

Diversidades alfa y beta en datos metagenomicos de shotgun de fresa

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Pre-procesamiento de los datos

DIVERSIDADES CON TODO EL CONJUNTO DE DATOS Y LOS DATOS FILTRADOS POR CALIDAD

Los datos fueron entregados en una carpeta de drive <https://drive.google.com/drive/folders/1x0106TYUr54gfqE6uod3g5qN8DQ5x>. Para poderlos usar en el servidor, se descargaron en mi maquina local y luego se pasaron por ssh al servidor.

En el equipo local

```
# $ scp Downloads/kraken_results-20230203T201634Z-001.zip camila@132.248.196.39:/home/camila/GIT/Tesis_Metagenomica/
# camila@132.248.196.39's password:
# kraken_results-20230203T201634Z-001.zip          100%   12MB   2.2MB/s  00:05
# $ scp Downloads/fastp_results-20230203T175936Z-001.zip camila@132.248.196.39:/home/camila/GIT/Tesis_Metagenomica/
# camila@132.248.196.39's password:
# fastp_results-20230203T175936Z-001.zip          100% 2728KB  2.1MB/s  00:01
# $ scp Downloads/bracken_results-20230203T203724Z-001.zip camila@132.248.196.39:/home/camila/GIT/Tesis_Metagenomica/
# camila@132.248.196.39's password:
# bracken_results-20230203T203724Z-001.zip         100% 3798KB  2.4MB/s  00:01
```

En el servidor

```
# $ unzip kraken_results-20230203T201634Z-001.zip
# $ unzip bracken_results-20230203T203724Z-001.zip
# $ unzip fastp_results-20230203T175936Z-001.zip
```

Luego de tener las muestras Kraken y bracken en el servidor con RStudio; es necesario generar el archivo **.biom**; para esto, se debe activar el ambiente de conda **metagenomics**, **kraken-biom** es un programa ampliamente utilizado para M a partir de la salida de kraken.

Con esto se obtiene una matriz de abundancia a partir de los archivos de salida de Kraken, que permite el uso de el paquete Phyloseq en R, que nos permiten analizar la diversidad y la abundancia mediante la manipulación de datos de asignación taxonómica.

```
# $ conda activate metagenomics
# $ kraken-biom kraken_results/* --fmt json -o fresa_kraken.biom
```

```

getwd()

## [1] "/home/camila/GIT/Tesis_Maestria/Analisis_Comparativo/Fresa_Solena"

# if (!requireNamespace("BiocManager", quietly = TRUE))
# +   install.packages("BiocManager")
# BiocManager::install("phyloseq")

# install.packages(c("ggplot2", "readr", "patchwork"))

```

Phyloseq es un paquete de Bioconductor (Open Source Software For Bioinformatics) para la manipulación y análisis (herramienta para importar, guardar, analizar y visualizar) de datos metagenómicos generados por metodologías de secuenciación de alto rendimiento.

```

library("phyloseq")
library("ggplot2")
library("igraph")

##
## Attaching package: 'igraph'

## The following objects are masked from 'package:stats':
## 
##     decompose, spectrum

## The following object is masked from 'package:base':
## 
##     union

library("readr")
library("patchwork")
library("vegan")

## Loading required package: permute

##
## Attaching package: 'permute'

## The following object is masked from 'package:igraph':
## 
##     permute

## Loading required package: lattice

## This is vegan 2.6-4

##
## Attaching package: 'vegan'

## The following object is masked from 'package:igraph':
## 
##     diversity

```

```

library("GUniFrac")
library("pbkrtest")

## Loading required package: lme4

## Loading required package: Matrix

#library("BiodiversityR")
library("kableExtra")

## Registered S3 method overwritten by 'httr':
##   method      from
##   print.response rmutil

library("RColorBrewer")

```

Datos kraken

Importamos los datos kraken en un archivo biom

```

setwd("/home/camila/GIT/Tesis_Maestria/Data/fresa_solenia")
fresa_kraken <- import_biom("fresa_kraken.biom")
class(fresa_kraken) # objeto phyloseq

## [1] "phyloseq"
## attr(,"package")
## [1] "phyloseq"

```

Queremos acceder a los datos que contiene nuestro objeto phyloseq **fresa_kraken**

Primero la tabla de taxonomia

```
#fresa_kraken@tax_table@.Data)
```

Cambiamos los nombres de las columnas a los niveles taxonomicos

```
colnames(fresa_kraken@tax_table@.Data) <- c("Kingdom", "Phylum", "Class", "Order", "Family", "Genus", "Species")
```

Queremos cortar la parte inicial nombres, ya que aparecen, por ejemplo: “Bacteria” y queremos que solo se vea “Bacteria” substring(). Este comando ayuda a extraer o reemplazar caracteres en un vector.

```
fresa_kraken@tax_table@.Data <- substr(fresa_kraken@tax_table@.Data, 4, 100)
#View(fresa_kraken@tax_table@.Data)
```

Queremos ver la tabla de OTUs, esta tabla contiene las abundancias de los otus de cada muestra

```
#fresa_kraken@otu_table@.Data
```

Queremos cortar los nombres de las muestras para que coincida con los metadatos

```
colnames(fresa_kraken@otu_table@.Data) <- substr(colnames(fresa_kraken@otu_table@.Data), 1, 6)
#View(fresa_kraken@otu_table@.Data)
```

Cargar los metadatos

Ya que hay un desface de dos muestras entre los metadatos y las muestras de la otu_table se deben ver cuales son y quitarlas del archivo de metadatos

```
# $ ls kraken_results |cut -d'.' -f1 > lista_kraken.txt
# $ ls metadata.csv |cut -d',' -f1 > lista_metadata.txt
# $ wc *txt
# $ cat lista_metadata.txt lista_kraken.txt | sort | uniq -c
# $ cat lista_metadata.txt lista_kraken.txt | sort | uniq -c | sort | head
# $ 1 MD2145
# $ 1 MD2146
# $ 2 MD2055
# $ 2 MD2056
```

Eliminamos las dos muestras que no estaban en nuestra otu_table y cargamos los metadatos

```
metadata_fresa <- read.csv2("/home/camila/GIT/Tesis_Maestria/Data/fresa_solenia/metadata.csv", header = 1)
#metadata_fresa <- sample_data(metadata_fresa)
```

luego hacemos que los metadatos pertenezcan al objeto phyloseq en la sección de **sam_data**

```
fresa_kraken@sam_data <- sample_data(metadata_fresa)
```

Creamos una columna extra en sam_data por necesidad de funcionamiento de mas adelante

```
fresa_kraken@sam_data$Sample <- row.names(fresa_kraken@sam_data)
colnames(fresa_kraken@sam_data) <- c('Treatment', 'Samples')
#View(fresa_kraken)
```

Ahora tenemos tambien la tabla de datos de calidad (fastp_kraken_summary),

para ver que muestra podemos eliminar de nuestro dataset, que no cumpla ciertos estandares de calidad + ID de la muestra + Reads_B - Reads_Before -> total de reads crudos + Reads_A - Reads_After -> total de reads despues del analisis de calidad + Reads_diff -> diferencia en tre Reads_B y Reads_A + Q30_B -> porcentaje arriba de 30 (escala fred) antes del analisis de calidad + Q30_A -> porcentaje arriba de 30 (escala fred) despues del analisis de calidad + LowQua -> reads de baja calidad + N_reads -> readas que contienen N y se descartan + too_short -> no pasan el tamaño minimo de calidad + Duplication -> porcentaje de duplicados + LengthR1 -> longitud promedio de los reads + LengthR2 -> longitud promedio de los reads + Classified -> porcentaje de clasificados del total despues del filtrado

ejemplo muestra MD2055 -> contienen 97millones de reads antes del filtrado de calidad, y despues queda con 79millones filtro usado para eliminar muestras, que luego del filtrado de calidad contengan menos de 25millones de reads los que nos da 5 muestras a eliminar (MP2079,MP2080,MP2088,MP2109,MP2137)

Eliminiar las muestras de baja calidad, usando filtro de menos de 25 millones de reads luego del análisis de calidad se eliminan las muestras por su nombre

```
samples_to_remove <- c("MP2079", "MP2080", "MP2088", "MP2109", "MP2137")
fresa_kraken_fil <- prune_samples(!(sample_names(fresa_kraken) %in% samples_to_remove), fresa_kraken)
```

podemos comprobar el número de muestras antes y después del filtrado

```
nsamples(fresa_kraken) # 58
```

```
## [1] 58
```

```
nsamples(fresa_kraken_fil) # 53
```

```
## [1] 53
```

Queremos hacer un análisis de diversidad de nuestras muestras, para esto las dos métricas las usadas son: Diversidad Alfa y Beta

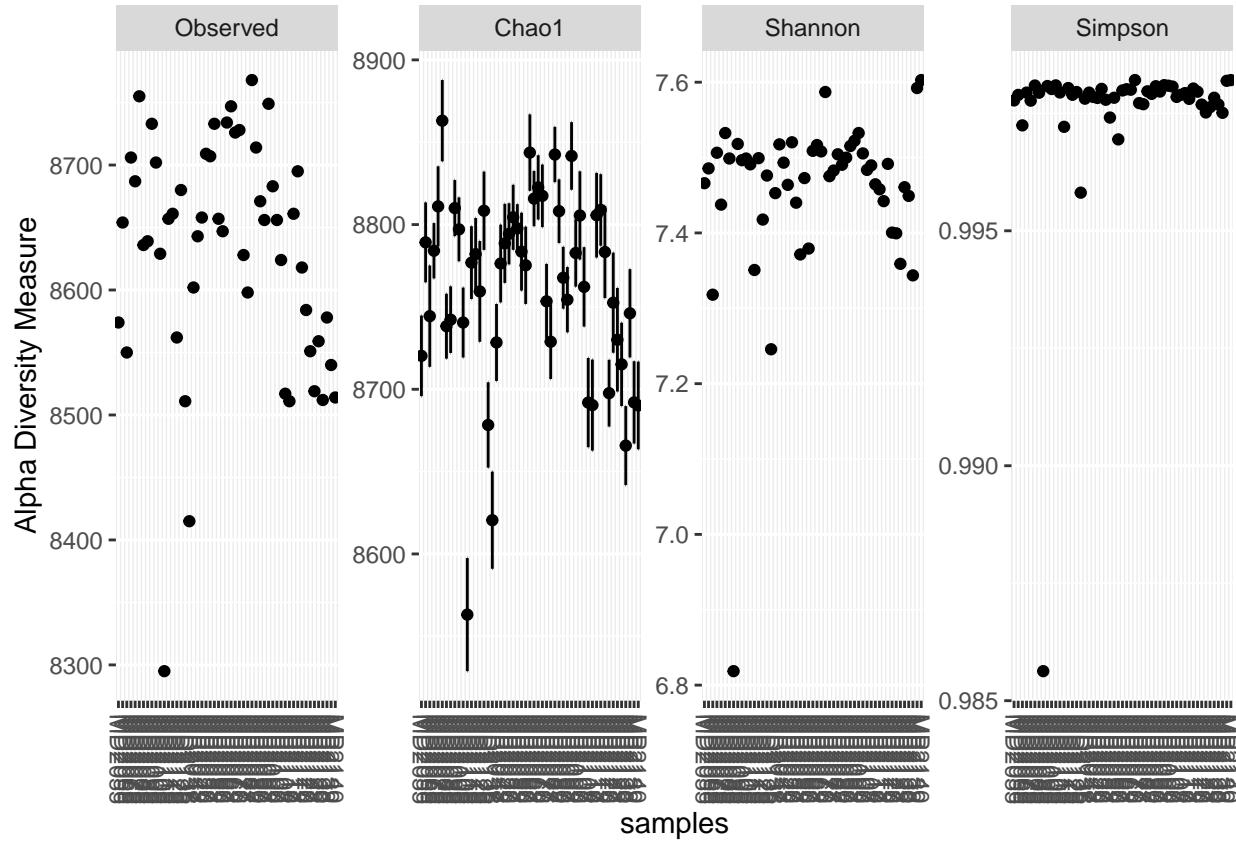
Diversidad Alfa

Esta representa la riqueza de las muestras, es decir el número de especies diferentes en ese ambiente o la abundancia de especies en ese ambiente. Para medir esta diversidad se tienen diferentes índices de medida, entre ellos los índices Shannon, Simpson, Chao1, entre otros.

```
index = estimate_richness(fresa_kraken_fil)
```

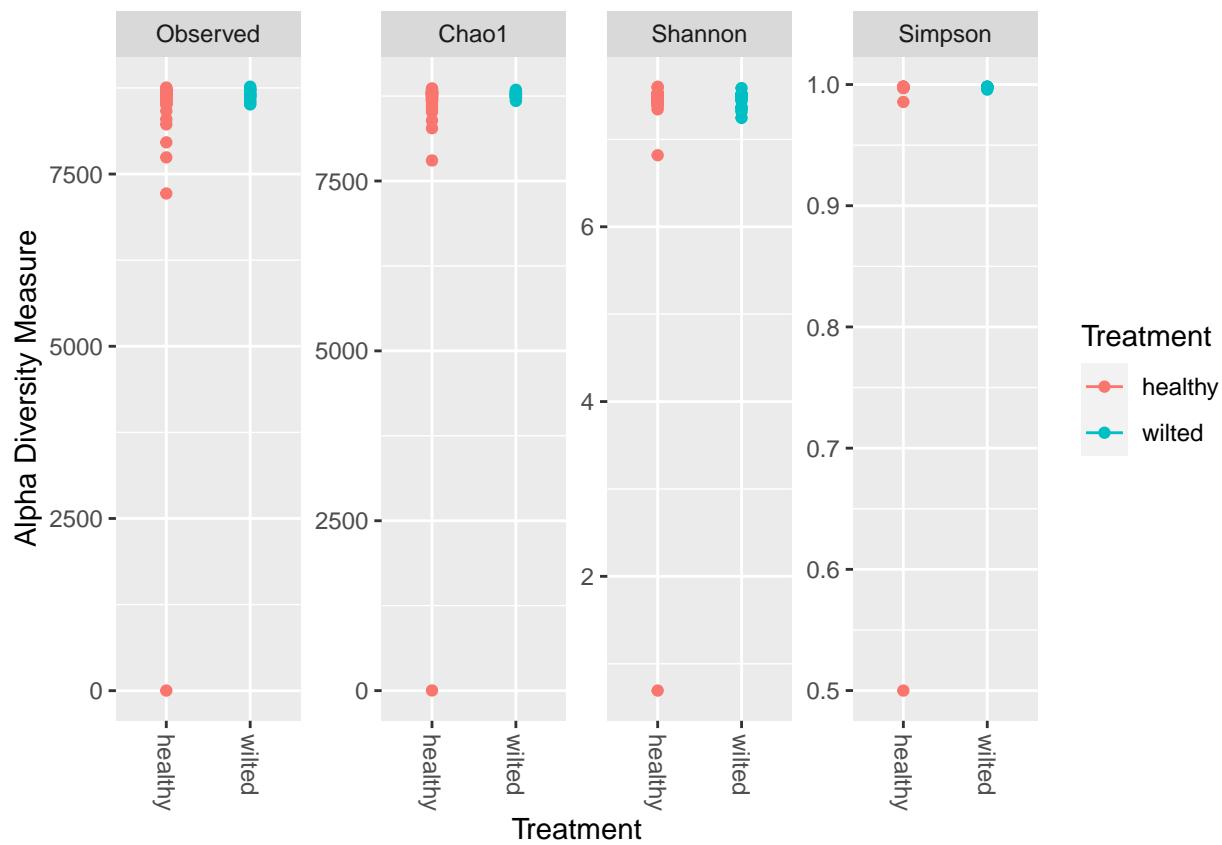
podemos ver una representación visual de la diversidad dentro de las muestras. Esta diversidad la podemos ver por muestra,

```
plot_richness(physeq = fresa_kraken_fil, measures = c("Observed", "Chao1", "Shannon", "simpson"))
```



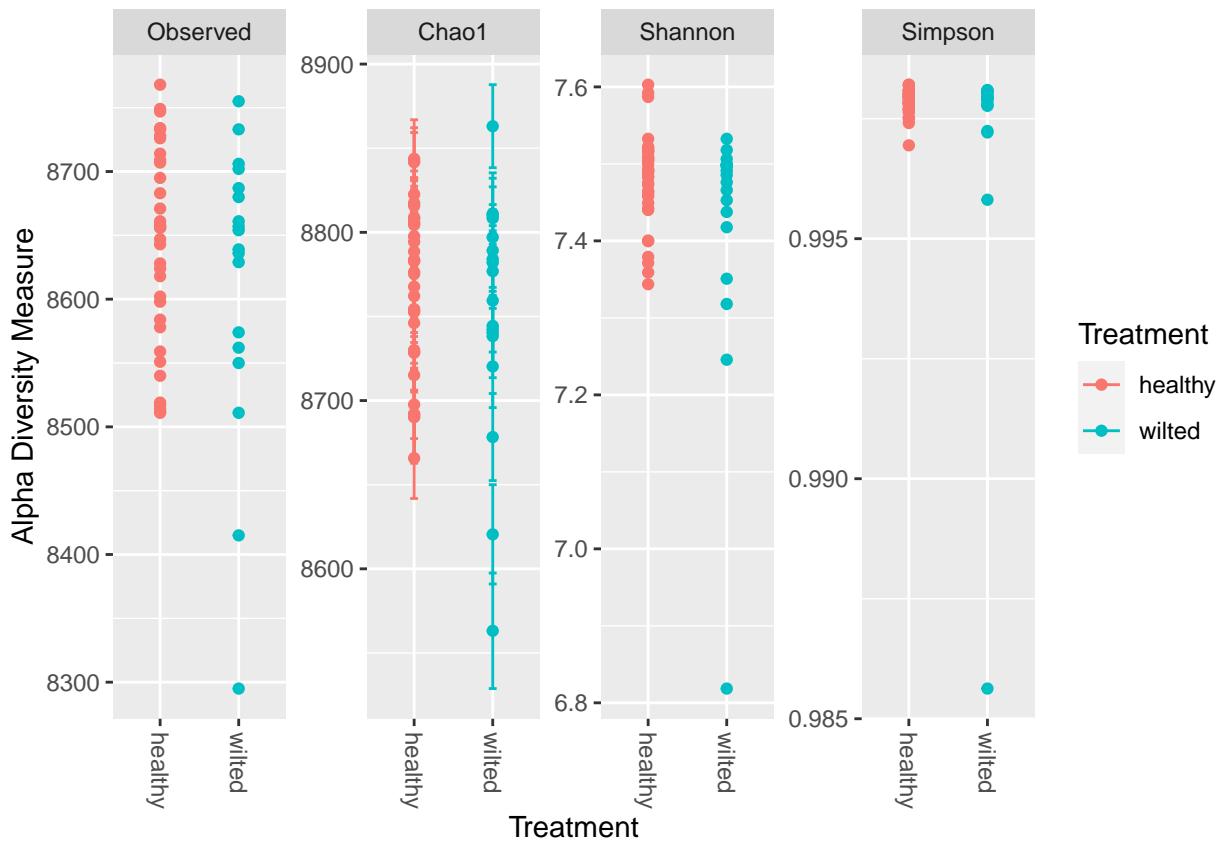
pero ya que queremos diferenciar entre plantas sanas y enfermas, tomamos estos dos conjuntos por separado para mostrar la diversidad

```
plot_richness(physeq = fresa_kraken, measures = c("Observed", "Chao1", "Shannon", "simpson"), x = "Treatment")
```



comparando entre los datos en crudo,y los datos filtrados por calidad

```
plot_richness(physeq = fresa_kraken_fil, measures = c("Observed", "Chao1", "Shannon", "simpson"), x = "Treatment")
```



Shannon

Simpson

Chao1

Aqui podemos ver cuantos reads tenemos por muestra

```
sample_sums(fresa_kraken)
```

```
##   MD2055    MD2056    MD2065    MD2066    MD2075    MD2076    MD2085    MD2086
## 9782432 12468526 11297600 15580959 12310781 16067839 10524919 9931297
##   MD2095    MD2096    MD2105    MD2106    MD2115    MD2116    MD2125    MD2126
## 18009912 14998268 11792397 4053295 13102554 12451637 8355853 14307309
##   MD2135    MD2136    MP2047    MP2048    MP2049    MP2050    MP2057    MP2058
## 7280751  6172369  9199079 12146967 13075806 16098757 17141427 20923502
##   MP2059    MP2060    MP2067    MP2068    MP2069    MP2070    MP2077    MP2078
```

```

## 14129981 13786630 16924218 20873789 15537530 12462356 9617847 7588787
## MP2079   MP2080   MP2087   MP2088   MP2089   MP2090   MP2097   MP2098
## 745830   3125701  20632320          2 16582404 11176782 11714000 16595897
## MP2099   MP2100   MP2107   MP2108   MP2109   MP2110   MP2117   MP2118
## 14844038 13342326 11014462 6728020 1405462 6901265 12624002 14711376
## MP2119   MP2120   MP2127   MP2128   MP2129   MP2130   MP2137   MP2138
## 9835326 10975712 7106567 9974861 8348307 6196725 2169734 8220431
## MP2139   MP2140
## 6158581 5267510

```

```
sample_sums(fresa_kraken_fil)
```

```

## MD2055   MD2056   MD2065   MD2066   MD2075   MD2076   MD2085   MD2086
## 9782432 12468526 11297600 15580959 12310781 16067839 10524919 9931297
## MD2095   MD2096   MD2105   MD2106   MD2115   MD2116   MD2125   MD2126
## 18009912 14998268 11792397 4053295 13102554 12451637 8355853 14307309
## MD2135   MD2136   MP2047   MP2048   MP2049   MP2050   MP2057   MP2058
## 7280751  6172369 9199079 12146967 13075806 16098757 17141427 20923502
## MP2059   MP2060   MP2067   MP2068   MP2069   MP2070   MP2077   MP2078
## 14129981 13786630 16924218 20873789 15537530 12462356 9617847 7588787
## MP2087   MP2089   MP2090   MP2097   MP2098   MP2099   MP2100   MP2107
## 20632320 16582404 11176782 11714000 16595897 14844038 13342326 11014462
## MP2108   MP2110   MP2117   MP2118   MP2119   MP2120   MP2127   MP2128
## 6728020  6901265 12624002 14711376 9835326 10975712 7106567 9974861
## MP2129   MP2130   MP2138   MP2139   MP2140
## 8348307 6196725 8220431 6158581 5267510

```

y con **summary** podemos darnos una idea de la uniformidad de los datos, ya que podemos ver datos estadísticos como el maximo, el minimo y la media

```
summary(fresa_kraken@otu_table@Data)
```

```

##      MD2055           MD2056           MD2065           MD2066
## Min.   :    0.0   Min.   :    0   Min.   :    0.0   Min.   :    0
## 1st Qu.: 14.0   1st Qu.: 18    1st Qu.: 13.0   1st Qu.: 24
## Median : 95.0   Median : 124   Median : 87.0   Median : 155
## Mean   : 1086.6  Mean   : 1385  Mean   : 1254.9  Mean   : 1731
## 3rd Qu.: 854.5   3rd Qu.: 1100  3rd Qu.: 846.5   3rd Qu.: 1412
## Max.   :309650.0  Max.   :369924  Max.   :382080.0  Max.   :470868
##      MD2075           MD2076           MD2085           MD2086
## Min.   :    0   Min.   :    0   Min.   :    0   Min.   :    0
## 1st Qu.: 22    1st Qu.: 40    1st Qu.: 18    1st Qu.: 18
## Median : 124   Median : 210   Median : 112   Median : 90
## Mean   : 1367  Mean   : 1785  Mean   : 1169  Mean   : 1103
## 3rd Qu.: 1027  3rd Qu.: 1470  3rd Qu.: 934   3rd Qu.: 675
## Max.   :347565  Max.   :391558  Max.   :290224  Max.   :1026738
##      MD2095           MD2096           MD2105           MD2106
## Min.   :    0.0   Min.   :    0   Min.   :    0   Min.   :    0.0
## 1st Qu.: 28.5   1st Qu.: 24    1st Qu.: 18    1st Qu.: 6.0
## Median : 188.0  Median : 153   Median : 115   Median : 43.0
## Mean   : 2000.4  Mean   : 1666  Mean   : 1310  Mean   : 450.2
## 3rd Qu.: 1667.0  3rd Qu.: 1333  3rd Qu.: 1056  3rd Qu.: 375.5

