

230306_Reporte1Exploracion

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```
library("phyloseq")
library("ggplot2")
library("vegan")
```

```
## Loading required package: permute
```

```
## Loading required package: lattice
```

```
## This is vegan 2.6-4
```

```
library("RColorBrewer")
library("stringi")
```

Cargado de datos originales

```
setwd("/home/camila/GIT/Tesis_Maestria/Data/fresa_solena/Data1")
outpath = "/home/camila/GIT/Tesis_Maestria/Analisis_Comparativo/Fresa_Solena/Results_img"

fresa_kraken <- import_biom("fresa_kraken.biom")
colnames(fresa_kraken@tax_table@.Data) <- c("Kingdom", "Phylum", "Class", "Order", "Family", "Genus", "Species")
fresa_kraken@tax_table@.Data <- substr(fresa_kraken@tax_table@.Data,4,100)
colnames(fresa_kraken@otu_table@.Data) <- substr(colnames(fresa_kraken@otu_table@.Data),1,6)
metadata_fresa <- read.csv2("/home/camila/GIT/Tesis_Maestria/Data/fresa_solena/Data1/metadata.csv",header=1)
fresa_kraken@sam_data <- sample_data(metadata_fresa)
fresa_kraken@sam_data$Sample <- row.names(fresa_kraken@sam_data)
colnames(fresa_kraken@sam_data) <- c('Treatment', 'Samples')
samples_to_remove <- c("MP2079", "MP2080", "MP2088", "MP2109", "MP2137")
fresa_kraken_fil <- prune_samples(!(sample_names(fresa_kraken) %in% samples_to_remove), fresa_kraken)
percentages_fil <- transform_sample_counts(fresa_kraken_fil, function(x) x*100 / sum(x) )
percentages_df <- psmelt(percentages_fil)
```

Subconjunto de “Eukaryota”

```
merge_Eukaryota <- subset_taxa(fresa_kraken_fil, Kingdom=="Eukaryota")
```

Subconjunto de “Bacteria”

```
merge_Bacteria<-subset_taxa(fresa_kraken_fil,Kingdom=="Bacteria")
```

Funciones

Crea los subconjuntos de datos input phy phyliseq total tax rango al que queremos recortar output

```
glomToGraph<-function(phy,tax){  
  ## creamos el subconjunto dependiendo del linaje taxonomico deseado  
  glom <- tax_glom(phy, taxrank = tax)  
  ## sacamos los porcentajes  
  percentages <- transform_sample_counts(glom, function(x) x*100 / sum(x) )  
  percentages_df <- psmelt(percentages)  
  return(list(glom,percentages,percentages_df))  
}
```

Graficar abundancias stackbar

input entra el percentages_df

```
Abundance_barras <- function(phy,tax,attribute,abundance_percentage){  
  ##llamar funcion de datos  
  Data <- glomToGraph(phy,tax)  
  glom <- Data[[1]] #phyloseq  
  percentages <- Data[[2]] #phyloseq  
  percentages_df <- Data[[3]] # dataframe  
  ## Graficamos para cada subconjunto las barras de abundancia  
  plot_barras <- ggplot(data=percentages_df, aes_string(x='Sample', y='Abundance', fill=tax ,color=attribute)) +  
    scale_colour_manual(values=c('white','black')) +  
    geom_bar(aes(), stat="identity", position="stack") +  
    labs(title = "Abundance", x='Sample', y='Abundance', color = tax) +  
    theme(legend.key.size = unit(0.2, "cm"),  
          legend.key.width = unit(0.25,"cm"),  
          legend.position = "bottom",  
          legend.direction = "horizontal",  
          legend.title=element_text(size=8, face = "bold"),  
          legend.text=element_text(size=6),  
          text = element_text(size=12),  
          axis.text.x = element_text(angle=90, size=5, hjust=1, vjust=0.5))  
  percentages_df$tax<-percentages_df[,ncol(percentages_df)]  
  percentages_df$tax[percentages_df$Abundance < abundance_percentage] <- "abundance_percentage"  
  percentages_df$tax <- as.factor(percentages_df$tax)  
  plot_percentages <- ggplot(data=percentages_df, aes_string(x='Sample', y='Abundance', fill='tax' ,color=attribute)) +  
    scale_colour_manual(values=c('white','black')) +  
    geom_bar(aes(), stat="identity", position="stack") +  
    labs(title = "Abundance", x='Sample', y='Abundance', color = tax) +  
    theme(legend.key.size = unit(0.3, "cm"),  
          legend.key.width = unit(0.5,"cm"),  
          legend.position = "bottom",
```

```

    legend.direction = "horizontal",
    legend.title=element_text(size=10, face = "bold"),
    legend.text=element_text(size=8),
    text = element_text(size=12),
    axis.text.x = element_text(angle=90, size=5, hjust=1, vjust=0.5))
  return(list(plot_barras,plot_percentages))
}

```

Graficar betadiversity

```

Beta_diversity <- function(phy,tax,attribute,distance){
  Data <- glomToGraph(phy,tax)
  glom <- Data[[1]]
  #CREAR UN GLOM AL 10%
  percentages <- Data[[2]]
  percentages_df <- Data[[3]]
  ## Beta diversidad
  meta_ord <- ordinate(physeq = percentages, method = "NMDS", distance = distance)
  plot_beta <- plot_ordination(physeq = percentages, ordination = meta_ord, color = attribute) +
    geom_text(mapping = aes(label = colnames(phy@otu_table@.Data)), size = 3, vjust = 1.5)
  return(plot_beta)
}

```

Graficar alphadiversity

```

Alpha_diversity <- function(phy,tax,attribute){
  ## llamamos la funcion que crea los dataset
  Data <- glomToGraph(phy,tax)
  glom <- Data[[1]]

  percentages <- Data[[2]]
  percentages_df <- Data[[3]]
  ## Alfa diversidad
  plot_alpha <- plot_richness(physeq = glom, measures = c("Observed","Chao1","Shannon","simpson"),x = a
  return(plot_alpha)
}

```

#———Eukarya by Phylum

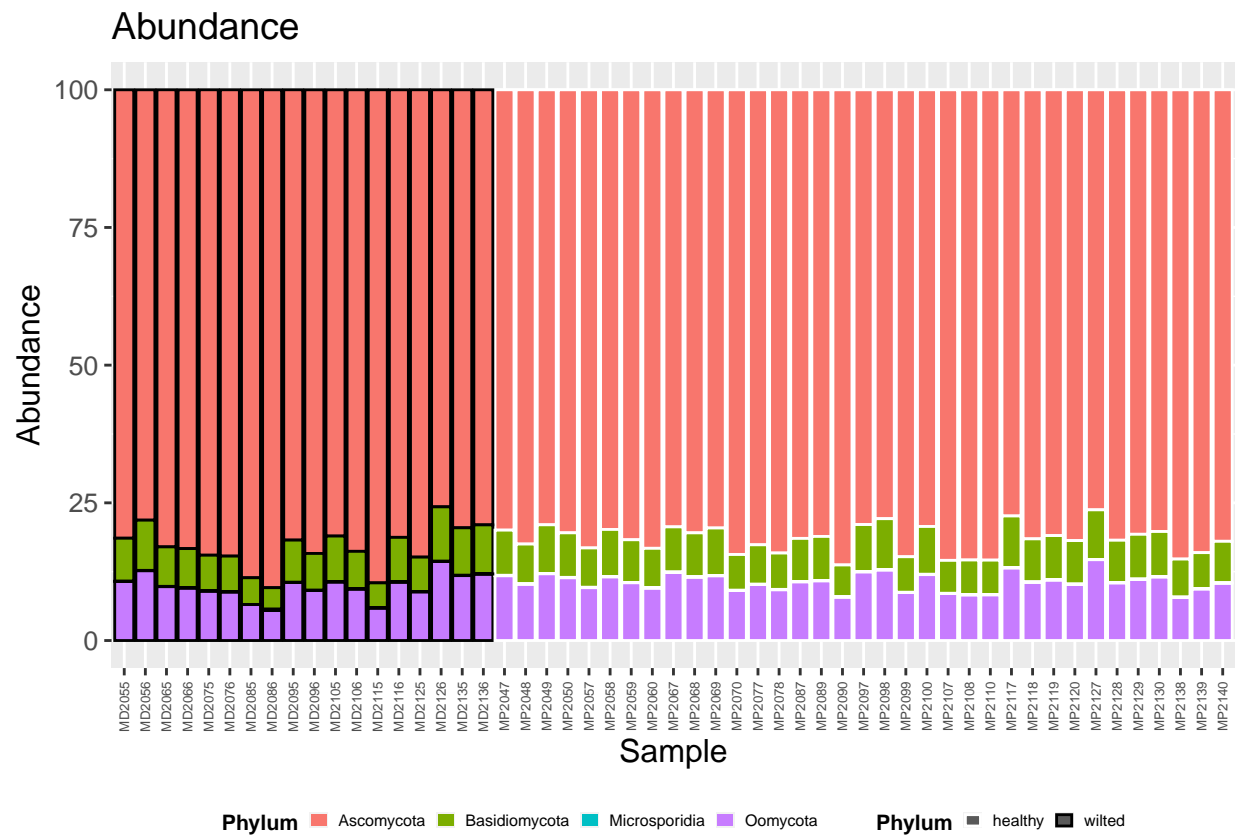
```
Barras_Phylum <- Abundance_barras(merge_Eukaryota,'Phylum' , 'Treatment', 10.0)
```

```
## Warning: 'aes_string()' was deprecated in ggplot2 3.0.0.
## i Please use tidy evaluation ideoms with 'aes()'

```

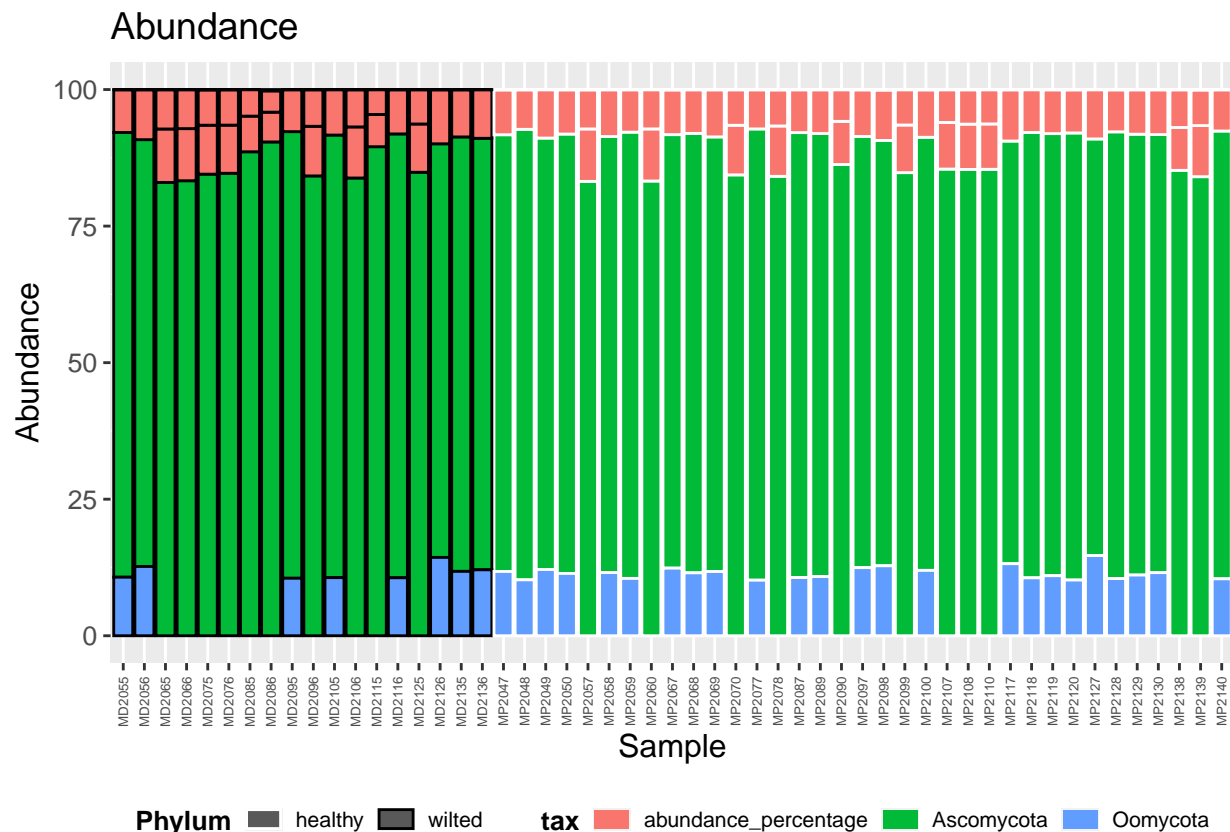
```
Barras_Phylum[1] # normal
```

```
## [[1]]
```



Barras_Phylum[2]

[[1]]



```
Beta_diversity(merge_Eukaryota , 'Phylum' , 'Treatment', 'bray')
```

```
## Square root transformation
## Wisconsin double standardization
## Run 0 stress 0.01148681
## Run 1 stress 9.641699e-05
## ... New best solution
## ... Procrustes: rmse 0.03460127 max resid 0.09064286
## Run 2 stress 9.692958e-05
## ... Procrustes: rmse 0.0001280624 max resid 0.0004094851
## ... Similar to previous best
## Run 3 stress 0.0009993135
## Run 4 stress 2.875625e-05
## ... New best solution
## ... Procrustes: rmse 0.0001075607 max resid 0.0002893054
## ... Similar to previous best
## Run 5 stress 7.419065e-05
## ... Procrustes: rmse 4.598373e-05 max resid 0.0001193756
## ... Similar to previous best
## Run 6 stress 7.803609e-05
## ... Procrustes: rmse 2.872485e-05 max resid 6.902742e-05
## ... Similar to previous best
## Run 7 stress 9.725895e-05
## ... Procrustes: rmse 0.0002168401 max resid 0.0005770855
## ... Similar to previous best
```

