon) % MT hits distal end
227     A(i+1,2,j)=Laxon; % Can't grow further
228     A(i+1,12,j)=0; % Switches to stable
229 end
230
231 % Update length of dynamic MTs
232 if(A(i,12,j)==0)
233     A(i+1,10,j)=A(i,10,j);
234 elseif(A(i,12,j)==1)
235     A(i+1,10,j)=A(i,10,j)+V_MTPoly*dt;
236     if(A(i+1,10,j)>Laxon)
237         A(i+1,10,j)=Laxon;
238     end
239 elseif(A(i,12,j)==2)
240     A(i+1,10,j)=A(i,10,j)-V_MTdePoly*dt;
241 end
242
243 % Apply conditions for if MT length shrinks to zero, nucleate new MT
244 if(A(i+1,10,j)<0)
245
246     A(i+1,3,j)=0.0; % Initial velocity (v)
247     A(i+1,4,j)=0.0; % Initial force (Fdp)
248     A(i+1,5,j)=0.0; % Initial force (Fdm)
249     A(i+1,6,j)= 0.0; % Initial dynein attachment number, forward pulling (Ndp)
250     A(i+1,7,j)=0.0; % Initial dynein attachment number, backward pulling (Ndm)
251     A(i+1,8,j)=0.0; % Initial cross-linker attachment number, parallel MTs (Nxpar)
252     A(i+1,9,j)=0.0; % Initial cross-linker attachment number, anti-parallel MTs (Nxanti)