```

```

## Max. :479760.0  Max. :408186  Max. :307824  Max. :100633.0
## MD2115          MD2116          MD2125          MD2126
## Min. : 0   Min. : 0   Min. : 0.0  Min. : 0
## 1st Qu.: 19  1st Qu.: 18  1st Qu.: 13.0 1st Qu.: 23
## Median : 126 Median : 123 Median : 80.0  Median : 146
## Mean   : 1455 Mean   : 1383 Mean   : 928.1 Mean   : 1589
## 3rd Qu.: 1112 3rd Qu.: 1106 3rd Qu.: 705.0 3rd Qu.: 1260
## Max.  :352456 Max.  :342009 Max.  :202939.0 Max.  :409587
## MD2135          MD2136          MP2047          MP2048
## Min. : 0.0  Min. : 0.0  Min. : 0.0  Min. : 0
## 1st Qu.: 11.0 1st Qu.: 9.0  1st Qu.: 15.0 1st Qu.: 18
## Median : 66.0 Median : 59.0  Median : 100.0 Median : 124
## Mean   : 808.7 Mean   : 685.6 Mean   : 1021.8 Mean   : 1349
## 3rd Qu.: 570.0 3rd Qu.: 540.5 3rd Qu.: 849.5 3rd Qu.: 1096
## Max.  :324929.0 Max.  :188357.0 Max.  :266580.0 Max.  :372843
## MP2049          MP2050          MP2057          MP2058
## Min. : 0   Min. : 0   Min. : 0   Min. : 0.0
## 1st Qu.: 19  1st Qu.: 28  1st Qu.: 23  1st Qu.: 28.5
## Median : 128 Median : 173 Median : 157 Median : 177.0
## Mean   : 1452 Mean   : 1788 Mean   : 1904 Mean   : 2324.1
## 3rd Qu.: 1120 3rd Qu.: 1444 3rd Qu.: 1422 3rd Qu.: 1613.5
## Max.  :405044 Max.  :475984 Max.  :527148 Max.  :750581.0
## MP2059          MP2060          MP2067          MP2068
## Min. : 0   Min. : 0   Min. : 0   Min. : 0
## 1st Qu.: 20  1st Qu.: 19  1st Qu.: 29  1st Qu.: 34
## Median : 134 Median : 119 Median : 184 Median : 220
## Mean   : 1570 Mean   : 1531 Mean   : 1880 Mean   : 2318
## 3rd Qu.: 1238 3rd Qu.: 1108 3rd Qu.: 1543 3rd Qu.: 1930
## Max.  :438466 Max.  :604749 Max.  :481017 Max.  :583698
## MP2069          MP2070          MP2077          MP2078
## Min. : 0.0  Min. : 0   Min. : 0.0  Min. : 0.0
## 1st Qu.: 27.5 1st Qu.: 27  1st Qu.: 17.0 1st Qu.: 14.0
## Median : 173.0 Median : 155 Median : 107.0 Median : 85.0
## Mean   : 1725.8 Mean   : 1384 Mean   : 1068.3 Mean   : 842.9
## 3rd Qu.: 1454.5 3rd Qu.: 1226 3rd Qu.: 819.5 3rd Qu.: 693.0
## Max.  :353252.0 Max.  :298480 Max.  :293779.0 Max.  :219233.0
## MP2079          MP2080          MP2087          MP2088
## Min. : 0.00  Min. : 0.0  Min. : 0   Min. : 0.0000000
## 1st Qu.: 1.00 1st Qu.: 5.0  1st Qu.: 35  1st Qu.: 0.0000000
## Median : 8.00 Median : 32.0 Median : 217 Median : 0.0000000
## Mean   : 82.84 Mean   : 347.2 Mean   : 2292 Mean   : 0.0002221
## 3rd Qu.: 66.00 3rd Qu.: 268.5 3rd Qu.: 1844 3rd Qu.: 0.0000000
## Max.  :24597.00 Max.  :96283.0 Max.  :591983 Max.  :1.0000000
## MP2089          MP2090          MP2097          MP2098
## Min. : 0   Min. : 0   Min. : 0   Min. : 0
## 1st Qu.: 26  1st Qu.: 19  1st Qu.: 21  1st Qu.: 31
## Median : 162 Median : 117 Median : 131 Median : 181
## Mean   : 1842 Mean   : 1241 Mean   : 1301 Mean   : 1843
## 3rd Qu.: 1466 3rd Qu.: 988  3rd Qu.: 1068 3rd Qu.: 1484
## Max.  :500426 Max.  :280636 Max.  :323878 Max.  :441849
## MP2099          MP2100          MP2107          MP2108
## Min. : 0   Min. : 0   Min. : 0   Min. : 0.0
## 1st Qu.: 24  1st Qu.: 20  1st Qu.: 16  1st Qu.: 10.0
## Median : 163 Median : 141 Median : 108 Median : 67.0

```

```

##  Mean   : 1649   Mean   : 1482   Mean   : 1223   Mean   : 747.3
## 3rd Qu.: 1404   3rd Qu.: 1211   3rd Qu.: 966    3rd Qu.: 605.0
## Max.   :381823   Max.   :347101   Max.   :340913   Max.   :196757.0
##          MP2109           MP2110           MP2117           MP2118
##  Min.   : 0.0    Min.   : 0.0    Min.   : 0     Min.   : 0
## 1st Qu.: 2.0    1st Qu.: 11.0   1st Qu.: 19     1st Qu.: 21
## Median : 15.0   Median : 68.0   Median : 131   Median : 143
## Mean   : 156.1  Mean   : 766.5  Mean   : 1402  Mean   : 1634
## 3rd Qu.: 129.0 3rd Qu.: 598.0  3rd Qu.: 1108  3rd Qu.: 1266
## Max.   :43535.0 Max.   :185697.0 Max.   :373971  Max.   :344457
##          MP2119           MP2120           MP2127           MP2128
##  Min.   : 0.0    Min.   : 0     Min.   : 0.0   Min.   : 0.0
## 1st Qu.: 16.0   1st Qu.: 14    1st Qu.: 11.0  1st Qu.: 12.0
## Median : 100.0  Median : 98    Median : 71.0  Median : 83.0
## Mean   : 1092.5 Mean   : 1219  Mean   : 789.4 Mean   : 1108.0
## 3rd Qu.: 882.5 3rd Qu.: 907   3rd Qu.: 599.0 3rd Qu.: 773.5
## Max.   :278616.0 Max.   :340307  Max.   :223557.0 Max.   :289910.0
##          MP2129           MP2130           MP2137           MP2138
##  Min.   : 0.0    Min.   : 0.0   Min.   : 0     Min.   : 0.0
## 1st Qu.: 12.0   1st Qu.: 10.0  1st Qu.: 3     1st Qu.: 12.0
## Median : 80.0   Median : 66.0  Median : 21    Median : 78.0
## Mean   : 927.3  Mean   : 688.3 Mean   : 241   Mean   : 913.1
## 3rd Qu.: 717.0 3rd Qu.: 539.0 3rd Qu.: 177   3rd Qu.: 650.5
## Max.   :249480.0 Max.   :190633.0 Max.   :81445  Max.   :203351.0
##          MP2139           MP2140
##  Min.   : 0.0    Min.   : 0.0
## 1st Qu.: 13.0   1st Qu.: 10.0
## Median : 81.0   Median : 68.0
## Mean   : 684.1  Mean   : 585.1
## 3rd Qu.: 622.5 3rd Qu.: 535.0
## Max.   :130999.0 Max.   :126168.0

```

```
summary(fresa_kraken_fil@otu_table@Data)
```

```

##          MD2055           MD2056           MD2065           MD2066
##  Min.   : 0.0    Min.   : 0     Min.   : 0.0   Min.   : 0
## 1st Qu.: 14.0   1st Qu.: 18    1st Qu.: 13.0  1st Qu.: 24
## Median : 95.0   Median : 124   Median : 87.0  Median : 155
## Mean   : 1086.6 Mean   : 1385  Mean   : 1254.9 Mean   : 1731
## 3rd Qu.: 854.5 3rd Qu.: 1100  3rd Qu.: 846.5 3rd Qu.: 1412
## Max.   :309650.0 Max.   :369924  Max.   :382080.0 Max.   :470868
##          MD2075           MD2076           MD2085           MD2086
##  Min.   : 0     Min.   : 0     Min.   : 0     Min.   : 0
## 1st Qu.: 22    1st Qu.: 40    1st Qu.: 18    1st Qu.: 18
## Median : 124   Median : 210   Median : 112   Median : 90
## Mean   : 1367  Mean   : 1785  Mean   : 1169  Mean   : 1103
## 3rd Qu.: 1027 3rd Qu.: 1470  3rd Qu.: 934   3rd Qu.: 675
## Max.   :347565 Max.   :391558  Max.   :290224  Max.   :1026738
##          MD2095           MD2096           MD2105           MD2106
##  Min.   : 0.0    Min.   : 0     Min.   : 0     Min.   : 0.0
## 1st Qu.: 28.5   1st Qu.: 24    1st Qu.: 18    1st Qu.: 6.0
## Median : 188.0  Median : 153   Median : 115   Median : 43.0
## Mean   : 2000.4 Mean   : 1666  Mean   : 1310  Mean   : 450.2
## 3rd Qu.: 1667.0 3rd Qu.: 1333  3rd Qu.: 1056  3rd Qu.: 375.5

```

```

## Max. :479760.0  Max. :408186  Max. :307824  Max. :100633.0
## MD2115          MD2116          MD2125          MD2126
## Min. : 0   Min. : 0   Min. : 0.0  Min. : 0
## 1st Qu.: 19  1st Qu.: 18  1st Qu.: 13.0 1st Qu.: 23
## Median : 126 Median : 123 Median : 80.0  Median : 146
## Mean   : 1455 Mean   : 1383 Mean   : 928.1 Mean   : 1589
## 3rd Qu.: 1112 3rd Qu.: 1106 3rd Qu.: 705.0 3rd Qu.: 1260
## Max.  :352456 Max.  :342009 Max.  :202939.0 Max.  :409587
## MD2135          MD2136          MP2047          MP2048
## Min. : 0.0  Min. : 0.0  Min. : 0.0  Min. : 0
## 1st Qu.: 11.0 1st Qu.: 9.0  1st Qu.: 15.0 1st Qu.: 18
## Median : 66.0 Median : 59.0  Median : 100.0 Median : 124
## Mean   : 808.7 Mean   : 685.6 Mean   : 1021.8 Mean   : 1349
## 3rd Qu.: 570.0 3rd Qu.: 540.5 3rd Qu.: 849.5 3rd Qu.: 1096
## Max.  :324929.0 Max.  :188357.0 Max.  :266580.0 Max.  :372843
## MP2049          MP2050          MP2057          MP2058
## Min. : 0   Min. : 0   Min. : 0   Min. : 0.0
## 1st Qu.: 19  1st Qu.: 28  1st Qu.: 23  1st Qu.: 28.5
## Median : 128 Median : 173 Median : 157 Median : 177.0
## Mean   : 1452 Mean   : 1788 Mean   : 1904 Mean   : 2324.1
## 3rd Qu.: 1120 3rd Qu.: 1444 3rd Qu.: 1422 3rd Qu.: 1613.5
## Max.  :405044 Max.  :475984 Max.  :527148 Max.  :750581.0
## MP2059          MP2060          MP2067          MP2068
## Min. : 0   Min. : 0   Min. : 0   Min. : 0
## 1st Qu.: 20  1st Qu.: 19  1st Qu.: 29  1st Qu.: 34
## Median : 134 Median : 119 Median : 184 Median : 220
## Mean   : 1570 Mean   : 1531 Mean   : 1880 Mean   : 2318
## 3rd Qu.: 1238 3rd Qu.: 1108 3rd Qu.: 1543 3rd Qu.: 1930
## Max.  :438466 Max.  :604749 Max.  :481017 Max.  :583698
## MP2069          MP2070          MP2077          MP2078
## Min. : 0.0  Min. : 0   Min. : 0.0  Min. : 0.0
## 1st Qu.: 27.5 1st Qu.: 27  1st Qu.: 17.0 1st Qu.: 14.0
## Median : 173.0 Median : 155 Median : 107.0 Median : 85.0
## Mean   : 1725.8 Mean   : 1384 Mean   : 1068.3 Mean   : 842.9
## 3rd Qu.: 1454.5 3rd Qu.: 1226 3rd Qu.: 819.5 3rd Qu.: 693.0
## Max.  :353252.0 Max.  :298480 Max.  :293779.0 Max.  :219233.0
## MP2087          MP2089          MP2090          MP2097
## Min. : 0   Min. : 0   Min. : 0   Min. : 0
## 1st Qu.: 35  1st Qu.: 26  1st Qu.: 19  1st Qu.: 21
## Median : 217 Median : 162 Median : 117 Median : 131
## Mean   : 2292 Mean   : 1842 Mean   : 1241 Mean   : 1301
## 3rd Qu.: 1844 3rd Qu.: 1466 3rd Qu.: 988 3rd Qu.: 1068
## Max.  :591983 Max.  :500426 Max.  :280636 Max.  :323878
## MP2098          MP2099          MP2100          MP2107
## Min. : 0   Min. : 0   Min. : 0   Min. : 0
## 1st Qu.: 31  1st Qu.: 24  1st Qu.: 20  1st Qu.: 16
## Median : 181 Median : 163 Median : 141 Median : 108
## Mean   : 1843 Mean   : 1649 Mean   : 1482 Mean   : 1223
## 3rd Qu.: 1484 3rd Qu.: 1404 3rd Qu.: 1211 3rd Qu.: 966
## Max.  :441849 Max.  :381823 Max.  :347101 Max.  :340913
## MP2108          MP2110          MP2117          MP2118
## Min. : 0.0  Min. : 0.0  Min. : 0   Min. : 0
## 1st Qu.: 10.0 1st Qu.: 11.0 1st Qu.: 19  1st Qu.: 21
## Median : 67.0 Median : 68.0 Median : 131 Median : 143

```

```

##   Mean    : 747.3   Mean    : 766.5   Mean    : 1402   Mean    : 1634
## 3rd Qu.: 605.0   3rd Qu.: 598.0   3rd Qu.: 1108   3rd Qu.: 1266
## Max.   :196757.0   Max.   :185697.0   Max.   :373971   Max.   :344457
##      MP2119          MP2120          MP2127          MP2128
## Min.   : 0.0       Min.   : 0       Min.   : 0.0       Min.   : 0.0
## 1st Qu.: 16.0      1st Qu.: 14       1st Qu.: 11.0      1st Qu.: 12.0
## Median : 100.0     Median : 98       Median : 71.0     Median : 83.0
## Mean   : 1092.5    Mean   : 1219     Mean   : 789.4    Mean   : 1108.0
## 3rd Qu.: 882.5    3rd Qu.: 907      3rd Qu.: 599.0    3rd Qu.: 773.5
## Max.   :278616.0   Max.   :340307     Max.   :223557.0   Max.   :289910.0
##      MP2129          MP2130          MP2138          MP2139
## Min.   : 0.0       Min.   : 0.0      Min.   : 0.0       Min.   : 0.0
## 1st Qu.: 12.0      1st Qu.: 10.0     1st Qu.: 12.0      1st Qu.: 13.0
## Median : 80.0      Median : 66.0     Median : 78.0     Median : 81.0
## Mean   : 927.3     Mean   : 688.3    Mean   : 913.1    Mean   : 684.1
## 3rd Qu.: 717.0     3rd Qu.: 539.0    3rd Qu.: 650.5    3rd Qu.: 622.5
## Max.   :249480.0   Max.   :190633.0   Max.   :203351.0   Max.   :130999.0
##      MP2140
## Min.   : 0.0
## 1st Qu.: 10.0
## Median : 68.0
## Mean   : 585.1
## 3rd Qu.: 535.0
## Max.   :126168.0

```

Con el siguiente comando podemos ver si tenemos muestras no identificadas taxonomicamente, esto se puede ver identificando los espacios en blanco (“ ”) en los diferentes niveles taxonomicos.

```
summary(fresa_kraken_fil@tax_table@Data== "")
```

```

##   Kingdom        Phylum        Class        Order
## Mode :logical  Mode :logical  Mode :logical  Mode :logical
## FALSE:9003    FALSE:9001    FALSE:8720    FALSE:8926
##           TRUE :2        TRUE :283      TRUE :77
##   Family        Genus        Species
## Mode :logical  Mode :logical  Mode :logical
## FALSE:8780    FALSE:8629    FALSE:7904
##           TRUE :223      TRUE :374      TRUE :1099

```

podemos ver los **TRUE** de cada nivel taxonomico, por ejemplo a nivel de “Phylum” tenemos solo 2 sin clasificar, y a nivel de “Species” tenemos 1099 sin clasificar

ESTO ES LO QUE QUEREMOS COMPARAR CON LAS SALIDAS DE BRACKEN, YA QUE PROMETE HACER UNA REASIGNACION DE TODO LO QUE QUEDA SIN CLASIFICAR CON KRAKEN Queremos convertir las abundancias absolutas a relativas, Calculamos las abundancias relativas con la funcion, tanto de los datos originales como delos filtrados

```
percentages <- transform_sample_counts(fresa_kraken, function(x) x*100 / sum(x) )
head(percentages@otu_table@Data)
```

```

##          MD2055    MD2056    MD2065    MD2066    MD2075    MD2076
## 2062 0.1531112 0.1337047 0.1277616 0.12826553 0.09536357 0.08823837

```

```

## 1883    3.1653683 2.9668623 3.3819572 3.02207329 2.82325711 2.43690517
## 2880933 0.2543846 0.3160277 0.1716471 0.19734986 0.07154704 0.11000235
## 1725411 0.1917928 0.1444277 0.1809853 0.22574349 0.16239425 0.06742039
## 2781734 0.1555646 0.1643498 0.1495185 0.15032451 0.20536471 0.04722477
## 659352   0.1322882 0.1013913 0.1062969 0.09431384 0.08101029 0.08015390
##          MD2085      MD2086      MD2095      MD2096      MD2105      MD2106
## 2062     0.13315067 0.09192153 0.12641372 0.12737471 0.12608124 0.12187615
## 1883     2.75749391 2.03547432 2.66386643 2.72155425 2.61035988 2.48274552
## 2880933 0.08798167 0.05986126 0.17062271 0.09673784 0.12733628 0.14141581
## 1725411 0.14190133 0.09567733 0.08915646 0.12301420 0.12184970 0.10542040
## 2781734 0.07582956 0.05173544 0.08305982 0.12376096 0.10120928 0.06261572
## 659352   0.12698435 0.10551492 0.08481441 0.09445091 0.08172215 0.07751718
##          MD2115      MD2116      MD2125      MD2126      MD2135      MD2136
## 2062     0.12157935 0.13369326 0.10235939 0.1249431 0.11316140 0.1356043
## 1883     2.49623089 2.74669909 2.42870477 2.8627815 2.88291689 3.0516160
## 2880933 0.19640446 0.14734609 0.15764997 0.1338337 0.12266592 0.2052372
## 1725411 0.14826117 0.16294243 0.15644124 0.2447490 0.27056275 0.2074082
## 2781734 0.07165779 0.10300654 0.13905223 0.1719331 0.23487962 0.1551592
## 659352   0.09626368 0.09998685 0.08516186 0.1146337 0.09656971 0.1267099
##          MP2047      MP2048      MP2049      MP2050      MP2057      MP2058
## 2062     0.12994779 0.13091334 0.1353569 0.1650252 0.13327945 0.1326403
## 1883     2.89789880 3.06943289 3.0976599 2.9566506 3.07528656 3.5872628
## 2880933 0.24140460 0.24960964 0.3567581 0.3820109 0.19714228 0.1649772
## 1725411 0.19986783 0.19189152 0.1822909 0.1163195 0.16083258 0.2897077
## 2781734 0.18310529 0.16248501 0.1854341 0.1073499 0.19543297 0.2227782
## 659352   0.08981334 0.09938283 0.1541549 0.1507321 0.09992167 0.1094893
##          MP2059      MP2060      MP2067      MP2068      MP2069      MP2070
## 2062     0.1623498 0.1957041 0.1248566 0.1398117 0.11208345 0.12093219
## 1883     3.1030898 4.3864889 2.8421815 2.7963203 2.27354026 2.39505275
## 2880933 0.2111680 0.2366568 0.1071719 0.1017879 0.09933368 0.07979230
## 1725411 0.1791156 0.3918289 0.1461338 0.1350162 0.08834094 0.09406729
## 2781734 0.1222224 0.2303246 0.1428249 0.1172284 0.07447130 0.05293542
## 659352   0.1068154 0.1145603 0.1185343 0.1010933 0.06839568 0.09067306
##          MP2077      MP2078      MP2079      MP2080      MP2087      MP2088      MP2089
## 2062     0.27124574 0.14217028 0.15566550 0.14435162 0.1344686 0 0.1361564
## 1883     3.05451937 2.88890702 3.29793653 3.08036501 2.8692023 0 3.0178133
## 2880933 0.08991617 0.08424271 0.13206763 0.11015129 0.1157698 0 0.1057929
## 1725411 0.16123151 0.12580403 0.16116273 0.14963043 0.1549171 0 0.1139521
## 2781734 0.06864322 0.06477979 0.09559819 0.09412289 0.1053299 0 0.1435256
## 659352   0.15777959 0.13256401 0.14882748 0.17304918 0.1344832 0 0.1050571
##          MP2090      MP2097      MP2098      MP2099      MP2100      MP2107
## 2062     0.11746673 0.1329179 0.12327746 0.12064103 0.13358241 0.1309823
## 1883     2.51088372 2.7648796 2.66239903 2.57223136 2.60150292 3.0951398
## 2880933 0.08851385 0.1573075 0.16191351 0.19151123 0.12725667 0.1974313
## 1725411 0.11094428 0.1426327 0.10347136 0.11859307 0.13364986 0.2122573
## 2781734 0.11140953 0.1237323 0.08825073 0.07174598 0.07075228 0.1302742
## 659352   0.09047327 0.1128820 0.07603687 0.08263924 0.11718347 0.1092200
##          MP2108      MP2109      MP2110      MP2117      MP2118      MP2119
## 2062     0.13983312 0.12956594 0.11584833 0.1359474 0.11563160 0.1307938
## 1883     2.92444137 3.09755796 2.69076756 2.9623807 2.34143292 2.8328090
## 2880933 0.20697025 0.16983739 0.21099030 0.2390446 0.16900527 0.2062158
## 1725411 0.14979147 0.15027087 0.18218399 0.1574857 0.12846521 0.1388464
## 2781734 0.08442305 0.09847296 0.13216418 0.2113118 0.11355158 0.1144649
## 659352   0.09848365 0.10139015 0.09909777 0.1119772 0.08738136 0.1002712

```

```

##          MP2120     MP2127     MP2128     MP2129     MP2130     MP2137     MP2138
## 2062    0.1378134 0.1460339 0.1412250 0.1368541 0.1352811 0.2033890 0.11853393
## 1883    3.1005460 3.1457805 2.9064064 2.9883903 3.0763508 3.7536859 2.47372674
## 2880933 0.2095718 0.3069133 0.1648945 0.3364155 0.3090665 0.1141615 0.06674832
## 1725411 0.1899285 0.1881781 0.2019477 0.1668242 0.1398803 0.1933878 0.20091404
## 2781734 0.1890265 0.1338902 0.1568944 0.1456942 0.1019571 0.1022245 0.12329037
## 659352   0.1210218 0.1863488 0.1247436 0.1123222 0.1972816 0.3263534 0.14036977
##          MP2139     MP2140
## 2062    0.11929696 0.12586592
## 1883    2.12709713 2.39521140
## 2880933 0.05186260 0.08951098
## 1725411 0.08297366 0.11445636
## 2781734 0.03882388 0.05412424
## 659352   0.09305715 0.10367327

percentages_fil <- transform_sample_counts(fresa_kraken_fil, function(x) x*100 / sum(x) )
head(percentages_fil@otu_table@Data)

##          MD2055     MD2056     MD2065     MD2066     MD2075     MD2076
## 2062    0.1531112 0.1337047 0.1277616 0.12826553 0.09536357 0.08823837
## 1883    3.1653683 2.9668623 3.3819572 3.02207329 2.82325711 2.43690517
## 2880933 0.2543846 0.3160277 0.1716471 0.19734986 0.07154704 0.11000235
## 1725411 0.1917928 0.1444277 0.1809853 0.22574349 0.16239425 0.06742039
## 2781734 0.1555646 0.1643498 0.1495185 0.15032451 0.20536471 0.04722477
## 659352   0.1322882 0.1013913 0.1062969 0.09431384 0.08101029 0.08015390
##          MD2085     MD2086     MD2095     MD2096     MD2105     MD2106
## 2062    0.13315067 0.09192153 0.12641372 0.12737471 0.12608124 0.12187615
## 1883    2.75749391 2.03547432 2.66386643 2.72155425 2.61035988 2.48274552
## 2880933 0.08798167 0.05986126 0.17062271 0.09673784 0.12733628 0.14141581
## 1725411 0.14190133 0.09567733 0.08915646 0.12301420 0.12184970 0.10542040
## 2781734 0.07582956 0.05173544 0.08305982 0.12376096 0.10120928 0.06261572
## 659352   0.12698435 0.10551492 0.08481441 0.09445091 0.08172215 0.07751718
##          MD2115     MD2116     MD2125     MD2126     MD2135     MD2136
## 2062    0.12157935 0.13369326 0.10235939 0.1249431 0.11316140 0.1356043
## 1883    2.49623089 2.74669909 2.42870477 2.8627815 2.88291689 3.0516160
## 2880933 0.19640446 0.14734609 0.15764997 0.1338337 0.12266592 0.2052372
## 1725411 0.14826117 0.16294243 0.15644124 0.2447490 0.27056275 0.2074082
## 2781734 0.07165779 0.10300654 0.13905223 0.1719331 0.23487962 0.1551592
## 659352   0.09626368 0.09998685 0.08516186 0.1146337 0.09656971 0.1267099
##          MP2047     MP2048     MP2049     MP2050     MP2057     MP2058
## 2062    0.12994779 0.13091334 0.1353569 0.1650252 0.13327945 0.1326403
## 1883    2.89789880 3.06943289 3.0976599 2.9566506 3.07528656 3.5872628
## 2880933 0.24140460 0.24960964 0.3567581 0.3820109 0.19714228 0.1649772
## 1725411 0.19986783 0.19189152 0.1822909 0.1163195 0.16083258 0.2897077
## 2781734 0.18310529 0.16248501 0.1854341 0.1073499 0.19543297 0.2227782
## 659352   0.08981334 0.09938283 0.1541549 0.1507321 0.09992167 0.1094893
##          MP2059     MP2060     MP2067     MP2068     MP2069     MP2070
## 2062    0.1623498 0.1957041 0.1248566 0.1398117 0.11208345 0.12093219
## 1883    3.1030898 4.3864889 2.8421815 2.7963203 2.27354026 2.39505275
## 2880933 0.2111680 0.2366568 0.1071719 0.1017879 0.09933368 0.07979230
## 1725411 0.1791156 0.3918289 0.1461338 0.1350162 0.08834094 0.09406729
## 2781734 0.1222224 0.2303246 0.1428249 0.1172284 0.07447130 0.05293542
## 659352   0.1068154 0.1145603 0.1185343 0.1010933 0.06839568 0.09067306
##          MP2077     MP2078     MP2087     MP2089     MP2090     MP2097