```

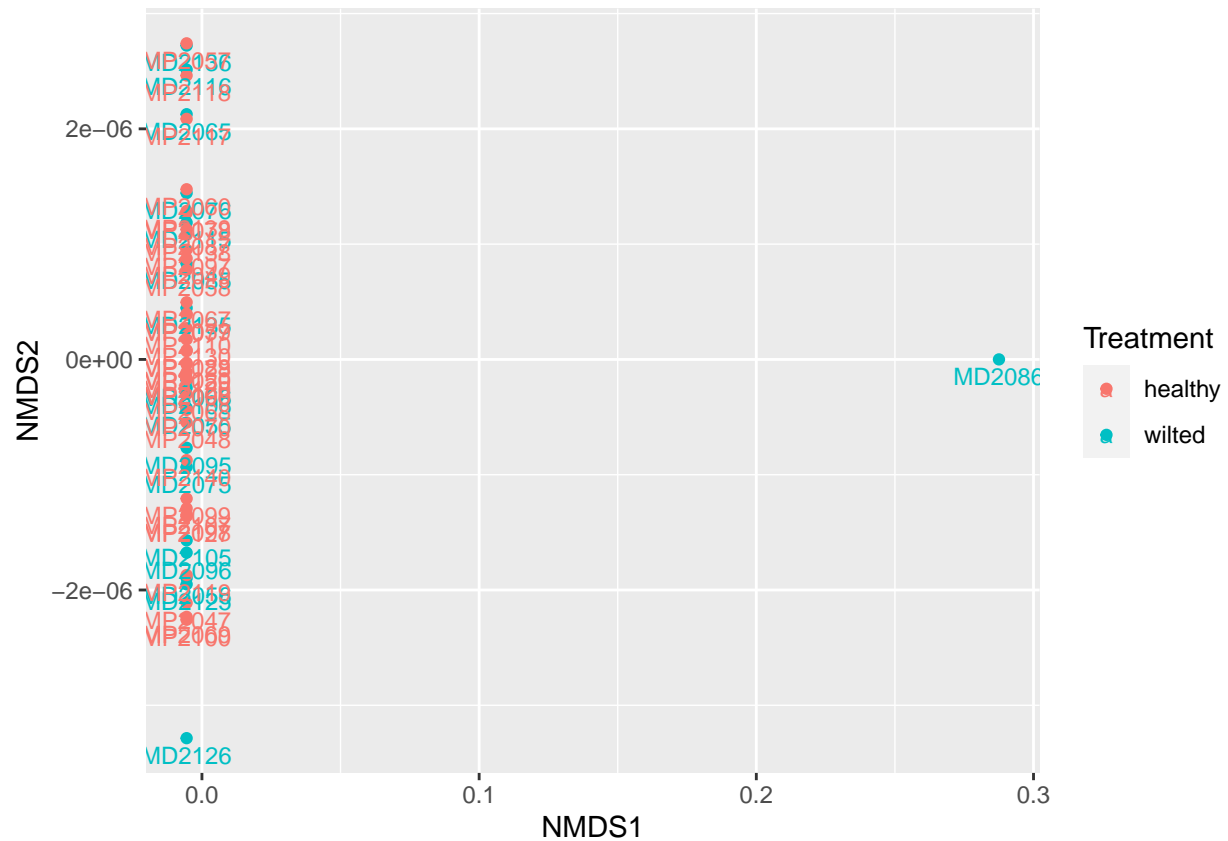
## Run 8 stress 8.900437e-05
## ... Procrustes: rmse 8.648394e-05  max resid 0.0002263783
## ... Similar to previous best
## Run 9 stress 9.712047e-05
## ... Procrustes: rmse 0.0001019901  max resid 0.0002717338
## ... Similar to previous best
## Run 10 stress 9.473721e-05
## ... Procrustes: rmse 0.0001079967  max resid 0.0002832011
## ... Similar to previous best
## Run 11 stress 9.169436e-05
## ... Procrustes: rmse 6.777917e-05  max resid 0.0001896028
## ... Similar to previous best
## Run 12 stress 0.0001125098
## ... Procrustes: rmse 0.000332578  max resid 0.0008723591
## ... Similar to previous best
## Run 13 stress 9.913374e-05
## ... Procrustes: rmse 0.000169905  max resid 0.0004530579
## ... Similar to previous best
## Run 14 stress 9.120442e-05
## ... Procrustes: rmse 2.554051e-05  max resid 0.0001256882
## ... Similar to previous best
## Run 15 stress 9.384635e-05
## ... Procrustes: rmse 7.33919e-05  max resid 0.0001795472
## ... Similar to previous best
## Run 16 stress 6.475988e-05
## ... Procrustes: rmse 3.209143e-05  max resid 7.189593e-05
## ... Similar to previous best
## Run 17 stress 0.0007519899
## Run 18 stress 0.0001067793
## ... Procrustes: rmse 0.0003143495  max resid 0.0008254772
## ... Similar to previous best
## Run 19 stress 9.906154e-05
## ... Procrustes: rmse 0.0001095915  max resid 0.0002853942
## ... Similar to previous best
## Run 20 stress 9.462542e-05
## ... Procrustes: rmse 0.0001393057  max resid 0.0003633612
## ... Similar to previous best
## *** Best solution repeated 16 times

```

```

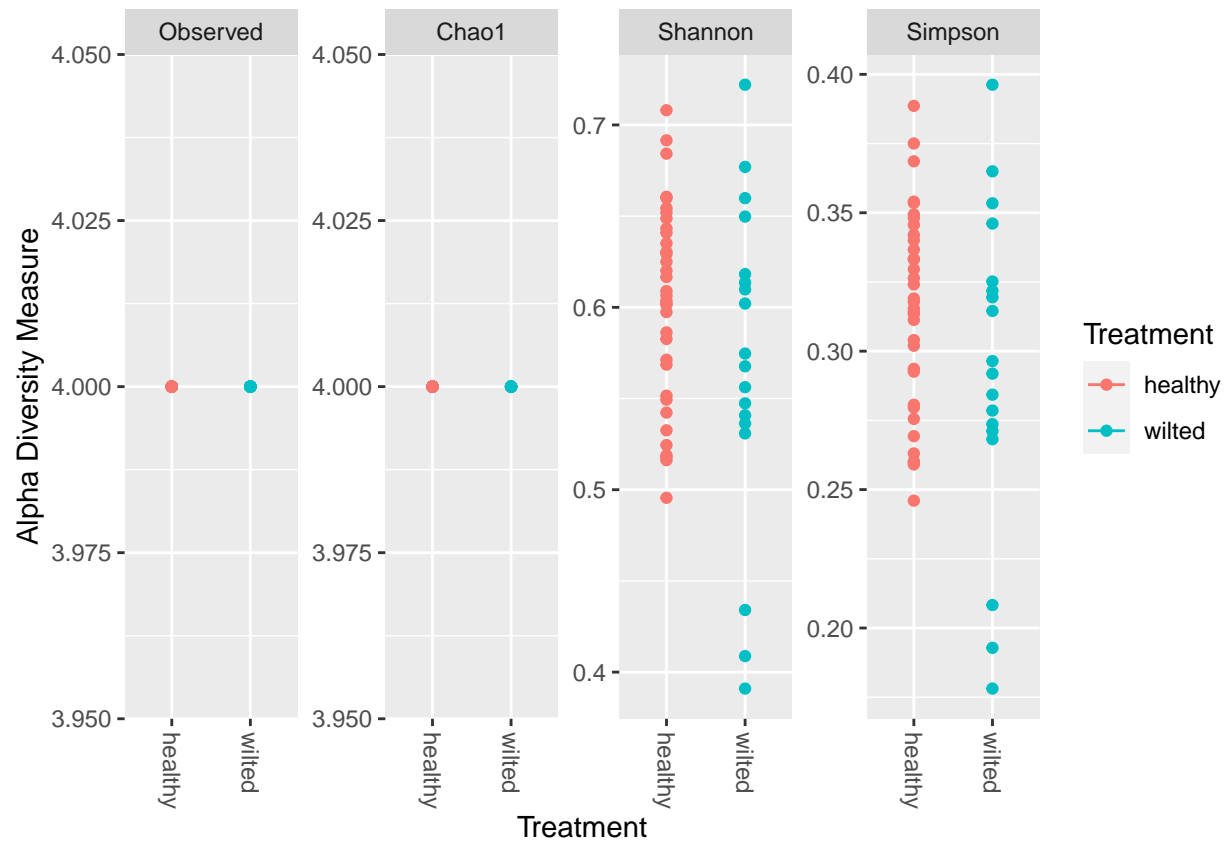
## Warning in metaMDS(veganifyOTU(physeq), distance, ...): stress is (nearly) zero:
## you may have insufficient data

```



```
Alpha_diversity(merge_Eukaryota , 'Phylum' , 'Treatment')
```

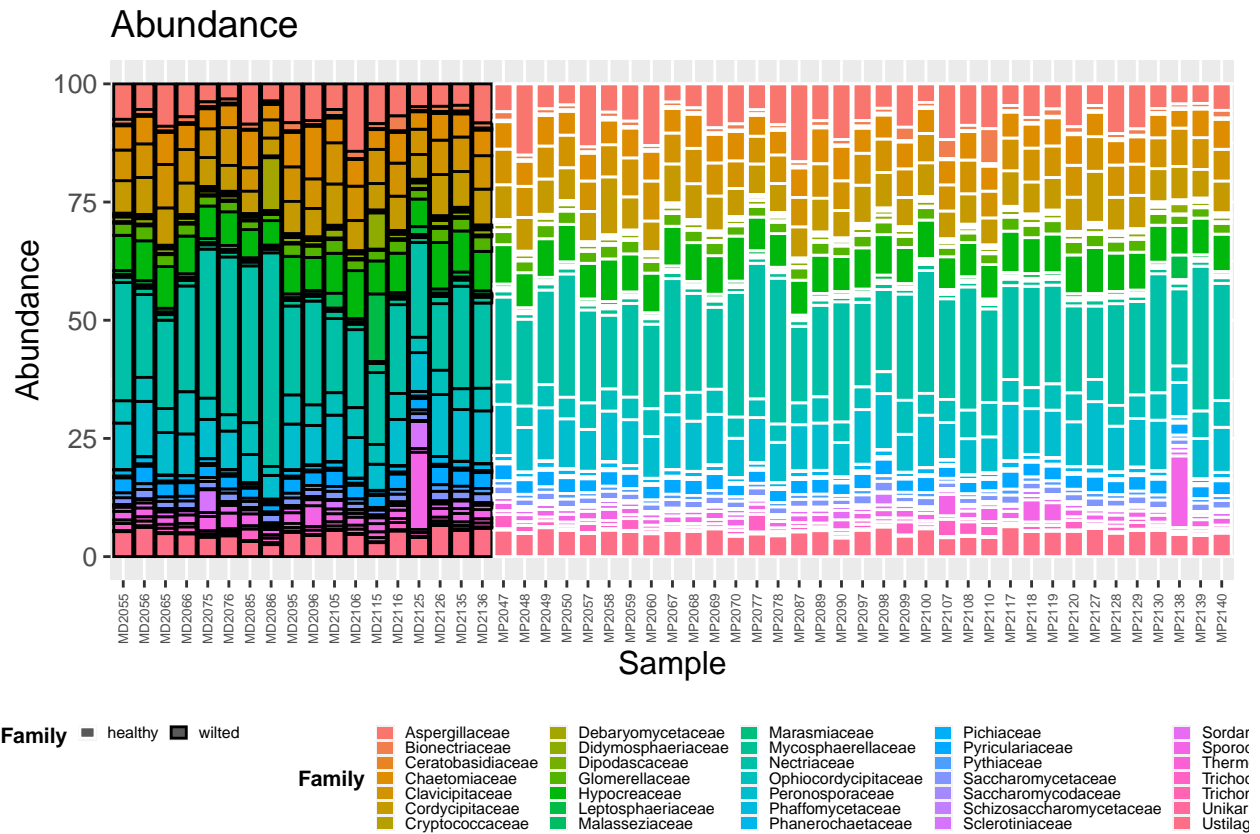
```
## Warning in estimate_richness(physeq, split = TRUE, measures = measures): The data you have provided
## any singletons. This is highly suspicious. Results of richness
## estimates (for example) are probably unreliable, or wrong, if you have already
## trimmed low-abundance taxa from the data.
##
## We recommended that you find the un-trimmed data and retry.
```



#—— Eukarya by Family

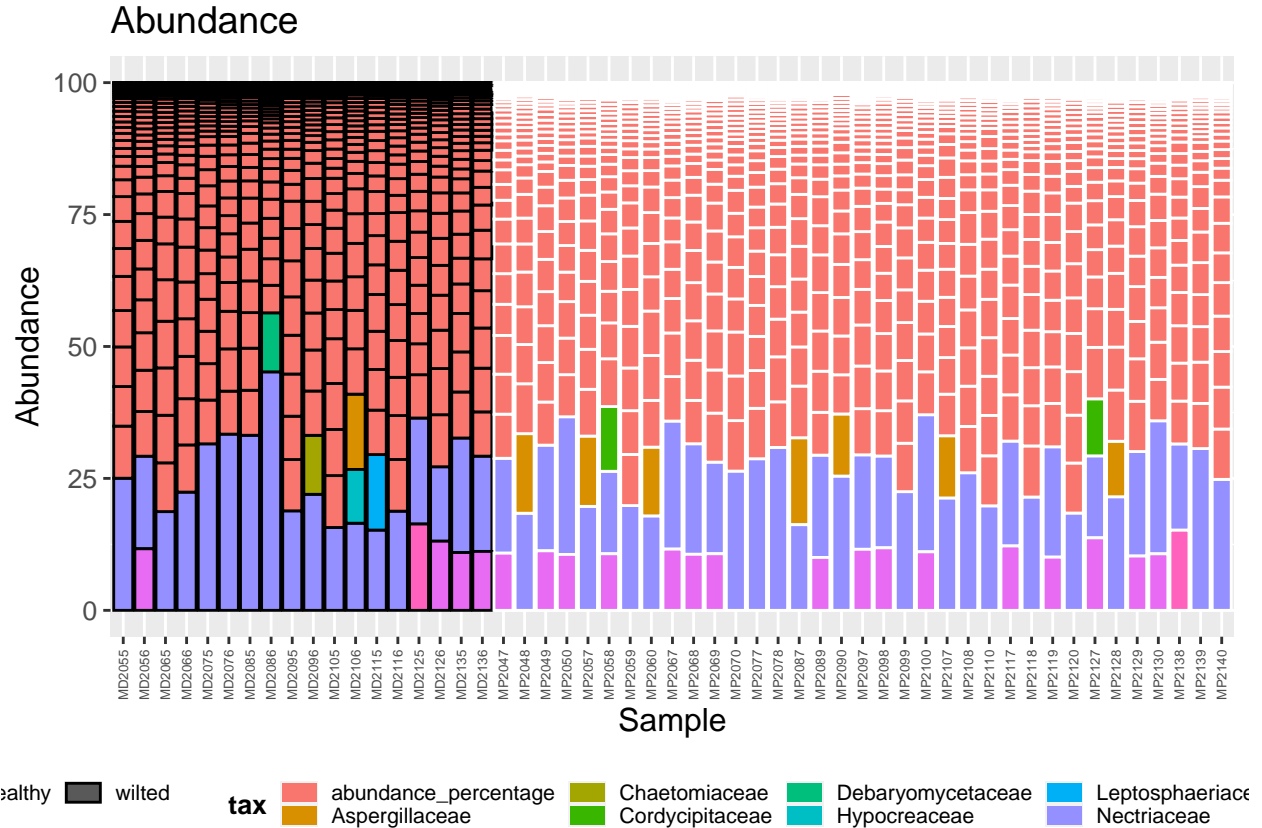
```
Barras_Species <- Abundance_barras(merge_Eukaryota, 'Family', 'Treatment', 10.0)
Barras_Species[1]
```

```
## [[1]]
```

Barras_Species [2]

[[1]]