253     A(i+1,10,j)=0.1; % Short initial length, 0.1micron for newly nucleated MT
254
255 %
256 A(i+1,2,j)=Laxon*rand([1,1,1]); % random location
```

Result:

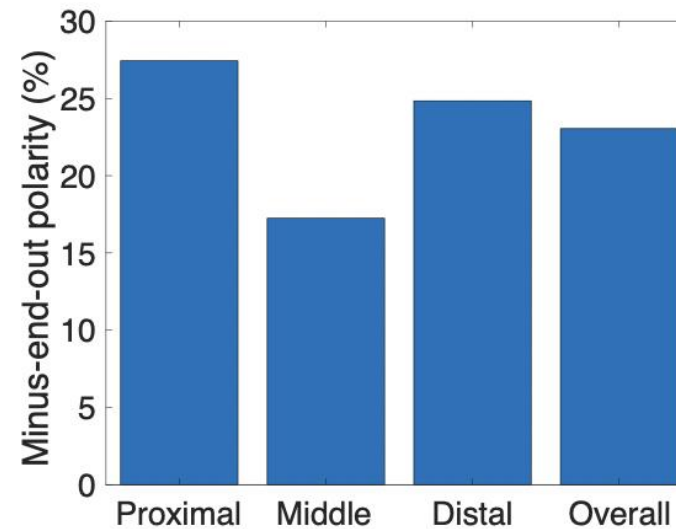
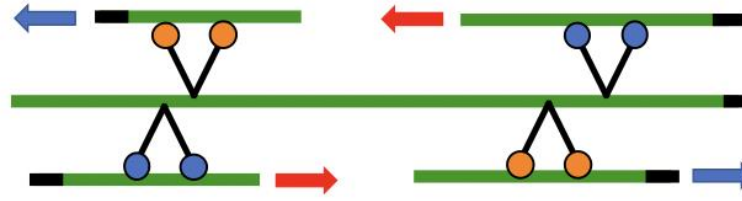


Kinesin prevents Dynein-based polarity sorting

(a) Dynein only



(b) Dynein and kinesin

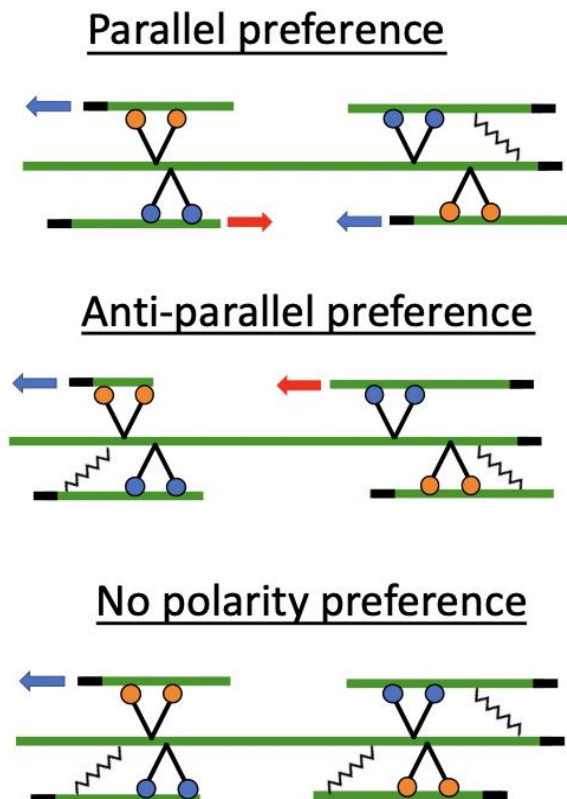


Result:

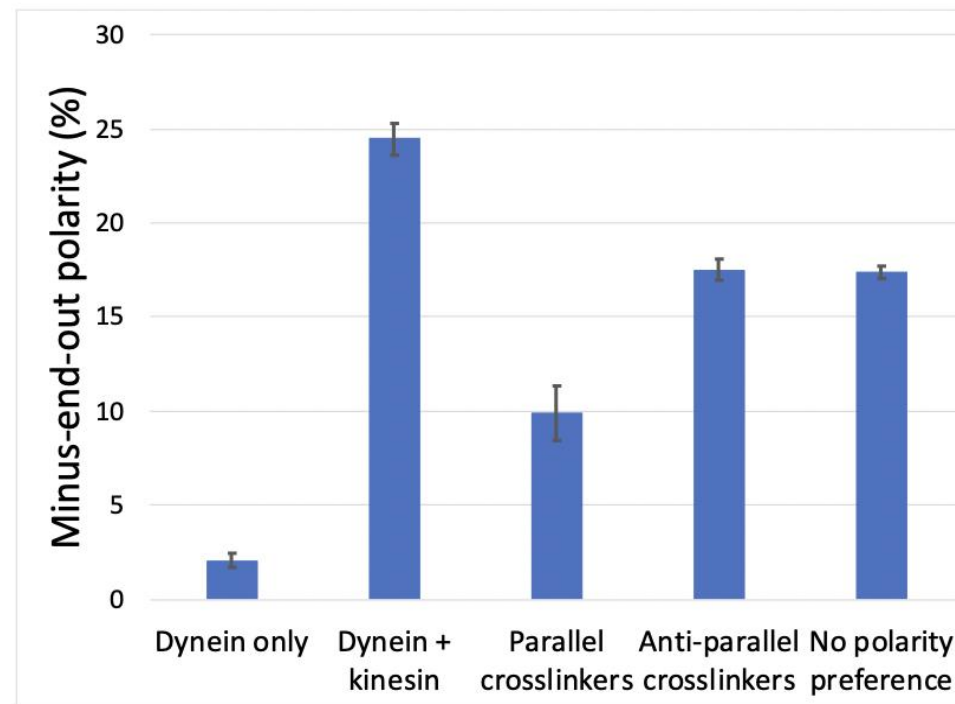


Certain types of crosslinkers impact polarity sorting more than others

