

Funciones automatizadas para visualizar las barras de abundancia, diversidades alfa y beta; a diferentes niveles taxonomicos

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```
library("phyloseq")
library("ggplot2")
library("vegan")
```

```
## Loading required package: permute
```

```
## Loading required package: lattice
```

```
## This is vegan 2.6-4
```

```
library("RColorBrewer")
library("stringi")
```

Cargado de datos originales

```
setwd("/home/camila/GIT/Tesis_Maestria/Data/fresa_solena/Data1")
outpath = "/home/camila/GIT/Tesis_Maestria/Analisis_Comparativo/Fresa_Solena/Results_img"

fresa_kraken <- import_biom("fresa_kraken.biom")
colnames(fresa_kraken@tax_table@.Data) <- c("Kingdom", "Phylum", "Class", "Order", "Family", "Genus", "Species")
fresa_kraken@tax_table@.Data <- substr(fresa_kraken@tax_table@.Data,4,100)
colnames(fresa_kraken@otu_table@.Data) <- substr(colnames(fresa_kraken@otu_table@.Data),1,6)
metadata_fresa <- read.csv2("/home/camila/GIT/Tesis_Maestria/Data/fresa_solena/Data1/metadata.csv",header=1)
fresa_kraken@sam_data <- sample_data(metadata_fresa)
fresa_kraken@sam_data$Sample <- row.names(fresa_kraken@sam_data)
colnames(fresa_kraken@sam_data) <- c('Treatment', 'Samples')
samples_to_remove <- c("MP2079", "MP2080", "MP2088", "MP2109", "MP2137")
fresa_kraken_fil <- prune_samples(!(sample_names(fresa_kraken) %in% samples_to_remove), fresa_kraken)
```

Subconjunto de “Eukaryota”

```
merge_Eukaryota <- subset_taxa(fresa_kraken_fil, Kingdom == "Eukaryota")
```

Subconjunto de “Bacteria”

```
merge_Bacteria<-subset_taxa(fresa_kraken_fil,Kingdom=="Bacteria")
```

Funciones

glomToGraph

Crea los subconjuntos de datos necesarios para más adelante

input phy <- es el objeto phyloseq base tax <- rango taxonomico al que queremos recortar output glom <- objeto phyloseq ya recortado percentages <- objeto phyloseq de porcentajes sobre el glom percentages_df <- data frame de porcentajes sobre el glom

```
glomToGraph<-function(phy,tax){  
  ## creamos el subconjunto dependiendo del linaje taxonomico deseado  
  glom <- tax_glom(phy, taxrank = tax)  
  ## sacamos los porcentajes  
  percentages <- transform_sample_counts(glom, function(x) x*100 / sum(x) )  
  percentages_df <- psmelt(percentages)  
  return(list(glom,percentages,percentages_df))  
}
```

Graficar abundancias (stackbar)

```
Abundance_barras <- function(phy,tax,attribute,abundance_percentage){  
  ##llamar funcion de datos  
  Data <- glomToGraph(phy,tax)  
  glom <- Data[[1]] #phyloseq  
  percentages <- Data[[2]] #phyloseq  
  percentages_df <- Data[[3]] # dataframe  
  ## Graficamos para cada subconjunto las barras de abundancia  
  plot_barras <- ggplot(data=percentages_df, aes_string(x='Sample', y='Abundance', fill=tax ,color=attribute)) +  
    scale_colour_manual(values=c('white','black')) +  
    geom_bar(aes(), stat="identity", position="stack") +  
    labs(title = "Abundance", x='Sample', y='Abundance', color = tax) +  
    theme(legend.key.size = unit(0.2, "cm"),  
          legend.key.width = unit(0.25,"cm"),  
          legend.position = "bottom",  
          legend.direction = "horizontal",  
          legend.title=element_text(size=8, face = "bold"),  
          legend.text=element_text(size=6),  
          text = element_text(size=12),  
          axis.text.x = element_text(angle=90, size=5, hjust=1, vjust=0.5))  
  percentages_df$tax<-percentages_df[,ncol(percentages_df)]  
  percentages_df$tax[percentages_df$Abundance < abundance_percentage] <- "Others"  
  percentages_df$tax <- as.factor(percentages_df$tax)  
  plot_percentages <- ggplot(data=percentages_df, aes_string(x='Sample', y='Abundance', fill='tax' ,color=attribute)) +  
    scale_colour_manual(values=c('white','black')) +
```

```

geom_bar(aes(), stat="identity", position="stack") +
labs(title = "Abundance", x='Sample', y='Abundance', color = tax) +
theme(legend.key.size = unit(0.3, "cm"),
      legend.key.width = unit(0.5,"cm"),
      legend.position = "bottom",
      legend.direction = "horizontal",
      legend.title=element_text(size=10, face = "bold"),
      legend.text=element_text(size=8),
      text = element_text(size=12),
      axis.text.x = element_text(angle=90, size=5, hjust=1, vjust=0.5))
return(list(plot_barras,plot_percentages))
}

```

Graficar diversidad beta

```

Beta_diversity <- function(phy,tax,attribute,distance){
  Data <- glomToGraph(phy,tax)
  glom <- Data[[1]]
  #CREAR UN GLOM AL 10%
  percentages <- Data[[2]]
  percentages_df <- Data[[3]]
  ## Beta diversidad
  meta_ord <- ordinate(physeq = percentages, method = "NMDS", distance = distance)
  plot_beta <- plot_ordination(physeq = percentages, ordination = meta_ord, color = attribute) +
    geom_text(mapping = aes(label = colnames(phy@otu_table@.Data)), size = 3, vjust = 1.5)
  return(plot_beta)
}

```

Graficar diversidad alfa

```

Alpha_diversity <- function(phy,tax,attribute){
  ## llamamos la funcion que crea los dataset
  Data <- glomToGraph(phy,tax)
  glom <- Data[[1]]

  percentages <- Data[[2]]
  percentages_df <- Data[[3]]
  ## Alfa diversidad
  plot_alpha <- plot_richness(physeq = glom, measures = c("Observed","Chao1","Shannon","simpson"),x = a
  return(plot_alpha)
}

```

Graficas

Graficamos las barras de abundancia, las barras de abundancia aglomerando el 10% menos abundantes, alfa y beta diversidad para los siguientes niveles taxonomicos.

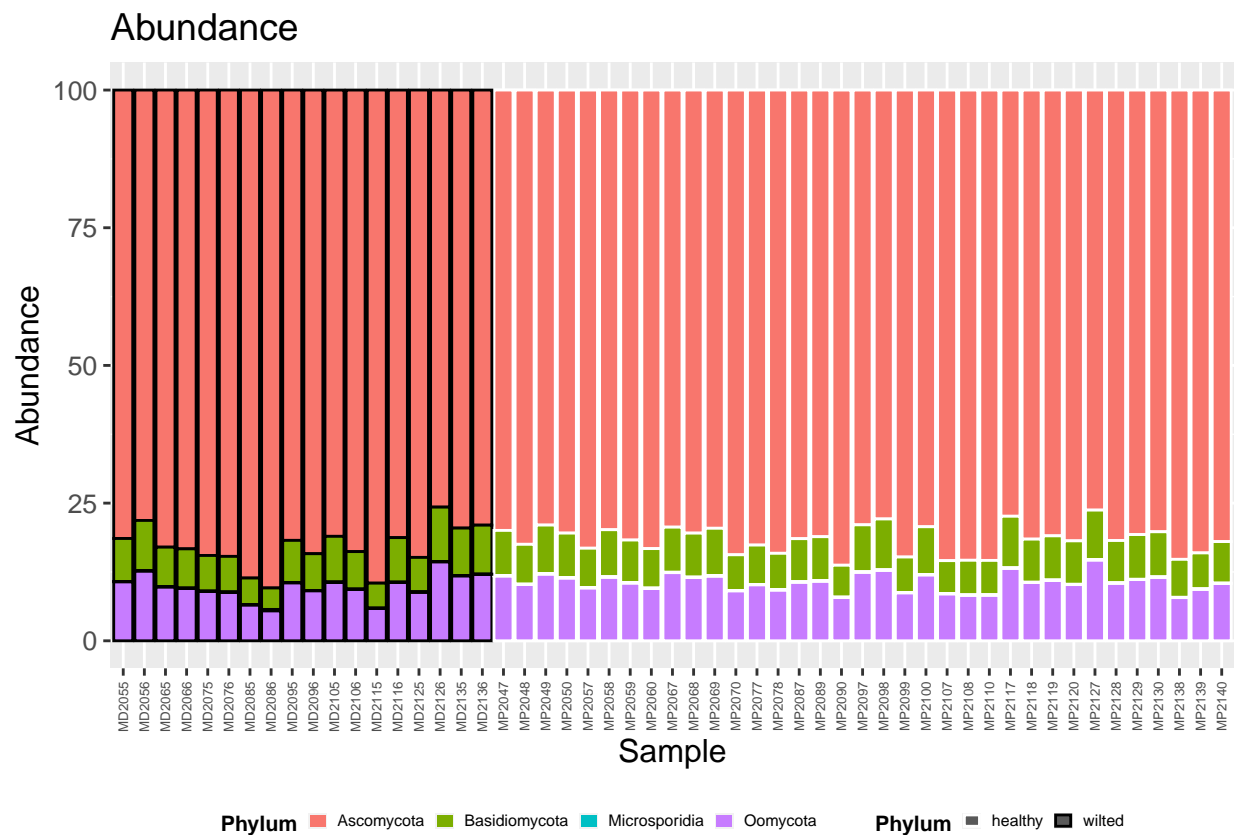
##———Eukarya by Phylum