```
Beta_diversity(merge_Eukaryota , 'Family' , 'Treatment' , 'bray')
```

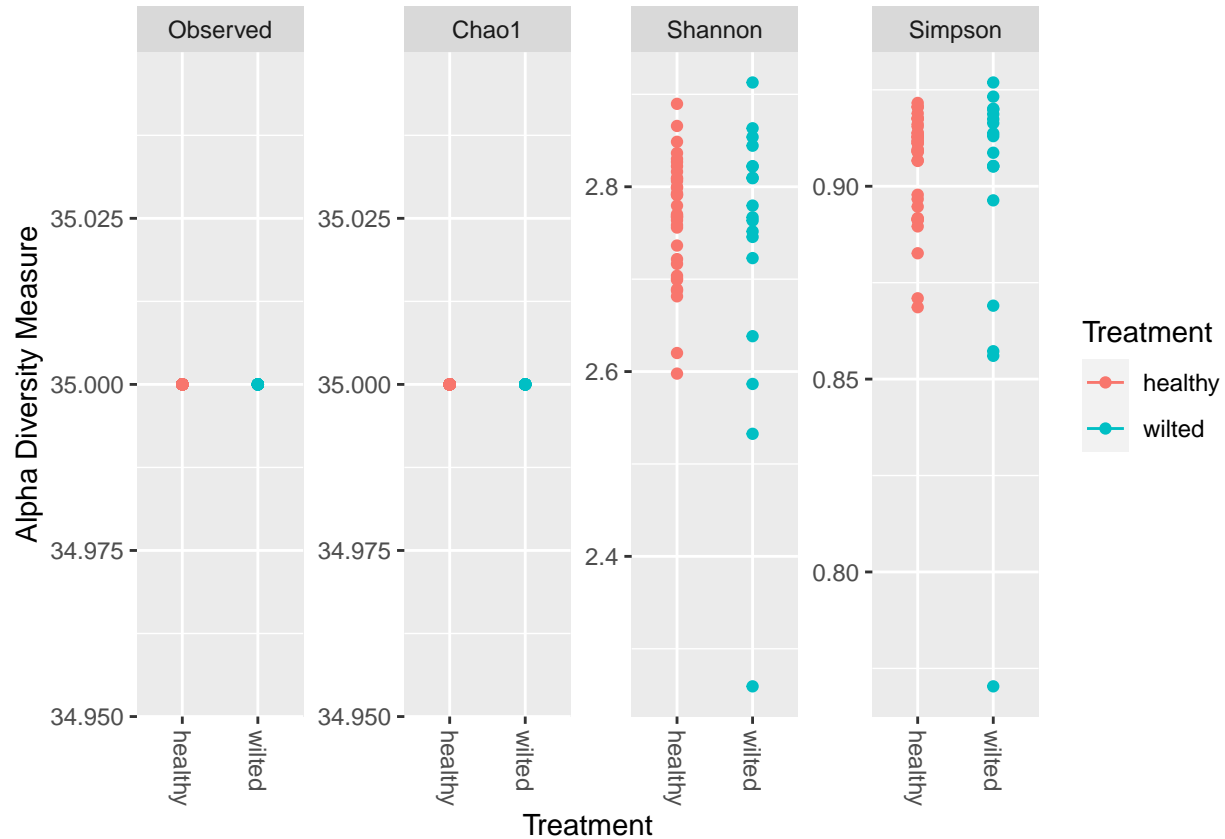
```
## Wisconsin double standardization
## Run 0 stress 0.1089081
## Run 1 stress 0.1088964
## ... New best solution
## ... Procrustes: rmse 0.002397465  max resid 0.009337499
## ... Similar to previous best
## Run 2 stress 0.1089479
## ... Procrustes: rmse 0.008318336  max resid 0.0543661
## Run 3 stress 0.1089298
## ... Procrustes: rmse 0.008610539  max resid 0.05495783
## Run 4 stress 0.1088985
## ... Procrustes: rmse 0.000636777  max resid 0.00368301
## ... Similar to previous best
## Run 5 stress 0.1352082
## Run 6 stress 0.109113
## ... Procrustes: rmse 0.01003345  max resid 0.05422836
## Run 7 stress 0.1088882
## ... New best solution
## ... Procrustes: rmse 0.00173353  max resid 0.009003435
## ... Similar to previous best
## Run 8 stress 0.1089152
## ... Procrustes: rmse 0.001735834  max resid 0.009378948
## ... Similar to previous best
```

```
## Run 9 stress 0.1185617
## Run 10 stress 0.1088982
## ... Procrustes: rmse 0.00155193 max resid 0.007565839
## ... Similar to previous best
## Run 11 stress 0.1089574
## ... Procrustes: rmse 0.00861713 max resid 0.05374258
## Run 12 stress 0.1089481
## ... Procrustes: rmse 0.008203471 max resid 0.05409664
## Run 13 stress 0.4042251
## Run 14 stress 0.1089316
## ... Procrustes: rmse 0.008488576 max resid 0.05456272
## Run 15 stress 0.1089084
## ... Procrustes: rmse 0.001691858 max resid 0.009408485
## ... Similar to previous best
## Run 16 stress 0.1113091
## Run 17 stress 0.1091144
## ... Procrustes: rmse 0.0100354 max resid 0.05392358
## Run 18 stress 0.1091131
## ... Procrustes: rmse 0.009953453 max resid 0.05403308
## Run 19 stress 0.1088877
## ... New best solution
## ... Procrustes: rmse 0.0001671813 max resid 0.0009656288
## ... Similar to previous best
## Run 20 stress 0.109126
## ... Procrustes: rmse 0.01001698 max resid 0.05420395
## *** Best solution repeated 1 times
```



```
Alpha_diversity(merge_Eukaryota , 'Family' , 'Treatment')
```

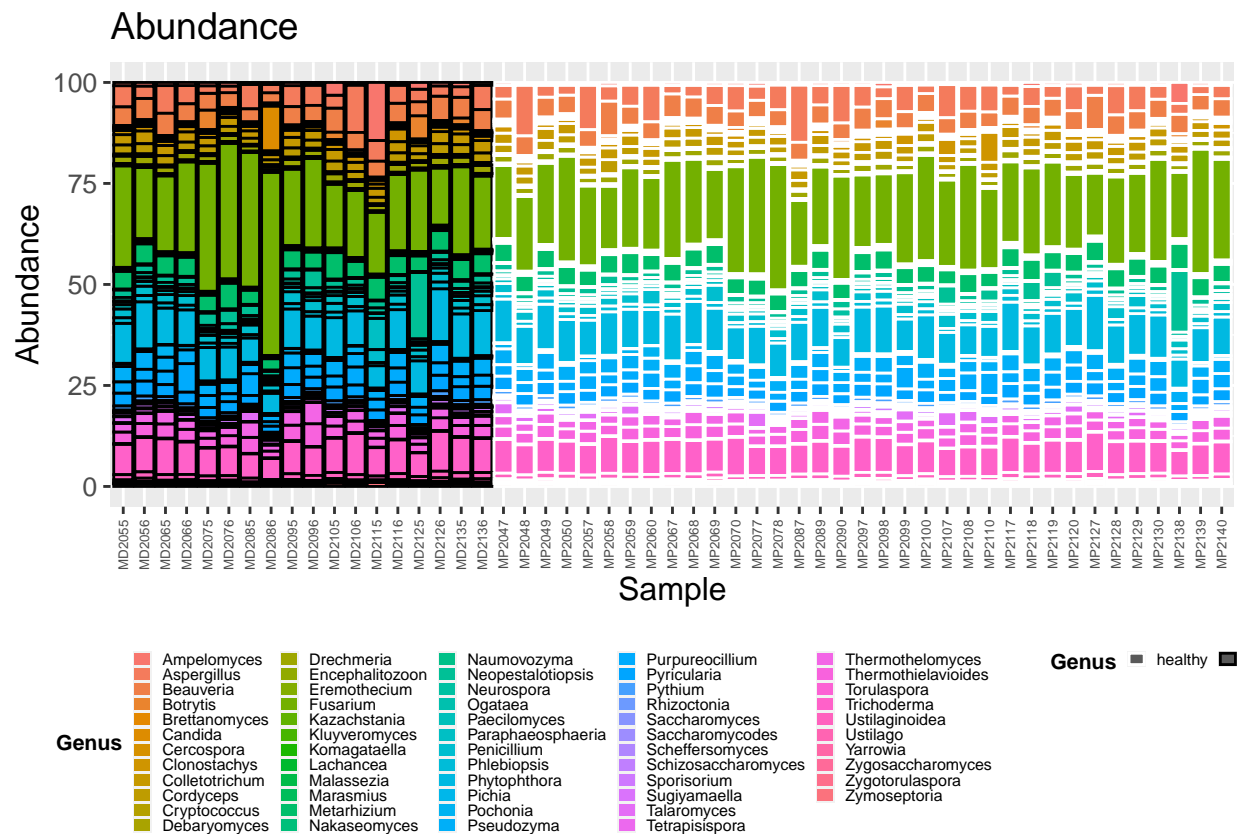
```
## Warning in estimate_richness(physeq, split = TRUE, measures = measures): The data you have provided
## any singletons. This is highly suspicious. Results of richness
## estimates (for example) are probably unreliable, or wrong, if you have already
## trimmed low-abundance taxa from the data.
##
## We recommended that you find the un-trimmed data and retry.
```



```
#———Eukarya by Genero
```

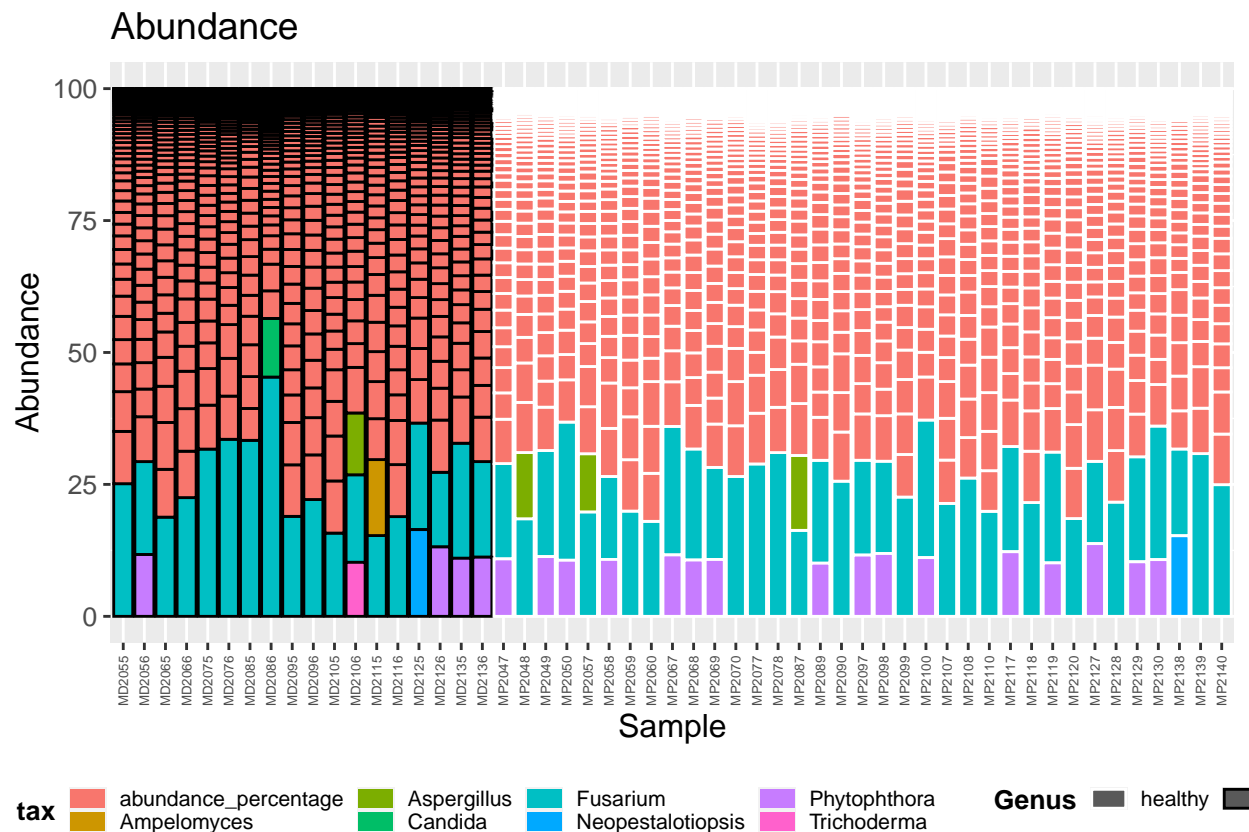
```
Barras_Species <- Abundance_barras(merge_Eukaryota, 'Genus', 'Treatment', 10.0)
Barras_Species[1]
```

```
## [[1]]
```



Barras_Species [2]

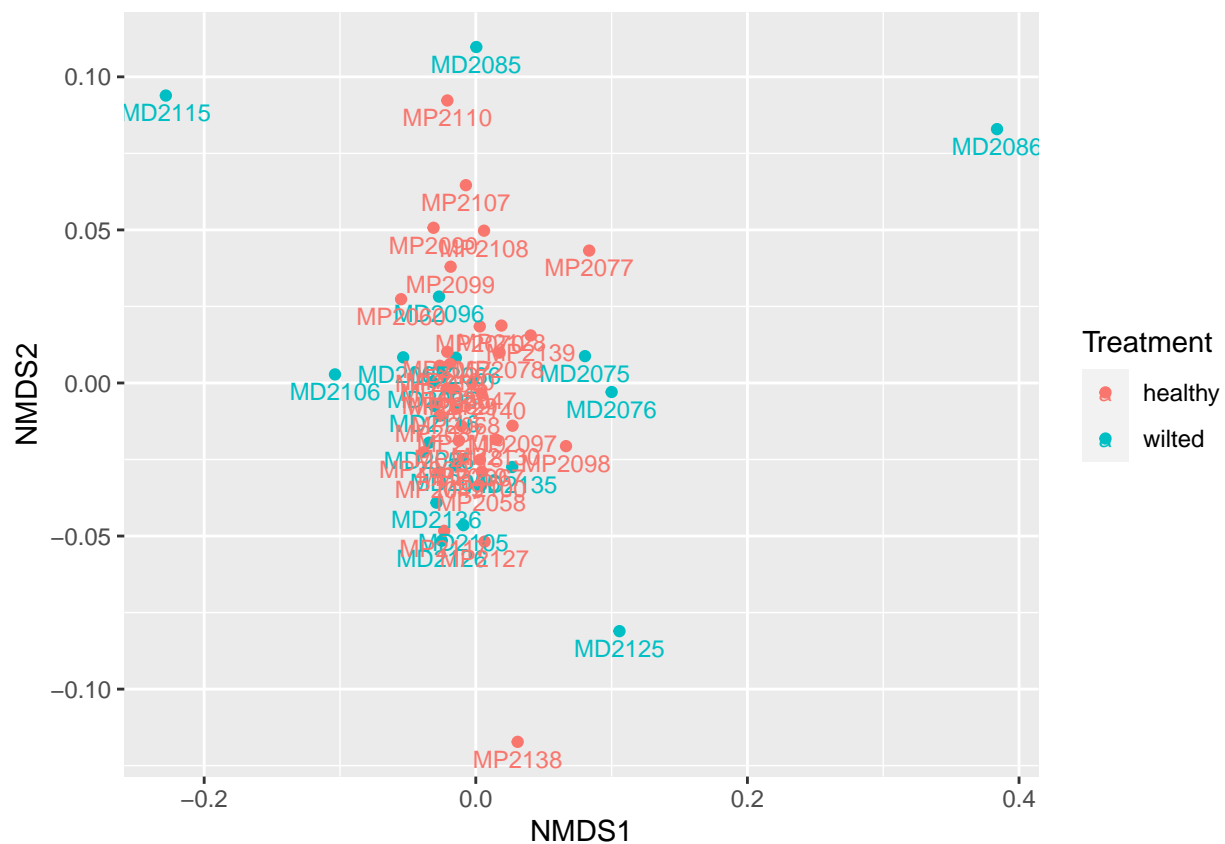
[[1]]