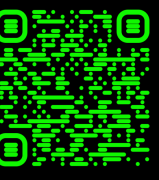
(c) Hypothetical Crosslinkers



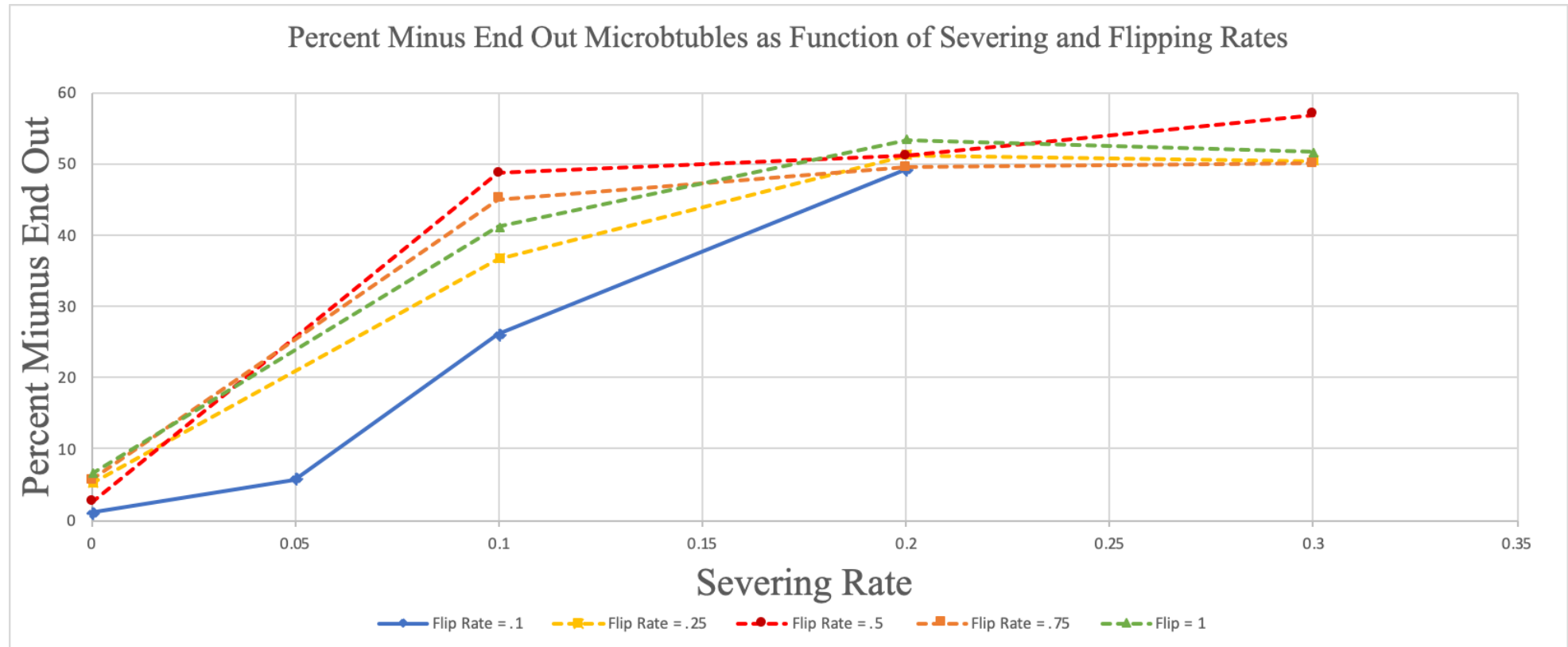
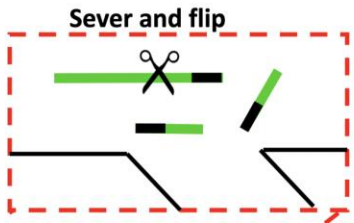
(d) Simulated distal polarity



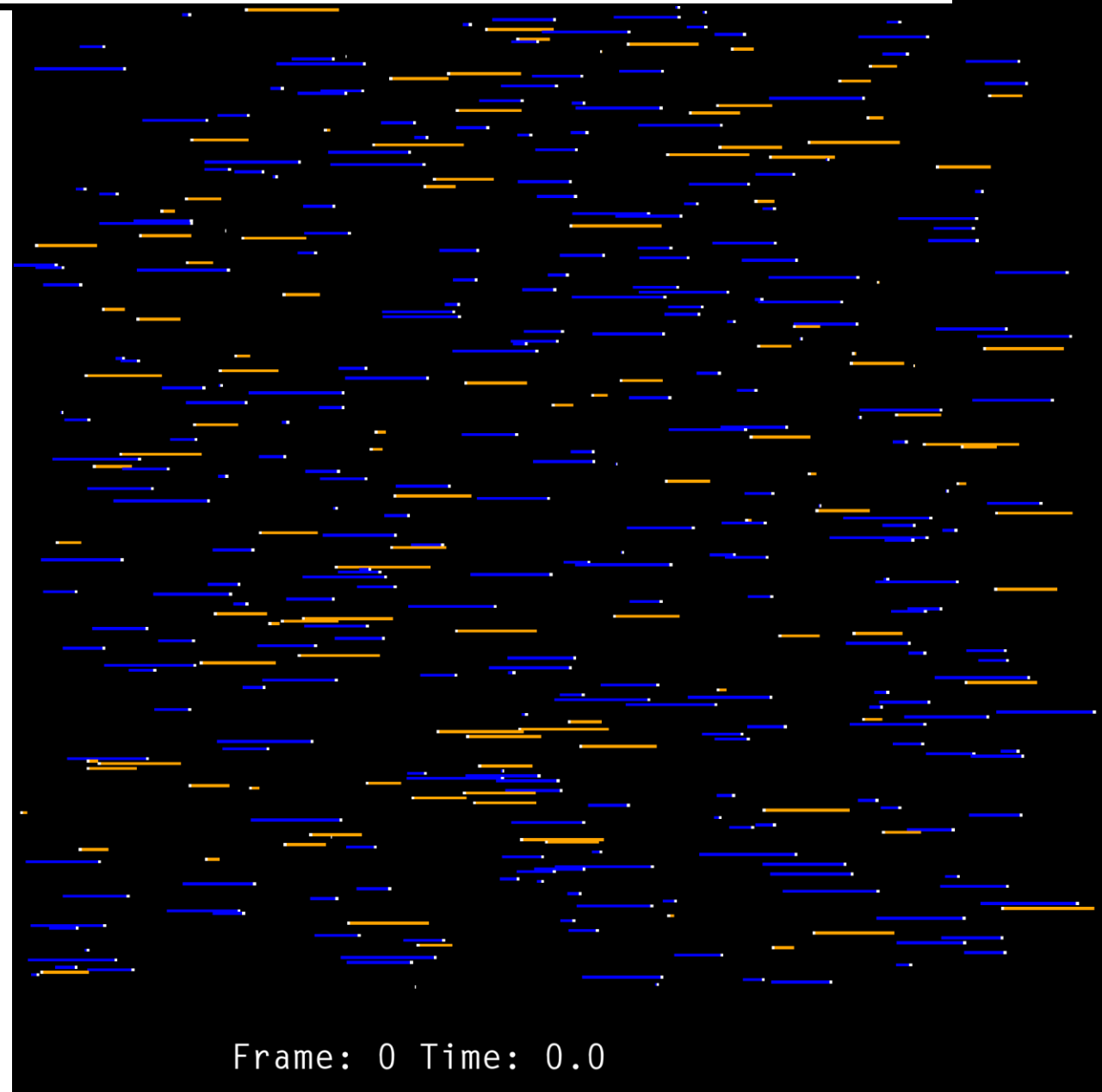
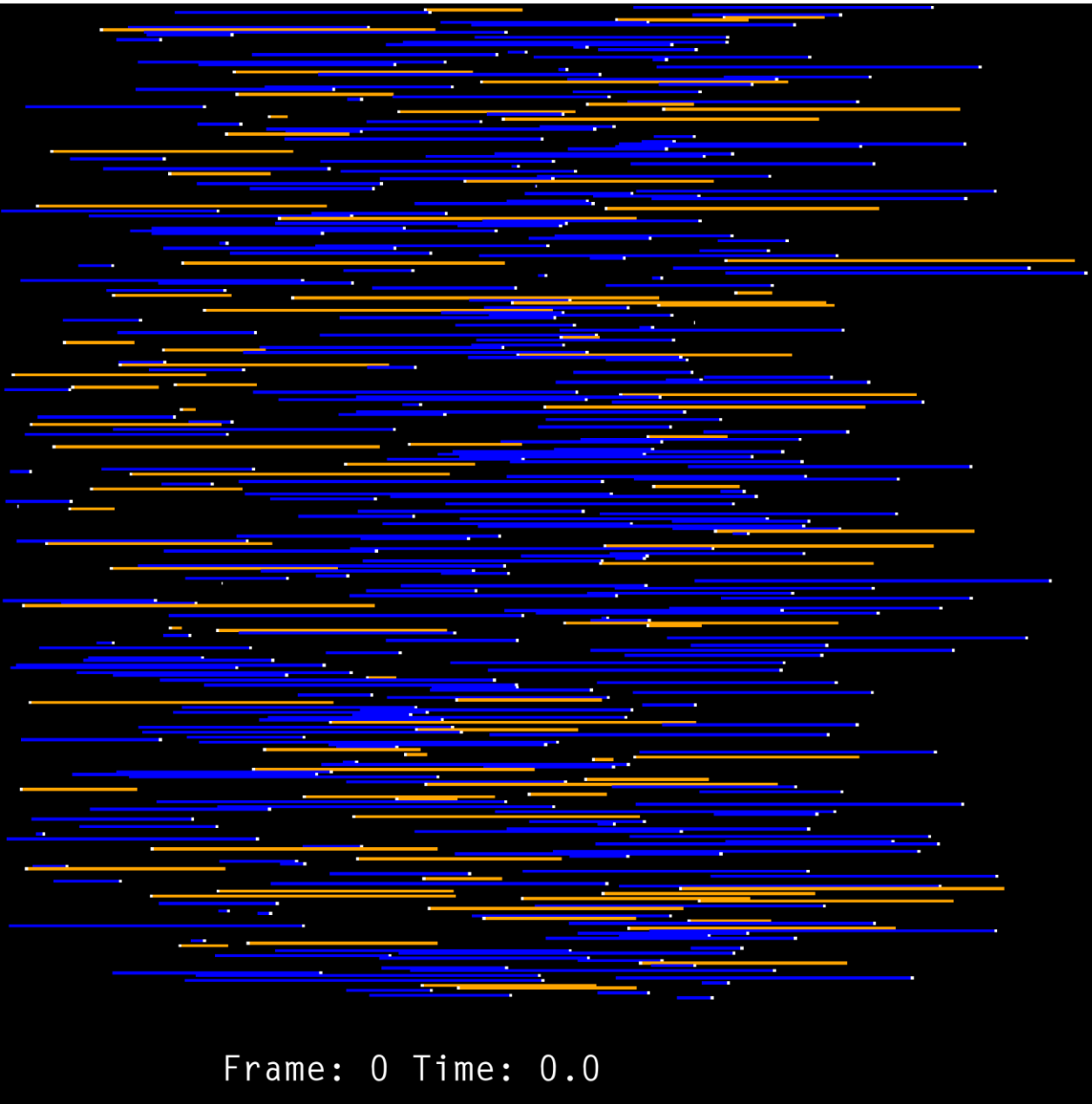
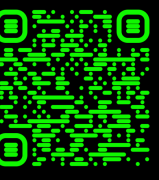
Result:



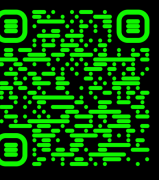
Severing and flipping are important in moderation for polarity sorting



Result:



Conclusion

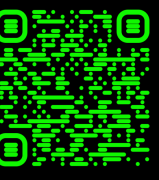