```
Barras_Phylum <- Abundance_barras(merge_Eukaryota, 'Phylum' , 'Treatment', 10.0)
```

```
## Warning: 'aes_string()' was deprecated in ggplot2 3.0.0.
## i Please use tidy evaluation idioms with 'aes()'.
## i See also 'vignette("ggplot2-in-packages")' for more information.
## This warning is displayed once every 8 hours.
## Call 'lifecycle::last_lifecycle_warnings()' to see where this warning was
## generated.
```

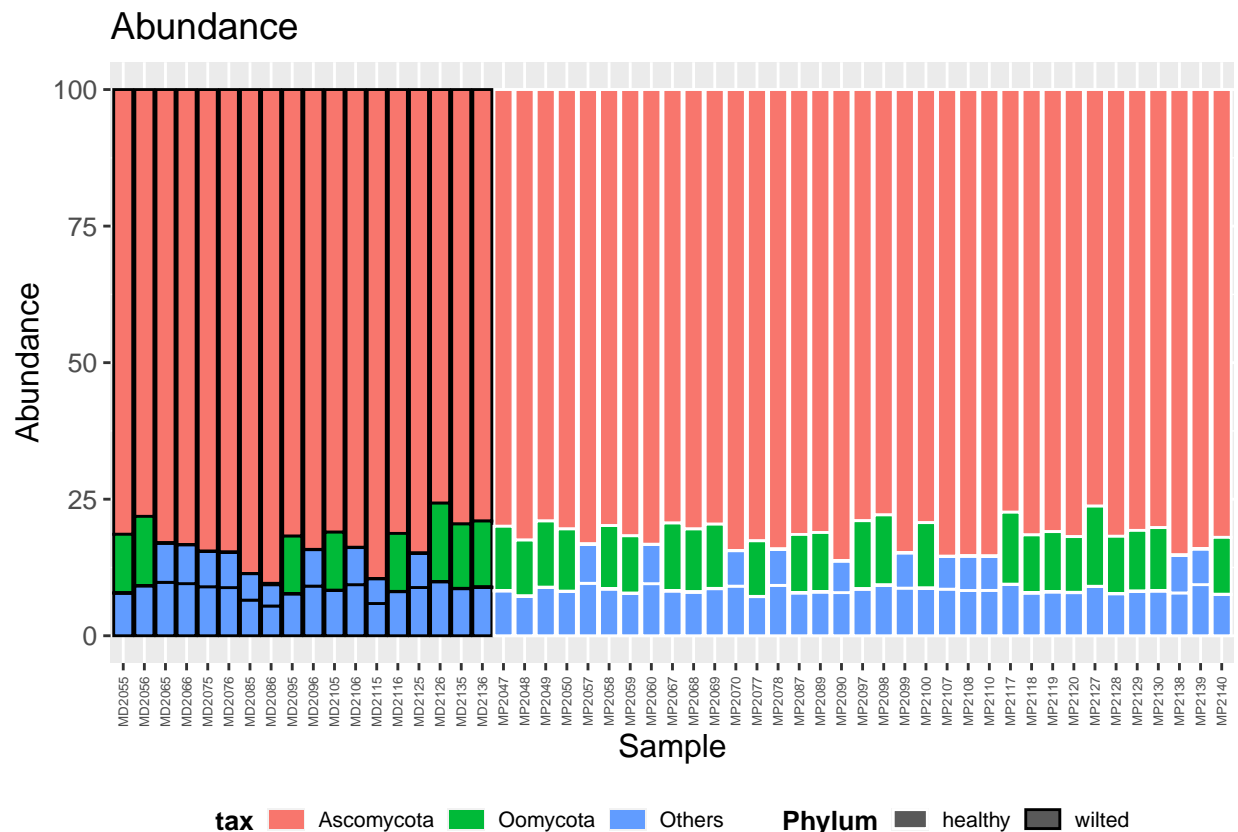
```
Barras_Phylum[1] # normal
```

```
## [[1]]
```



```
Barras_Phylum[2]
```

```
## [[1]]
```



```
Beta_diversity(merge_Eukaryota , 'Phylum' , 'Treatment', 'bray')
```

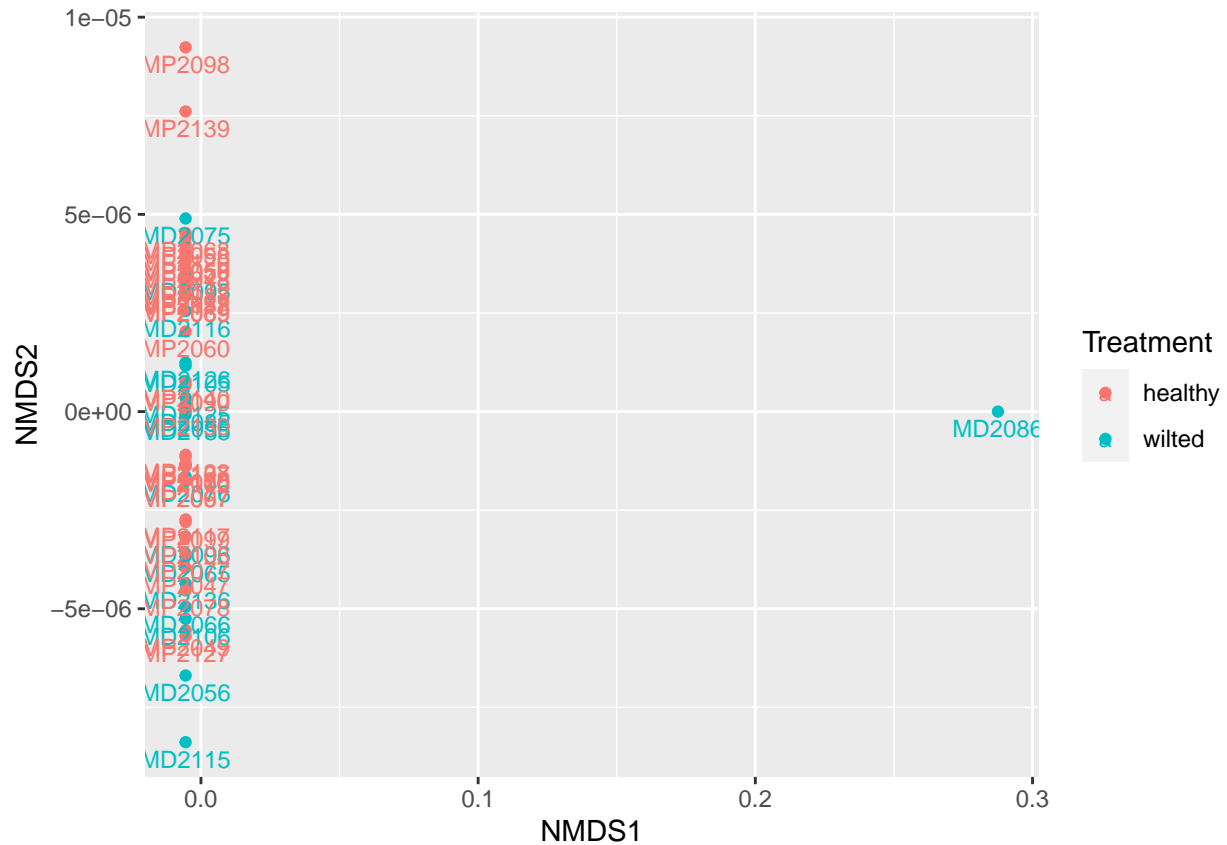
```
## Square root transformation
## Wisconsin double standardization
## Run 0 stress 0.01148681
## Run 1 stress 0.0001012286
## ... New best solution
## ... Procrustes: rmse 0.03438264 max resid 0.09018208
## Run 2 stress 8.080954e-05
## ... New best solution
## ... Procrustes: rmse 0.0002593369 max resid 0.0006111869
## ... Similar to previous best
## Run 3 stress 9.943482e-05
## ... Procrustes: rmse 0.0001287111 max resid 0.0003746192
## ... Similar to previous best
## Run 4 stress 5.946927e-05
## ... New best solution
## ... Procrustes: rmse 9.047211e-05 max resid 0.0002198202
## ... Similar to previous best
## Run 5 stress 9.885507e-05
## ... Procrustes: rmse 0.0001702104 max resid 0.0004297977
## ... Similar to previous best
## Run 6 stress 9.544256e-05
## ... Procrustes: rmse 0.000104487 max resid 0.0002663839
## ... Similar to previous best
```