```

```

## 2062    0.27124574 0.14217028 0.1344686 0.1361564 0.11746673 0.1329179
## 1883    3.05451937 2.88890702 2.8692023 3.0178133 2.51088372 2.7648796
## 2880933  0.08991617 0.08424271 0.1157698 0.1057929 0.08851385 0.1573075
## 1725411  0.16123151 0.12580403 0.1549171 0.1139521 0.11094428 0.1426327
## 2781734  0.06864322 0.06477979 0.1053299 0.1435256 0.11140953 0.1237323
## 659352   0.15777959 0.13256401 0.1344832 0.1050571 0.09047327 0.1128820
##          MP2098     MP2099     MP2100     MP2107     MP2108     MP2110
## 2062    0.12327746 0.12064103 0.13358241 0.1309823 0.13983312 0.11584833
## 1883    2.66239903 2.57223136 2.60150292 3.0951398 2.92444137 2.69076756
## 2880933  0.16191351 0.19151123 0.12725667 0.1974313 0.20697025 0.21099030
## 1725411  0.10347136 0.11859307 0.13364986 0.2122573 0.14979147 0.18218399
## 2781734  0.08825073 0.07174598 0.07075228 0.1302742 0.08442305 0.13216418
## 659352   0.07603687 0.08263924 0.11718347 0.1092200 0.09848365 0.09909777
##          MP2117     MP2118     MP2119     MP2120     MP2127     MP2128     MP2129
## 2062    0.1359474 0.11563160 0.1307938 0.1378134 0.1460339 0.1412250 0.1368541
## 1883    2.9623807 2.34143292 2.8328090 3.1005460 3.1457805 2.9064064 2.9883903
## 2880933  0.2390446 0.16900527 0.2062158 0.2095718 0.3069133 0.1648945 0.3364155
## 1725411  0.1574857 0.12846521 0.1388464 0.1899285 0.1881781 0.2019477 0.1668242
## 2781734  0.2113118 0.11355158 0.1144649 0.1890265 0.1338902 0.1568944 0.1456942
## 659352   0.1119772 0.08738136 0.1002712 0.1210218 0.1863488 0.1247436 0.1123222
##          MP2130     MP2138     MP2139     MP2140
## 2062    0.1352811 0.11853393 0.11929696 0.12586592
## 1883    3.0763508 2.47372674 2.12709713 2.39521140
## 2880933  0.3090665 0.06674832 0.05186260 0.08951098
## 1725411  0.1398803 0.20091404 0.08297366 0.11445636
## 2781734  0.1019571 0.12329037 0.03882388 0.05412424
## 659352   0.1972816 0.14036977 0.09305715 0.10367327

```

Ahora ya es posible una buena comparación de abundancias dadas por porcentajes con índices beta

Diversidad Beta

La diversidad beta mide la diferencia entre dos o más entornos. Se puede medir con métricas como la disimilitud de Bray-Curtis, la distancia Jaccard o la distancia UniFrac.

Mide que tan similares o diferentes son un par de especies, muestras o conjuntos de muestras. Aquí podemos ver una lista de distancias disponibles, que Phyloseq puede usar

```
distanceMethodList
```

```

## $UniFrac
## [1] "unifrac"  "wunifrac"
##
## $DPCoA
## [1] "dpcoa"
##
## $JSD
## [1] "jsd"
##
## $vegdist
## [1] "manhattan"  "euclidean"  "canberra"   "bray"       "kulczynski"
## [6] "jaccard"    "gower"      "altGower"   "morisita"   "horn"
## [11] "mountford"  "raup"       "binomial"   "chao"      "cao"

```

```

## 
## $betadiver
## [1] "w"    "-1"   "c"    "wb"   "r"    "I"    "e"    "t"    "me"   "j"    "sor"  "m"
## [13] "-2"   "co"   "cc"   "g"    "-3"   "l"    "19"   "hk"   "rlb"  "sim"  "gl"   "z"
##
## $dist
## [1] "maximum"   "binary"     "minkowski"
##
## $designdist
## [1] "ANY"

```

Siendo las siguientes las mas usadas: + Disimilitud de Bray-Curtis

- Distancia Jaccard
- UniFrac

```
#vegdist(meta_ord_fil, "bray")# usa la libreria Vegan, REVISAR COMO FUNCIONA PARA IMPRIMIR LA TABLA
```

Usamos “ordinate” para asignar las distancias entre muestras, usando “Bray-Curtis”, ya que es una de las metricas mas completas y mayormente utilizadas para medir la diversidad beta

Hay diferentes formas de trazar y mostrar los resultados de dicho análisis. Entre otros, se utilizan ampliamente los análisis PCA, PCoA o NMDS.

```
meta_ord <- ordinate(physeq = percentages, method = "NMDS", distance = "bray")
```

```

## Wisconsin double standardization
## Run 0 stress 8.456249e-05
## Run 1 stress 8.730444e-05
## ... Procrustes: rmse 5.846526e-05 max resid 0.0002085156
## ... Similar to previous best
## Run 2 stress 9.346884e-05
## ... Procrustes: rmse 5.528965e-05 max resid 0.0001282282
## ... Similar to previous best
## Run 3 stress 6.626938e-05
## ... New best solution
## ... Procrustes: rmse 3.757661e-05 max resid 0.0001847309
## ... Similar to previous best
## Run 4 stress 9.35411e-05
## ... Procrustes: rmse 6.408873e-05 max resid 0.00020861
## ... Similar to previous best
## Run 5 stress 9.460451e-05
## ... Procrustes: rmse 5.258747e-05 max resid 0.0001477321
## ... Similar to previous best
## Run 6 stress 9.416872e-05
## ... Procrustes: rmse 4.99617e-05 max resid 0.0001476149
## ... Similar to previous best
## Run 7 stress 9.861422e-05
## ... Procrustes: rmse 5.588722e-05 max resid 0.0001753003
## ... Similar to previous best
## Run 8 stress 9.20651e-05
## ... Procrustes: rmse 4.040013e-05 max resid 0.0001907229

```

```

## ... Similar to previous best
## Run 9 stress 9.359642e-05
## ... Procrustes: rmse 6.307103e-05 max resid 0.0002722143
## ... Similar to previous best
## Run 10 stress 9.300692e-05
## ... Procrustes: rmse 5.323925e-05 max resid 0.0001645599
## ... Similar to previous best
## Run 11 stress 8.874184e-05
## ... Procrustes: rmse 5.797053e-05 max resid 0.000195058
## ... Similar to previous best
## Run 12 stress 9.706551e-05
## ... Procrustes: rmse 4.587631e-05 max resid 0.0001442516
## ... Similar to previous best
## Run 13 stress 9.835124e-05
## ... Procrustes: rmse 5.595098e-05 max resid 0.0001620877
## ... Similar to previous best
## Run 14 stress 9.513722e-05
## ... Procrustes: rmse 6.185097e-05 max resid 0.0002543875
## ... Similar to previous best
## Run 15 stress 9.767676e-05
## ... Procrustes: rmse 4.769726e-05 max resid 0.0001773738
## ... Similar to previous best
## Run 16 stress 9.037714e-05
## ... Procrustes: rmse 6.313906e-05 max resid 0.0001957713
## ... Similar to previous best
## Run 17 stress 9.720465e-05
## ... Procrustes: rmse 6.969577e-05 max resid 0.0002828839
## ... Similar to previous best
## Run 18 stress 9.669563e-05
## ... Procrustes: rmse 6.232526e-05 max resid 0.0001738937
## ... Similar to previous best
## Run 19 stress 9.570455e-05
## ... Procrustes: rmse 5.551977e-05 max resid 0.0001644574
## ... Similar to previous best
## Run 20 stress 9.363498e-05
## ... Procrustes: rmse 5.804502e-05 max resid 0.0001658404
## ... Similar to previous best
## *** Best solution repeated 18 times

## Warning in metaMDS(veganifyOTU(physeq), distance, ...): stress is (nearly) zero:
## you may have insufficient data

meta_ord_fil <- ordinate(physeq = percentages_fil, method = "NMDS", distance = "bray")

```

```

## Wisconsin double standardization
## Run 0 stress 0.1686115
## Run 1 stress 0.1720959
## Run 2 stress 0.2021837
## Run 3 stress 0.1646876
## ... New best solution
## ... Procrustes: rmse 0.05972879 max resid 0.2657492
## Run 4 stress 0.1665887
## Run 5 stress 0.1668561

```

```

## Run 6 stress 0.1939999
## Run 7 stress 0.1639277
## ... New best solution
## ... Procrustes: rmse 0.0442278 max resid 0.1798766
## Run 8 stress 0.1668603
## Run 9 stress 0.170307
## Run 10 stress 0.1639272
## ... New best solution
## ... Procrustes: rmse 0.003506486 max resid 0.016453
## Run 11 stress 0.1798568
## Run 12 stress 0.1635176
## ... New best solution
## ... Procrustes: rmse 0.03481015 max resid 0.1533124
## Run 13 stress 0.1768353
## Run 14 stress 0.1788165
## Run 15 stress 0.1697222
## Run 16 stress 0.1636949
## ... Procrustes: rmse 0.0165617 max resid 0.0518228
## Run 17 stress 0.1663009
## Run 18 stress 0.1800728
## Run 19 stress 0.16661
## Run 20 stress 0.1637885
## ... Procrustes: rmse 0.007302641 max resid 0.04717434
## *** Best solution was not repeated -- monoMDS stopping criteria:
##     4: no. of iterations >= maxit
##     16: stress ratio > sratmax

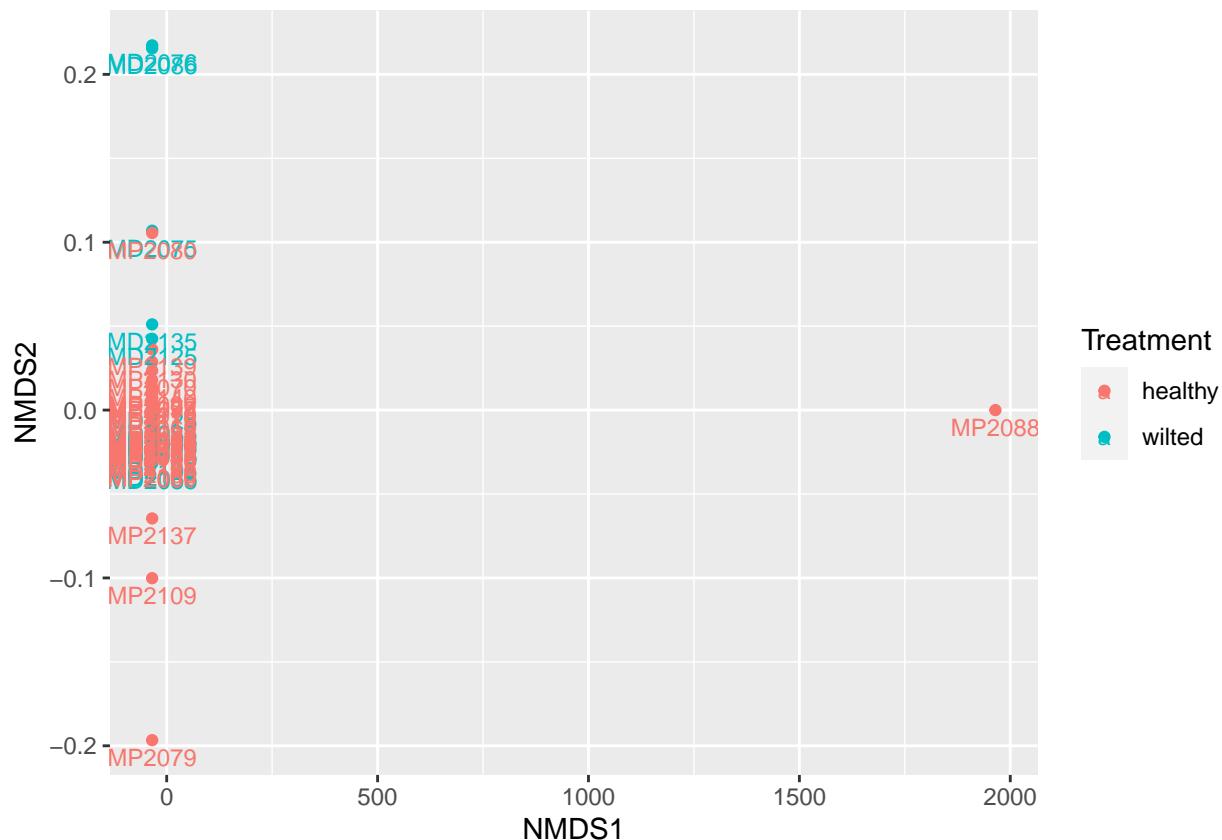
```

Ademas lo queremos diferenciar por color entre plantas sanas y enfermas,

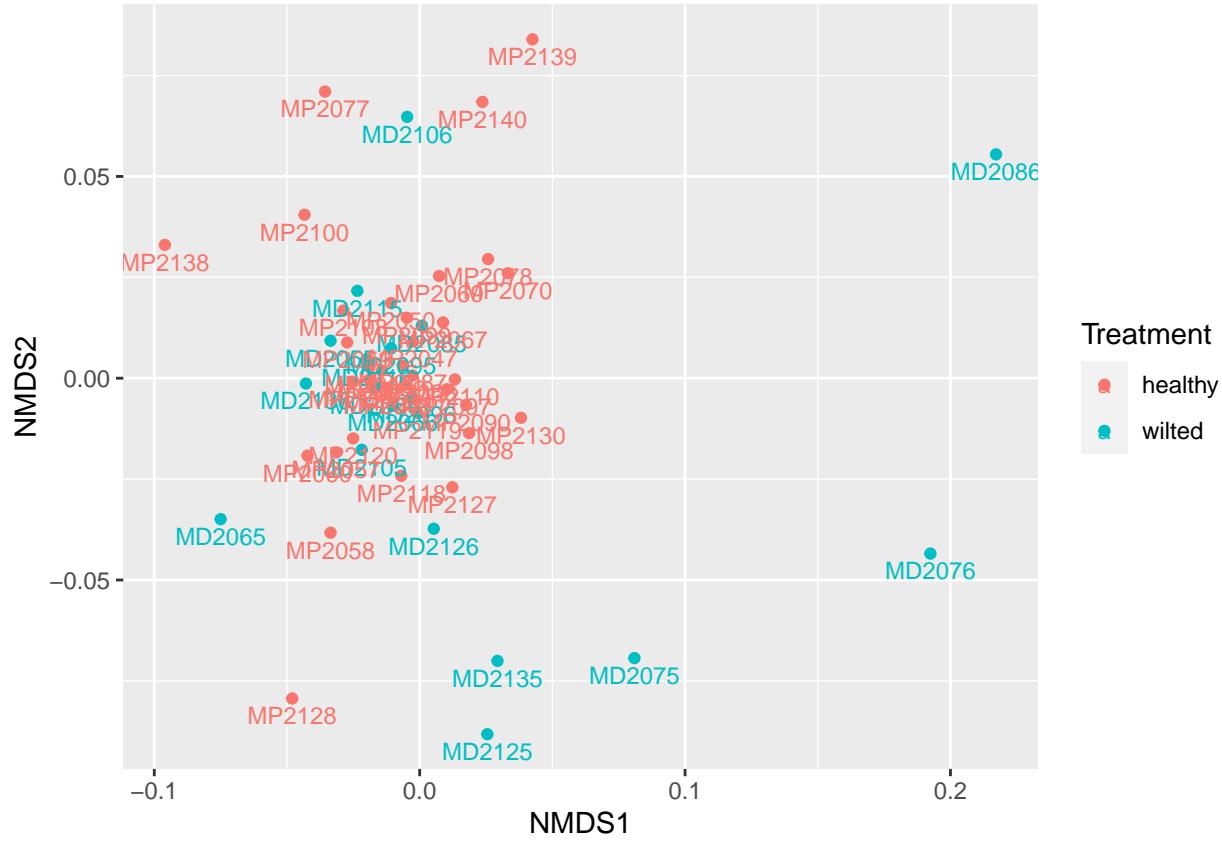
```

plot_ordination(physeq = percentages, ordination = meta_ord, color = "Treatment") +
  geom_text(mapping = aes(label = colnames(fresa_kraken@otu_table@Data)), size = 3, vjust = 1.5)

```



```
plot_ordination(physeq = percentages_fil, ordination = meta_ord_fil, color = "Treatment") +
  geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data)), size = 3, vjust = 1.5)
```



En este gráfico NMDS, cada punto representa la abundancia combinada de todas sus OTU. No se puede ver una diferencia entre los conjuntos de muestras de plantas sanas y muestras enfermas, por lo tanto: Probaremos varias distancias ya con los datos filtrados podemos ver varios ejemplos y llegar a la posibilidad que se diferencien un poco los dos conjuntos de datos

Usando

```
meta_ord_fil_2 <- ordinate(physeq = percentages_fil, method = "NMDS", distance = "jsd")

## Run 0 stress 9.277469e-05
## Run 1 stress 9.453648e-05
## ... Procrustes: rmse 6.849104e-05 max resid 0.0002878395
## ... Similar to previous best
## Run 2 stress 9.236093e-05
## ... New best solution
## ... Procrustes: rmse 5.475432e-05 max resid 0.0001708646
## ... Similar to previous best
## Run 3 stress 9.508889e-05
## ... Procrustes: rmse 7.576757e-05 max resid 0.0003657582
## ... Similar to previous best
## Run 4 stress 9.373873e-05
## ... Procrustes: rmse 7.313094e-05 max resid 0.0002071405
## ... Similar to previous best
## Run 5 stress 9.787758e-05
```

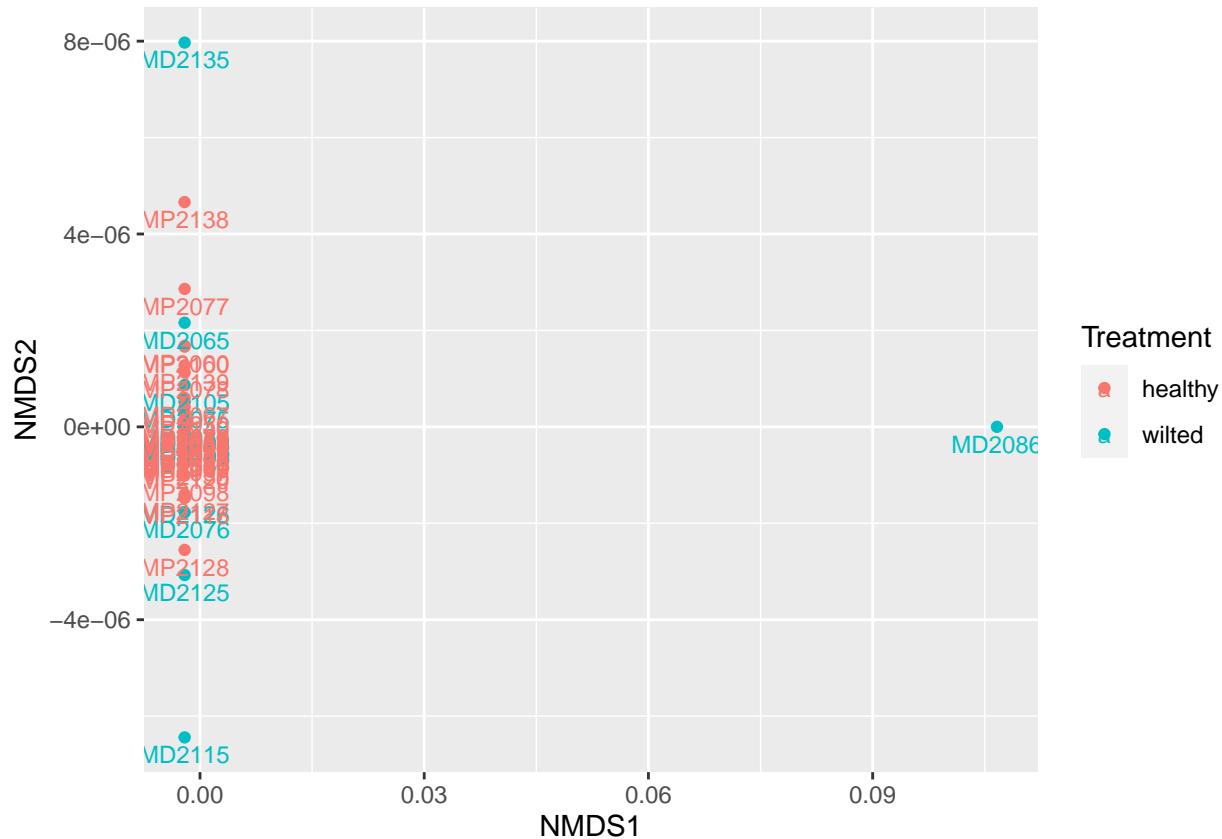
```

## ... Procrustes: rmse 8.831411e-05 max resid 0.0002825702
## ... Similar to previous best
## Run 6 stress 9.463442e-05
## ... Procrustes: rmse 9.062191e-05 max resid 0.0003763376
## ... Similar to previous best
## Run 7 stress 9.154183e-05
## ... New best solution
## ... Procrustes: rmse 8.821241e-05 max resid 0.0003372005
## ... Similar to previous best
## Run 8 stress 0.4030179
## Run 9 stress 9.354169e-05
## ... Procrustes: rmse 8.064795e-05 max resid 0.0003218632
## ... Similar to previous best
## Run 10 stress 9.821504e-05
## ... Procrustes: rmse 7.190303e-05 max resid 0.0002683282
## ... Similar to previous best
## Run 11 stress 6.322483e-05
## ... New best solution
## ... Procrustes: rmse 6.264448e-05 max resid 0.0002140281
## ... Similar to previous best
## Run 12 stress 9.429718e-05
## ... Procrustes: rmse 5.499954e-05 max resid 0.0001841124
## ... Similar to previous best
## Run 13 stress 9.102691e-05
## ... Procrustes: rmse 6.85014e-05 max resid 0.0002222926
## ... Similar to previous best
## Run 14 stress 9.927494e-05
## ... Procrustes: rmse 5.268635e-05 max resid 0.0001937807
## ... Similar to previous best
## Run 15 stress 8.927591e-05
## ... Procrustes: rmse 6.346448e-05 max resid 0.0002516416
## ... Similar to previous best
## Run 16 stress 8.888119e-05
## ... Procrustes: rmse 5.620376e-05 max resid 0.0001832829
## ... Similar to previous best
## Run 17 stress 9.732597e-05
## ... Procrustes: rmse 6.883855e-05 max resid 0.0002839787
## ... Similar to previous best
## Run 18 stress 9.821188e-05
## ... Procrustes: rmse 8.005894e-05 max resid 0.0002698677
## ... Similar to previous best
## Run 19 stress 9.052606e-05
## ... Procrustes: rmse 6.720451e-05 max resid 0.0002285255
## ... Similar to previous best
## Run 20 stress 9.063654e-05
## ... Procrustes: rmse 5.500878e-05 max resid 0.0001959764
## ... Similar to previous best
## *** Best solution repeated 10 times

## Warning in metaMDS(ps.dist): stress is (nearly) zero: you may have insufficient
## data

```

```
plot_ordination(physeq = percentages_fil, ordination = meta_ord_fil_2, color = "Treatment") +
  geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data)), size = 3, vjust = 1.5)
```