```
Beta_diversity(merge_Eukaryota , 'Genus' , 'Treatment', 'bray')
```

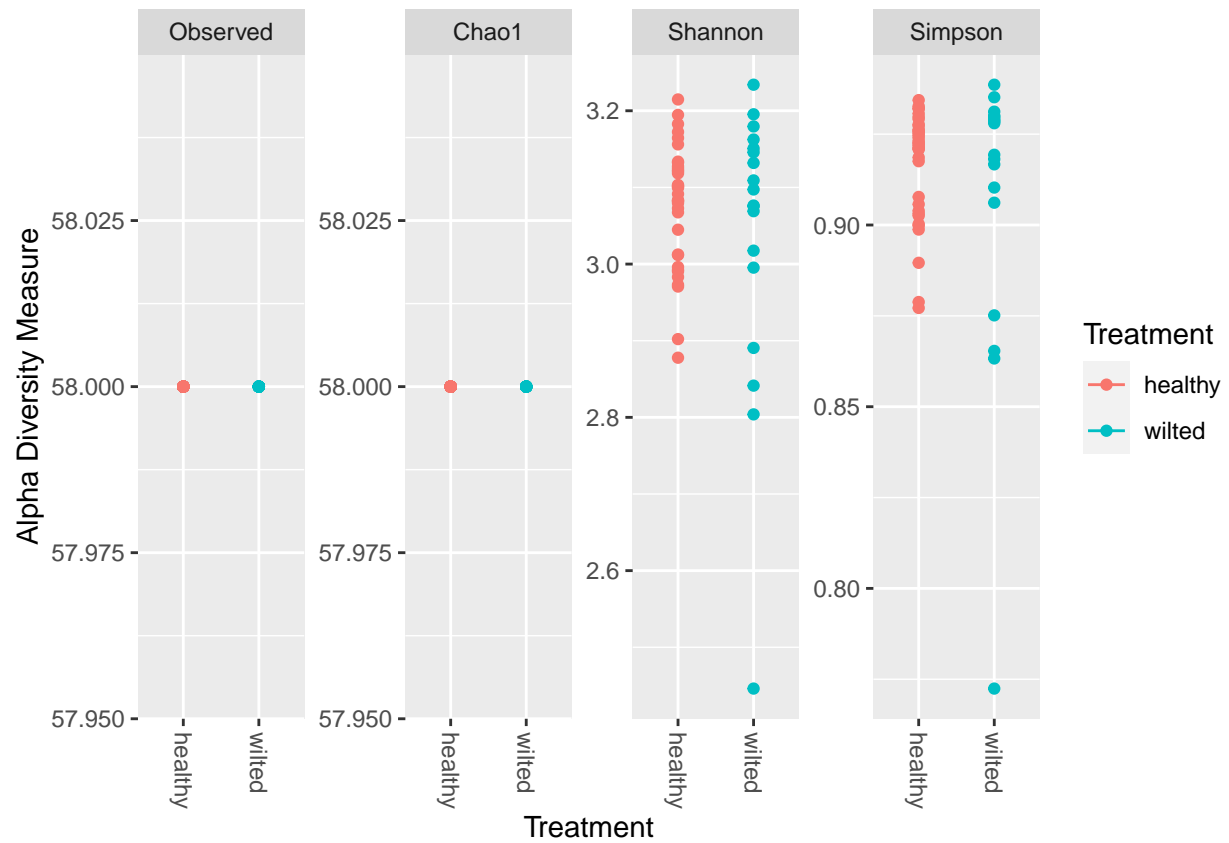
```
## Wisconsin double standardization
## Run 0 stress 0.110709
## Run 1 stress 0.1111514
## ... Procrustes: rmse 0.01374689 max resid 0.06906551
## Run 2 stress 0.1235012
## Run 3 stress 0.1119814
## Run 4 stress 0.1107094
## ... Procrustes: rmse 0.001118204 max resid 0.006393413
## ... Similar to previous best
## Run 5 stress 0.123502
## Run 6 stress 0.1235014
## Run 7 stress 0.1223845
## Run 8 stress 0.1107098
## ... Procrustes: rmse 0.0002377637 max resid 0.001329702
## ... Similar to previous best
## Run 9 stress 0.1236307
## Run 10 stress 0.1236309
## Run 11 stress 0.110709
## ... New best solution
## ... Procrustes: rmse 1.991006e-05 max resid 0.0001234856
## ... Similar to previous best
## Run 12 stress 0.1235025
## Run 13 stress 0.1236304
```

```
## Run 14 stress 0.1107086
## ... New best solution
## ... Procrustes: rmse 0.000252225  max resid 0.001539016
## ... Similar to previous best
## Run 15 stress 0.1107097
## ... Procrustes: rmse 0.0009435275  max resid 0.005267492
## ... Similar to previous best
## Run 16 stress 0.1223877
## Run 17 stress 0.1107172
## ... Procrustes: rmse 0.002093797  max resid 0.01195467
## Run 18 stress 0.1235015
## Run 19 stress 0.1223843
## Run 20 stress 0.1107093
## ... Procrustes: rmse 0.000821115  max resid 0.00457383
## ... Similar to previous best
## *** Best solution repeated 3 times
```



```
Alpha_diversity(merge_Eukaryota , 'Genus' , 'Treatment')
```

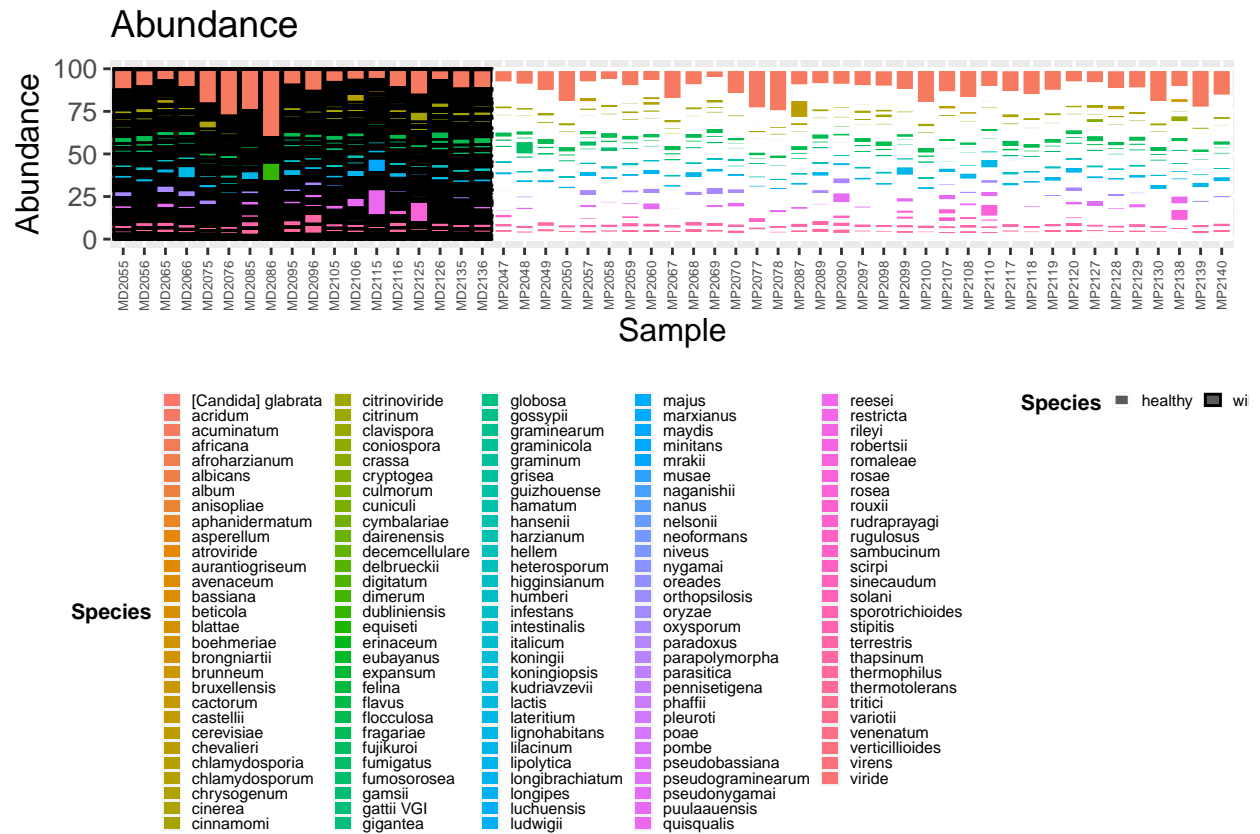
```
## Warning in estimate_richness(physeq, split = TRUE, measures = measures): The data you have provided contains
## any singletons. This is highly suspicious. Results of richness
## estimates (for example) are probably unreliable, or wrong, if you have already
## trimmed low-abundance taxa from the data.
##
## We recommended that you find the un-trimmed data and retry.
```



#———Eukarya by Species

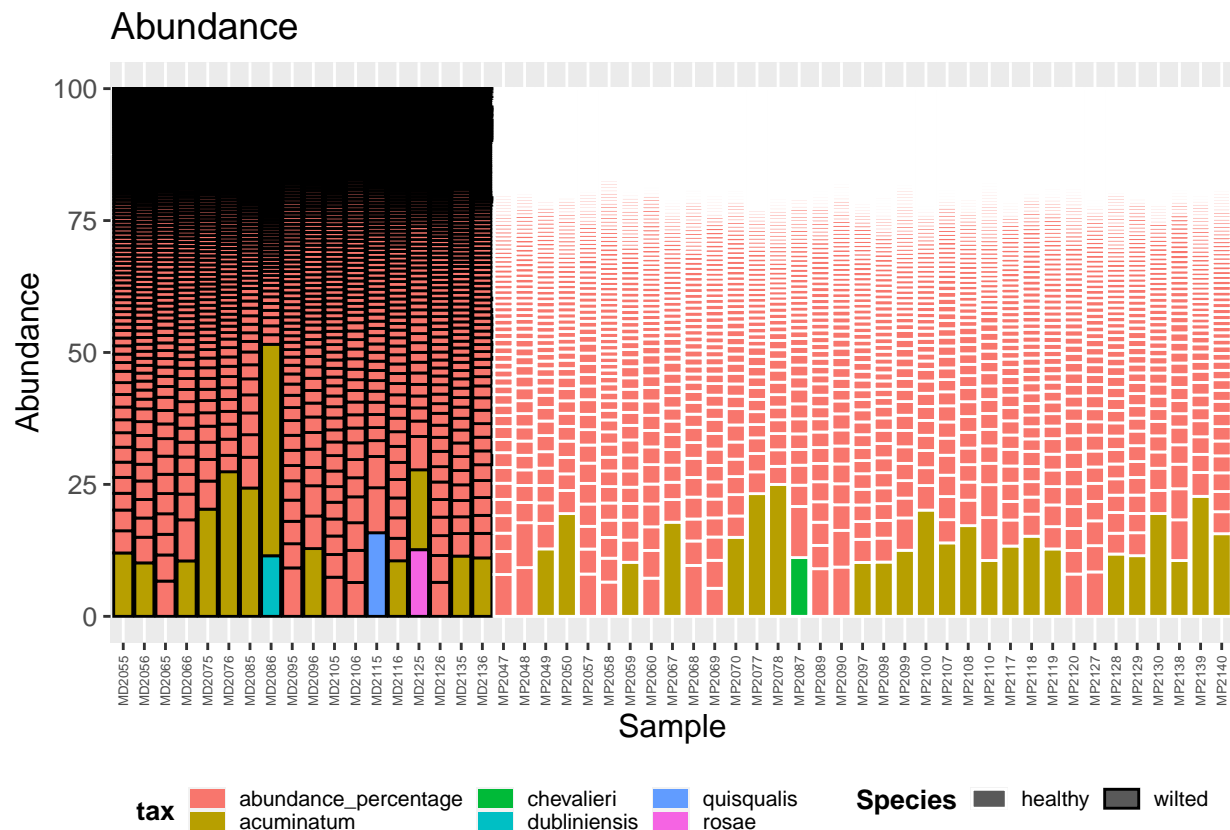
```
Barras_Species <- Abundance_barras(merge_Eukaryota, 'Species', 'Treatment', 10.0)
Barras_Species[1]
```

```
## [[1]]
```

Barras_Species [2]

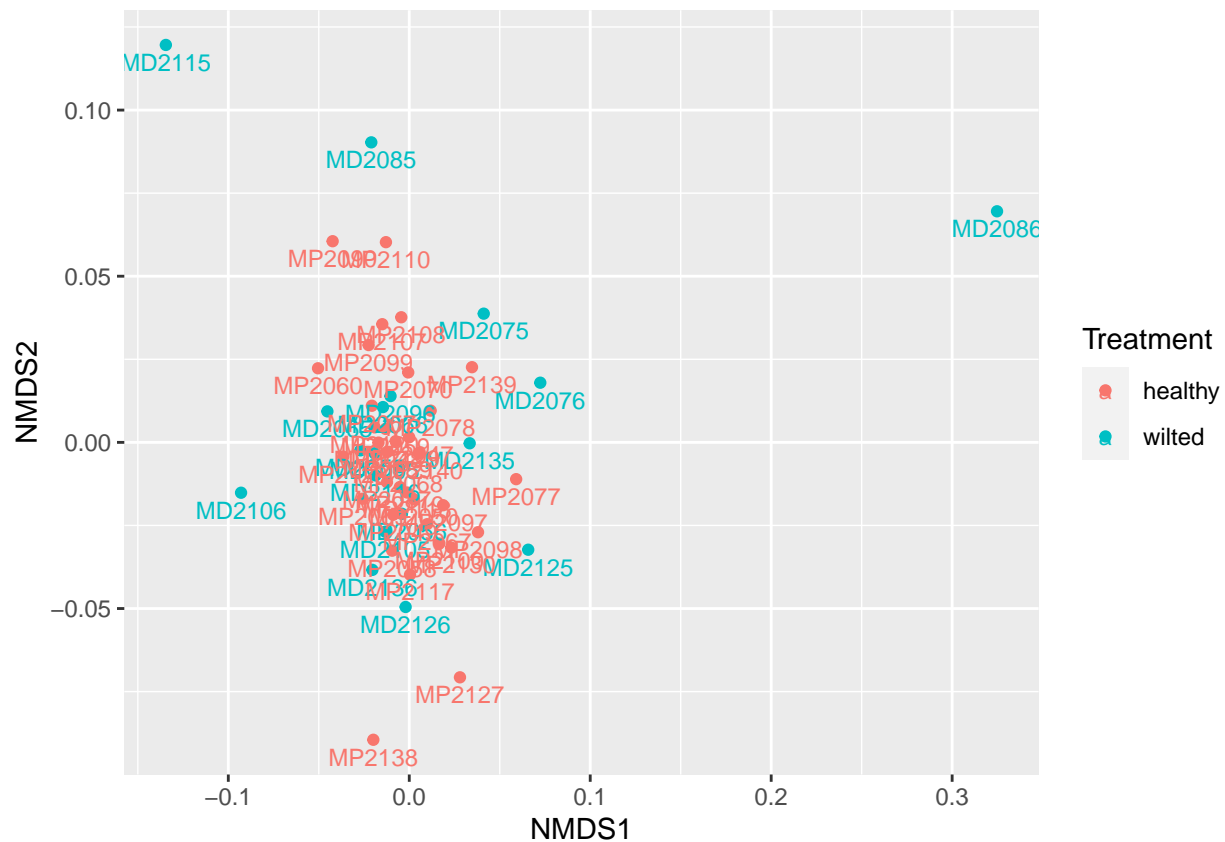
[[1]]

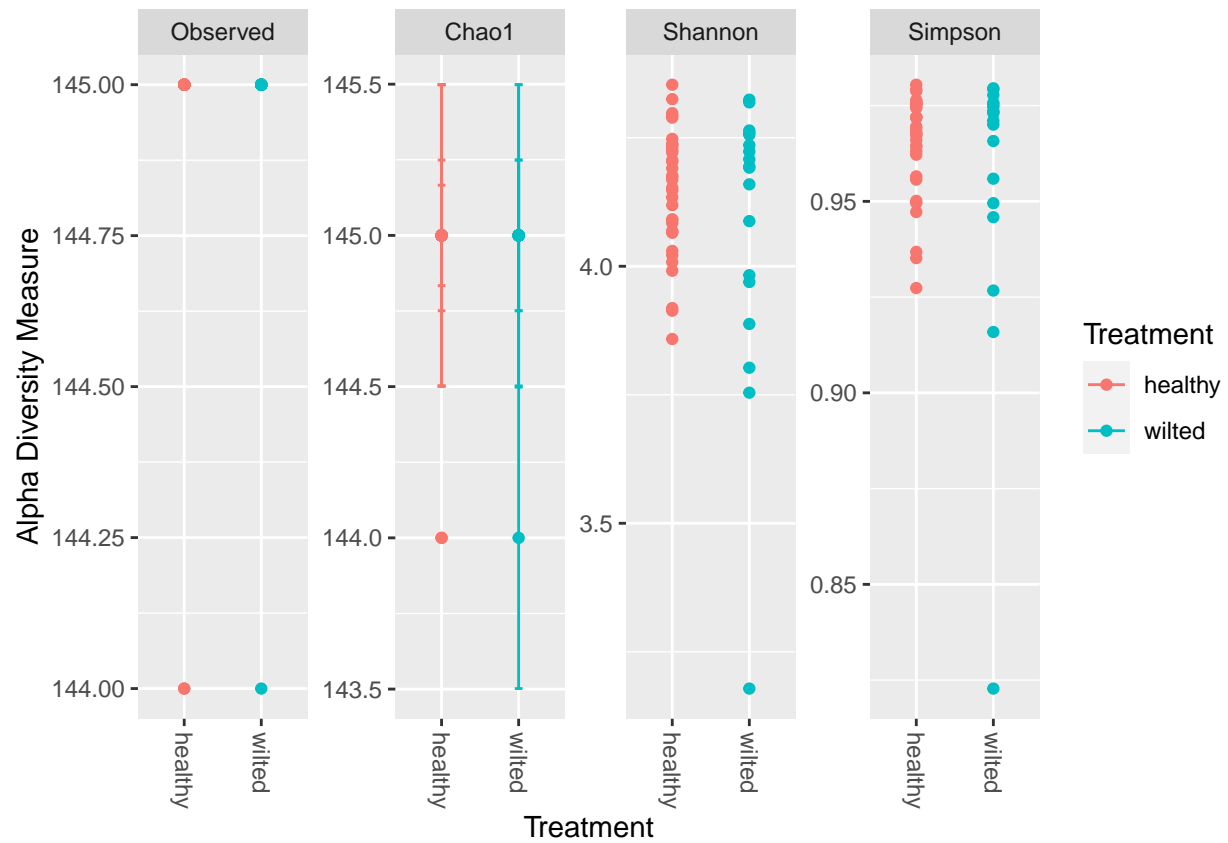