```

## Run 7 stress 9.956639e-05
## ... Procrustes: rmse 8.390127e-05  max resid 0.0002437379
## ... Similar to previous best
## Run 8 stress 9.896738e-05
## ... Procrustes: rmse 0.0002583407  max resid 0.0006581007
## ... Similar to previous best
## Run 9 stress 7.056306e-05
## ... Procrustes: rmse 5.259222e-05  max resid 0.0001351559
## ... Similar to previous best
## Run 10 stress 9.479563e-05
## ... Procrustes: rmse 6.431556e-05  max resid 0.000134783
## ... Similar to previous best
## Run 11 stress 9.797854e-05
## ... Procrustes: rmse 0.0001674142  max resid 0.0004164753
## ... Similar to previous best
## Run 12 stress 0.0009492233
## Run 13 stress 0.0004211999
## ... Procrustes: rmse 0.0007288265  max resid 0.001932309
## ... Similar to previous best
## Run 14 stress 0.0001019576
## ... Procrustes: rmse 0.0002993734  max resid 0.000761613
## ... Similar to previous best
## Run 15 stress 9.067076e-05
## ... Procrustes: rmse 9.471571e-05  max resid 0.0002264973
## ... Similar to previous best
## Run 16 stress 9.6781e-05
## ... Procrustes: rmse 0.0001265178  max resid 0.0003112381
## ... Similar to previous best
## Run 17 stress 8.654003e-05
## ... Procrustes: rmse 0.0001074085  max resid 0.0002850164
## ... Similar to previous best
## Run 18 stress 6.269106e-05
## ... Procrustes: rmse 2.68101e-05  max resid 5.602508e-05
## ... Similar to previous best
## Run 19 stress 9.655239e-05
## ... Procrustes: rmse 0.0001244309  max resid 0.0003037819
## ... Similar to previous best
## Run 20 stress 0.0003162956
## ... Procrustes: rmse 0.0009368401  max resid 0.002436478
## ... Similar to previous best
## *** Best solution repeated 16 times

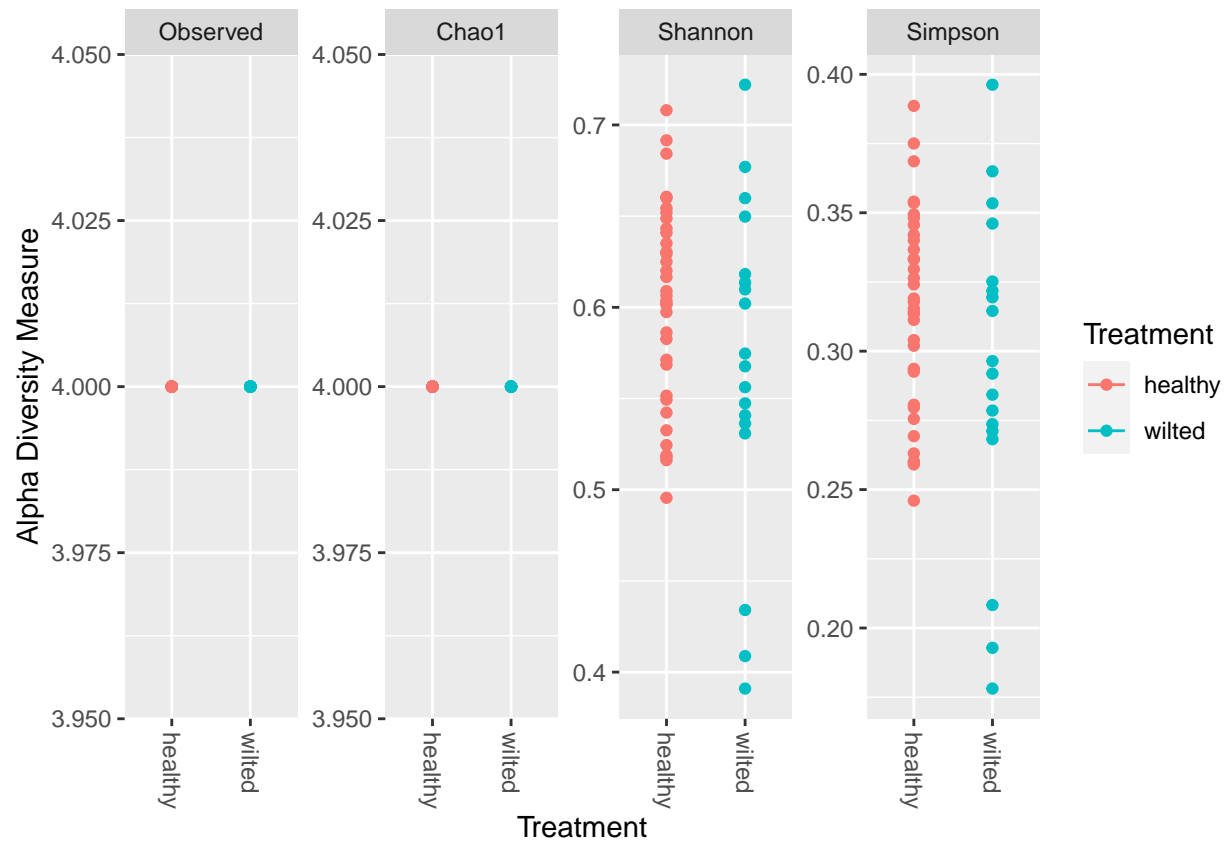
## Warning in metaMDS(veganifyOTU(physeq), distance, ...): stress is (nearly)
## zero: you may have insufficient data

```