Usando

```
meta_ord_fil_3 <- ordinate(physeq = percentages_fil, method = "NMDS", distance = "euclidean")

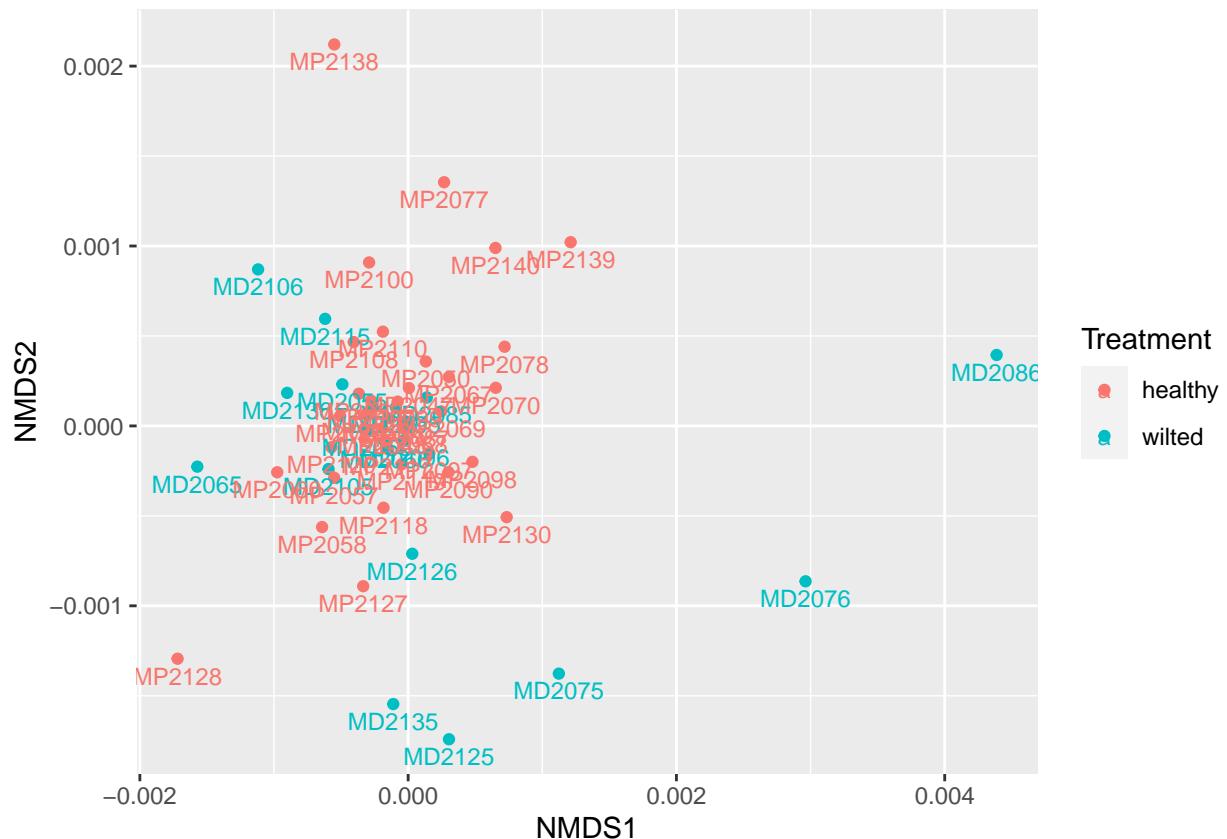
## Wisconsin double standardization
## Run 0 stress 0.1866907
## Run 1 stress 0.1961377
## Run 2 stress 0.1979378
## Run 3 stress 0.1876519
## Run 4 stress 0.1810222
## ... New best solution
## ... Procrustes: rmse 0.09243898 max resid 0.4393109
## Run 5 stress 0.1898393
## Run 6 stress 0.1779441
## ... New best solution
## ... Procrustes: rmse 0.05095054 max resid 0.2104657
## Run 7 stress 0.1891111
## Run 8 stress 0.1901222
## Run 9 stress 0.1771694
## ... New best solution
## ... Procrustes: rmse 0.044194 max resid 0.2059813
## Run 10 stress 0.1867635
## Run 11 stress 0.2014674
```

```

## Run 12 stress 0.2079774
## Run 13 stress 0.2065766
## Run 14 stress 0.2050305
## Run 15 stress 0.1914003
## Run 16 stress 0.1779186
## Run 17 stress 0.1812592
## Run 18 stress 0.1836356
## Run 19 stress 0.1765558
## ... New best solution
## ... Procrustes: rmse 0.02898278 max resid 0.1375525
## Run 20 stress 0.1913343
## *** Best solution was not repeated -- monoMDS stopping criteria:
##      2: no. of iterations >= maxit
##      18: stress ratio > sratmax

plot_ordination(physeq = percentages_fil, ordination = meta_ord_fil_3, color = "Treatment") +
  geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data)), size = 3, vjust = 1.5)

```



```

## Usando

meta_ord_fil_4 <- ordinate(physeq = percentages_fil, method = "NMDS", distance = "jaccard")

## Wisconsin double standardization
## Run 0 stress 0.168611
## Run 1 stress 0.169946
## Run 2 stress 0.1677187

```

```

## ... New best solution
## ... Procrustes: rmse 0.03554263  max resid 0.1588795
## Run 3 stress 0.1676929
## ... New best solution
## ... Procrustes: rmse 0.04220422  max resid 0.2179595
## Run 4 stress 0.1665993
## ... New best solution
## ... Procrustes: rmse 0.08434842  max resid 0.3423987
## Run 5 stress 0.1797529
## Run 6 stress 0.1781481
## Run 7 stress 0.1651799
## ... New best solution
## ... Procrustes: rmse 0.04704966  max resid 0.2148221
## Run 8 stress 0.1665887
## Run 9 stress 0.1705237
## Run 10 stress 0.1675346
## Run 11 stress 0.163308
## ... New best solution
## ... Procrustes: rmse 0.03803754  max resid 0.1521945
## Run 12 stress 0.171941
## Run 13 stress 0.1657738
## Run 14 stress 0.1680284
## Run 15 stress 0.1766145
## Run 16 stress 0.1737255
## Run 17 stress 0.1746764
## Run 18 stress 0.1704801
## Run 19 stress 0.1649569
## Run 20 stress 0.1678345
## *** Best solution was not repeated -- monoMDS stopping criteria:
##      1: no. of iterations >= maxit
##      19: stress ratio > sratmax

plot_ordination(physeq = percentages_fil, ordination = meta_ord_fil_4, color = "Treatment") +
  geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data)), size = 3, vjust = 1.5)

```



Usando

```
meta_ord_fil_5 <- ordinate(physeq = percentages_fil, method = "NMDS", distance = "manhattan")

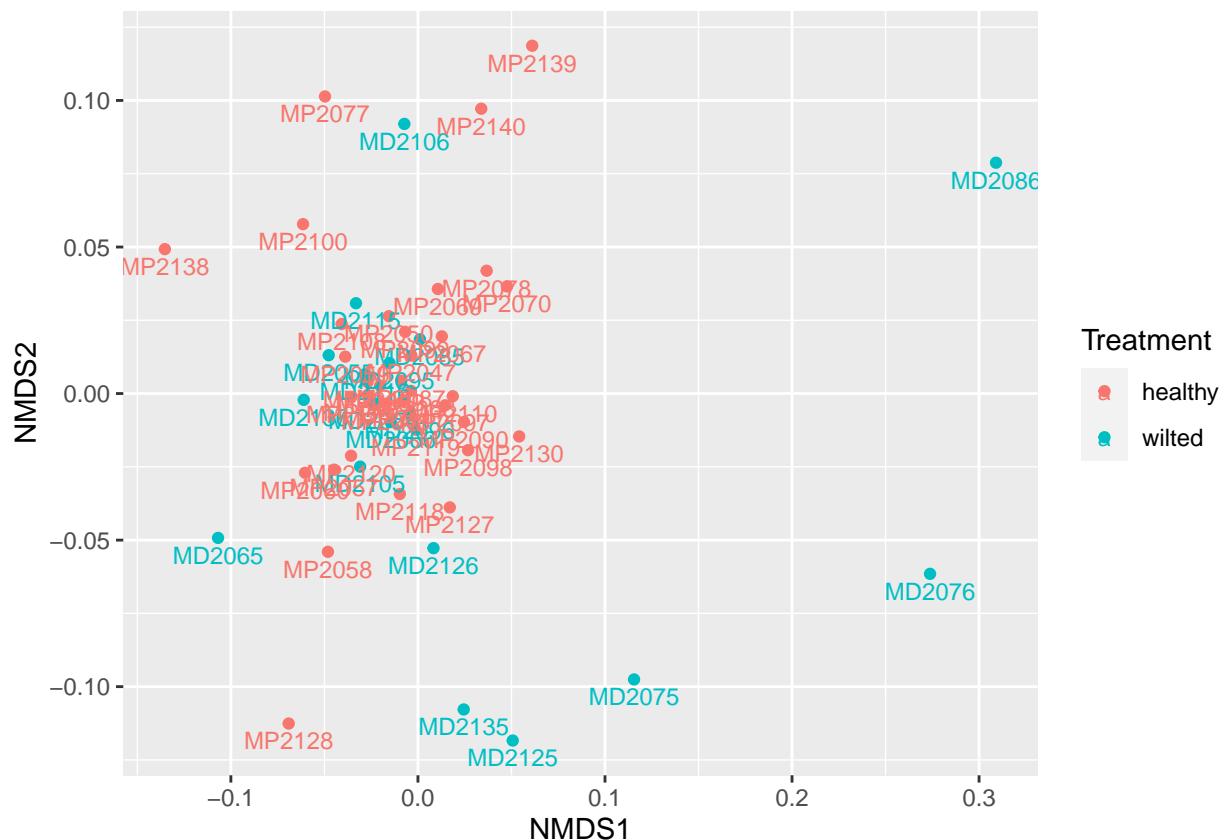
## Wisconsin double standardization
## Run 0 stress 0.1686118
## Run 1 stress 0.1660961
## ... New best solution
## ... Procrustes: rmse 0.065342 max resid 0.3487895
## Run 2 stress 0.2078356
## Run 3 stress 0.1701508
## Run 4 stress 0.1707507
## Run 5 stress 0.1777985
## Run 6 stress 0.1674247
## Run 7 stress 0.1775214
## Run 8 stress 0.1665992
## Run 9 stress 0.1666117
## Run 10 stress 0.1634885
## ... New best solution
## ... Procrustes: rmse 0.06032723 max resid 0.3426419
## Run 11 stress 0.1790928
## Run 12 stress 0.1687607
## Run 13 stress 0.167619
## Run 14 stress 0.1693476
## Run 15 stress 0.1955763
## Run 16 stress 0.1688576
```

```

## Run 17 stress 0.1723263
## Run 18 stress 0.1824605
## Run 19 stress 0.1775322
## Run 20 stress 0.1734123
## *** Best solution was not repeated -- monoMDS stopping criteria:
##      2: no. of iterations >= maxit
##      18: stress ratio > sratmax

plot_ordination(physeq = percentages_fil, ordination = meta_ord_fil_5, color = "Treatment") +
  geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data)), size = 3, vjust = 1.5)

```



Con ninguna distancia vemos una diferenciacion clara, por lo que lo veremos por los distintos niveles taxonomicos

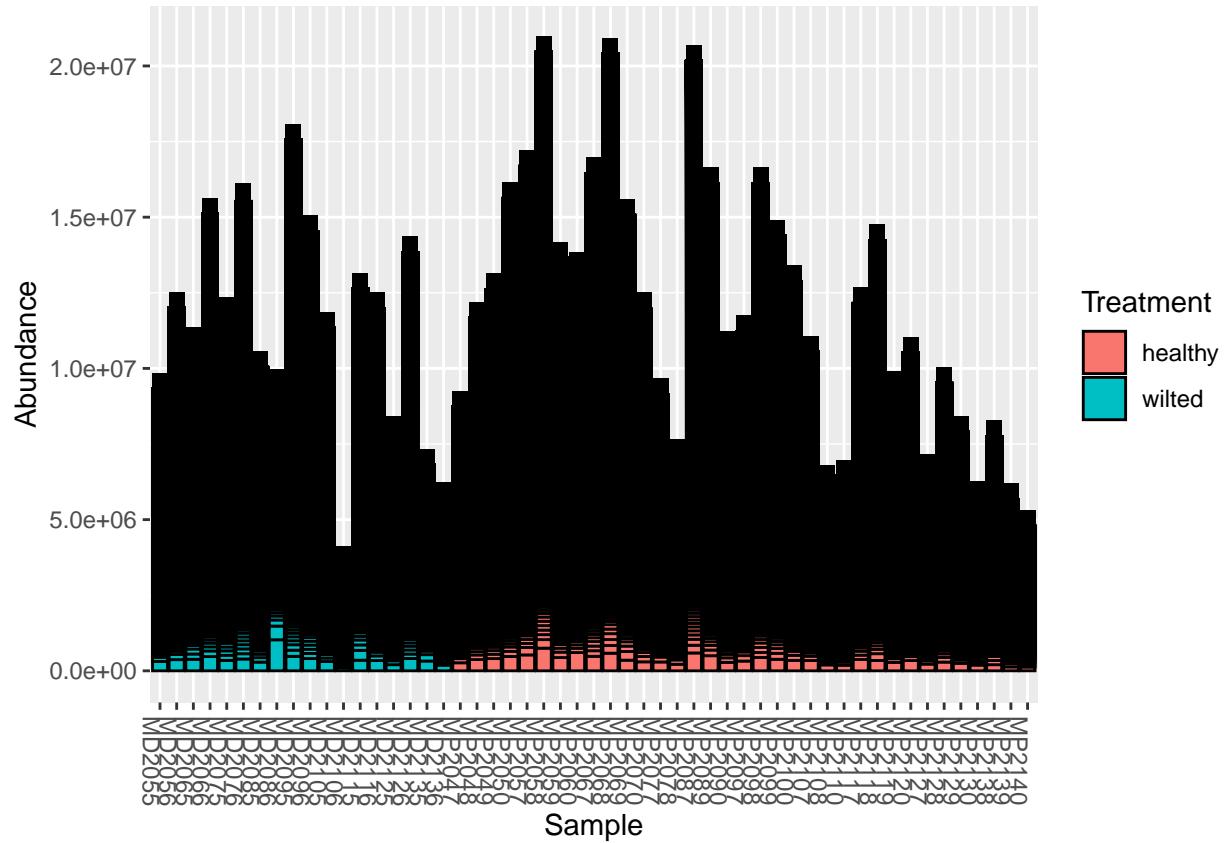
AHORA QUEREMOS VER DIFERENCIAS ENTRE DIFERENTES NIVELES TAXONOMICOS

De aqui en adelante solo trabajaremos con los datos filtrados,

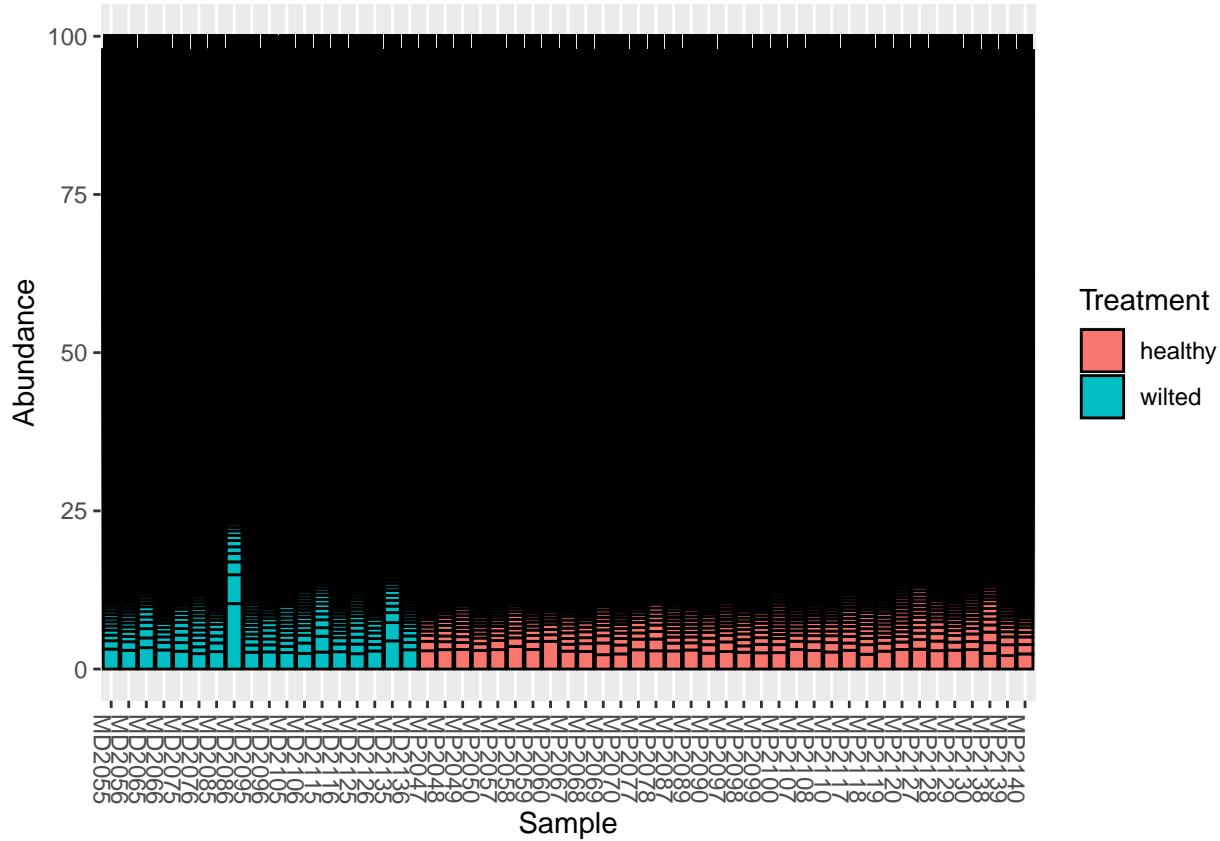
Grafico de barras de abundancia

Trazando las muestras en el eje x y las abundancias en el eje y Los valores de abundancia para cada OTUen cada muestra se apilan en el orden de mayor a menor, separados por una fina linea horizontal.

```
plot_bar(fresa_kraken_fil,fill="Treatment")
```



```
plot_bar(percentages_fil,fill="Treatment")
```



Queremos explorar nuestras muestras a diferentes niveles taxonómicos específicos
 ## A nivel de Kingdom

```
percentages_glom_kingdom <- tax_glom(percentages_fil, taxrank = 'Kingdom')
##View(percentages_glom_kingdom@tax_table@.Data)
```

A nivel de Phylum

```
percentages_glom_phylum <- tax_glom(percentages_fil, taxrank = 'Phylum')
##View(percentages_glom_phylum@tax_table@.Data)
```

Creamos un dataframe con los porcentajes de phylum

```
percentages_df_phylum <- psmelt(percentages_glom_phylum)
str(percentages_df_phylum)
```

```
## 'data.frame': 2279 obs. of 7 variables:
## $ OTU      : chr "1883" "1883" "1883" "1883" ...
## $ Sample   : chr "MP2060" "MP2058" "MP2059" "MP2049" ...
## $ Abundance: num 56.3 52.7 52.7 52.7 52.4 ...
## $ Treatment: chr "healthy" "healthy" "healthy" "healthy" ...
## $ Samples  : chr "MP2060" "MP2058" "MP2059" "MP2049" ...
## $ Kingdom  : chr "Bacteria" "Bacteria" "Bacteria" "Bacteria" ...
## $ Phylum   : chr "Actinobacteria" "Actinobacteria" "Actinobacteria" "Actinobacteria" ...
```

```

absolute_grom_phylum <- tax_grom(physeq = fresa_kraken_fil, taxrank = "Phylum")
absolute_df_phylum <- psmelt(absolute_grom_phylum)
str(absolute_df_phylum)

## 'data.frame': 2279 obs. of 7 variables:
## $ OTU      : chr "1883" "1883" "1883" "374" ...
## $ Sample   : chr "MP2058" "MP2087" "MP2068" "MP2068" ...
## $ Abundance: num 11028897 10342054 9967507 9190530 8805431 ...
## $ Treatment: chr "healthy" "healthy" "healthy" "healthy" ...
## $ Samples  : chr "MP2058" "MP2087" "MP2068" "MP2068" ...
## $ Kingdom  : chr "Bacteria" "Bacteria" "Bacteria" "Bacteria" ...
## $ Phylum   : chr "Actinobacteria" "Actinobacteria" "Actinobacteria" "Proteobacteria" ...

absolute_df_phylum$Phylum <- as.factor(absolute_df_phylum$Phylum)
phylum_colors_abs <- colorRampPalette(brewer.pal(8,"Dark2")) (length(levels(absolute_df_phylum$Phylum)))

absolute_plot <- ggplot(data= absolute_df_phylum, aes(x=Sample, y=Abundance, fill=Phylum))+  

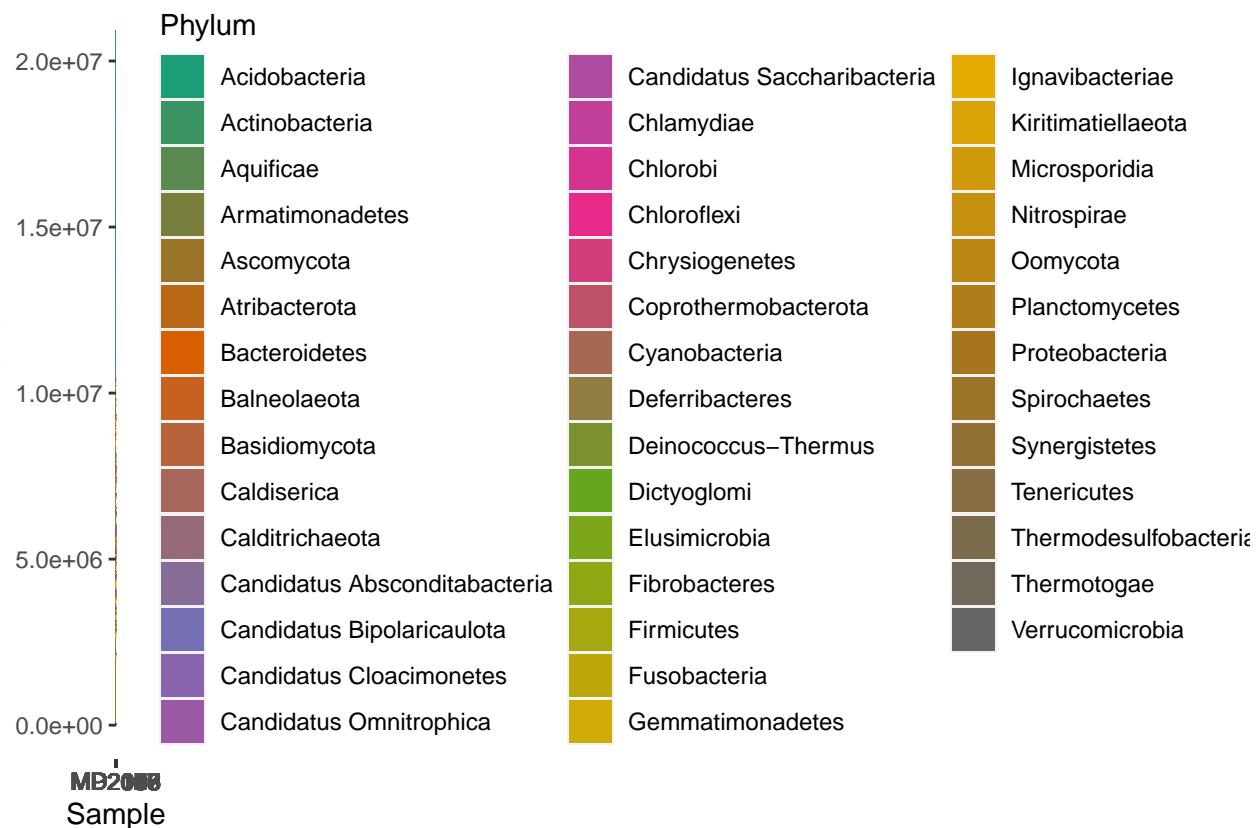
  geom_bar(aes(), stat="identity", position="stack")+
  scale_fill_manual(values = phylum_colors_abs)

percentages_df_phylum$Phylum <- as.factor(percentages_df_phylum$Phylum)
phylum_colors_rel<- colorRampPalette(brewer.pal(8,"Dark2")) (length(levels(percentages_df_phylum$Phylum)))
relative_plot <- ggplot(data=percentages_df_phylum, aes(x=Sample, y=Abundance, fill=Phylum))+  

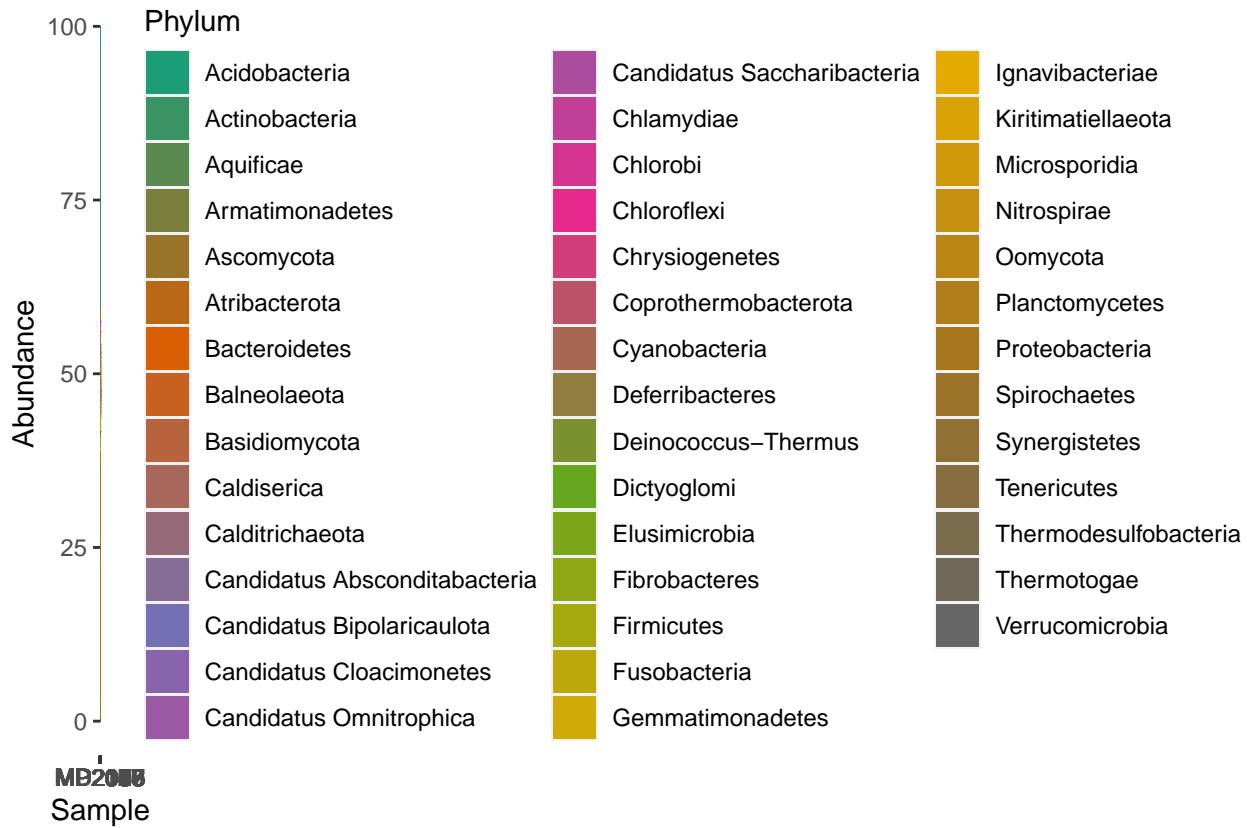
  geom_bar(aes(), stat="identity", position="stack")+
  scale_fill_manual(values = phylum_colors_rel)

absolute_plot