```
Beta_diversity(merge_Eukaryota , 'Species' , 'Treatment', 'bray')
```

```
## Wisconsin double standardization
## Run 0 stress 0.1226512
## Run 1 stress 0.1226515
## ... Procrustes: rmse 0.0009565164 max resid 0.004818758
## ... Similar to previous best
## Run 2 stress 0.1233302
## Run 3 stress 0.1218539
## ... New best solution
## ... Procrustes: rmse 0.02026332 max resid 0.1282693
## Run 4 stress 0.1278289
## Run 5 stress 0.1218549
## ... Procrustes: rmse 0.0006613939 max resid 0.003265407
## ... Similar to previous best
## Run 6 stress 0.1227526
## Run 7 stress 0.1226511
## Run 8 stress 0.1218544
## ... Procrustes: rmse 0.00050139 max resid 0.002464929
## ... Similar to previous best
## Run 9 stress 0.1218543
## ... Procrustes: rmse 0.0002429245 max resid 0.001192044
## ... Similar to previous best
## Run 10 stress 0.1233307
## Run 11 stress 0.1272847
```

```
Alpha_diversity(merge_Eukaryota , 'Species' , 'Treatment')
```

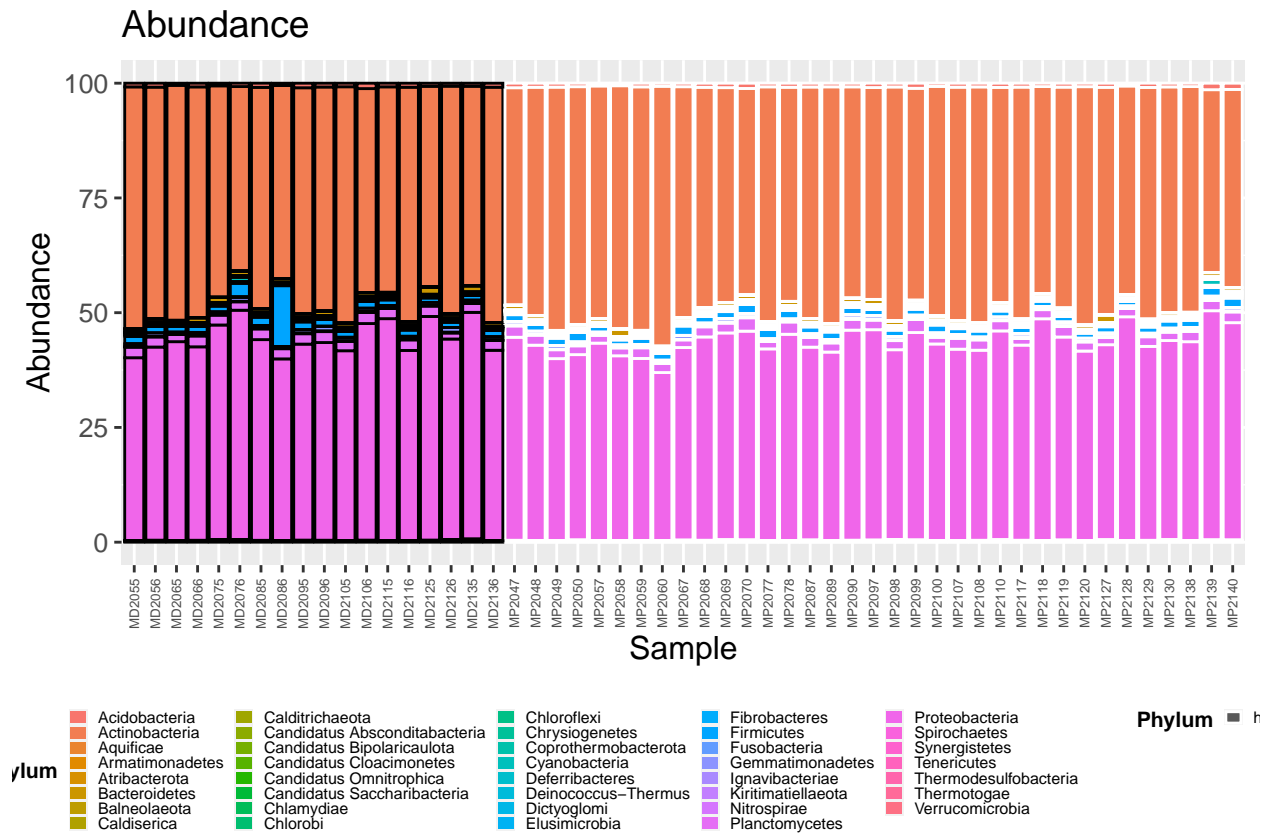




#——Bacteria by Phylum

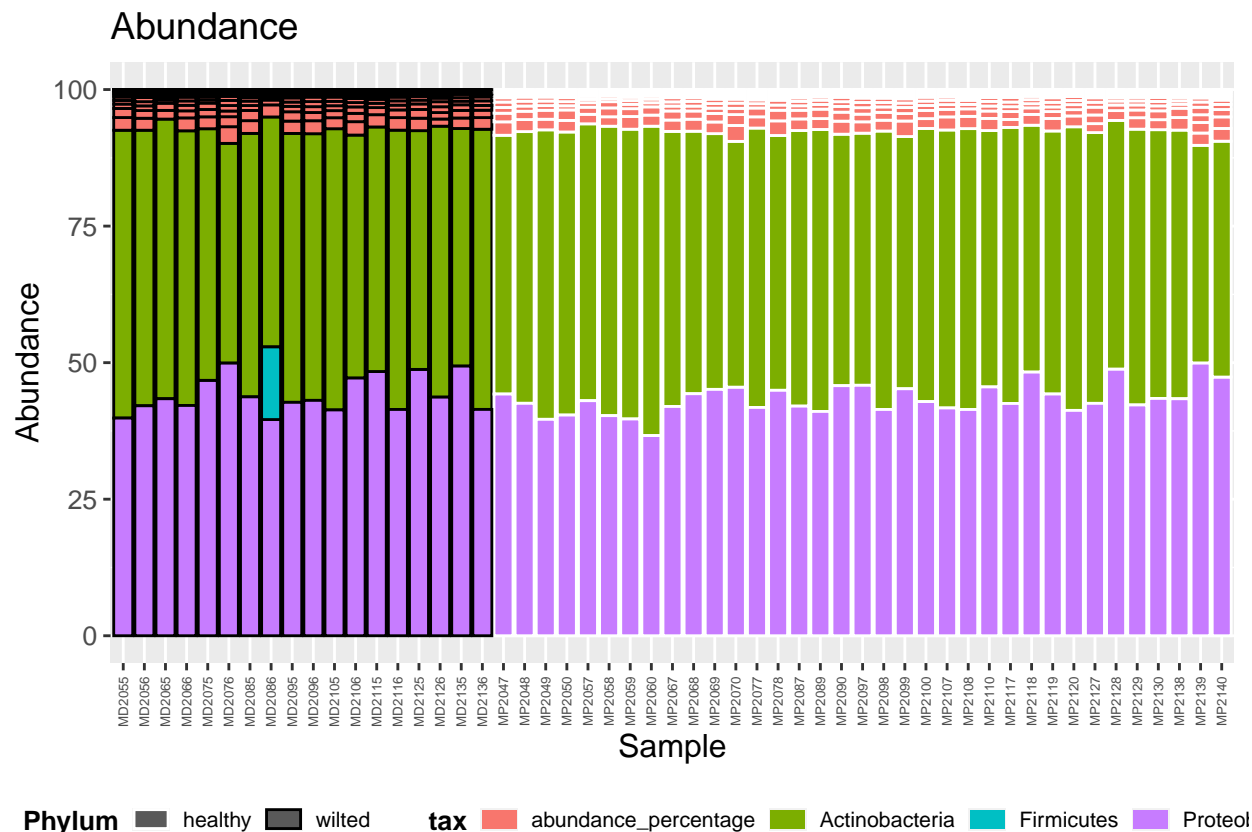
```
Barras_Species <- Abundance_barras(merge_Bacteria, 'Phylum', 'Treatment', 10.0)
Barras_Species[1]
```

```
## [[1]]
```



Barras_Species[2]

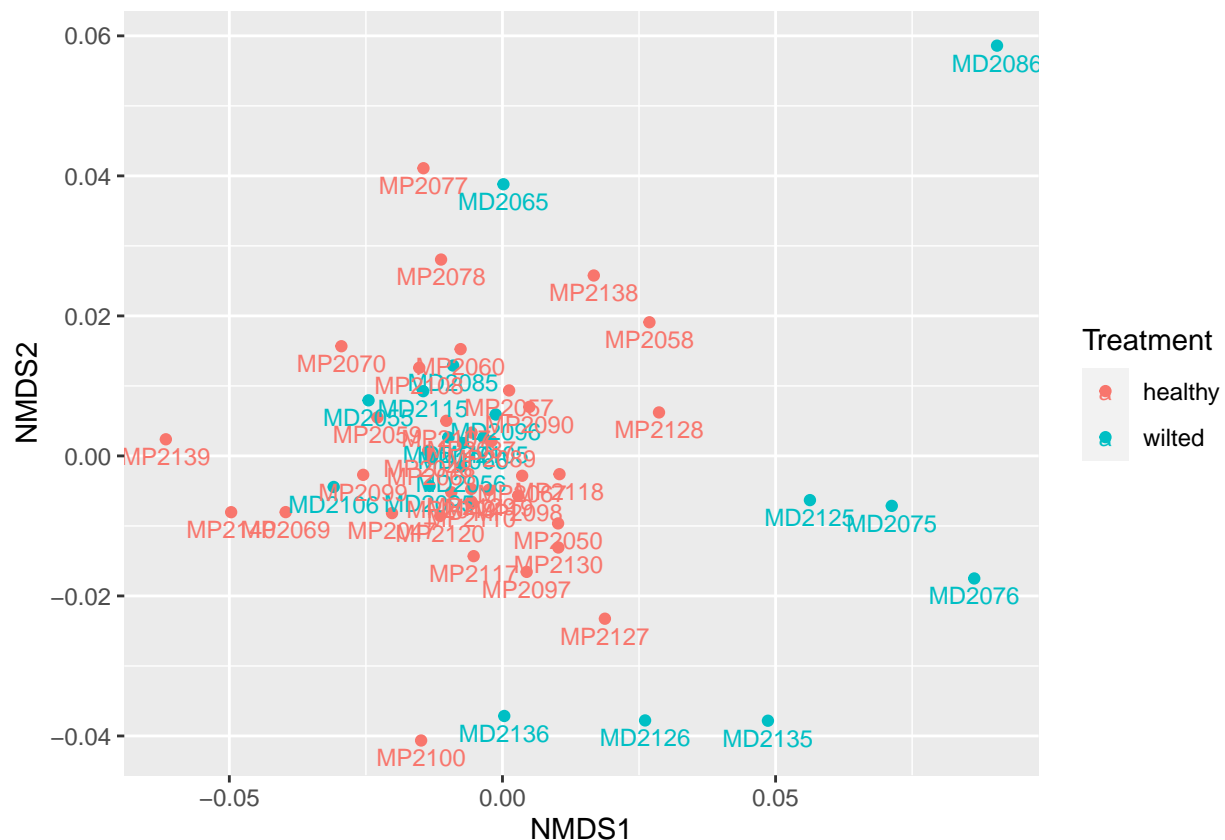
[[1]]