```
Alpha_diversity(merge_Eukaryota , 'Phylum' , 'Treatment')
```

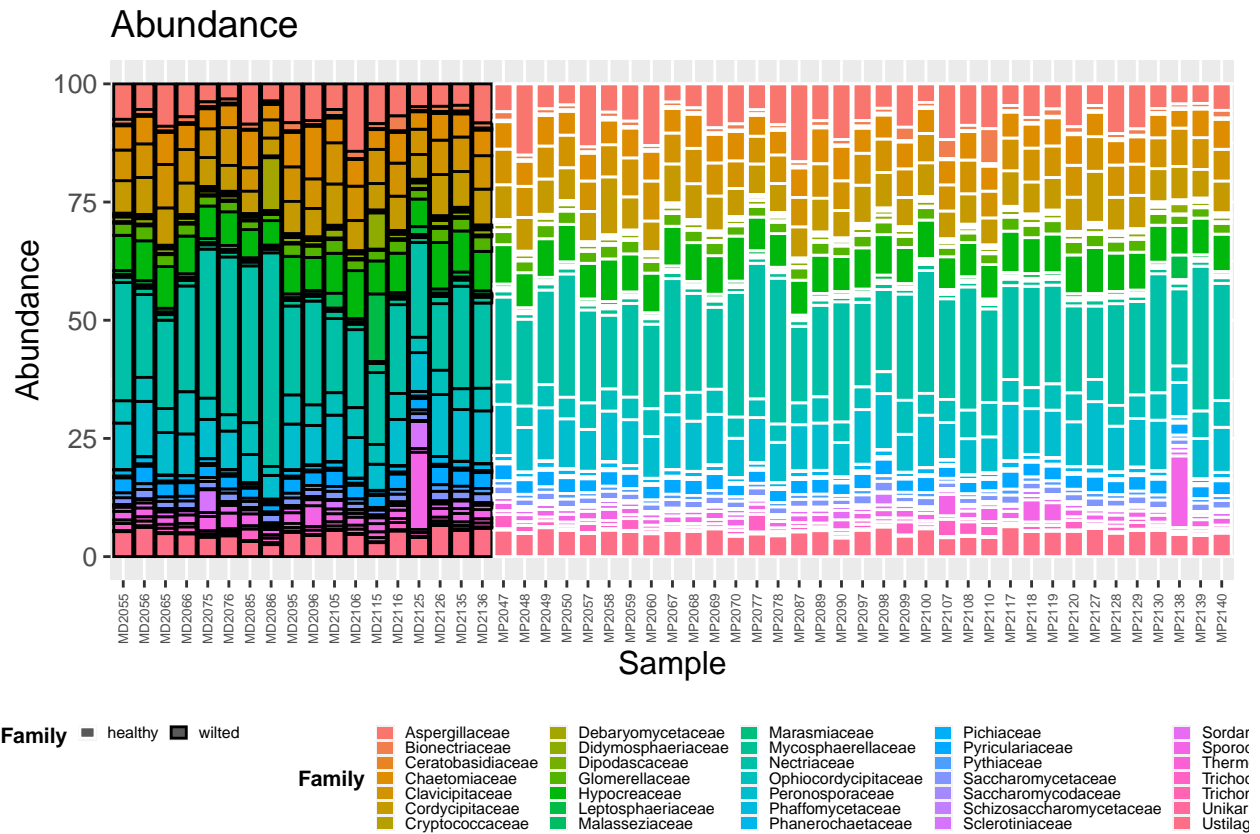
```
## Warning in estimate_richness(physeq, split = TRUE, measures = measures): The data you have provided contains
## any singletons. This is highly suspicious. Results of richness
## estimates (for example) are probably unreliable, or wrong, if you have already
## trimmed low-abundance taxa from the data.
##
## We recommended that you find the un-trimmed data and retry.
```



— Eukarya by Family

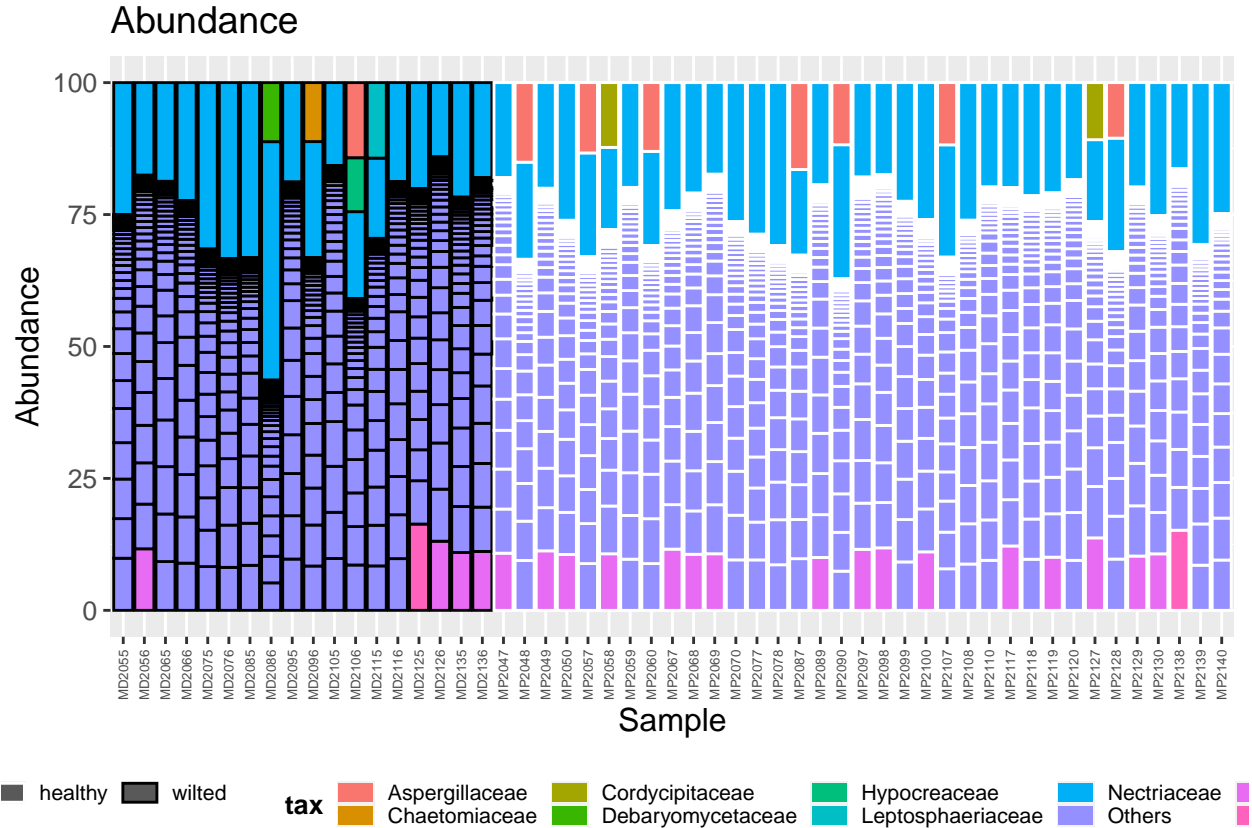
```
Barras_Species <- Abundance_barras(merge_Eukaryota, 'Family', 'Treatment', 10.0)
Barras_Species[1]
```

[[1]]



Barras_Species [2]

[[1]]



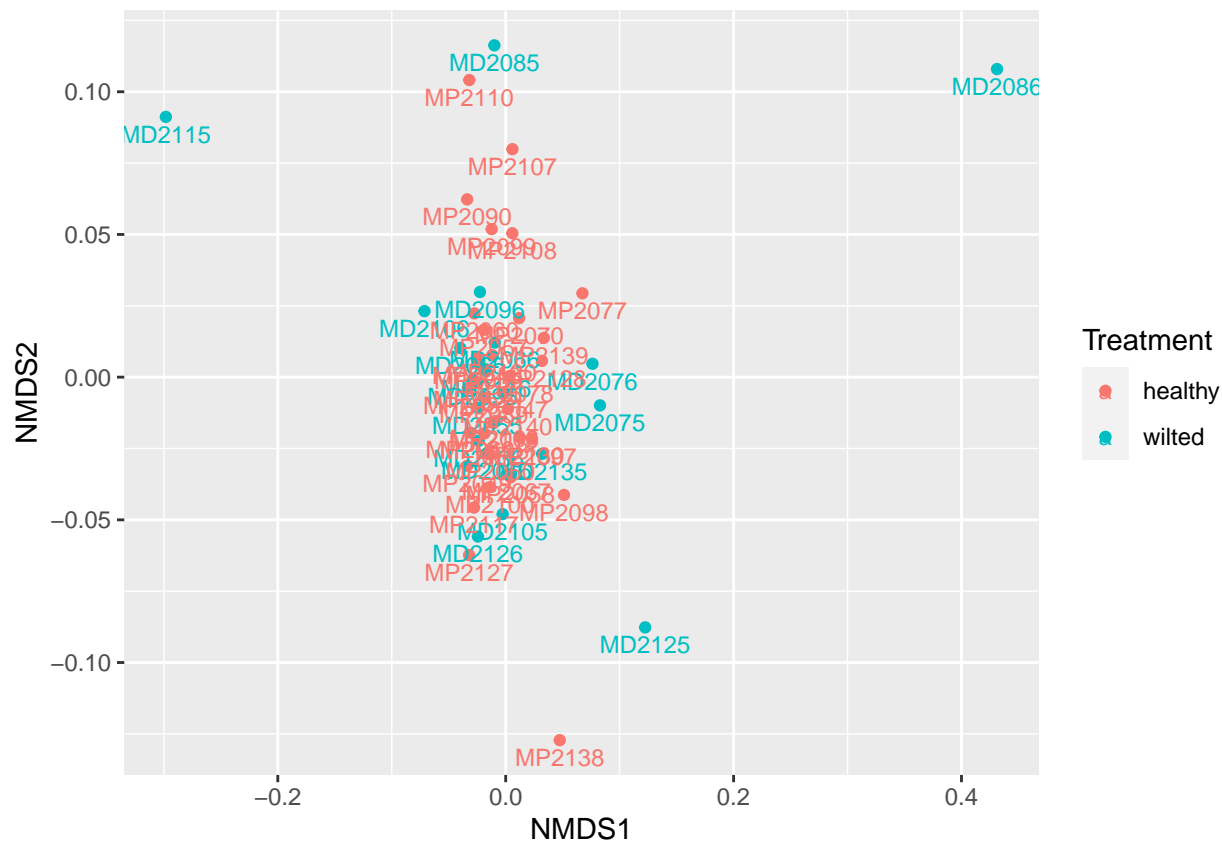
```
Beta_diversity(merge_Eukaryota , 'Family' , 'Treatment', 'bray')
```

```
## Wisconsin double standardization
## Run 0 stress 0.1089081
## Run 1 stress 0.1088881
## ... New best solution
## ... Procrustes: rmse 0.00169424 max resid 0.009378103
## ... Similar to previous best
## Run 2 stress 0.1088884
## ... Procrustes: rmse 0.001265836 max resid 0.007170168
## ... Similar to previous best
## Run 3 stress 0.1089292
## ... Procrustes: rmse 0.008427579 max resid 0.05473371
## Run 4 stress 0.1089076
## ... Procrustes: rmse 0.001717778 max resid 0.009428019
## ... Similar to previous best
## Run 5 stress 0.1088922
## ... Procrustes: rmse 0.0007890617 max resid 0.004563198
## ... Similar to previous best
## Run 6 stress 0.1091132
## ... Procrustes: rmse 0.009868536 max resid 0.05417681
## Run 7 stress 0.1089292
## ... Procrustes: rmse 0.008430896 max resid 0.05472563
## Run 8 stress 0.1091137
## ... Procrustes: rmse 0.01001222 max resid 0.05396838
```