```



relative_plot



Usaremos un comando llamado unique() para explorar cuántos filos y reinos tenemos.

```
unique(fresa_kraken_fil@tax_table@Data[, "Kingdom"])
```

```
## [1] "Bacteria" "Eukaryota"
```

```
unique(fresa_kraken_fil@tax_table@Data[, "Phylum"])
```

```
## [1] "Actinobacteria"           "Firmicutes"
## [3] "Deinococcus-Thermus"      "Cyanobacteria"
## [5] "Chloroflexi"              "Armatimonadetes"
## [7] "Tenericutes"               "Proteobacteria"
## [9] "Planctomycetes"            "Verrucomicrobia"
## [11] "Kiritimatiellaeota"        "Chlamydiae"
## [13] "Candidatus Omnitrophica"   "Bacteroidetes"
## [15] "Chlorobi"                  "Balneolaeota"
## [17] "Ignavibacteriae"           "Gemmatimonadetes"
## [19] "Fibrobacteres"             "Candidatus Cloacimonetes"
## [21] "Acidobacteria"              "Nitrospirae"
## [23] "Spirochaetes"               "Synergistetes"
## [25] "Candidatus Bipolaricaulota" "Candidatus Saccharibacteria"
## [27] ""                           "Candidatus Absconditabacteria"
## [29] "Thermotogae"                "Aquifcae"
## [31] "Thermodesulfobacteria"       "Deferrribacteres"
## [33] "Fusobacteria"                 "Chrysioctenes"
```

```

## [35] "Calditrichaeota"           "Elusimicrobia"
## [37] "Caldiserica"              "Coprothermobacterota"
## [39] "Atribacterota"             "Dictyoglomi"
## [41] "Ascomycota"                "Basidiomycota"
## [43] "Microsporidia"              "Oomycota"

```

con esto podemos ver cuantos “Eukaryota” tenemos en “Kingdom”

```
sum(fresa_kraken_fil@tax_table@Data[,"Kingdom"] == "Eukaryota")
```

```
## [1] 181
```

y cuantos “Bacteria”

```
sum(fresa_kraken_fil@tax_table@Data[,"Kingdom"] == "Bacteria")
```

```
## [1] 8822
```

Diversidad Beta

veremos aqui solo el reino Eucariota

```
merge_Eukaryota <- subset_taxa(fresa_kraken_fil,Kingdom=="Eukaryota")
```

sacamos las abundancias relativas

```
percentages_Eukaryota <- transform_sample_counts(merge_Eukaryota, function(x) x*100 / sum(x) )
percentages_Eukaryota_df <- psmelt(percentages_Eukaryota)
```

beta diversidad de Eukaryota

```

meta_ord_Eukaryota <- ordinate(physeq = percentages_Eukaryota, method = "NMDS", distance = "bray")

## Wisconsin double standardization
## Run 0 stress 0.1341389
## Run 1 stress 0.1558514
## Run 2 stress 0.1636495
## Run 3 stress 0.1368379
## Run 4 stress 0.1392503
## Run 5 stress 0.1611309
## Run 6 stress 0.1711303
## Run 7 stress 0.1348056
## Run 8 stress 0.1394637
## Run 9 stress 0.1394555
## Run 10 stress 0.1392747
## Run 11 stress 0.1365988
## Run 12 stress 0.1359144
## Run 13 stress 0.1644409

```

```

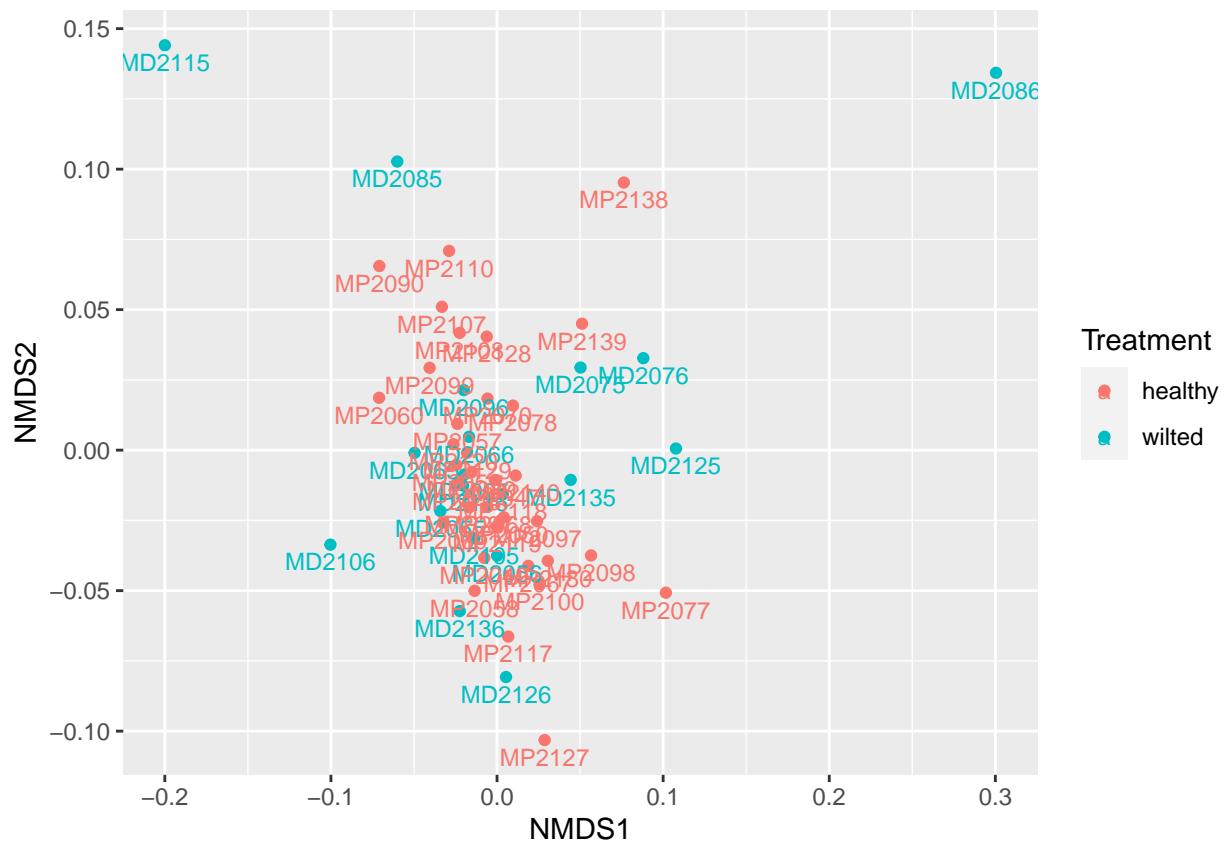
## Run 14 stress 0.1341975
## ... Procrustes: rmse 0.01193448 max resid 0.08063325
## Run 15 stress 0.1394645
## Run 16 stress 0.1490905
## Run 17 stress 0.1398812
## Run 18 stress 0.1657792
## Run 19 stress 0.1616037
## Run 20 stress 0.1708762
## *** Best solution was not repeated -- monoMDS stopping criteria:
##      3: no. of iterations >= maxit
##      17: stress ratio > sratmax

```

```

plot_ordination(physeq = percentages_Eukaryota, ordination = meta_ord_Eukaryota, color = "Treatment") +
  geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data)), size = 3, vjust = 1.5)

```



```

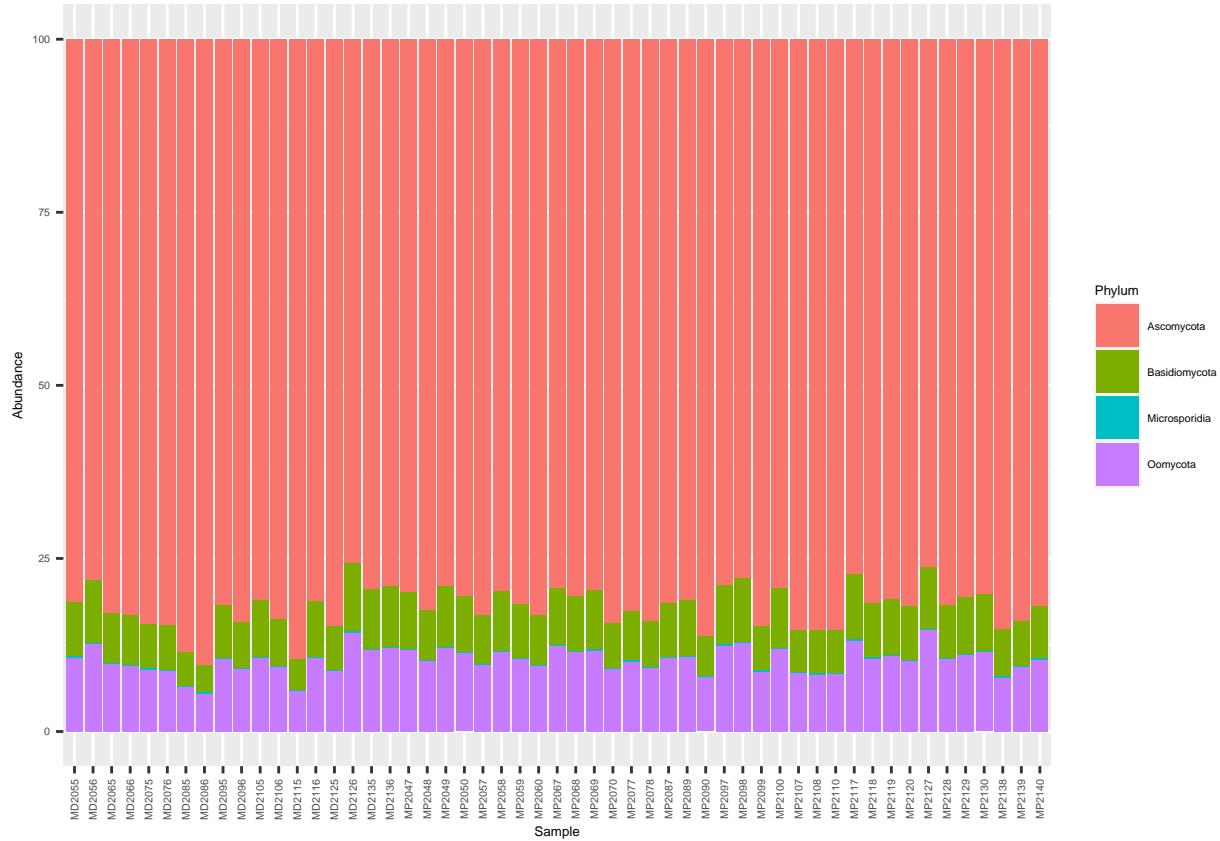
## Eukaryota por Phylum

```

```

ggplot(data= percentages_Eukaryota_df, aes(x=Sample, y=Abundance, fill=Phylum))+
  geom_bar(aes(), stat="identity", position="stack") +
  theme(text = element_text(size = 5),
        axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5))

```



```
merge_Eukaryota_Phylum <- tax_glom(merge_Eukaryota, taxrank = "Phylum")
```

sacamos las abundancias relativas

```
percentages_Eukaryota_Phylum <- transform_sample_counts(merge_Eukaryota_Phylum, function(x) x*100 / sum(x))
percentages_Eukaryota_Phylum_df <- psmelt(percentages_Eukaryota_Phylum)
meta_ord_Eukaryota_Phylum <- ordinate(physeq = percentages_Eukaryota_Phylum, method = "NMDS", distance = "euclidean")

## Square root transformation
## Wisconsin double standardization
## Run 0 stress 0.01148681
## Run 1 stress 9.833358e-05
## ... New best solution
## ... Procrustes: rmse 0.03458091 max resid 0.0906527
## Run 2 stress 9.964059e-05
## ... Procrustes: rmse 0.0001593859 max resid 0.0003895394
## ... Similar to previous best
## Run 3 stress 0.0001807176
## ... Procrustes: rmse 0.000450952 max resid 0.00112692
## ... Similar to previous best
## Run 4 stress 8.208215e-05
## ... New best solution
## ... Procrustes: rmse 0.0001064449 max resid 0.0003268099
## ... Similar to previous best
## Run 5 stress 9.186741e-05
```

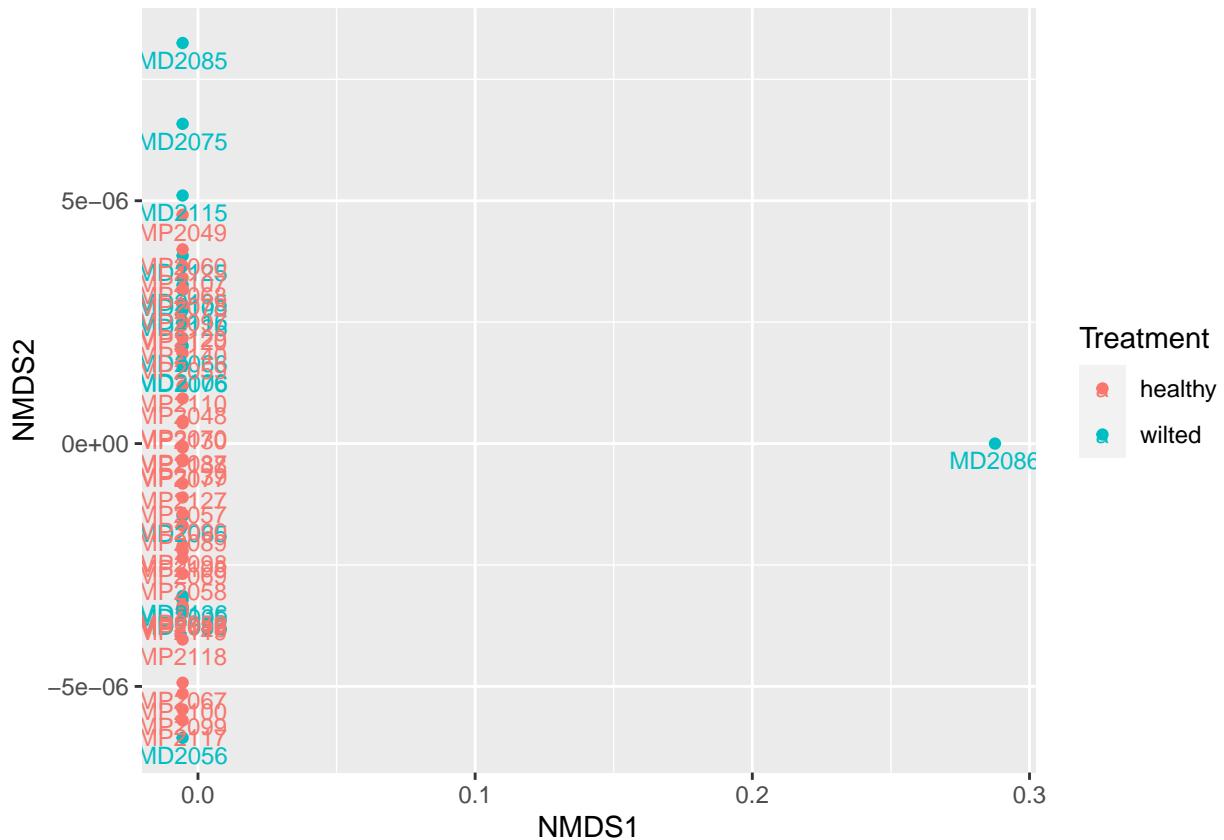
```

## ... Procrustes: rmse 0.0001412208 max resid 0.0004309775
## ... Similar to previous best
## Run 6 stress 7.462886e-05
## ... New best solution
## ... Procrustes: rmse 3.211791e-05 max resid 8.559031e-05
## ... Similar to previous best
## Run 7 stress 0.0001048768
## ... Procrustes: rmse 0.0002993268 max resid 0.0007891489
## ... Similar to previous best
## Run 8 stress 7.996713e-05
## ... Procrustes: rmse 5.233094e-05 max resid 0.0001331529
## ... Similar to previous best
## Run 9 stress 7.32546e-05
## ... New best solution
## ... Procrustes: rmse 4.60509e-05 max resid 0.0001679312
## ... Similar to previous best
## Run 10 stress 6.222568e-05
## ... New best solution
## ... Procrustes: rmse 4.352739e-05 max resid 0.0001077313
## ... Similar to previous best
## Run 11 stress 0.001023353
## Run 12 stress 9.984771e-05
## ... Procrustes: rmse 0.0001075881 max resid 0.0003100759
## ... Similar to previous best
## Run 13 stress 9.721e-05
## ... Procrustes: rmse 0.0001820262 max resid 0.0004477894
## ... Similar to previous best
## Run 14 stress 8.409742e-05
## ... Procrustes: rmse 3.708914e-05 max resid 0.0001118275
## ... Similar to previous best
## Run 15 stress 9.869134e-05
## ... Procrustes: rmse 0.0001608863 max resid 0.0004522027
## ... Similar to previous best
## Run 16 stress 9.797609e-05
## ... Procrustes: rmse 0.0001113431 max resid 0.0003237375
## ... Similar to previous best
## Run 17 stress 9.955898e-05
## ... Procrustes: rmse 0.0001629702 max resid 0.0004561932
## ... Similar to previous best
## Run 18 stress 9.530357e-05
## ... Procrustes: rmse 9.445489e-05 max resid 0.0002854999
## ... Similar to previous best
## Run 19 stress 4.705652e-05
## ... New best solution
## ... Procrustes: rmse 3.033326e-05 max resid 7.031517e-05
## ... Similar to previous best
## Run 20 stress 0.001748362
## *** Best solution repeated 1 times

## Warning in metaMDS(veganifyOTU(physeq), distance, ...): stress is (nearly) zero:
## you may have insufficient data

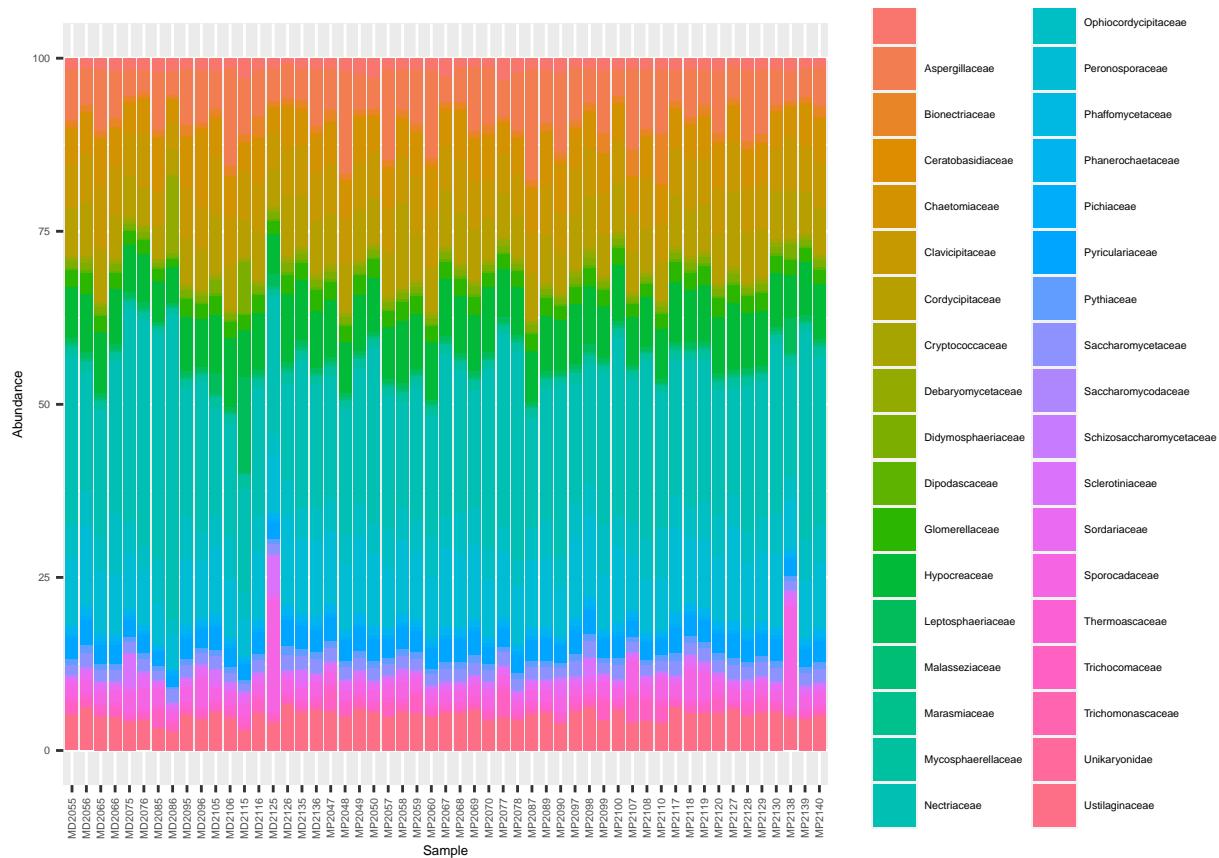
```

```
plot_ordination(physeq = percentages_Eukaryota_Phylum, ordination = meta_ord_Eukaryota_Phylum, color =
  geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data))), size = 3, vjust = 1.5)
```



```
## Eukaryota por Family
```

```
ggplot(data= percentages_Eukaryota_df, aes(x=Sample, y=Abundance, fill=Family))+  
  geom_bar(aes(), stat="identity", position="stack") +  
  theme(text = element_text(size = 5),  
        axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5))
```



```
merge_Eukaryota_Family<-tax_glom(merge_Eukaryota, taxrank="Family")
```

sacamos las abundancias relativas

```
percentages_Eukaryota_Family <- transform_sample_counts(merge_Eukaryota_Family, function(x) x*100 / sum(x))
percentages_Eukaryota_Family_df <- psmelt(percentages_Eukaryota_Family)
meta_ord_Eukaryota_Family <- ordinate(physeq = percentages_Eukaryota_Family, method = "NMDS", distance =
```