```
Beta_diversity(merge_Bacteria , 'Phylum' , 'Treatment', 'bray')
```

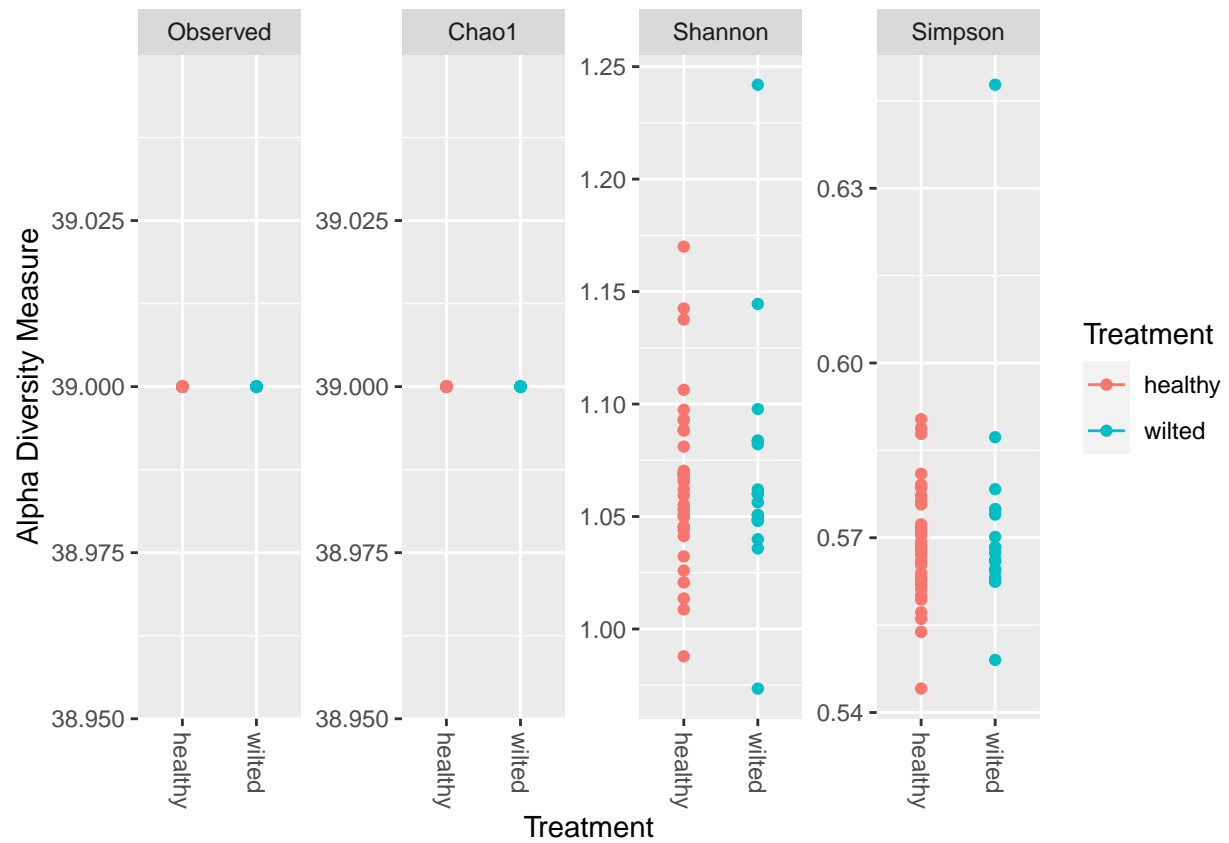
```
## Square root transformation
## Wisconsin double standardization
## Run 0 stress 0.1520004
## Run 1 stress 0.1636958
## Run 2 stress 0.1700011
## Run 3 stress 0.1519724
## ... New best solution
## ... Procrustes: rmse 0.008285382 max resid 0.04681388
## Run 4 stress 0.1552698
## Run 5 stress 0.1519014
## ... New best solution
## ... Procrustes: rmse 0.04885038 max resid 0.3152661
## Run 6 stress 0.1614796
## Run 7 stress 0.1513921
## ... New best solution
## ... Procrustes: rmse 0.04582075 max resid 0.3024696
## Run 8 stress 0.1779282
## Run 9 stress 0.1548791
## Run 10 stress 0.1629442
## Run 11 stress 0.16964
## Run 12 stress 0.1524009
## Run 13 stress 0.1662298
## Run 14 stress 0.1613925
```

```
## Run 15 stress 0.1652755
## Run 16 stress 0.1633312
## Run 17 stress 0.1607191
## Run 18 stress 0.1672632
## Run 19 stress 0.1513905
## ... New best solution
## ... Procrustes: rmse 0.0004428603 max resid 0.002388818
## ... Similar to previous best
## Run 20 stress 0.1513906
## ... Procrustes: rmse 3.760188e-05 max resid 0.0002150336
## ... Similar to previous best
## *** Best solution repeated 2 times
```



```
Alpha_diversity(merge_Bacteria , 'Phylum' , 'Treatment')
```

```
## Warning in estimate_richness(physeq, split = TRUE, measures = measures): The data you have provided contains
## any singletons. This is highly suspicious. Results of richness
## estimates (for example) are probably unreliable, or wrong, if you have already
## trimmed low-abundance taxa from the data.
##
## We recommended that you find the un-trimmed data and retry.
```



#——Bacteria by Familia

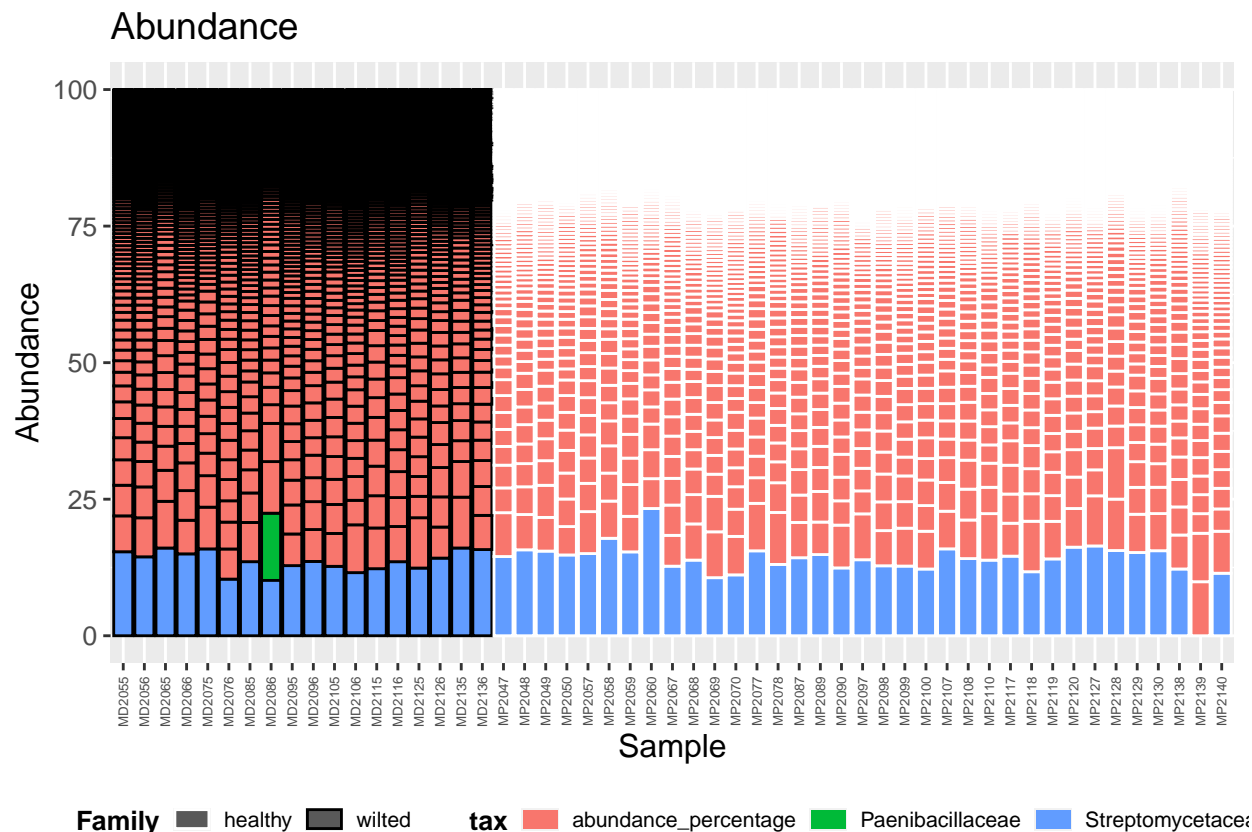
```
Barras_Species <- Abundance_barras(merge_Bacteria, 'Family', 'Treatment', 10.0)
Barras_Species[1]
```

```
## [[1]]
```


Actinomycesaceae	Chloromonadaceae	Halobacterioviraceae	Nocardiaceae	St
Actinopolymorphaceae	Clostridiaceae	Halobacteroidaceae	Nocardioidaceae	St
Actinopolysporaceae	Clostridiales Family XVI. Incertae Sedis	Halomonadaceae	Nocardiopsaceae	St
Aerococcaceae	Clostridiales Family XVII. Incertae Sedis	Halothiobacillaceae	Nostocaceae	St
Aeromonadaceae	Cohaesibacteraceae	Hapalosiphonaceae	Oceanospirillaceae	St
Akkermansiaceae	Coleofasciculaceae	Helicobacteraceae	Oculatellaceae	St
Alcaligenaceae	Colwelliaceae	Heliothecaceae	Odoribacteraceae	St
Alcanivoracaceae	Comamonadaceae	Holospiraceae	Oleiphilaceae	St
Alicyclobacillaceae	Conexibacteraceae	Hydrogenimonaceae	Opiritaceae	St
Alteromonadaceae	Coprothermobacteraceae	Hydrogenophilaceae	Orbaceae	St
Amoebophilaceae	Coriobacteriaceae	Hydrogenothermaceae	Ornithinimicrobiaceae	Su
Amorphaceae	Corynebacteriaceae	Hyellaceae	Oscillatoriaceae	Su
Anaerohalospiraaceae	Coxiellaceae	Hymenobacteraceae	Oscillospiraceae	Su
Anaerolineaceae	Crocinitomicaceae	Hyphomicrobiaceae	Oxalobacteraceae	Su
Anaeromyxobacteraceae	Cyclobacteriaceae	Hyphomonadaceae	Paenibacillaceae	Su
Anaplasmataceae	Cytophagaceae	Iamiaceae	Paludibacteraceae	Sy
Aphanizomenonaceae	Deferribacteraceae	Ichthyobacteriaceae	Parachiamydiaceae	Sy
Aphanothecaceae	Dehalococcoidaceae	Idiomarinaceae	Parvibaculaceae	Sy
Aquificaceae	Deinococcaceae	Ignavibacteriaceae	Parvularculaceae	Sy
Archangiaceae	Demeguinaceae	Ilumatobacteraceae	Pasteurellaceae	Sy
Arcobacteraceae	Dermabacteraceae	Immundisolibacteraceae	Pectobacteriaceae	Sy
Ardenticatenaceae	Dermacoccaceae	Intrasporangiaceae	Pelagibacteraceae	Sy
Atopobiaceae	Dermatophilaceae	Isosphaeraceae	Peptococcaceae	Ta
Atribacteraceae	Dermocarpellaceae	Jatrophihabitantaceae	Peptoniphilaceae	Te
Aurantimonadaceae	Desulfallaceae	Jiangellaceae	Peptostreptococcaceae	Te
Azonexaceae	Desulfarculaceae	Jonesiaceae	Petrotoxiaceae	Te
Azospirillaceae	Desulfatobacteriaceae	Kaistiaceae	Phototrophicaceae	Te
Bacillaceae	Desulfobacteriaceae	Kangiellaceae	Phreatobacteraceae	Te
Bacterioviraceae	Desulfobulbaceae	Kineosporiaceae	Phycisphaeraceae	Tr
Bacteroidaceae	Desulfocapsaceae	Kiritimatiellaceae	Phyllobacteriaceae	Tr
Baekduiaceae	Desulfohalobiaceae	Koferiaceae	Pirellulaceae	Tr
Barnesiellaceae	Desulfomicrobiaceae	Koleobacteraceae	Piscirickettsiaceae	Tr
Bartonellaceae	Desulfosudaceae	Kordiimonadaceae	Planctomycetaceae	Tr
Bdellovibrionaceae	Desulfotomaculaceae	Kosmotogaceae	Planococcaceae	Tr
Beijerinckiaceae	Desulfotomaculaceae	Kribbellaceae	Pleomorphomonadaceae	Tr
Bernardetiaceae	Desulfurellaceae	Ktedonosporobacteraceae	Polyangiaceae	Tr
Beutenbergiaceae	Desulfurobacteriaceae	Kytococcaceae	Porphyromonadaceae	Tr
Bifidobacteriaceae	Desulfuromonadaceae	Labilillicaceae	Prevotellaceae	Tr
Blastochloridaceae	Devosiaceae	Lachnospiraceae	Prochlorococcaceae	Tr
Blattabacteriaceae	Dictyoglomaceae	Lacipirellulaceae	Prochlorotrichaceae	Tr
Bogoriellaceae	Dietziaceae	Lactobacillaceae	Prolixibacteraceae	Tr
Borrelliaceae	Dissulfurispiraceae	Lawsonellaceae	Promicromonosporaceae	Tr
Boseaceae	Dysgonomonadaceae	Legionellaceae	Propionibacteriaceae	Tr
Brachyspiraceae	Ectothiorhodospiraceae	Leptolyngbyaceae	Proteinivoraceae	Tr
Bradymonadaceae	Eggerthellaceae	Leptospiaceae	Pseudanabaenaceae	Tr
Bradyrhizobiaceae	Egibacteraceae	Leptotrichiaceae	Pseudoalteromonadaceae	Tr
Breoghanaceae	Egicoccaceae	Lichenihabitantaceae	Pseudomonadaceae	Tr
Brevibacteriaceae	Elioraeaceae	Limnochordaceae	Pseudonocardaceae	Tr
Brucellaceae	Elusimicrobiaceae	Listeriaceae	Psychromonadaceae	Tr
Bruguierivoracaceae	Emcibacteraceae	Litoricolaceae	Puniceicoccaceae	Tr
Bryobacteraceae	Endomicrobiaceae	Magnetococcaceae	Rhabdochlamydiaceae	Tr
Budviciaceae	Endozoicomonadaceae	Mangrovivirgaceae	Rhizobiaceae	Tr
Burkholderiaceae	Enterobacteriaceae	Maricaulaceae	Rhodanobacteraceae	Ti
Caedimonadaceae	Enterococcaceae	Marinifiliaceae	Rhodobacteraceae	Ti
Caldilineaceae	Entomoplasmataceae	Marinifiliaceae	Rhodocyclaceae	Tc
Caldiseriaceae	Erwinaceae	Marinobacteraceae	Rhodospirillaceae	Tr
Calditrichaceae	Erysipelotrichaceae	Mariprofundaceae	Rhodothermaceae	Tr
Calothrixaceae	Erythrobacteraceae	Marinimaceae	Rickettsiaceae	Tr

Barras_Species [2]

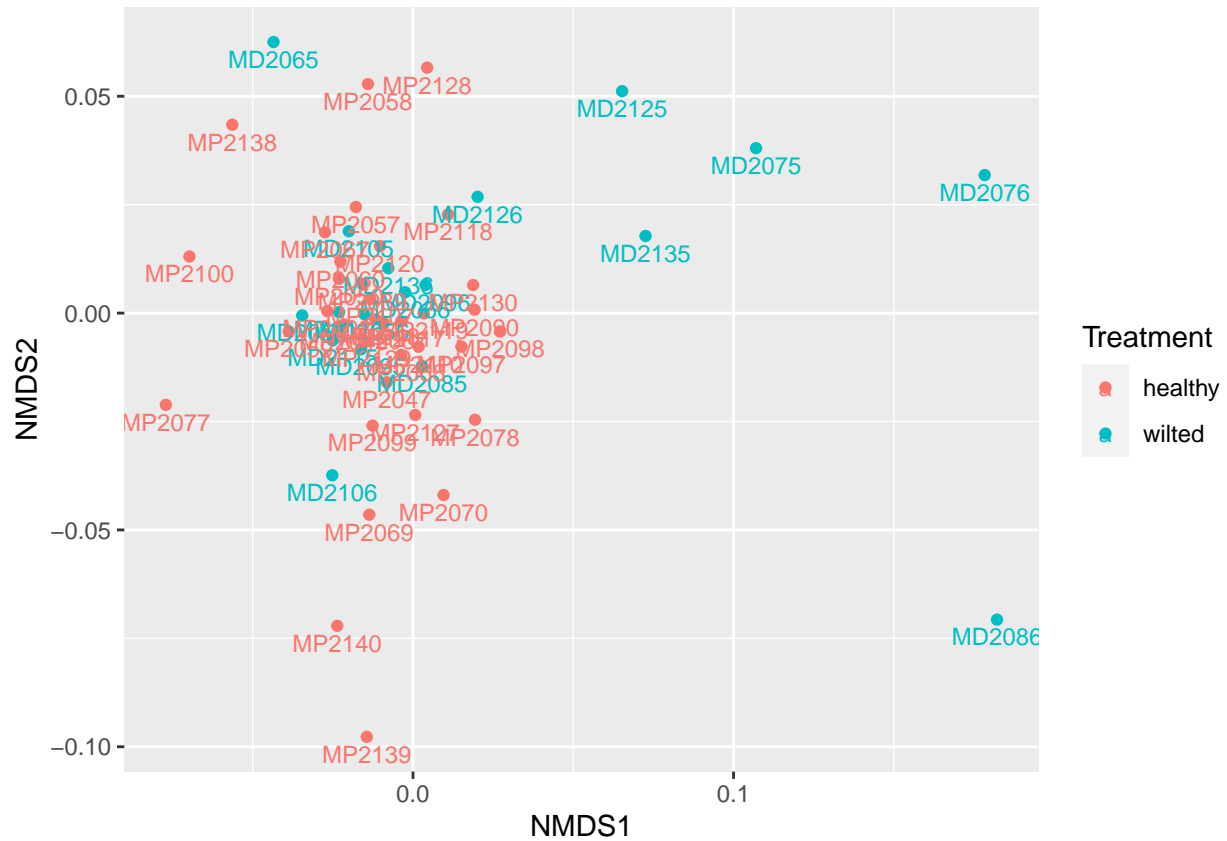
[[1]]



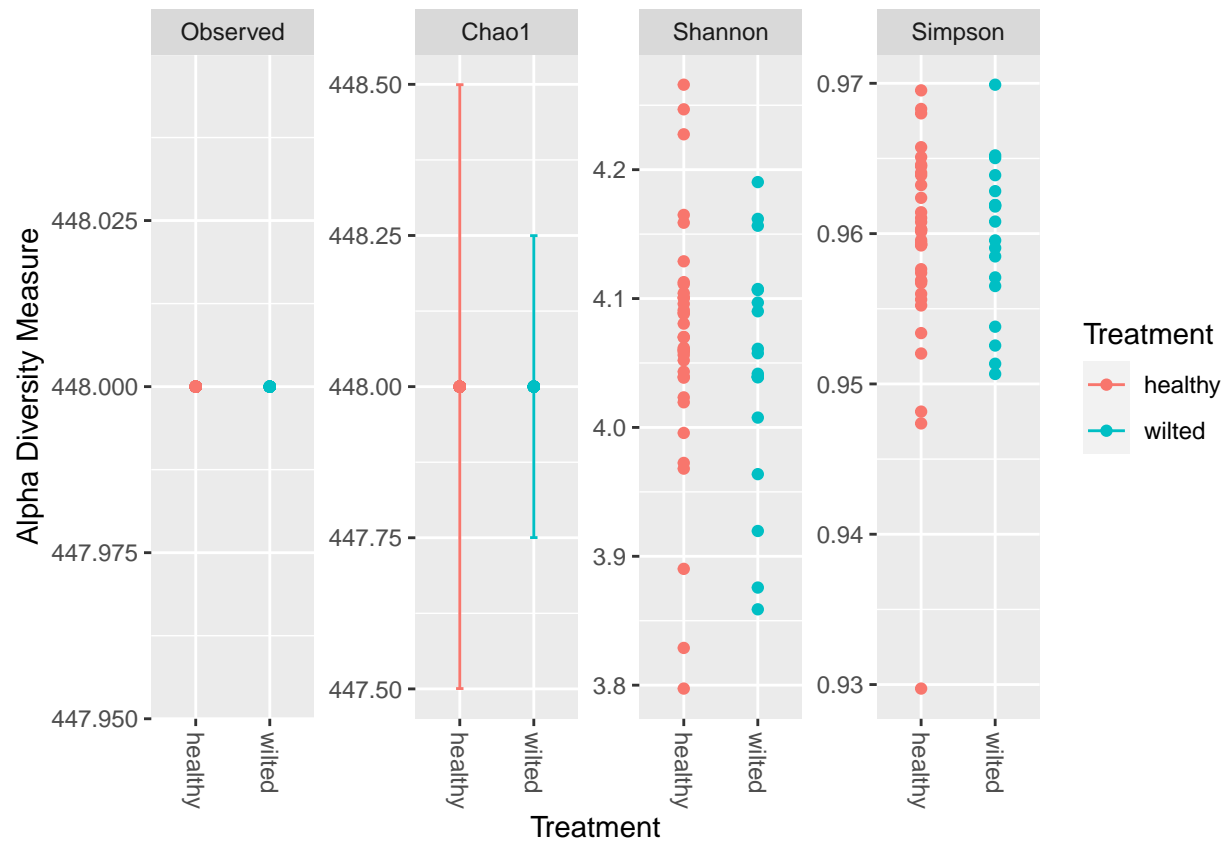
```
Beta_diversity(merge_Bacteria , 'Family' , 'Treatment', 'bray')
```