```

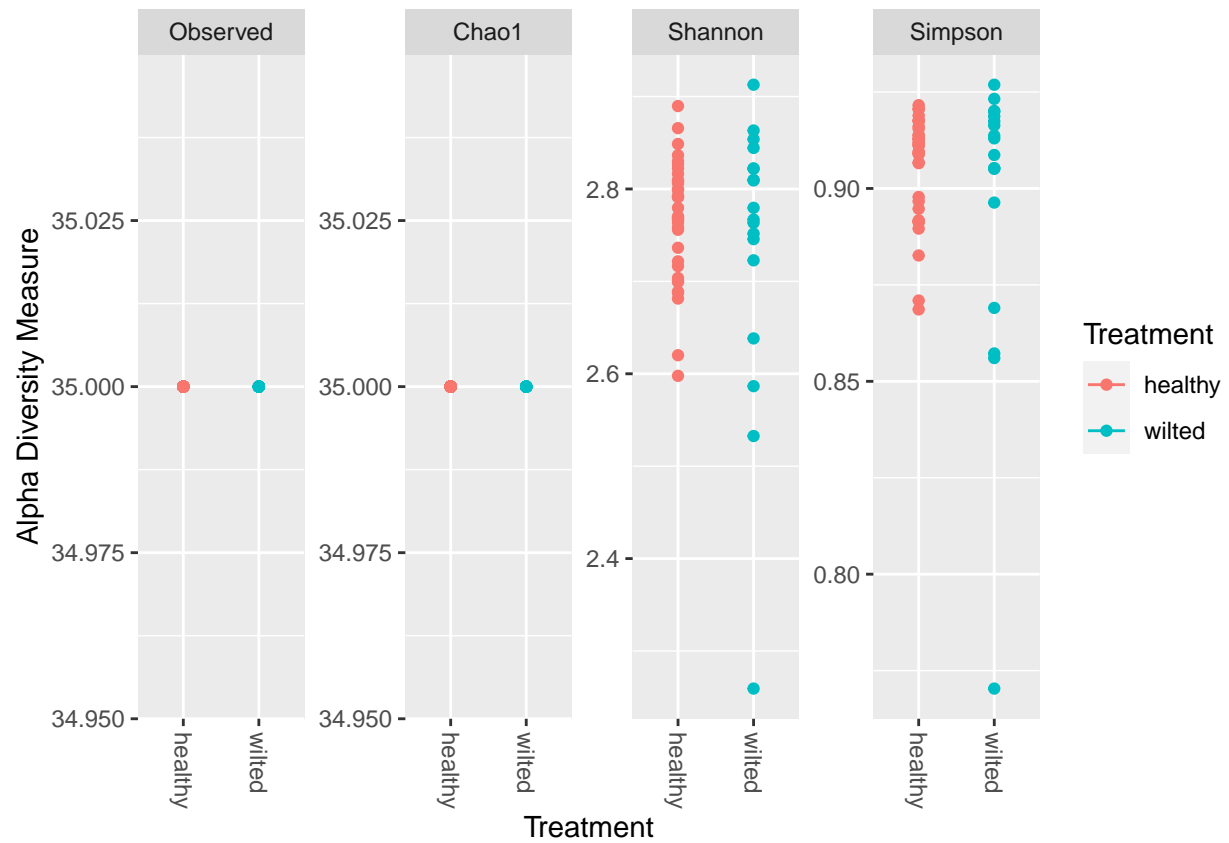
## Run 9 stress 0.1089299
## ... Procrustes: rmse 0.008416469 max resid 0.054788
## Run 10 stress 0.1089293
## ... Procrustes: rmse 0.008426162 max resid 0.05474377
## Run 11 stress 0.1089218
## ... Procrustes: rmse 0.000974449 max resid 0.005158194
## ... Similar to previous best
## Run 12 stress 0.1088884
## ... Procrustes: rmse 0.001341409 max resid 0.007533249
## ... Similar to previous best
## Run 13 stress 0.1089242
## ... Procrustes: rmse 0.002627083 max resid 0.0150801
## Run 14 stress 0.1088881
## ... Procrustes: rmse 5.634936e-06 max resid 2.393439e-05
## ... Similar to previous best
## Run 15 stress 0.1089298
## ... Procrustes: rmse 0.008417301 max resid 0.05478119
## Run 16 stress 0.1089185
## ... Procrustes: rmse 0.002321269 max resid 0.009035555
## ... Similar to previous best
## Run 17 stress 0.1088968
## ... Procrustes: rmse 0.002055205 max resid 0.009041251
## ... Similar to previous best
## Run 18 stress 0.1091401
## ... Procrustes: rmse 0.009638276 max resid 0.05345771
## Run 19 stress 0.108949
## ... Procrustes: rmse 0.00826652 max resid 0.05401651
## Run 20 stress 0.1089489
## ... Procrustes: rmse 0.008264563 max resid 0.05402098
## *** Best solution repeated 9 times

```