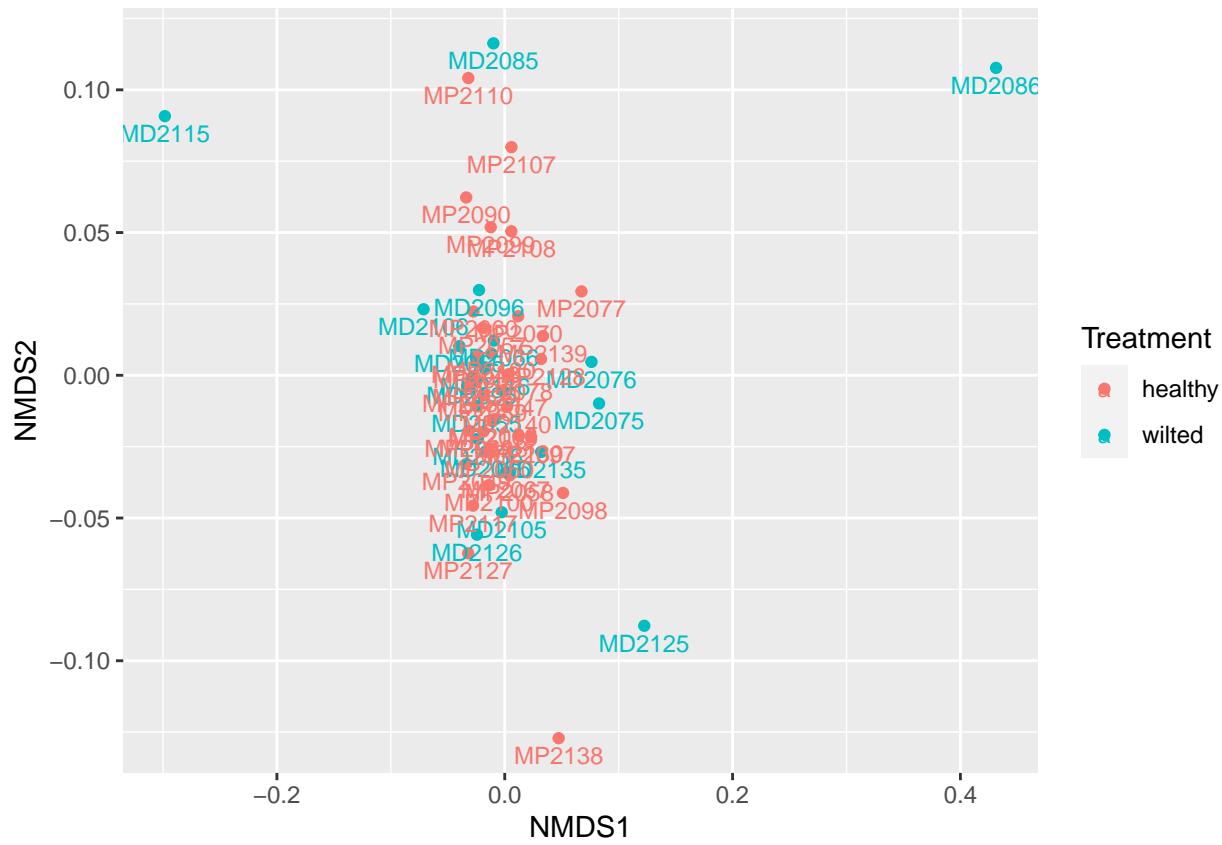
```
## Wisconsin double standardization
## Run 0 stress 0.1089081
## Run 1 stress 0.1088888
## ... New best solution
## ... Procrustes: rmse 0.001692112 max resid 0.009361591
## ... Similar to previous best
## Run 2 stress 0.1089296
## ... Procrustes: rmse 0.008412076 max resid 0.05476102
## Run 3 stress 0.1089294
## ... Procrustes: rmse 0.008515315 max resid 0.05452947
## Run 4 stress 0.10893
## ... Procrustes: rmse 0.008409098 max resid 0.05478302
## Run 5 stress 0.1089298
## ... Procrustes: rmse 0.008410544 max resid 0.05477326
## Run 6 stress 0.1352079
## Run 7 stress 0.1352087
## Run 8 stress 0.1089299
```

```

## ... Procrustes: rmse 0.008409391 max resid 0.05478159
## Run 9 stress 0.108949
## ... Procrustes: rmse 0.008246563 max resid 0.05400406
## Run 10 stress 0.1089484
## ... Procrustes: rmse 0.008204904 max resid 0.05405395
## Run 11 stress 0.1092107
## ... Procrustes: rmse 0.01213051 max resid 0.05289255
## Run 12 stress 0.1090933
## ... Procrustes: rmse 0.005793809 max resid 0.0287062
## Run 13 stress 0.1089087
## ... Procrustes: rmse 0.001706961 max resid 0.009327453
## ... Similar to previous best
## Run 14 stress 0.1089293
## ... Procrustes: rmse 0.008417598 max resid 0.05473245
## Run 15 stress 0.109113
## ... Procrustes: rmse 0.00989047 max resid 0.0541286
## Run 16 stress 0.1352077
## Run 17 stress 0.1089296
## ... Procrustes: rmse 0.008412506 max resid 0.05475463
## Run 18 stress 0.1089489
## ... Procrustes: rmse 0.008233157 max resid 0.05401687
## Run 19 stress 0.1088897
## ... Procrustes: rmse 0.00156262 max resid 0.008901338
## ... Similar to previous best
## Run 20 stress 0.1352081
## *** Best solution repeated 3 times

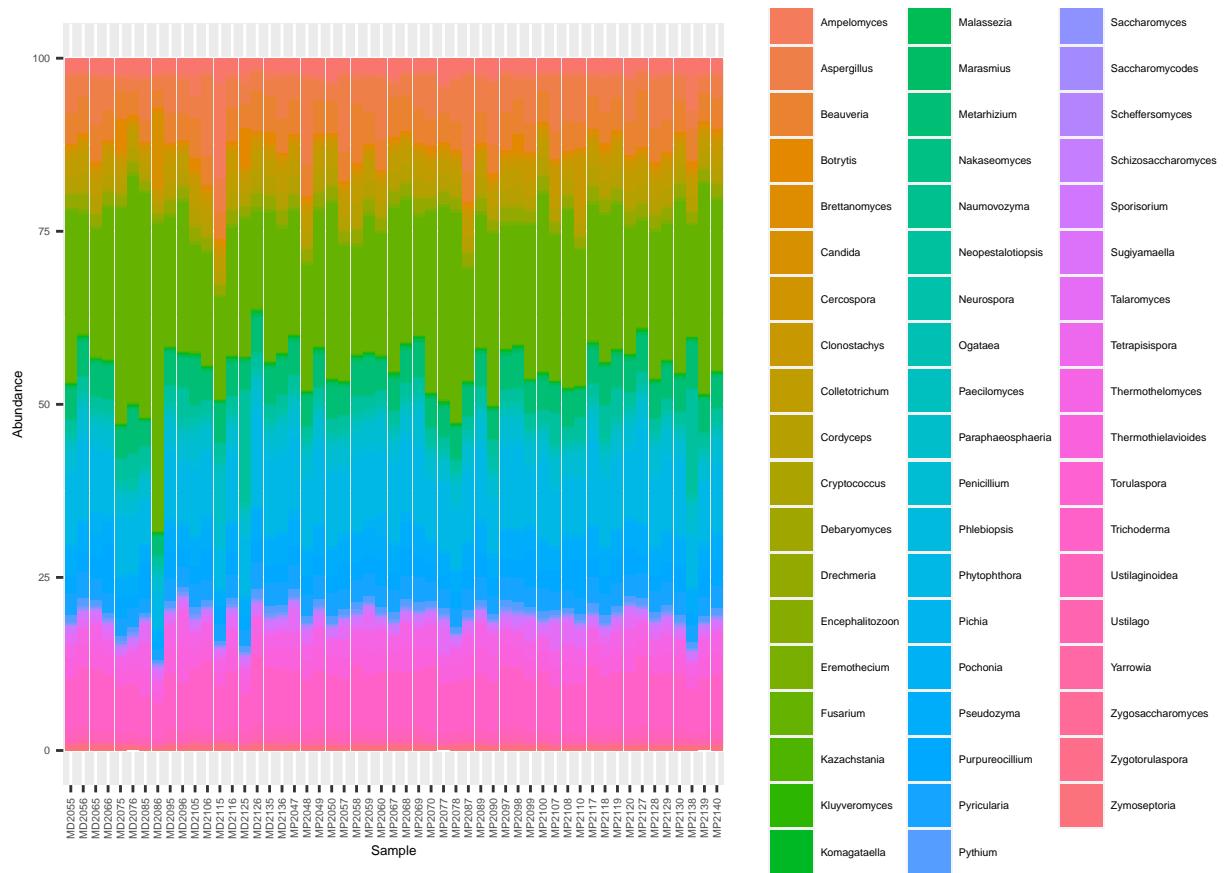
plot_ordination(physeq = percentages_Eukaryota_Family, ordination = meta_ord_Eukaryota_Family, color =
  geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data))), size = 3, vjust = 1.5)

```



Eukaryota por Genus

```
ggplot(data= percentages_Eukaryota_df, aes(x=Sample, y=Abundance, fill=Genus))+  
  geom_bar(aes(), stat="identity", position="stack") +  
  theme(text = element_text(size = 5),  
        axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5))
```



```
merge_Eukaryota_Genus <- tax_glom(merge_Eukaryota, taxrank = "Genus")
```

sacamos las abundancias relativas

```
percentages_Eukaryota_Genus <- transform_sample_counts(merge_Eukaryota_Genus, function(x) x*100 / sum(x))
percentages_Eukaryota_Genus_df <- psmelt(percentages_Eukaryota_Genus)
meta_ord_Eukaryota_Genus <- ordinate(physeq = percentages_Eukaryota_Genus, method = "NMDS", distance =
```

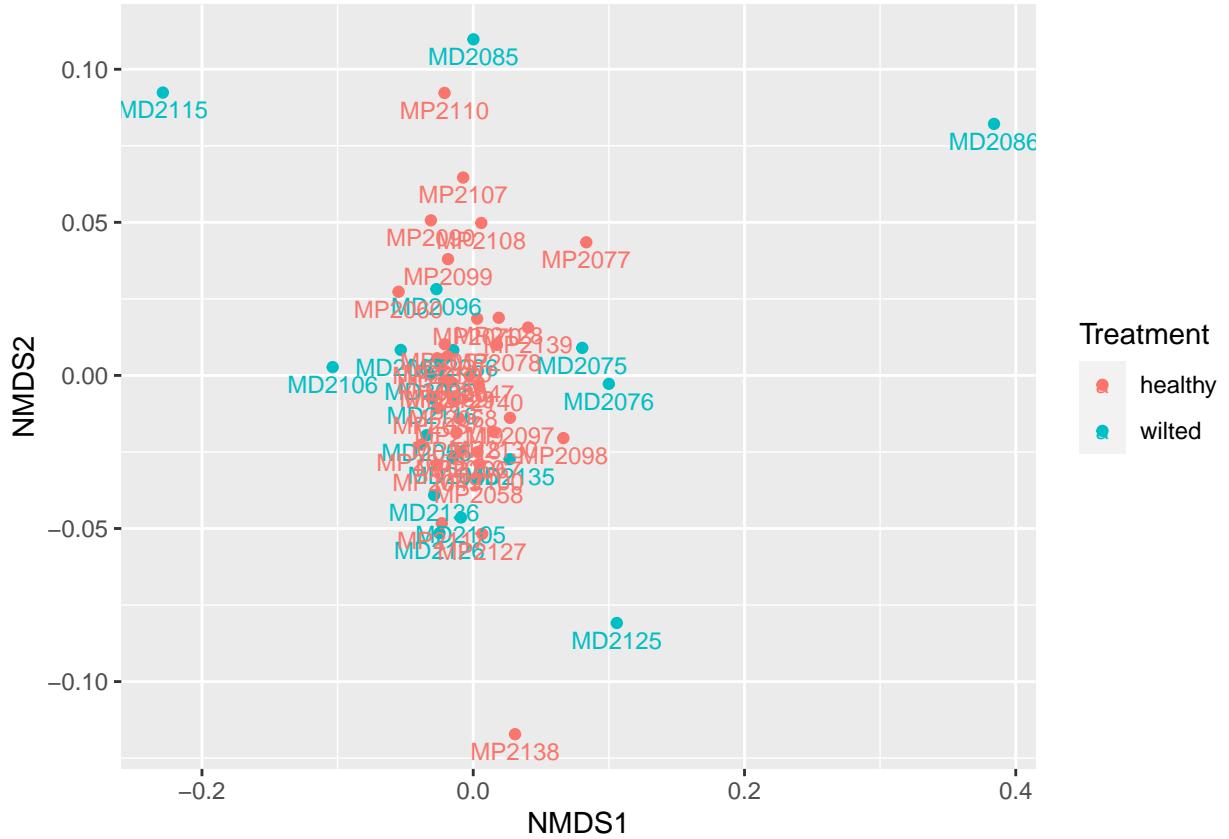
```
## Wisconsin double standardization
## Run 0 stress 0.110709
## Run 1 stress 0.1119817
## Run 2 stress 0.1176789
## Run 3 stress 0.1107094
## ... Procrustes: rmse 0.001110875 max resid 0.006357406
## ... Similar to previous best
## Run 4 stress 0.1107123
## ... Procrustes: rmse 0.0006081088 max resid 0.003382023
## ... Similar to previous best
## Run 5 stress 0.1119822
## Run 6 stress 0.111982
## Run 7 stress 0.1107085
## ... New best solution
## ... Procrustes: rmse 0.0006905962 max resid 0.003953207
## ... Similar to previous best
## Run 8 stress 0.1107097
```

```

## ... Procrustes: rmse 0.000498978 max resid 0.00285821
## ... Similar to previous best
## Run 9 stress 0.1107094
## ... Procrustes: rmse 0.0004042803 max resid 0.002318586
## ... Similar to previous best
## Run 10 stress 0.1145078
## Run 11 stress 0.1236307
## Run 12 stress 0.1119823
## Run 13 stress 0.1107093
## ... Procrustes: rmse 0.0003766442 max resid 0.002156754
## ... Similar to previous best
## Run 14 stress 0.11071
## ... Procrustes: rmse 0.0009845455 max resid 0.005603379
## ... Similar to previous best
## Run 15 stress 0.1119828
## Run 16 stress 0.1119826
## Run 17 stress 0.1107099
## ... Procrustes: rmse 0.000956729 max resid 0.005442851
## ... Similar to previous best
## Run 18 stress 0.1107096
## ... Procrustes: rmse 0.0008634493 max resid 0.004912192
## ... Similar to previous best
## Run 19 stress 0.1119812
## Run 20 stress 0.1119789
## *** Best solution repeated 7 times

plot_ordination(physeq = percentages_Eukaryota_Genus, ordination = meta_ord_Eukaryota_Genus, color = "T"
  geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data)), size = 3, vjust = 1.5)

```



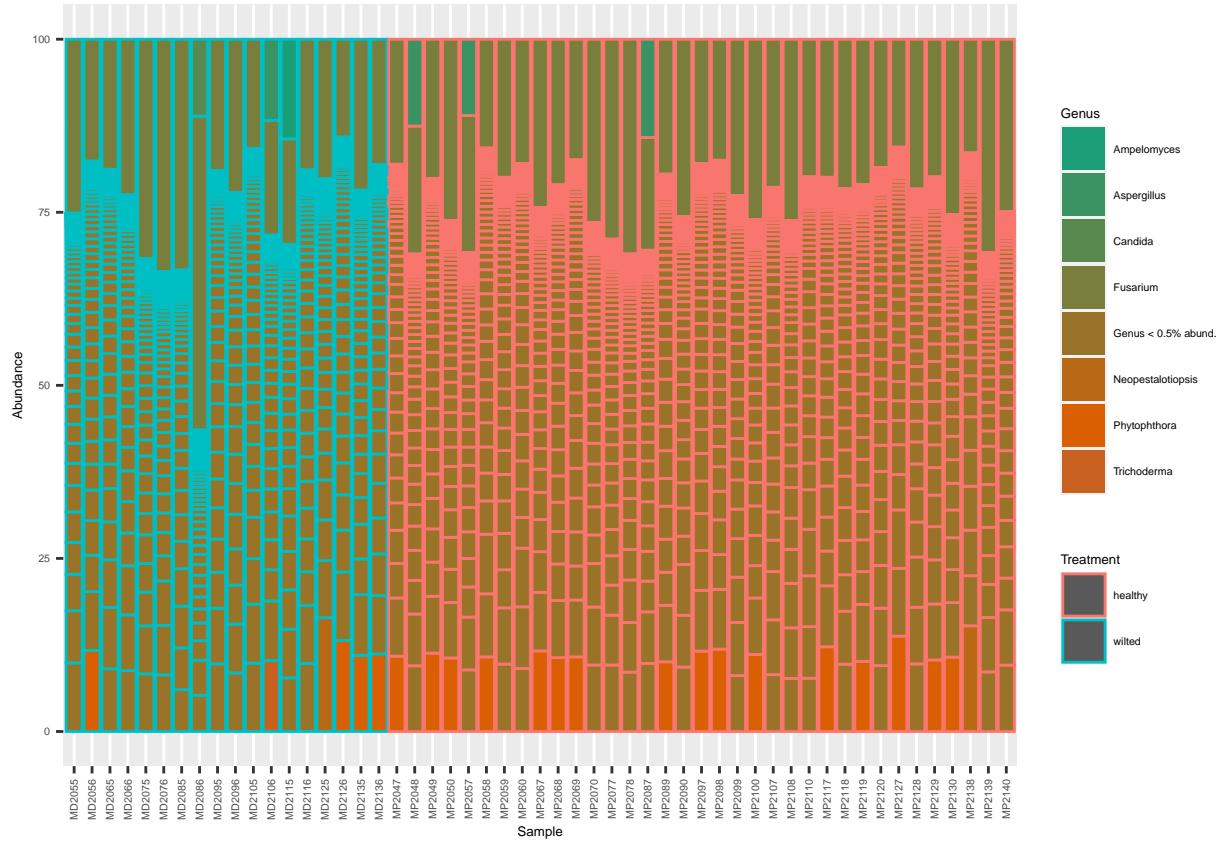
tomando diferentes porcentajes de abundancia

```

percentages_Eukaryota_Genus_df$Genus[percentages_Eukaryota_Genus_df$Abundance < 10.0] <- "Genus < 0.5%"
percentages_Eukaryota_Genus_df$Genus <- as.factor(percentages_Eukaryota_Genus_df$Genus)

genus_colors_rel <- colorRampPalette(brewer.pal(8, "Dark2")) (length(levels(percentages_Eukaryota_Genus_df$Genus))
relative_plot <- ggplot(data=percentages_Eukaryota_Genus_df, aes(x=Sample, y=Abundance, fill=Genus ,color=Genus))
  geom_bar(aes(), stat="identity", position="stack") +
  scale_fill_manual(values = phylum_colors_rel) +
  theme(text = element_text(size = 5),
        axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5))

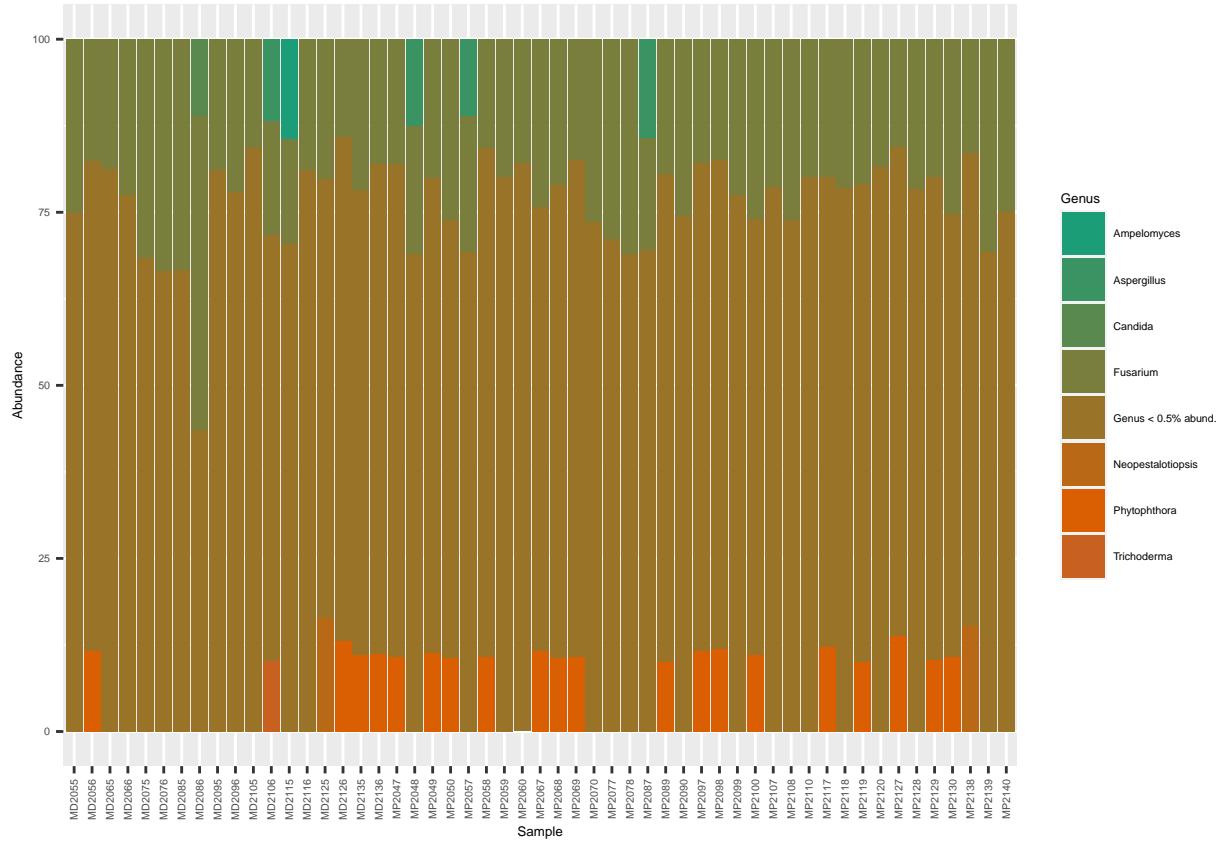
relative_plot
  
```



```

relative_plot <- ggplot(data=percentages_Eukaryota_Genus_df, aes(x=Sample, y=Abundance, fill=Genus))+
  geom_bar(aes(), stat="identity", position="stack") +
  scale_fill_manual(values = phylum_colors_rel) +
  theme(text = element_text(size = 5),
        axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5))
relative_plot

```



sacando la beta diversidad con aglomerado de 10%

```
meta_ord_Eukaryota_Genus <- ordinate(physeq = percentages_Eukaryota_Genus, method = "NMDS", distance = 

## Wisconsin double standardization
## Run 0 stress 0.110709
## Run 1 stress 0.1223848
## Run 2 stress 0.1119784
## Run 3 stress 0.1107097
## ... Procrustes: rmse 0.0002124151 max resid 0.001183418
## ... Similar to previous best
## Run 4 stress 0.1315555
## Run 5 stress 0.1107095
## ... Procrustes: rmse 0.000161435 max resid 0.0008925069
## ... Similar to previous best
## Run 6 stress 0.1223841
## Run 7 stress 0.1222462
## Run 8 stress 0.1119813
## Run 9 stress 0.1119815
## Run 10 stress 0.1235025
## Run 11 stress 0.1119824
## Run 12 stress 0.1236308
## Run 13 stress 0.122247
## Run 14 stress 0.1341447
## Run 15 stress 0.1235034
## Run 16 stress 0.1119786
```

```

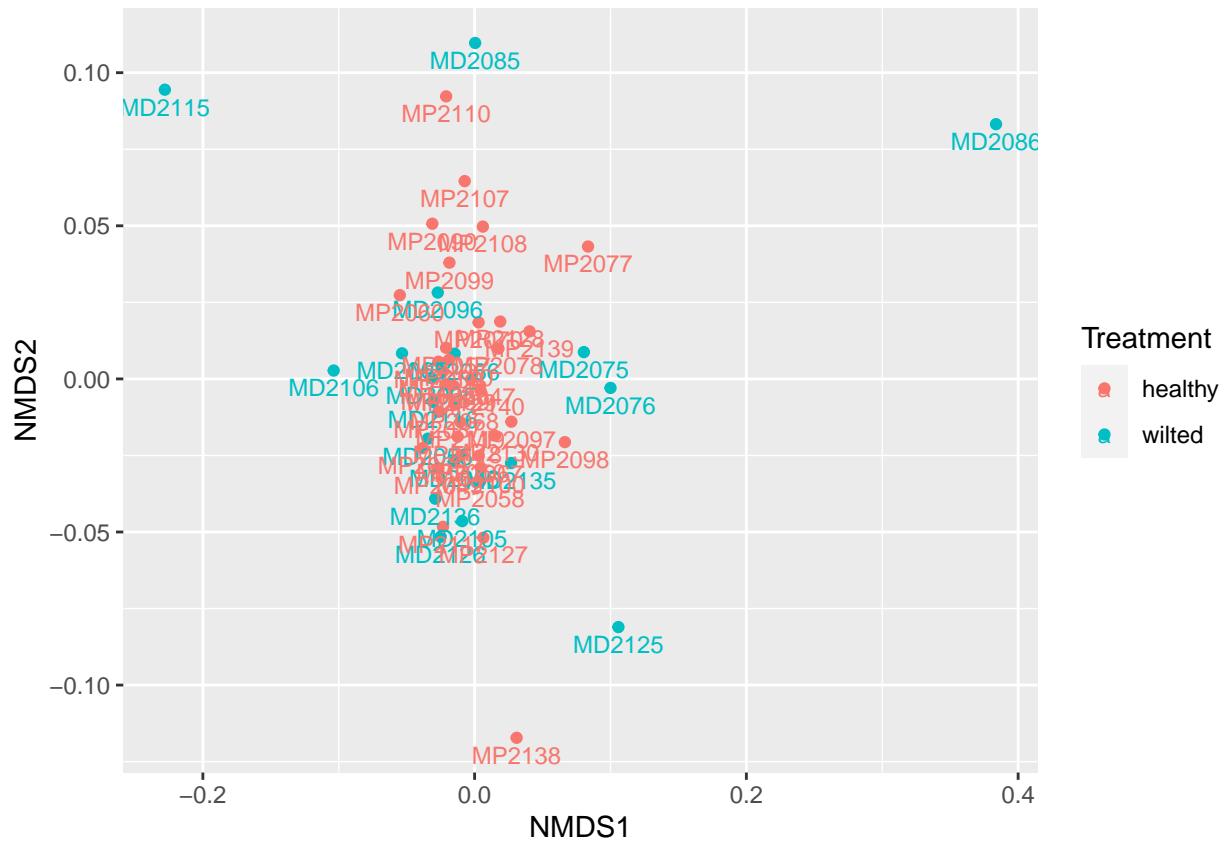
## Run 17 stress 0.1235018
## Run 18 stress 0.1116557
## Run 19 stress 0.1107088
## ... New best solution
## ... Procrustes: rmse 0.0001141873 max resid 0.0006636301
## ... Similar to previous best
## Run 20 stress 0.1119817
## *** Best solution repeated 1 times