- Our models show how severing and flipping impact polarity sorting, being necessary but only to an extent
 - We can visualize this model to gain further intuition and demonstrate experimentally testable predictions
-

Next Steps

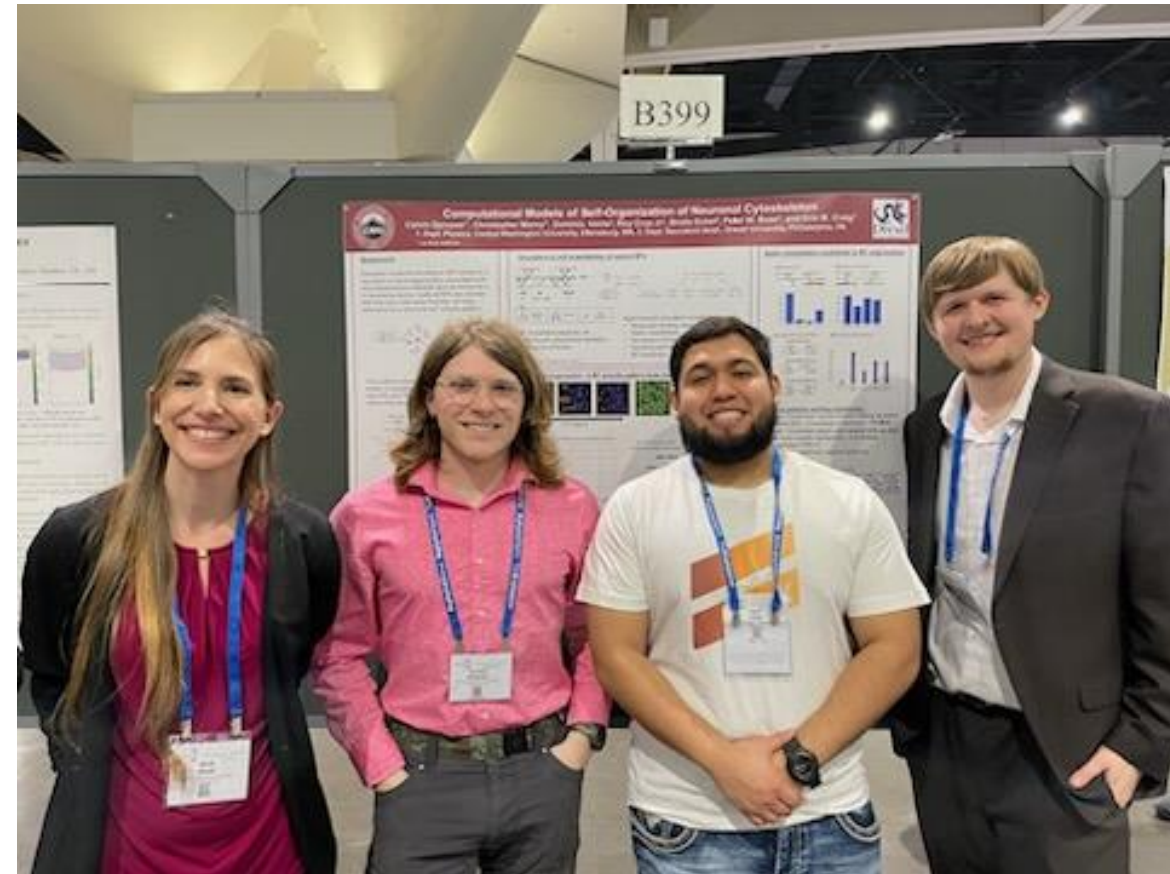
- Experimental test of model predictions.
Related study: Rao et al., 2017.
- Investigate the impact of polarity flaws on neuronal function.
Related study:
Eckel et al., 2022.

Acknowledgements

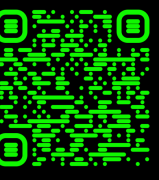


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References



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2. Craig et al. (2017), Polarity sorting of axonal microtubules: a computational study, Mol. Biol. Cell, 28(23):3271–3285.
3. Rao et al. (2017), Cytoplasmic dynein transports axonal microtubules in a polarity-sorting manner, Cell Reports, 19:2210-2219.
4. Eckel et al. (2022). Microtubule polarity flaws as a treatable driver of neurodegeneration., Brain Research Bulletin, 192:208-215.

See more animations here!