```
Alpha_diversity(merge_Eukaryota , 'Family' , 'Treatment')
```

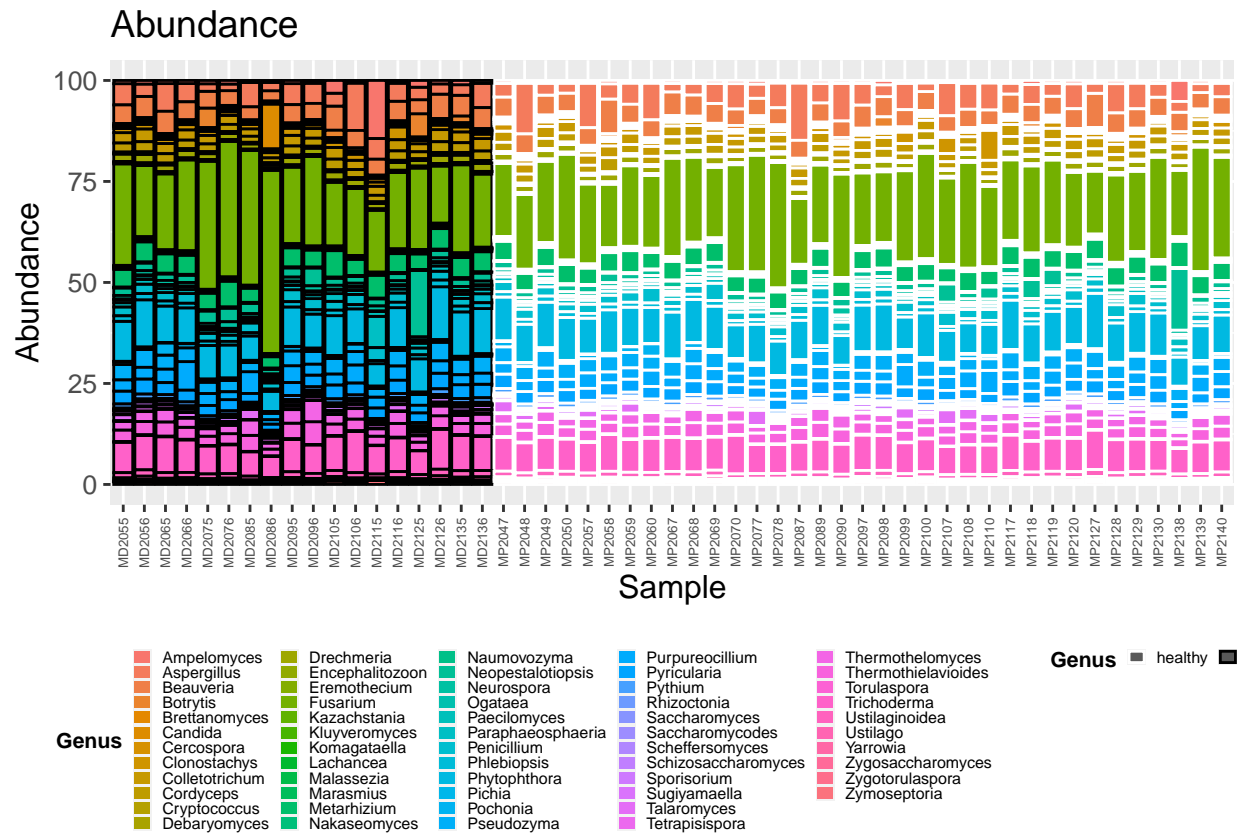
```
## Warning in estimate_richness(physeq, split = TRUE, measures = measures): The data you have provided
## any singletons. This is highly suspicious. Results of richness
## estimates (for example) are probably unreliable, or wrong, if you have already
## trimmed low-abundance taxa from the data.
##
## We recommended that you find the un-trimmed data and retry.
```



— Eukarya by Genero

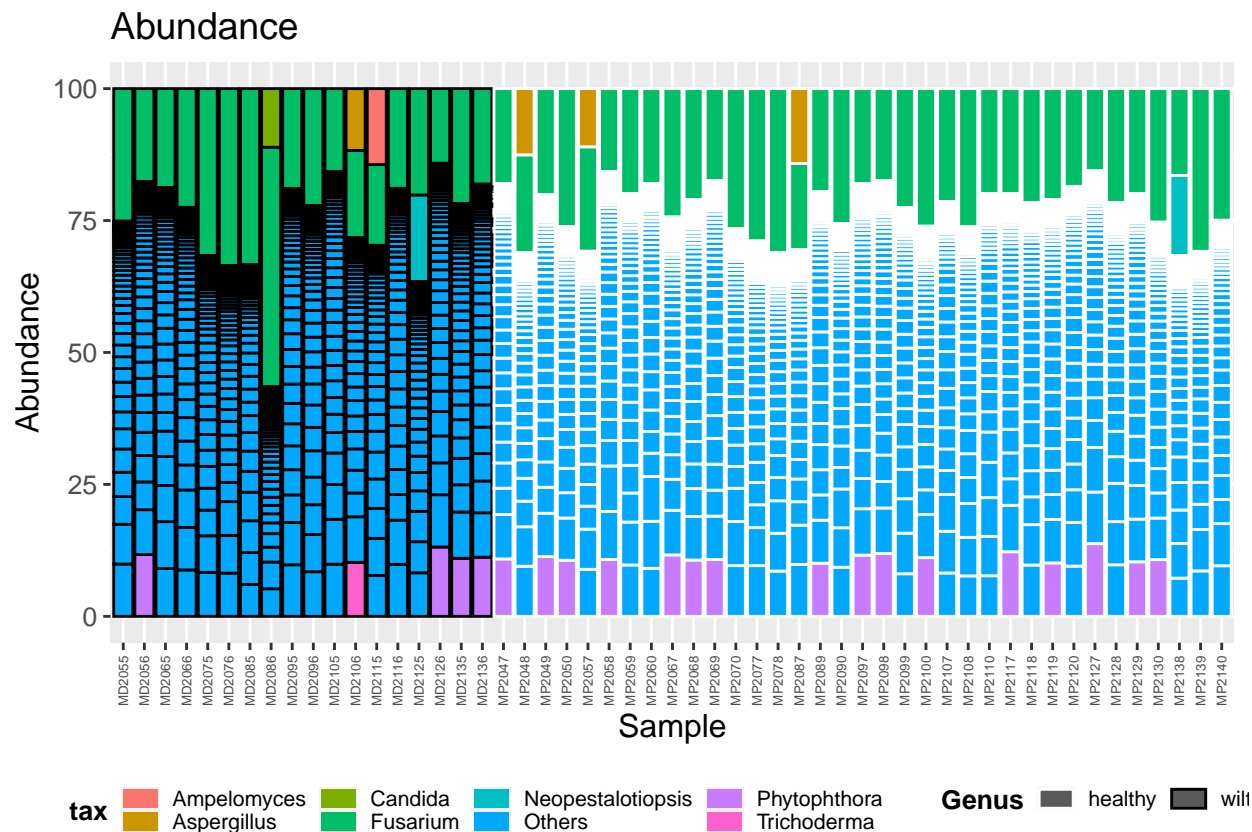
```
Barras_Species <- Abundance_barras(merge_Eukaryota, 'Genus', 'Treatment', 10.0)
Barras_Species[1]
```

[[1]]



Barras_Species [2]

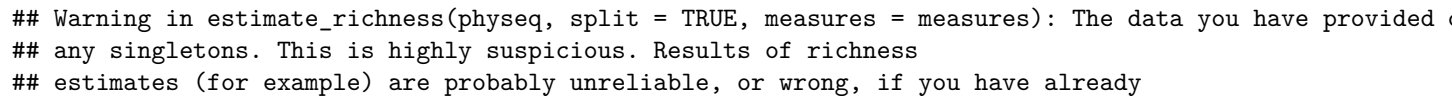
[[1]]



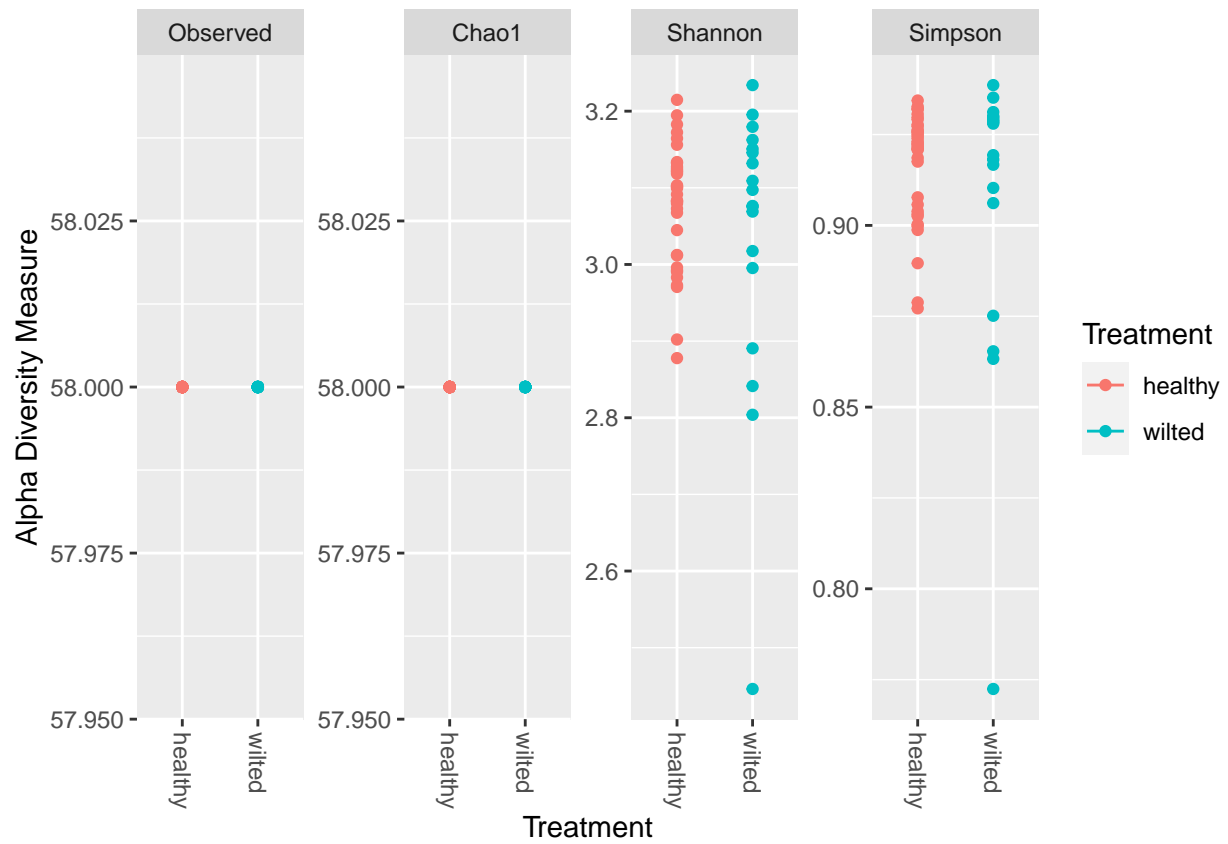
```
Beta_diversity(merge_Eukaryota , 'Genus' , 'Treatment', 'bray')
```

```
## Wisconsin double standardization
## Run 0 stress 0.110709
## Run 1 stress 0.1107087
## ... New best solution
## ... Procrustes: rmse 0.0007840091 max resid 0.004488343
## ... Similar to previous best
## Run 2 stress 0.1223849
## Run 3 stress 0.4035873
## Run 4 stress 0.1223838
## Run 5 stress 0.1107094
## ... Procrustes: rmse 0.0003365426 max resid 0.00192326
## ... Similar to previous best
## Run 6 stress 0.1107089
## ... Procrustes: rmse 0.000132357 max resid 0.0007613431
## ... Similar to previous best
## Run 7 stress 0.1223844
## Run 8 stress 0.1107097
## ... Procrustes: rmse 0.0004018827 max resid 0.002457857
## ... Similar to previous best
## Run 9 stress 0.1107095
## ... Procrustes: rmse 0.000934129 max resid 0.005321928
## ... Similar to previous best
## Run 10 stress 0.110709
```