```

```

plot_ordination(physeq = percentages_Eukaryota_Genus, ordination = meta_ord_Eukaryota_Genus, color = "Treatment"
geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data)), size = 3, vjust = 1.5)

```



Veremos aqui solo el reino Bacteriano

```
merge_Bacteria <- subset_taxa(fresa_kraken_fil, Kingdom == "Bacteria")
```

sacamos las abundancias relativas

```
percentages_Bacteria <- transform_sample_counts(merge_Bacteria, function(x) x * 100 / sum(x))
percentages_Bacteria_df <- psmelt(percentages_Bacteria)
```

beta diversidad de Bacteria

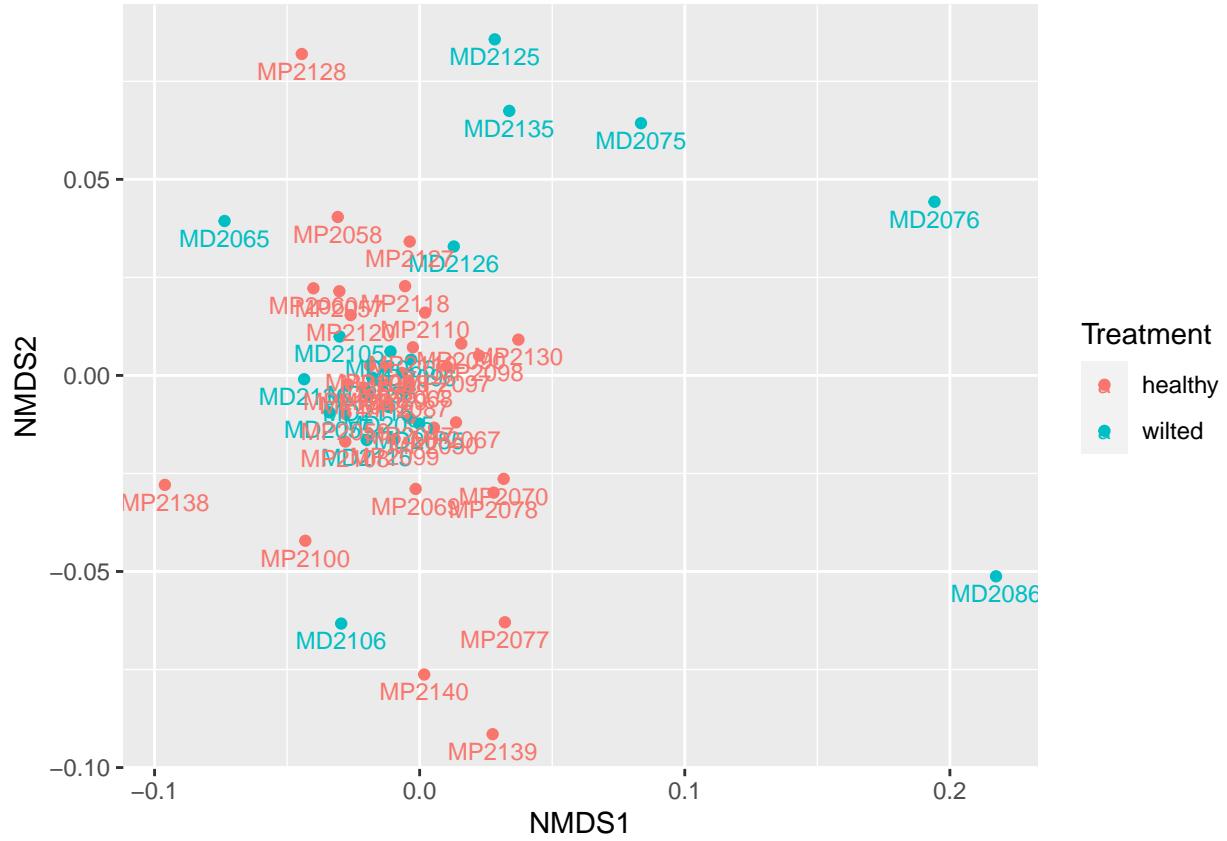
```

meta_ord_Bacteria <- ordinate(physeq = percentages_Bacteria, method = "NMDS", distance = "bray")

## Wisconsin double standardization
## Run 0 stress 0.1680232
## Run 1 stress 0.1658268
## ... New best solution
## ... Procrustes: rmse 0.06625423 max resid 0.3497755
## Run 2 stress 0.1687377
## Run 3 stress 0.1691068
## Run 4 stress 0.1686396
## Run 5 stress 0.1665102
## Run 6 stress 0.1797448
## Run 7 stress 0.1742406
## Run 8 stress 0.1692049
## Run 9 stress 0.1992948
## Run 10 stress 0.1661233
## ... Procrustes: rmse 0.07083586 max resid 0.3407577
## Run 11 stress 0.1708893
## Run 12 stress 0.1658877
## ... Procrustes: rmse 0.06560222 max resid 0.3474303
## Run 13 stress 0.1689047
## Run 14 stress 0.1657577
## ... New best solution
## ... Procrustes: rmse 0.01952603 max resid 0.1283628
## Run 15 stress 0.1717097
## Run 16 stress 0.1710615
## Run 17 stress 0.1800279
## Run 18 stress 0.1680232
## Run 19 stress 0.1640079
## ... New best solution
## ... Procrustes: rmse 0.05965582 max resid 0.3561727
## Run 20 stress 0.1799721
## *** Best solution was not repeated -- monoMDS stopping criteria:
##      2: no. of iterations >= maxit
##      18: stress ratio > sratmax

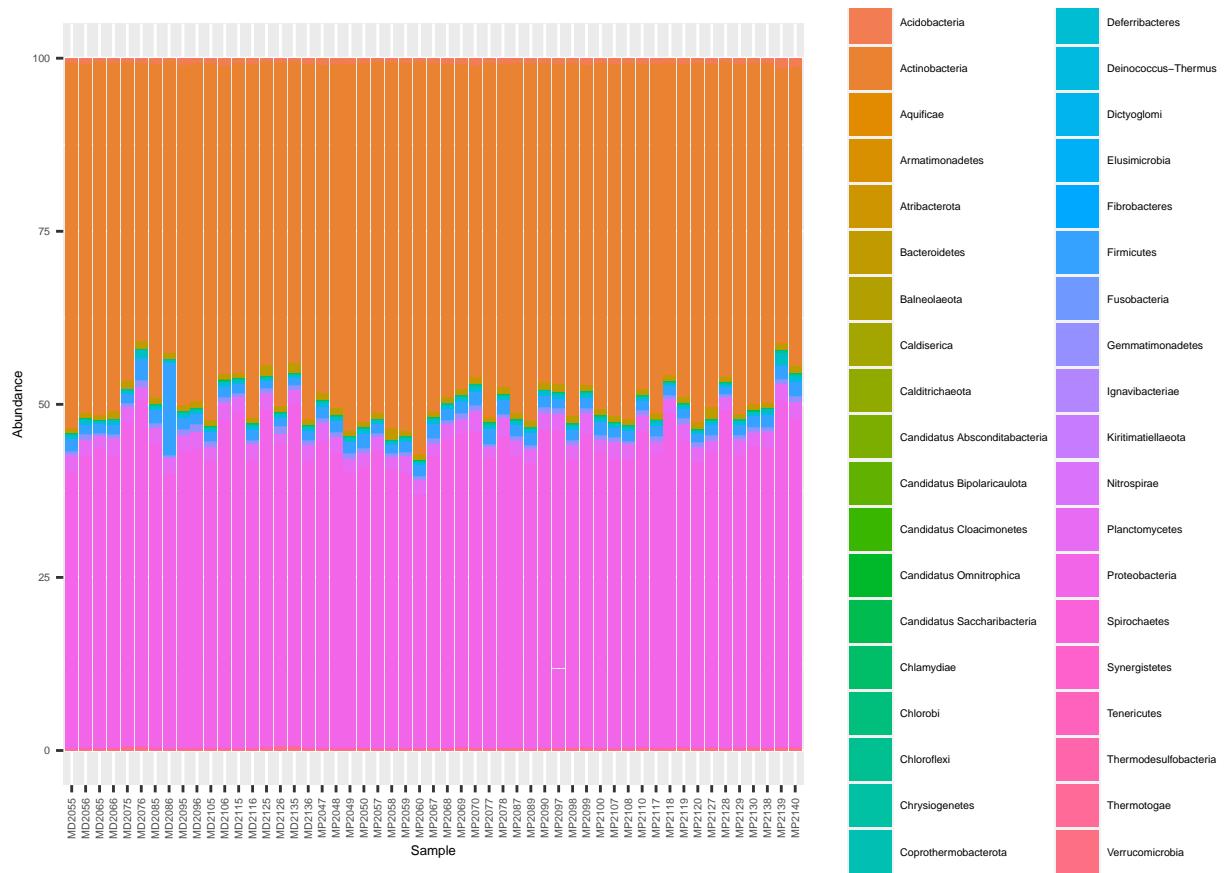
plot_ordination(physeq = percentages_Bacteria, ordination = meta_ord_Bacteria, color = "Treatment") +
  geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data)), size = 3, vjust = 1.5)

```



Bacterias por Phylum

```
ggplot(data= percentages_Bacteria_df, aes(x=Sample, y=Abundance, fill=Phylum))+  
  geom_bar(aes(), stat="identity", position="stack") +  
  theme(text = element_text(size = 5),  
        axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5))
```



```
merge_Bacteria_Phylum<-tax_glm(merge_Bacteria, taxrank="Phylum")
```

sacamos las abundancias relativas

```
percentages_Bacteria_Phylum <- transform_sample_counts(merge_Bacteria_Phylum, function(x) x*100 / sum(x))
percentages_Bacteria_Phylum_df <- psmelt(percentages_Bacteria_Phylum)
meta_ord_Bacteria_Phylum <- ordinate(phylseq = percentages_Bacteria_Phylum, method = "NMDS", distance =
```

```
## Square root transformation
## Wisconsin double standardization
## Run 0 stress 0.1520004
## Run 1 stress 0.1524009
## ... Procrustes: rmse 0.04800039 max resid 0.315377
## Run 2 stress 0.1646847
## Run 3 stress 0.1519998
## ... New best solution
## ... Procrustes: rmse 0.0002224863 max resid 0.001257028
## ... Similar to previous best
## Run 4 stress 0.1550553
## Run 5 stress 0.1597146
## Run 6 stress 0.1550552
## Run 7 stress 0.1619444
## Run 8 stress 0.155422
## Run 9 stress 0.174961
## Run 10 stress 0.1548791
```

```

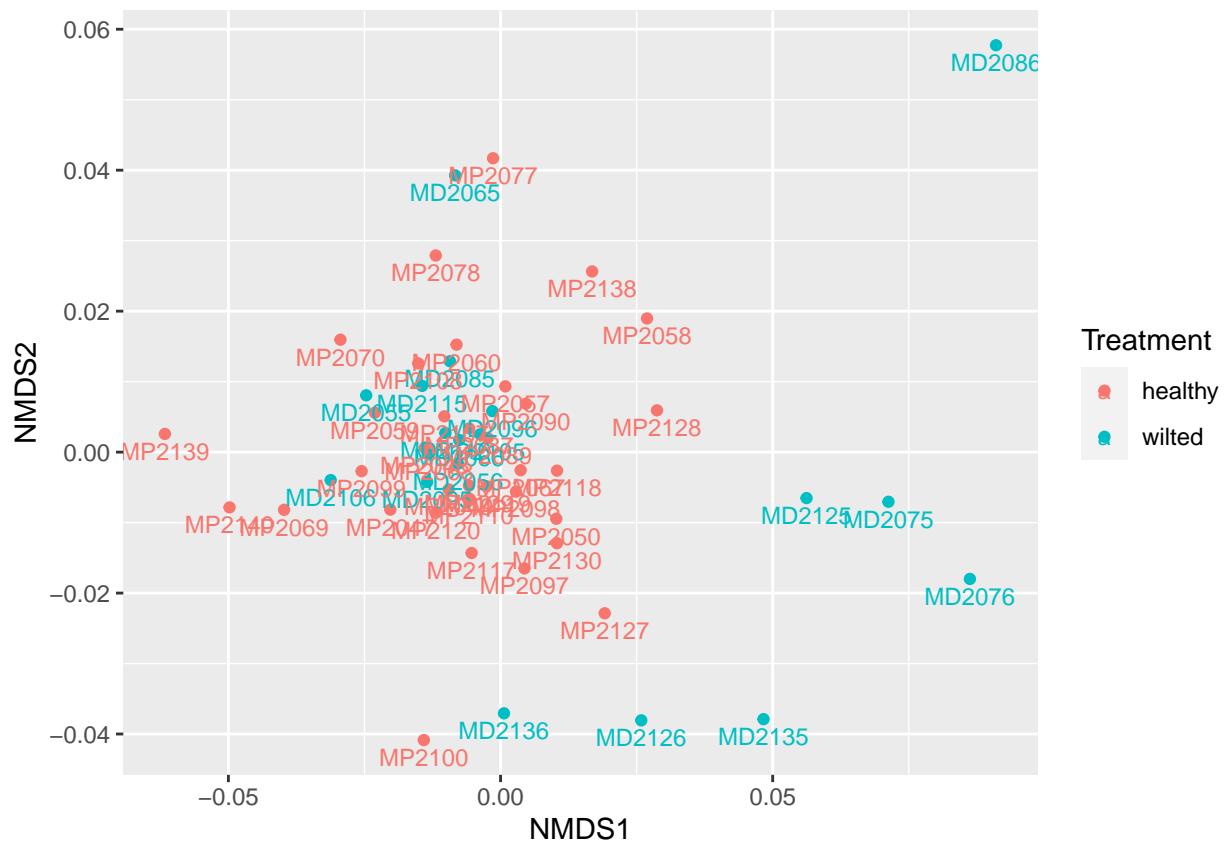
## Run 11 stress 0.1749744
## Run 12 stress 0.1652839
## Run 13 stress 0.1587859
## Run 14 stress 0.1578492
## Run 15 stress 0.1710106
## Run 16 stress 0.1597147
## Run 17 stress 0.1572805
## Run 18 stress 0.1693713
## Run 19 stress 0.164222
## Run 20 stress 0.1630501
## *** Best solution repeated 1 times

```

```

plot_ordination(physeq = percentages_Bacteria_Phylum, ordination = meta_ord_Bacteria_Phylum, color = "Treatment"
geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data)), size = 3, vjust = 1.5)

```



```

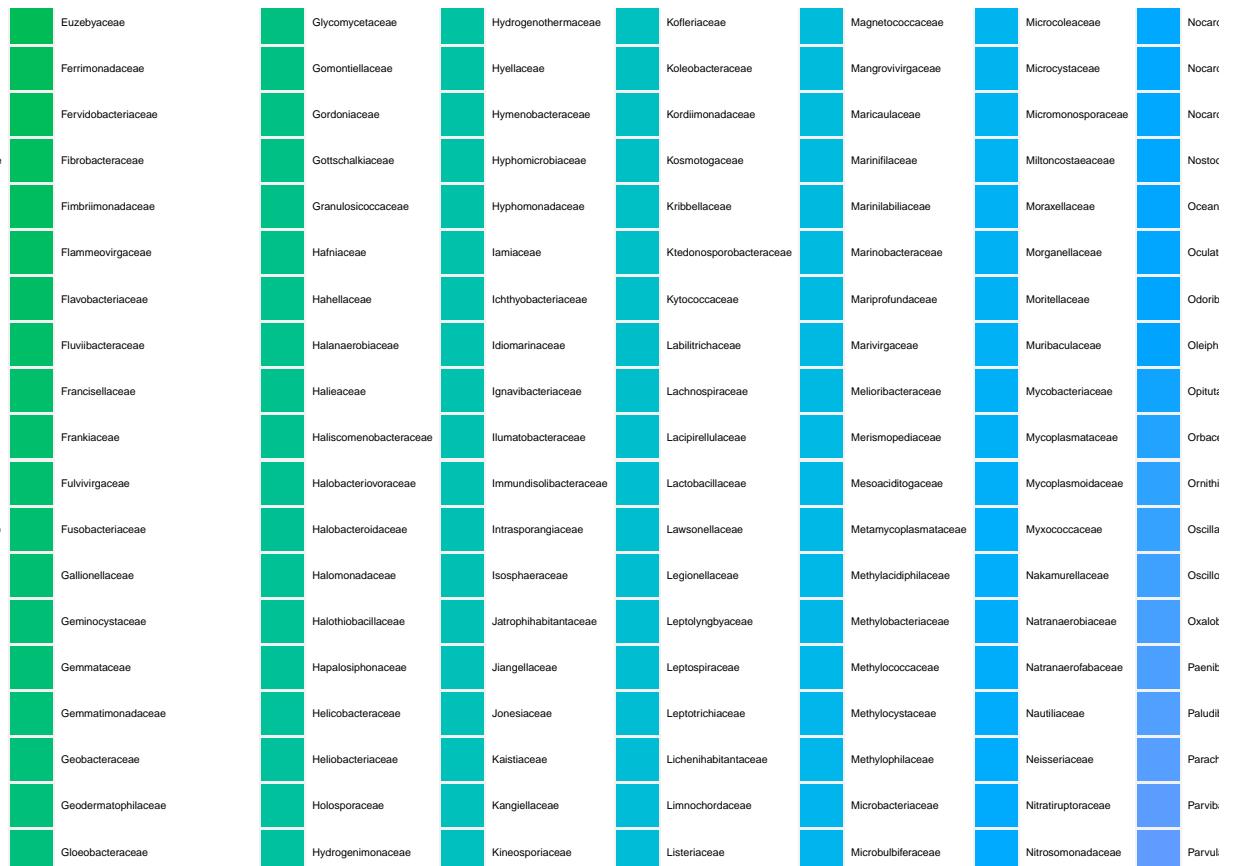
## Bacterias por Family

```

```

ggplot(data= percentages_Bacteria_df, aes(x=Sample, y=Abundance, fill=Family))+ 
  geom_bar(aes(), stat="identity", position="stack") + 
  theme(text = element_text(size = 5),
        axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5))

```



```
merge_Bacteria_Family <- tax_glom(merge_Bacteria, taxrank = "Family")
```

sacamos las abundancias relativas

```
percentages_Bacteria_Family <- transform_sample_counts(merge_Bacteria_Family, function(x) x*100 / sum(x))
percentages_Bacteria_Family_df <- psmelt(percentages_Bacteria_Family)
meta_ord_Bacteria_Family <- ordinate(physeq = percentages_Bacteria_Family, method = "NMDS", distance = "euclidean")
```

```
## Wisconsin double standardization
## Run 0 stress 0.1412073
## Run 1 stress 0.1841839
## Run 2 stress 0.171039
## Run 3 stress 0.1373771
## ... New best solution
## ... Procrustes: rmse 0.06365222 max resid 0.3610578
## Run 4 stress 0.1396571
## Run 5 stress 0.1569678
## Run 6 stress 0.1423481
## Run 7 stress 0.137038
## ... New best solution
## ... Procrustes: rmse 0.01628313 max resid 0.05956157
## Run 8 stress 0.1369861
## ... New best solution
```

```

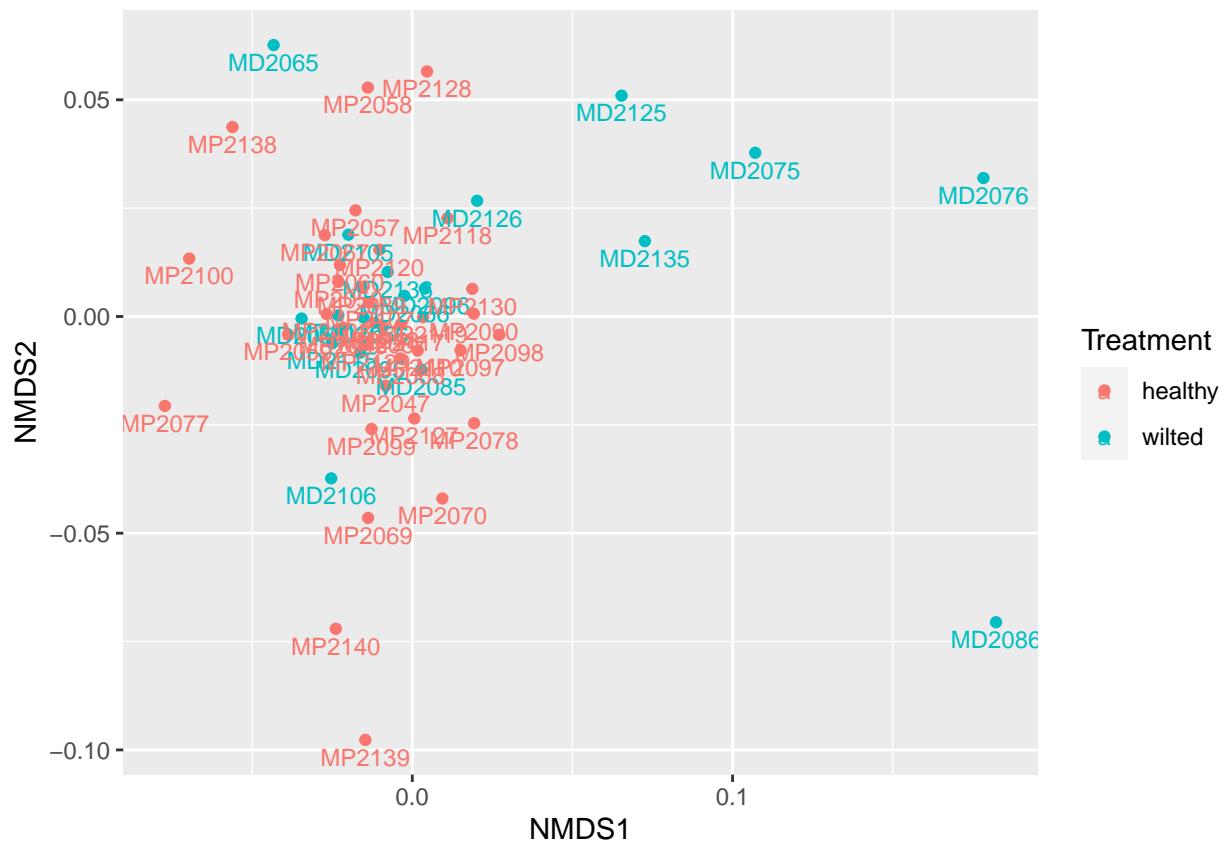
## ... Procrustes: rmse 0.00239762 max resid 0.01201976
## Run 9 stress 0.137038
## ... Procrustes: rmse 0.002397831 max resid 0.0120207
## Run 10 stress 0.1731639
## Run 11 stress 0.1412064
## Run 12 stress 0.1374744
## ... Procrustes: rmse 0.0155282 max resid 0.05889792
## Run 13 stress 0.1799551
## Run 14 stress 0.1370382
## ... Procrustes: rmse 0.002410986 max resid 0.01199184
## Run 15 stress 0.1522704
## Run 16 stress 0.1373768
## ... Procrustes: rmse 0.01527212 max resid 0.05900154
## Run 17 stress 0.1401715
## Run 18 stress 0.139046
## Run 19 stress 0.139501
## Run 20 stress 0.1373772
## ... Procrustes: rmse 0.01518985 max resid 0.05894204
## *** Best solution was not repeated -- monoMDS stopping criteria:
##      3: no. of iterations >= maxit
##      17: stress ratio > sratmax

```

```

plot_ordination(physeq = percentages_Bacteria_Family, ordination = meta_ord_Bacteria_Family, color = "Treatment",
geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data)), size = 3, vjust = 1.5)