```
## Wisconsin double standardization
## Run 0 stress 0.1412073
## Run 1 stress 0.1418145
## Run 2 stress 0.1390466
## ... New best solution
## ... Procrustes: rmse 0.02128683 max resid 0.1331159
## Run 3 stress 0.1405626
## Run 4 stress 0.145559
## Run 5 stress 0.1863488
## Run 6 stress 0.1412066
## Run 7 stress 0.1369866
## ... New best solution
## ... Procrustes: rmse 0.06315741 max resid 0.3787318
## Run 8 stress 0.1551948
## Run 9 stress 0.142348
## Run 10 stress 0.1405103
## Run 11 stress 0.1372656
## ... Procrustes: rmse 0.01219911 max resid 0.0572946
## Run 12 stress 0.1369866
## ... Procrustes: rmse 5.037779e-05 max resid 0.0001538958
## ... Similar to previous best
## Run 13 stress 0.1412071
## Run 14 stress 0.1402344
```

```
## Run 15 stress 0.1373769
## ... Procrustes: rmse 0.01547261  max resid 0.05919644
## Run 16 stress 0.1412072
## Run 17 stress 0.1373766
## ... Procrustes: rmse 0.01563171  max resid 0.05932202
## Run 18 stress 0.1441744
## Run 19 stress 0.1552141
## Run 20 stress 0.1625797
## *** Best solution repeated 1 times
```



```
Alpha_diversity(merge_Bacteria , 'Family' , 'Treatment')
```



#——Bacteria by Genero

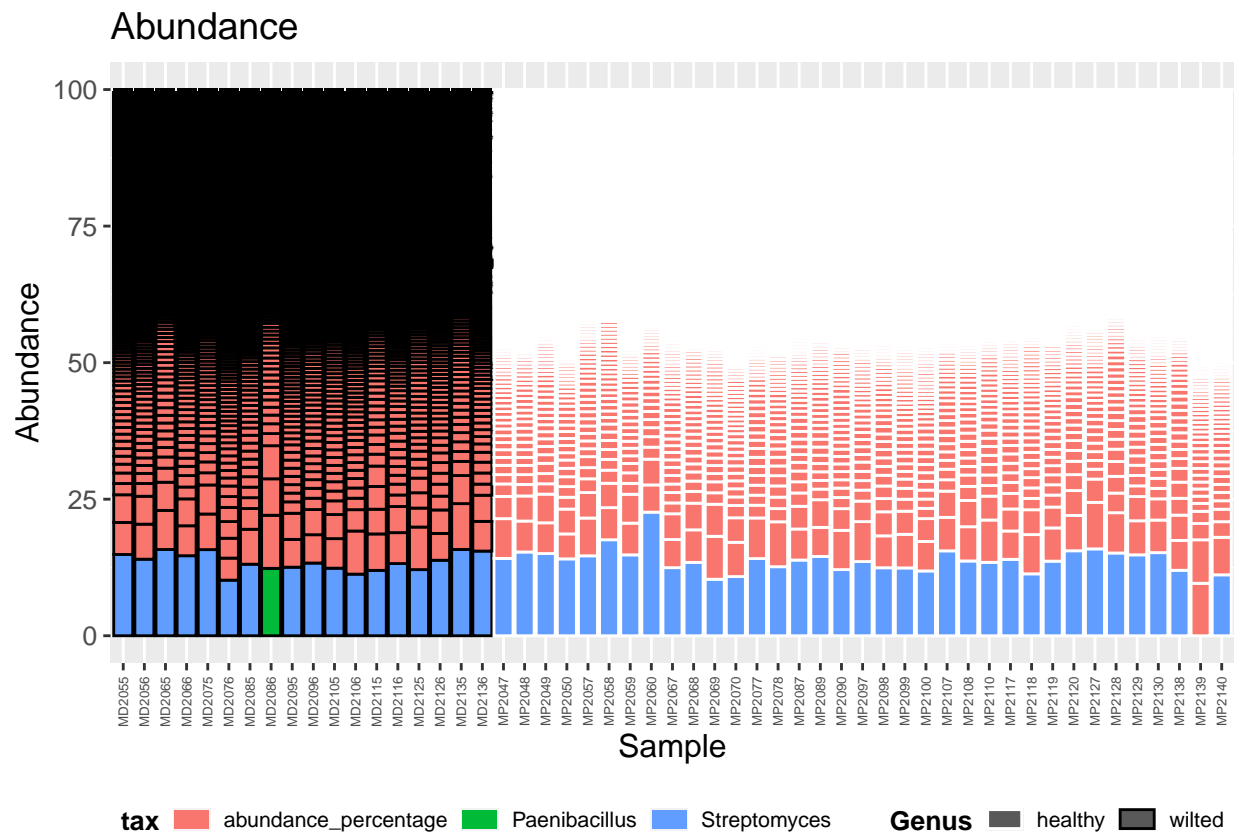
```
Barras_Species <- Abundance_barras(merge_Bacteria, 'Genus', 'Treatment', 10.0)
Barras_Species[1]
```

```
## [[1]]
```

lex	Desulfuricoccus	Labyrinthula	Paramecium	Sulfolobus
flexum	Desulfatibacillum	Labrys	Paroceanicella	Succinivibrio
habitans	Desulfatibacterium	Laceyella	Parolsenella	Suicoccus
luna	Desulfobacca	Lachnoanaerobaculum	Parvibaculum	Sulfidibacter
marina	Desulfobacter	Lachnoclostridium	Parvimonas	Sulfitobacter
ncola	Desulfobacula	Lachnospira	Parvularcula	Sulfuricaulis
rhabdus	Desulfobulbus	Lacibacter	Pasteurella	Sulfuricella
rufa	Desulfocapsa	Lacimicrobium	Paucibacter	Sulfuricurvum
salimonas	Desulfococcus	Lacinutrix	Paucilactobacillus	Sulfuriferula
salinus	Desulfocurvibacter	Lacipirellula	Pauljensenia	Sulfuriflexus
sediminimonas	Desulfofarcimen	Lacrimispora	Pectinatus	Sulfurifustis
sphaera	Desulfoglaeba	Lacticaseibacillus	Pectobacterium	Sulfurihydrogenibium
talea	Desulfoglobium	Lactiplantibacillus	Pediococcus	Sulfurimicrobium
ibacter	Desulfoluna	Lactobacillus	Pedobacter	Sulfurimonas
hidicoccus	Desulfolutivibrio	Lactococcus	Pedococcus	Sulfuriroseicoccus
hnia	Desulfomarina	Lacunisphaera	Pelagerythrobacter	Sulfuritalea
nobacterium	Desulfomicrobium	Lancefieldella	Pelagibacterium	Sulfuritortus
angium	Desulfomonile	Lapidilactobacillus	Pelagovum	Sulfurivermis
bacter	Desulfonema	Laribacter	Pelolistega	Sulfurospirillum
cibacterium	Desulforamulus	Larkinella	Pelobacter	Sulfurovum
ibacter	Desulforapulum	Latlactobacillus	Pelodictyon	Sutcliffeella
imonas	Desulfosarcina	Lautropia	Pelolinea	Sutterella
ratoleum	Desulfoscipio	Lawsonella	Pelosinus	Suttonella
nicrococcus	Desulfosediminicola	Lawsonia	Pengzhengrongella	Swingsia
nophonus	Desulfosporosinus	Leadbetterella	Peptacetobacter	Symbiobacterium
obacter	Desulfosudis	Leclercia	Peptoclostridium	Symmachella
a	Desulfotalea	Lederbergia	Peptoniphilus	Synechococcus
acaulis	Desulfotomaculum	Leeuwenhoekella	Peribacillus	Synechocystis
lassotoga	Desulfovibrio	Legionella	Periweissella	Syntrophobacter
tibacter	Desulfurispirillum	Leifsonia	Permianibacter	Syntrophobotulus
obium	Desulfurivibrio	Leisingera	Persephonella	Syntrophomonas
acter	Desulfurobacterium	Lelliottia	Persicimonas	Syntrophotalea
ntiibacter	Desulfuromonas	Leminorella	Peteryoungia	Syntrophothermus
ntimicrobium	Devosia	Lentibacillus	Petrimonas	Syntrophus
ntimonas	Devriesea	Lentilactobacillus	Petrocella	Tabrificola
ticoccus	Dialister	Lentilitoribacter	Petrotoga	Tamiana
imonas	Diaminobutyricimonas	Lentzea	Phaeobacter	Tannerella
liella	Diaphorobacter	Leptodesmis	Phascolarctobacterium	Tardibacter
idibacter	Dichelobacter	Leptolyngbya	Pherylobacterium	Tardiphaga
wickia	Dickeya	Leptosira	Phnomibacter	Tateyamaria
acterium	Dictyoglomus	Leptospirillum	Phocaeicola	Tatlockia
rcus	Dietzia	Leptothermofonsia	Phoenicibacter	Tatumella
hizobium	Dinoroseobacter	Leptothrix	Photobacterium	Tautonia
pira	Dissulfurimicrobium	Leptotrichia	Photorhabdus	Taylorella
pirillum	Dissulfurispira	Leucobacter	Phototrophicus	Telmatocola
obacter	Dokdonella	Leuconostoc	Phreatobacter	Tenacibaculum
llus	Dokdonia	Levilactobacillus	Phycococcus	Tenuifilum
eriplanes	Dolichospermum	Liberibacter	Phycisphaera	Tepidanaerobacter
eriovorax	Dolosigranulum	Lichenicola	Phyllobacterium	Tepidibacter
eroides	Dongshaia	Lichenihabitans	Phytobacter	Tepidiforma
duia	Dorea	Ligilactobacillus	Phytohabetans	Tepidimonas
esiella	Draconibacterium	Lignipirellula	Pigmentiphaga	Tepiditoga
onella	Duganella	Limihaloglobus	Pikeienueella	Terasakiella
lea	Duncaniella	Limnobacter	Pimelobacter	Teredinibacter
lovibrio	Dyadobacter	Limnobaculum	Pirellula	Terribacillus
giatota	Dyella	Limnochorda	Pirellulimonas	Terricaulis
irickia	Dysgonomonas	Limnoglobus	Piscirickettsia	Terriglobus
	Dysnomobacter	Limnolobus	Piscirickettsia	Terrirhabdus

Barras_Species [2]

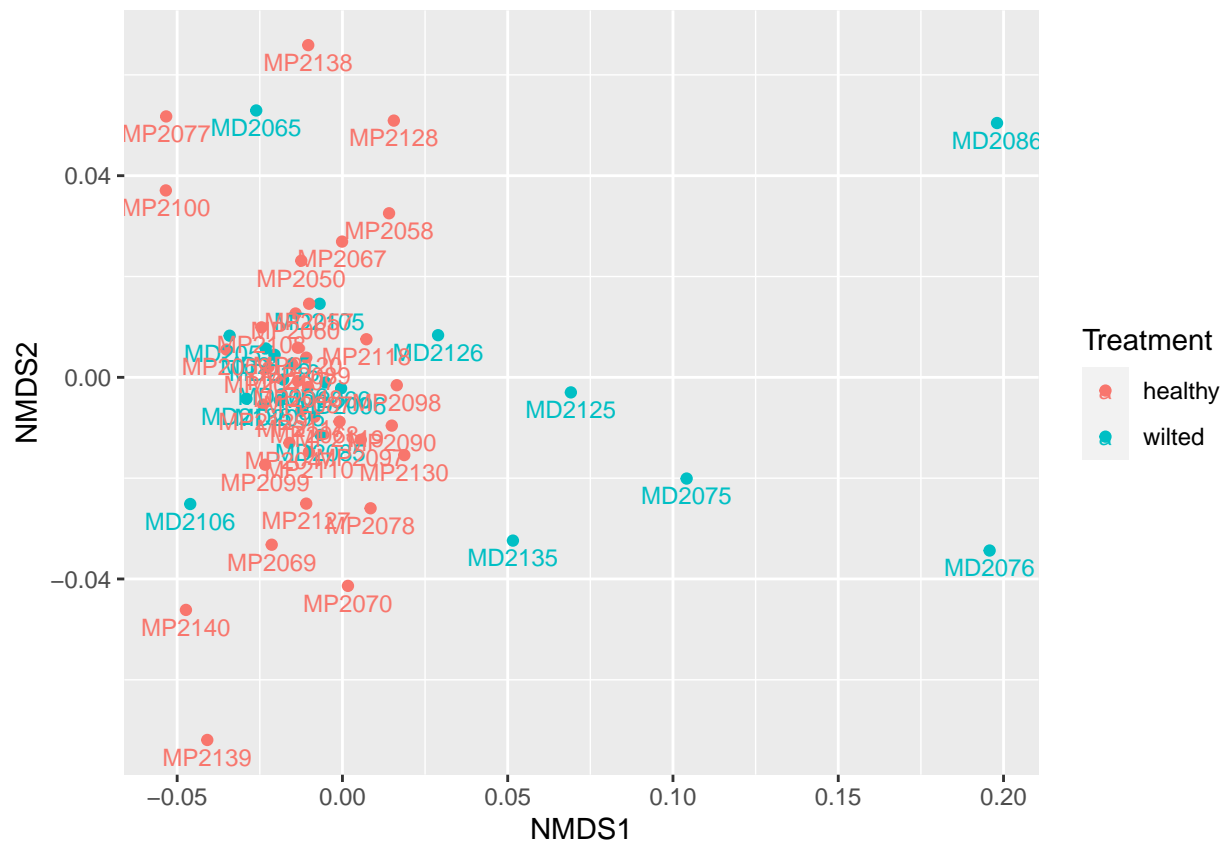
[[1]]