```
Alpha_diversity(merge_Eukaryota , 'Genus' , 'Treatment')
```



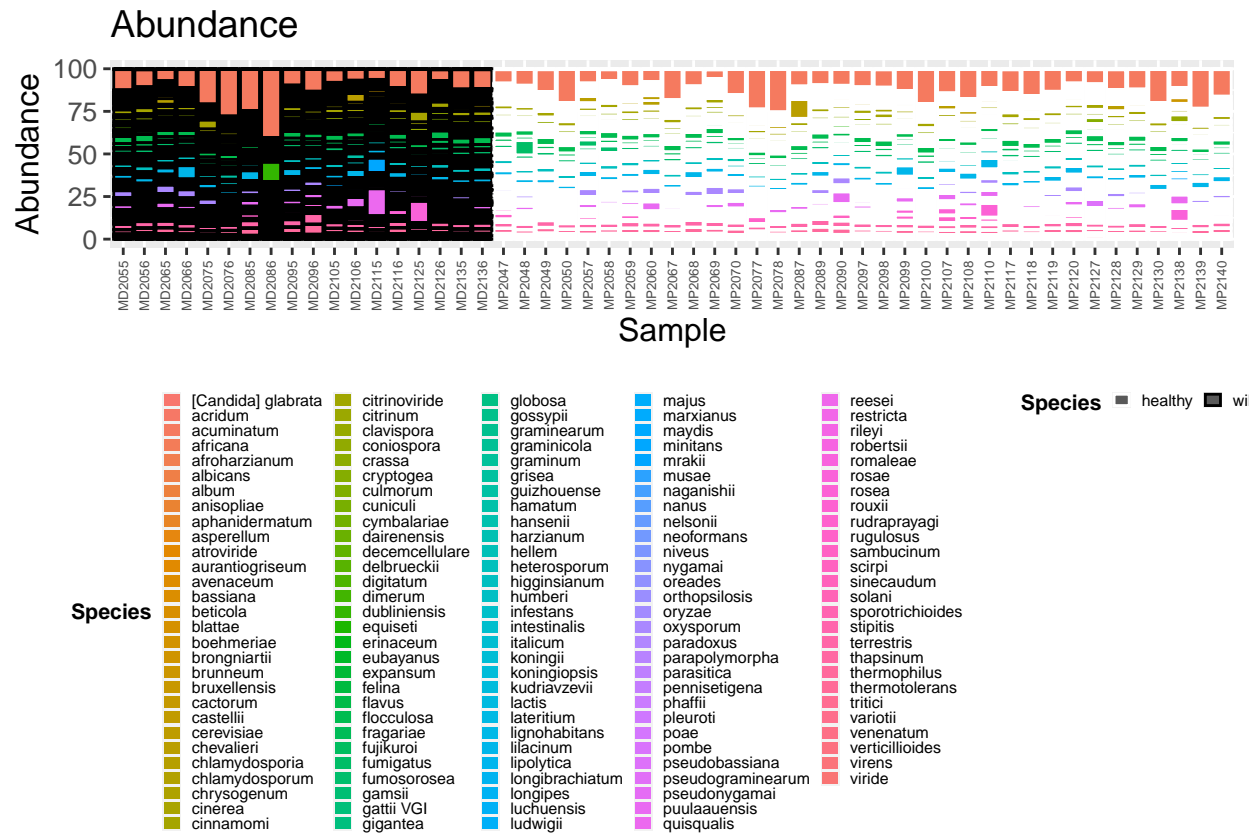

```
## trimmed low-abundance taxa from the data.
##
## We recommended that you find the un-trimmed data and retry.
```



```
##———Eukarya by Species
```

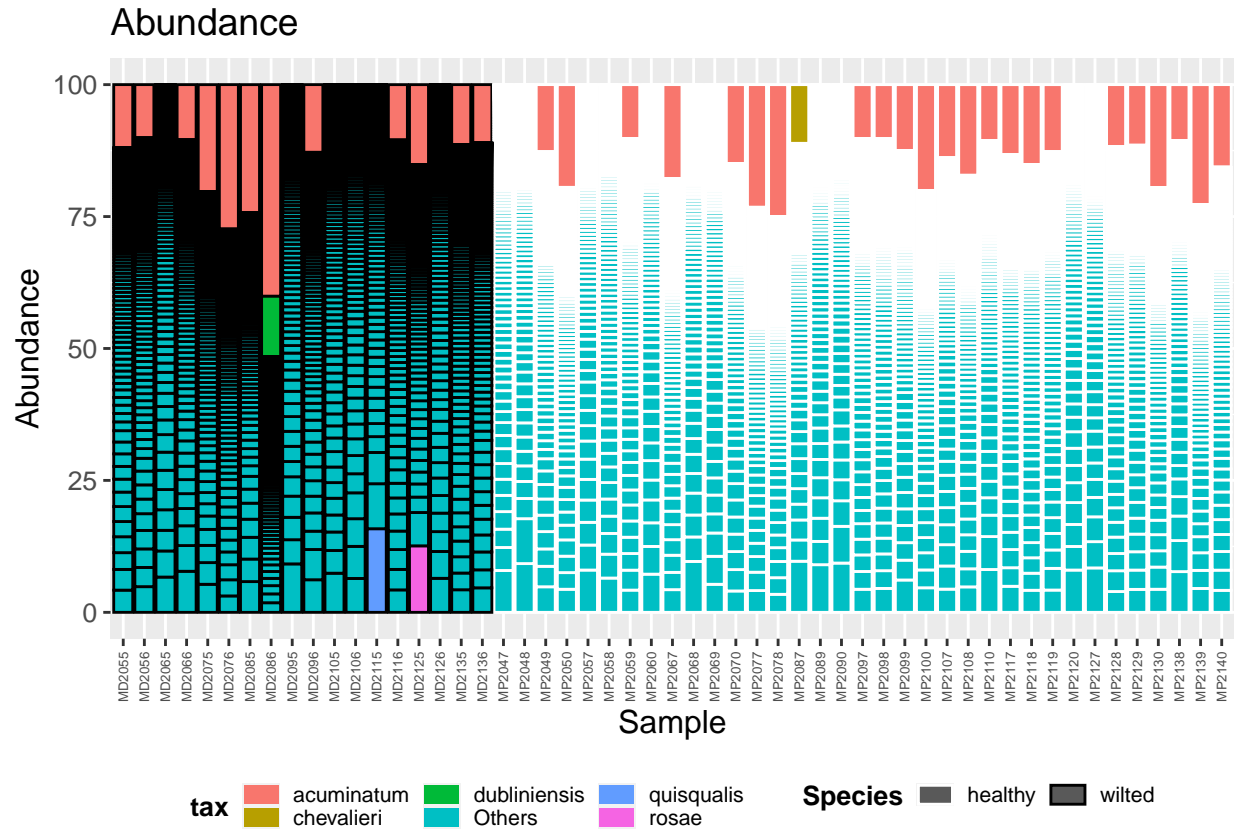
```
Barras_Species <- Abundance_barras(merge_Eukaryota, 'Species', 'Treatment', 10.0)
Barras_Species[1]
```