```

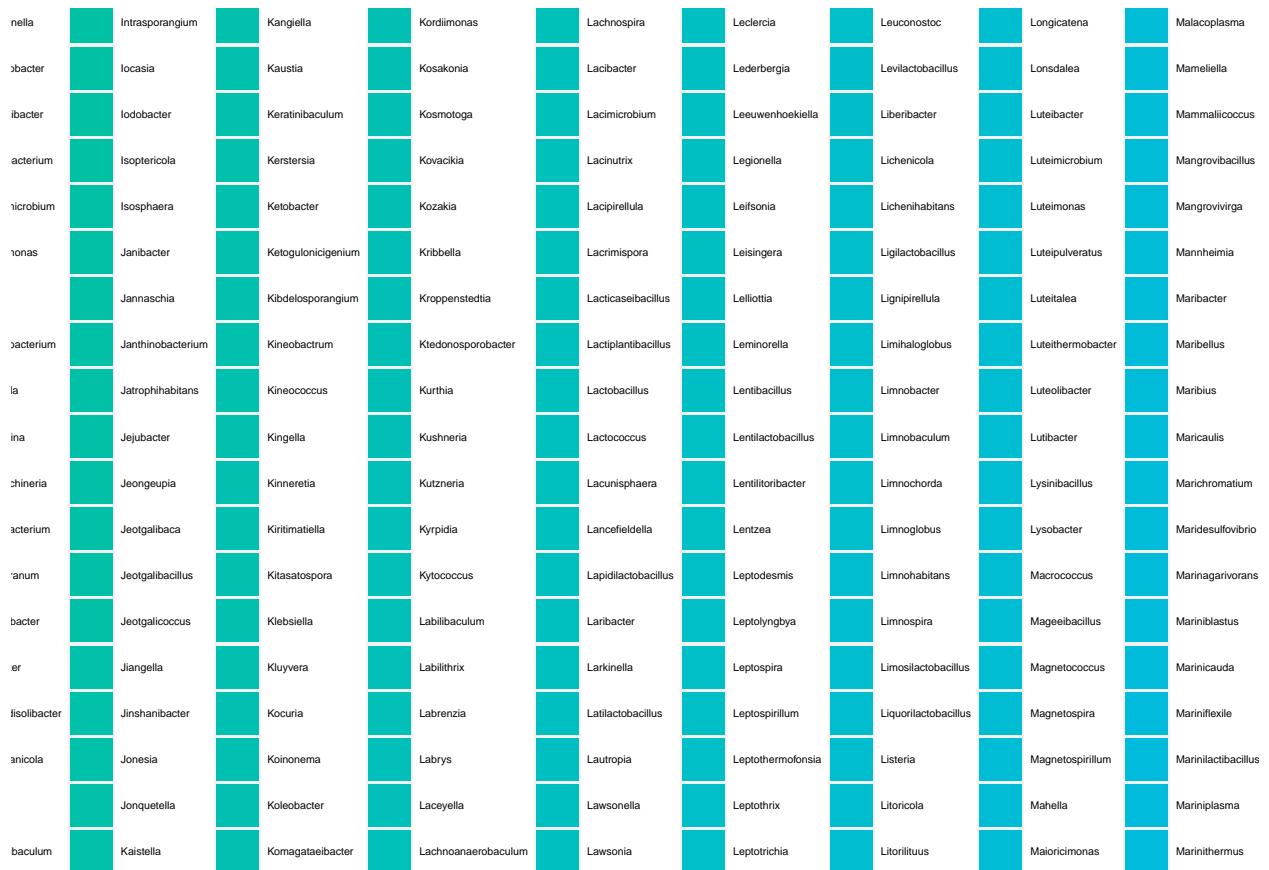


```

## Bacterias por Genero

```

```
ggplot(data= percentages_Bacteria_df, aes(x=Sample, y=Abundance, fill=Genus))+  
  geom_bar(aes(), stat="identity", position="stack") +  
  theme(text = element_text(size = 5),  
        axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5))
```



```
merge_Bacteria_Genus<-tax_glom(merge_Bacteria,taxrank="Genus")
```

sacamos las abundancias relativas

```
percentages_Bacteria_Genus <- transform_sample_counts(merge_Bacteria_Genus, function(x) x*100 / sum(x))  
percentages_Bacteria_Genus_df <- psmelt(percentages_Bacteria_Genus)  
meta_ord_Bacteria_Genus <- ordinate(physeq = percentages_Bacteria_Genus, method = "NMDS", distance = "bray")
```

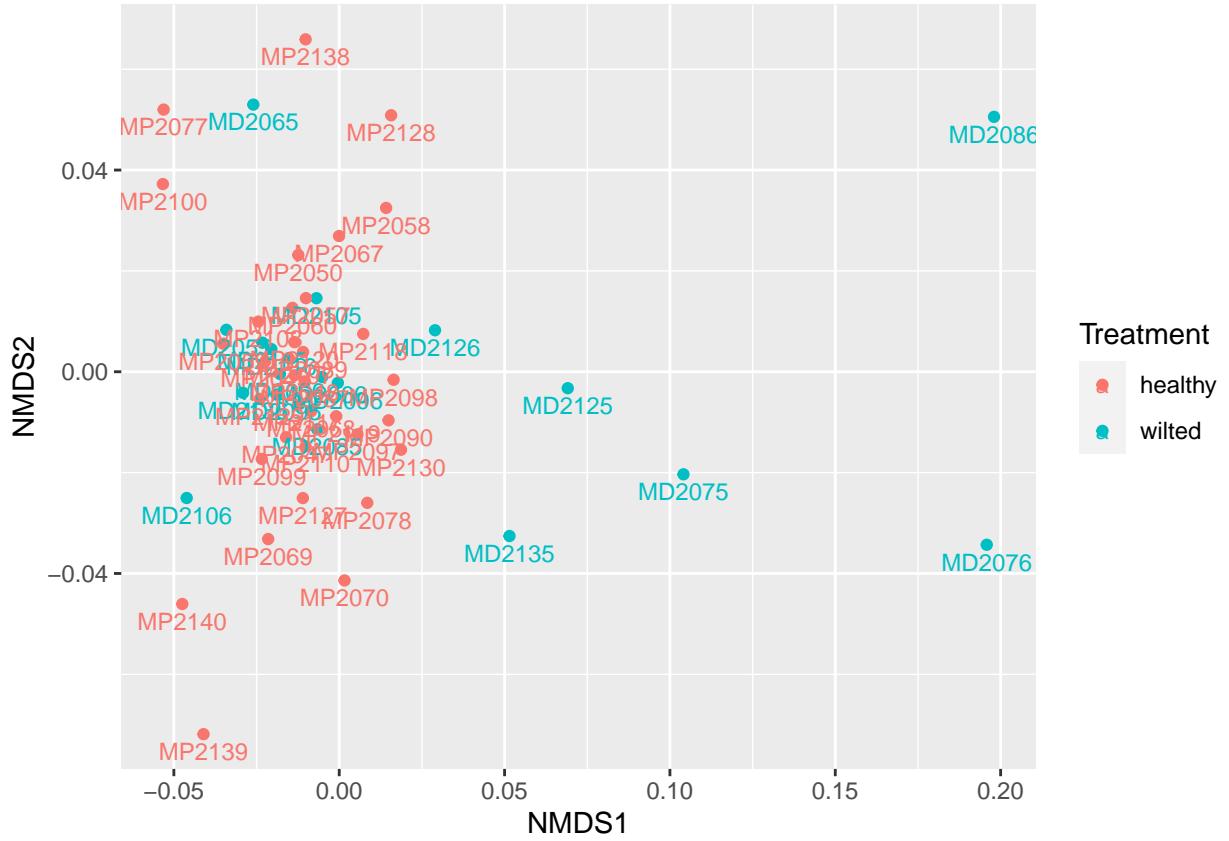
```
## Wisconsin double standardization  
## Run 0 stress 0.1467945  
## Run 1 stress 0.1467691  
## ... New best solution  
## ... Procrustes: rmse 0.002700535 max resid 0.01690071  
## Run 2 stress 0.1660091  
## Run 3 stress 0.1514  
## Run 4 stress 0.1527708  
## Run 5 stress 0.1453935  
## ... New best solution  
## ... Procrustes: rmse 0.08909263 max resid 0.2984409
```

```

## Run 6 stress 0.1439574
## ... New best solution
## ... Procrustes: rmse 0.02430511 max resid 0.1319898
## Run 7 stress 0.1504571
## Run 8 stress 0.1456405
## Run 9 stress 0.1502732
## Run 10 stress 0.1538257
## Run 11 stress 0.143814
## ... New best solution
## ... Procrustes: rmse 0.05706939 max resid 0.3103235
## Run 12 stress 0.143284
## ... New best solution
## ... Procrustes: rmse 0.05080514 max resid 0.3015335
## Run 13 stress 0.1432845
## ... Procrustes: rmse 0.001290418 max resid 0.005619273
## ... Similar to previous best
## Run 14 stress 0.154184
## Run 15 stress 0.1490354
## Run 16 stress 0.1482214
## Run 17 stress 0.1484507
## Run 18 stress 0.1432838
## ... New best solution
## ... Procrustes: rmse 6.320535e-05 max resid 0.0003107312
## ... Similar to previous best
## Run 19 stress 0.1433099
## ... Procrustes: rmse 0.003956295 max resid 0.02546086
## Run 20 stress 0.1438152
## *** Best solution repeated 1 times

plot_ordination(physeq = percentages_Bacteria_Genus, ordination = meta_ord_Bacteria_Genus, color = "Tre
  geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data)), size = 3, vjust = 1.5)

```



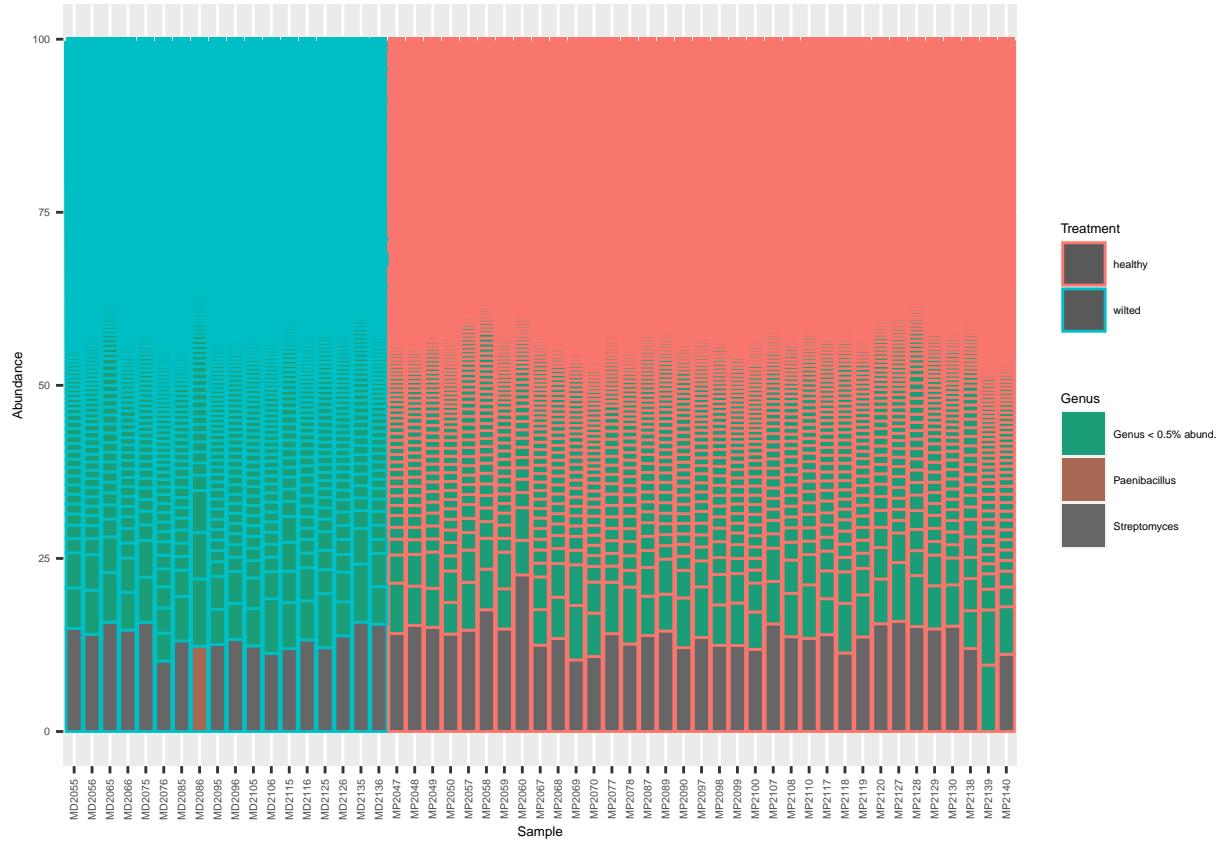
tomando diferentes porcentajes de abundancia

```

percentages_Bacteria_Genus_df$Genus [percentages_Bacteria_Genus_df$Abundance < 10.0] <- "Genus < 0.5% abundance"
percentages_Bacteria_Genus_df$Genus <- as.factor(percentages_Bacteria_Genus_df$Genus)

genus_colors_rel <- colorRampPalette(brewer.pal(8,"Dark2")) (length(levels(percentages_Bacteria_Genus_df$Genus)))
relative_plot <- ggplot(data=percentages_Bacteria_Genus_df, aes(x=Sample, y=Abundance, fill=Genus ,color=Genus))
  geom_bar(aes(), stat="identity", position="stack") +
  scale_fill_manual(values = genus_colors_rel) +
  theme(text = element_text(size = 5),
        axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5))

relative_plot
  
```

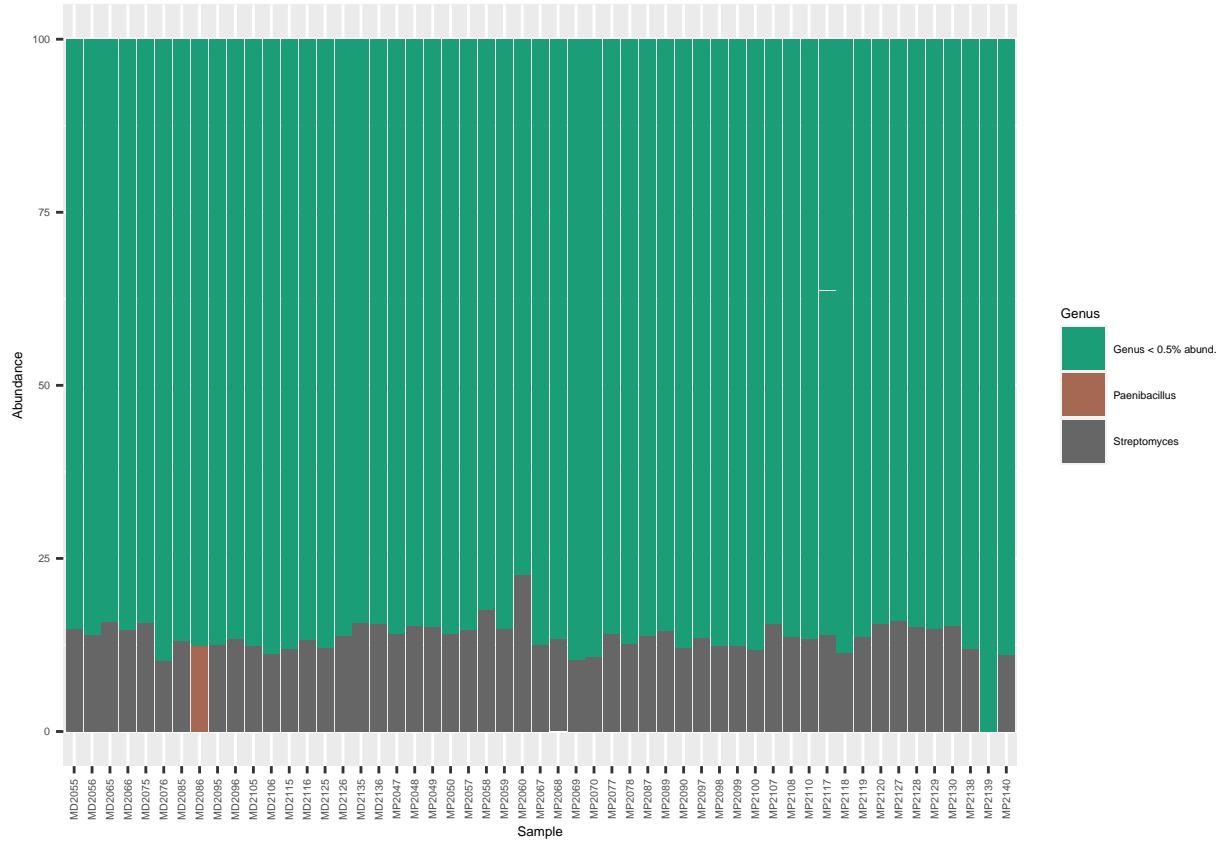


```

relative_plot <- ggplot(data=percentages_Bacteria_Genus_df, aes(x=Sample, y=Abundance, fill=Genus)) +
  geom_bar(aes(), stat="identity", position="stack") +
  scale_fill_manual(values = genus_colors_rel) +
  theme(text = element_text(size = 5),
        axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5))

relative_plot

```



sacando la beta diversidad con aglomerado de 10%

```
meta_ord_Bacteria_Genus <- ordinate(physeq = percentages_Bacteria_Genus, method = "NMDS", distance = "b
```

```
## Wisconsin double standardization
## Run 0 stress 0.1467945
## Run 1 stress 0.1561836
## Run 2 stress 0.1497397
## Run 3 stress 0.1482215
## Run 4 stress 0.1458785
## ... New best solution
## ... Procrustes: rmse 0.05080048 max resid 0.2810505
## Run 5 stress 0.1433354
## ... New best solution
## ... Procrustes: rmse 0.07338854 max resid 0.3700134
## Run 6 stress 0.1541723
## Run 7 stress 0.1502742
## Run 8 stress 0.1476415
## Run 9 stress 0.1544768
## Run 10 stress 0.1432845
## ... New best solution
## ... Procrustes: rmse 0.005055307 max resid 0.03013918
## Run 11 stress 0.1513999
## Run 12 stress 0.1438163
## Run 13 stress 0.1437857
## Run 14 stress 0.1437856
```

```

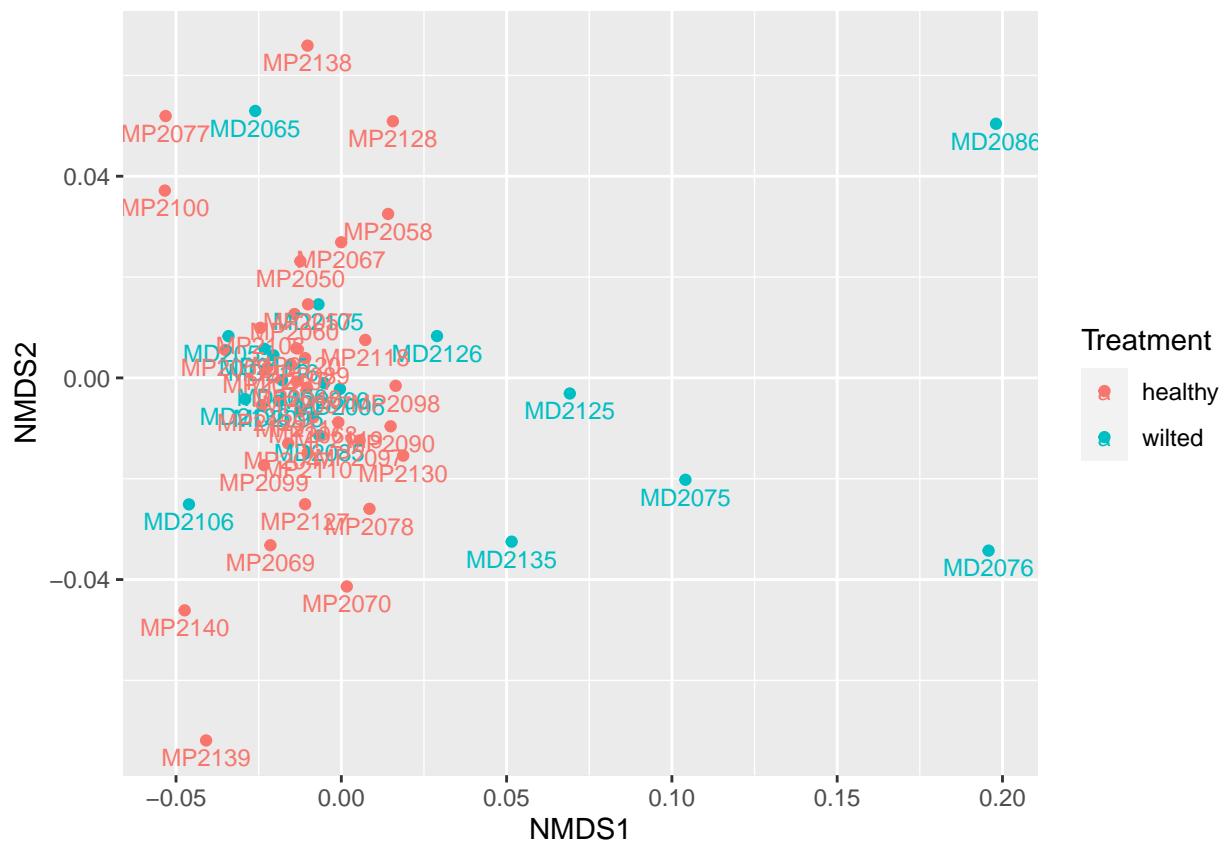
## Run 15 stress 0.1492682
## Run 16 stress 0.1432835
## ... New best solution
## ... Procrustes: rmse 0.0003142473 max resid 0.001426629
## ... Similar to previous best
## Run 17 stress 0.1505676
## Run 18 stress 0.4024306
## Run 19 stress 0.1454658
## Run 20 stress 0.1441635
## *** Best solution repeated 1 times

```

```

plot_ordination(physeq = percentages_Bacteria_Genus, ordination = meta_ord_Bacteria_Genus, color = "Treatment"
  geom_text(mapping = aes(label = colnames(fresa_kraken_fil@otu_table@Data)), size = 3, vjust = 1.5)

```



REDES podemos hacer un data.frame uniendo toda la informacion del objeto phyloseq

```
df <- psmelt(fresa_kraken_fil)
```

Hay dos funciones en el paquete phyloseq para trazar la red del microbioma usando “ggplot2”: `plot_network()` y `plot_net()`

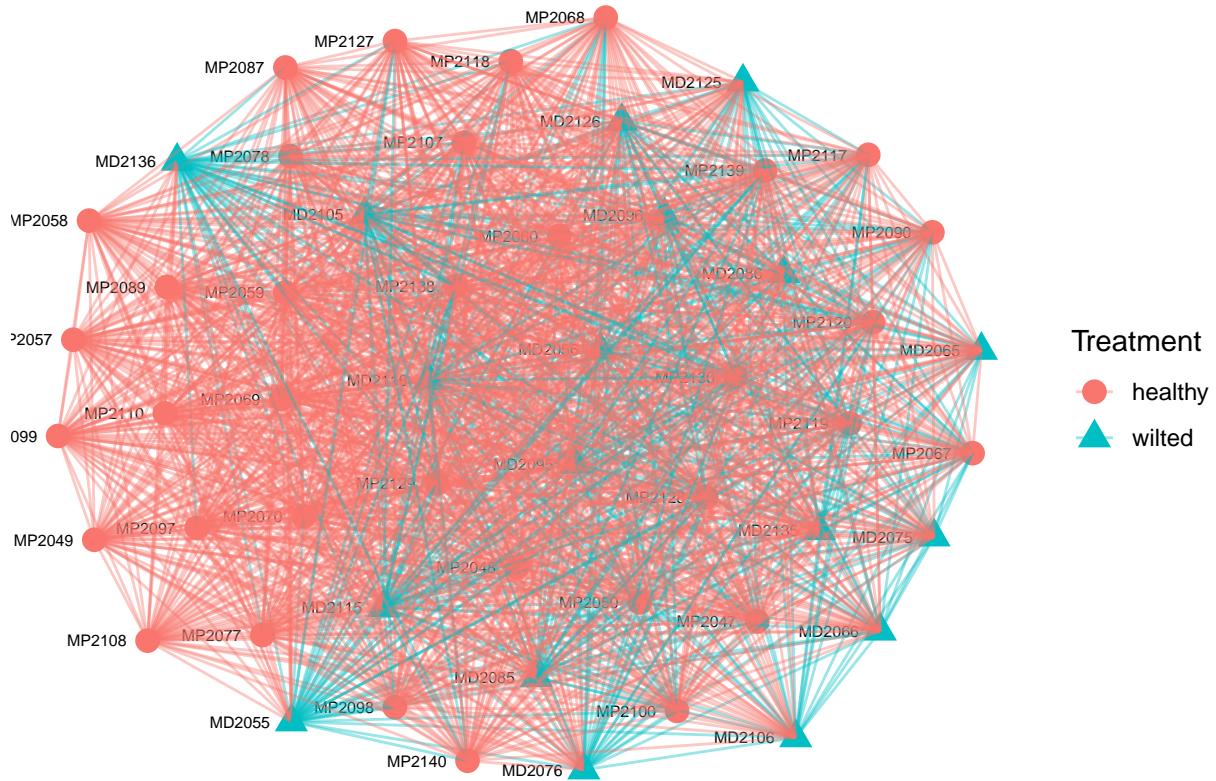
Se crean un grafo basado en igraph, basado en el método de distancia por defecto, Jaccard y una distancia máxima entre nodos conectados de 0,8. El “Treatment” se utiliza para los mapeos de color y forma para visualizar la estructura de las muestras

Hacemos una gráfica a partir de el objeto phyloseq

```
ig <- make_network(fresa_kraken_fil, max.dist=0.8)
```

Graficamos

```
plot_network(ig, fresa_kraken_fil, color="Treatment", shape="Treatment")
```



Los siguientes códigos crean una red basada en una distancia máxima entre los nodos conectados de 0,5
Graficamos sin necesidad de hacer el objeto anterior

```
plot_net(fresa_kraken_fil, maxdist = 0.5, color = "Treatment", shape="Treatment")
```