```
Beta_diversity(merge_Bacteria , 'Genus' , 'Treatment', 'bray')
```

```
## Wisconsin double standardization
## Run 0 stress 0.1467945
## Run 1 stress 0.145778
## ... New best solution
## ... Procrustes: rmse 0.04626627 max resid 0.2220378
## Run 2 stress 0.1433318
## ... New best solution
## ... Procrustes: rmse 0.08271459 max resid 0.3946675
## Run 3 stress 0.1467201
## Run 4 stress 0.1531013
## Run 5 stress 0.1467425
## Run 6 stress 0.146185
## Run 7 stress 0.1505679
## Run 8 stress 0.1500532
## Run 9 stress 0.145519
## Run 10 stress 0.151215
## Run 11 stress 0.1512369
## Run 12 stress 0.1490812
## Run 13 stress 0.148721
## Run 14 stress 0.1432834
## ... New best solution
## ... Procrustes: rmse 0.004246344 max resid 0.02586992
## Run 15 stress 0.1454653
```

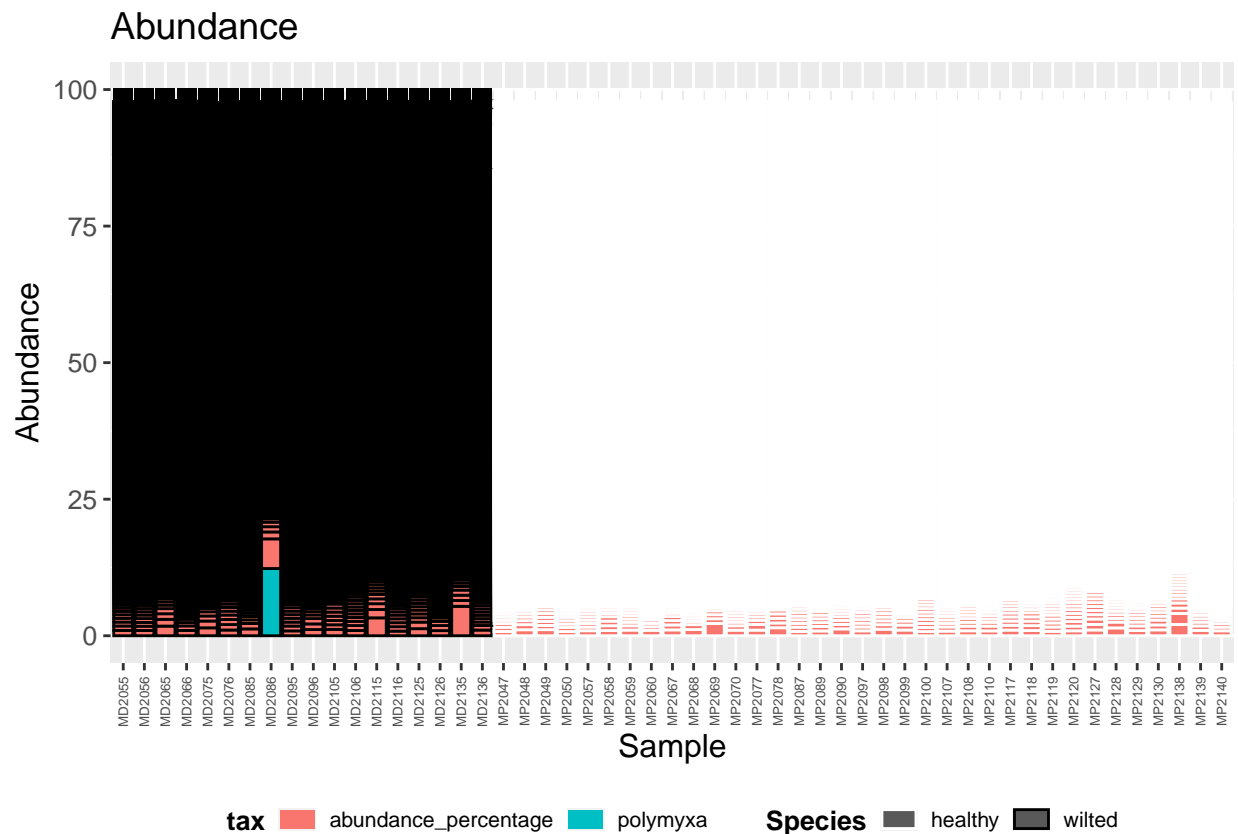
```
Alpha_diversity(merge_Bacteria , 'Genus' , 'Treatment')
```



e	<div></div> lactamica	<div></div> sp. 'Soap Lake #7'	<div></div> sp. KSB-15	<div></div> sp. X
	<div></div> lactarius	<div></div> sp. (Blaberus giganteus)	<div></div> sp. KSM-R2A25	<div></div> sp. X
	<div></div> lactatifermentans	<div></div> sp. (Blatta orientalis)	<div></div> sp. KSM-R2A30	<div></div> sp. X
	<div></div> lactatiformans	<div></div> sp. (Blattella germanica)	<div></div> sp. KT 15	<div></div> sp. X
	<div></div> lactis	<div></div> sp. (Cryptocercus kyebangensis)	<div></div> sp. KT25b	<div></div> sp. X
	<div></div> lactucae	<div></div> sp. (Cryptocercus punctulatus) str. Cpu	<div></div> sp. KTR9	<div></div> sp. X
	<div></div> lacunae	<div></div> sp. (ex Adelges kitamiensis)	<div></div> sp. KUDC0405	<div></div> sp. X
	<div></div> lacus	<div></div> sp. (ex Biomphalaria glabrata)	<div></div> sp. KUDC0406	<div></div> sp. X
	<div></div> lacuslunae	<div></div> sp. (Mastotermes darwiniensis)	<div></div> sp. KUDC1026	<div></div> sp. X
	<div></div> lacustris	<div></div> sp. (Nauphoeta cinerea)	<div></div> sp. KUDC1714	<div></div> sp. X
	<div></div> laguerreae	<div></div> sp. (Periplaneta americana)	<div></div> sp. KUIN-1	<div></div> sp. X
	<div></div> laidlawii	<div></div> sp. 001	<div></div> sp. Kuro-4	<div></div> sp. X
	<div></div> laixinhei	<div></div> sp. 007	<div></div> sp. KW1	<div></div> sp. X
	<div></div> laikuanensis	<div></div> sp. 008	<div></div> sp. KX20019	<div></div> sp. X
	<div></div> laminatus	<div></div> sp. 02C 26	<div></div> sp. KY-GH-1	<div></div> sp. X
	<div></div> lanienae	<div></div> sp. 09C 129	<div></div> sp. KY-YJ-3	<div></div> sp. X
	<div></div> lanii	<div></div> sp. 09RB8471	<div></div> sp. KY3	<div></div> sp. Y
	<div></div> lansingensis	<div></div> sp. 09RB8910	<div></div> sp. KY5	<div></div> sp. Y
	<div></div> lanthieri	<div></div> sp. 1_2014MBL_MicDiv	<div></div> sp. KY70	<div></div> sp. Y
	<div></div> lapagei	<div></div> sp. 1_2015MBL_MicDiv	<div></div> sp. KY75	<div></div> sp. Y
	<div></div> lapidicaptus	<div></div> sp. 1-1C	<div></div> sp. L-07	<div></div> sp. Y
	<div></div> lapsinanis	<div></div> sp. 1.5R	<div></div> sp. L-2-11	<div></div> sp. Y
	<div></div> lari	<div></div> sp. 1(2017)	<div></div> sp. L-8-10	<div></div> sp. Y
	<div></div> lariciata	<div></div> sp. 103DPR2	<div></div> sp. L-8-3	<div></div> sp. Y
	<div></div> larrymorei	<div></div> sp. 1063	<div></div> sp. L-A4	<div></div> sp. Y
	<div></div> larvae	<div></div> sp. 107-1	<div></div> sp. L12M9	<div></div> sp. Y
	<div></div> lata	<div></div> sp. 1070	<div></div> sp. L1A13	<div></div> sp. Y
	<div></div> latens	<div></div> sp. 10FS3-1	<div></div> sp. L1A34	<div></div> sp. Y
	<div></div> laterosporus	<div></div> sp. 10M-3C3	<div></div> sp. L1A9	<div></div> sp. Y
	<div></div> laumondii	<div></div> sp. 10RB9215	<div></div> sp. L1139	<div></div> sp. Y
	<div></div> lautus	<div></div> sp. 11-1-2	<div></div> sp. L1SW	<div></div> sp. Y
	<div></div> lavamentivorans	<div></div> sp. 11-B-312	<div></div> sp. L2A1	<div></div> sp. Y
	<div></div> lavendulae	<div></div> sp. 113-1-2	<div></div> sp. L2A11	<div></div> sp. Y
	<div></div> leachii	<div></div> sp. 113-3-3	<div></div> sp. L3-i22	<div></div> sp. Y
	<div></div> leadbetteri	<div></div> sp. 113-3-9	<div></div> sp. L3-i23	<div></div> sp. Y
	<div></div> leeuwenhoekii	<div></div> sp. 1137	<div></div> sp. L3A3	<div></div> sp. Y
	<div></div> leguminosarum	<div></div> sp. 113P3	<div></div> sp. L3A6	<div></div> sp. Y
	<div></div> lehensis	<div></div> sp. 114	<div></div> sp. L3A8	<div></div> sp. Y
	<div></div> lekithochrous	<div></div> sp. 11515TR	<div></div> sp. L5	<div></div> sp. Y
	<div></div> lemovicicum	<div></div> sp. 116-D4	<div></div> sp. L51/94	<div></div> sp. Y
	<div></div> lemorum	<div></div> sp. 119287	<div></div> sp. L5B5	<div></div> sp. Y
	<div></div> lenghuensis	<div></div> sp. 11B	<div></div> sp. L6-1	<div></div> sp. Y
	<div></div> lenta	<div></div> sp. 11kri321	<div></div> sp. L9-4	<div></div> sp. Y
	<div></div> lentiflavum	<div></div> sp. 12200R-103	<div></div> sp. LA-2-3-30-S1-D2	<div></div> sp. Y
	<div></div> lentis	<div></div> sp. 13-15	<div></div> sp. LA112445	<div></div> sp. Y
	<div></div> lentocellum	<div></div> sp. 131	<div></div> sp. LA31	<div></div> sp. Y
	<div></div> lentus	<div></div> sp. 131-2-1	<div></div> sp. LAB-08	<div></div> sp. Y
	<div></div> leopoldii	<div></div> sp. 131-2-5	<div></div> sp. LADL05-105	<div></div> sp. Y
	<div></div> leprae	<div></div> sp. 131-3-5	<div></div> sp. LAS2	<div></div> sp. Y
	<div></div> lepromatosis	<div></div> sp. 13159349	<div></div> sp. LB1	<div></div> sp. Y
	<div></div> leptomitiformis	<div></div> sp. 135	<div></div> sp. LBG001	<div></div> sp. Y
	<div></div> lettingae	<div></div> sp. 140616W15	<div></div> sp. LBUM 1475	<div></div> sp. Y
andiella	<div></div> liangguodongii	<div></div> sp. 14171R-50	<div></div> sp. LBUM 1480	<div></div> sp. Y
ibbosus	<div></div> liangshanensis	<div></div> sp. 14181154	<div></div> sp. LBUM 1482	<div></div> sp. Y
ifera	<div></div> liaowanqingii	<div></div> sp. 143-21	<div></div> sp. LBUM920	<div></div> sp. Y
	<div></div> libanensis	<div></div> sp. 144S4	<div></div> sp. LC2018020214	<div></div> sp. Y
	<div></div> licheniformis	<div></div> sp. 15-184	<div></div> sp. LC6	<div></div> sp. Y

Barras_Species[2]

[[1]]



```
Beta_diversity(merge_Bacteria , 'Species' , 'Treatment' , 'bray')
```

```
## Wisconsin double standardization
## Run 0 stress 0.1597841
## Run 1 stress 0.1616467
## Run 2 stress 0.1596992
## ... New best solution
## ... Procrustes: rmse 0.05927183 max resid 0.3719294
## Run 3 stress 0.1703226
## Run 4 stress 0.1633768
## Run 5 stress 0.1622795
## Run 6 stress 0.166837
## Run 7 stress 0.1577449
## ... New best solution
## ... Procrustes: rmse 0.02327626 max resid 0.1144114
## Run 8 stress 0.1676656
## Run 9 stress 0.1603708
## Run 10 stress 0.1596212
## Run 11 stress 0.1635446
## Run 12 stress 0.1605549
## Run 13 stress 0.1637696
## Run 14 stress 0.1641623
## Run 15 stress 0.1602436
## Run 16 stress 0.1603294
## Run 17 stress 0.1587399
```

```
Alpha_diversity(merge_Bacteria , 'Species' , 'Treatment')
```