```
## [[1]]
```



Barras_Species [2]

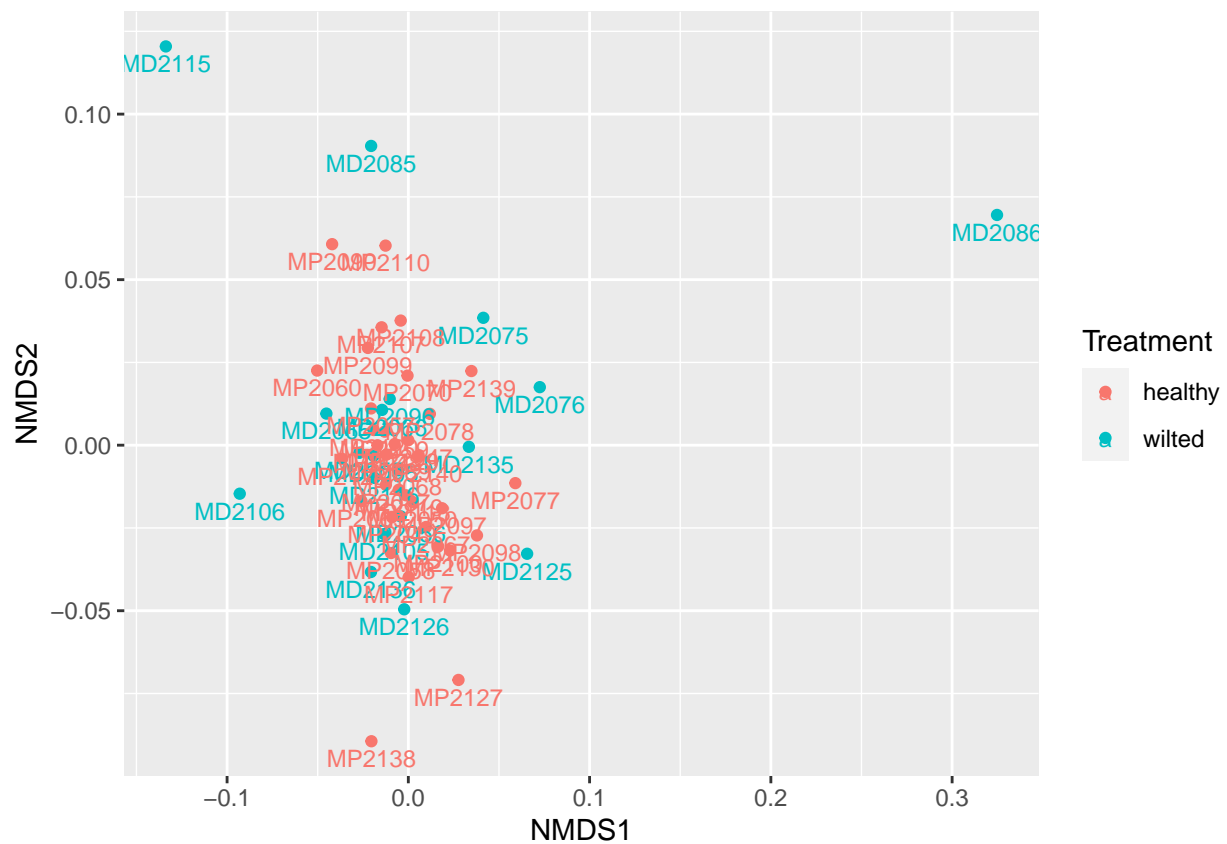
[[1]]

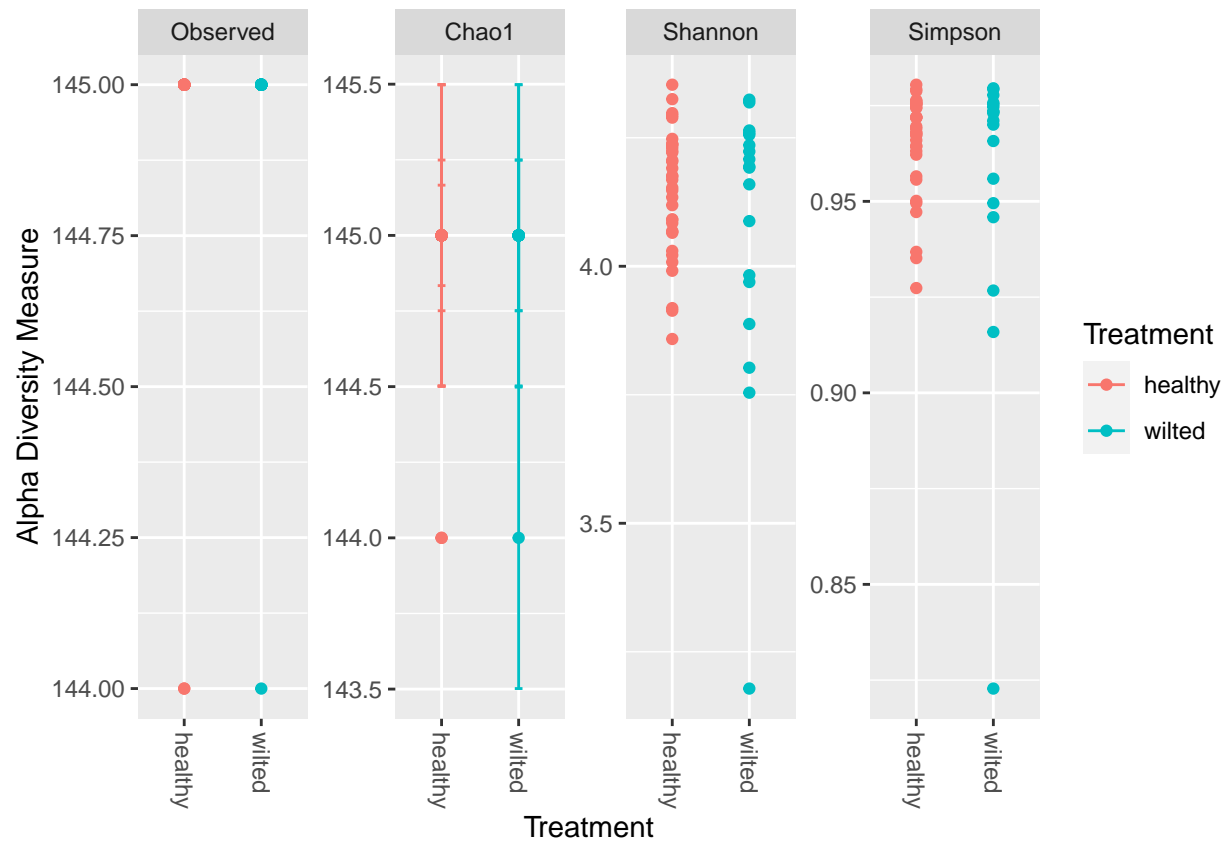


```
Beta_diversity(merge_Eukaryota , 'Species' , 'Treatment', 'bray')
```

```
## Wisconsin double standardization
## Run 0 stress 0.1226512
## Run 1 stress 0.1227531
## ... Procrustes: rmse 0.02315755 max resid 0.1274509
## Run 2 stress 0.1590668
## Run 3 stress 0.1272832
## Run 4 stress 0.1226511
## ... New best solution
## ... Procrustes: rmse 3.546733e-05 max resid 0.0001841906
## ... Similar to previous best
## Run 5 stress 0.1350911
## Run 6 stress 0.1226516
## ... Procrustes: rmse 0.0001415113 max resid 0.0007263343
## ... Similar to previous best
## Run 7 stress 0.1546168
## Run 8 stress 0.1345886
## Run 9 stress 0.135126
## Run 10 stress 0.1520428
## Run 11 stress 0.122651
## ... New best solution
## ... Procrustes: rmse 3.145075e-05 max resid 0.0001960645
## ... Similar to previous best
## Run 12 stress 0.1218544
```

```
Alpha_diversity(merge_Eukaryota , 'Species' , 'Treatment')
```

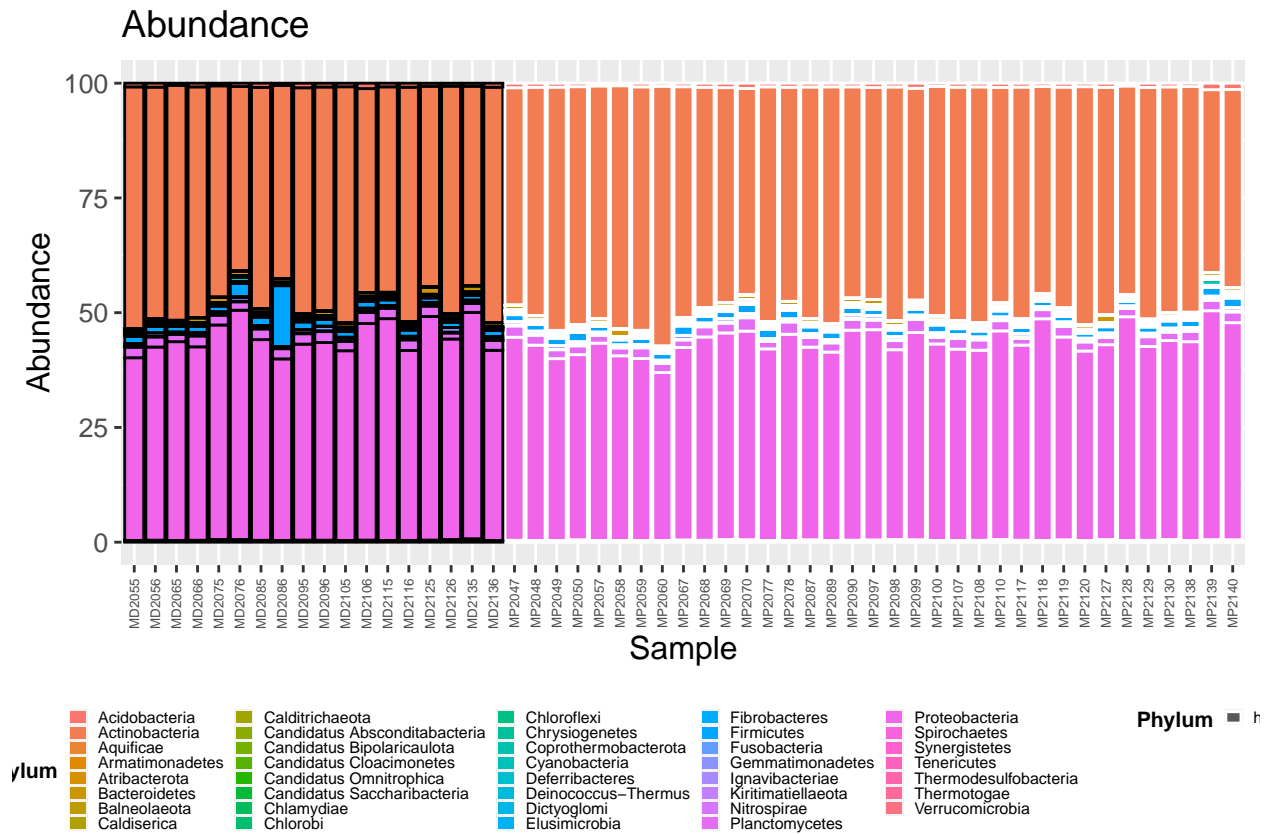




```
## ----- Bacteria by Phylum
```

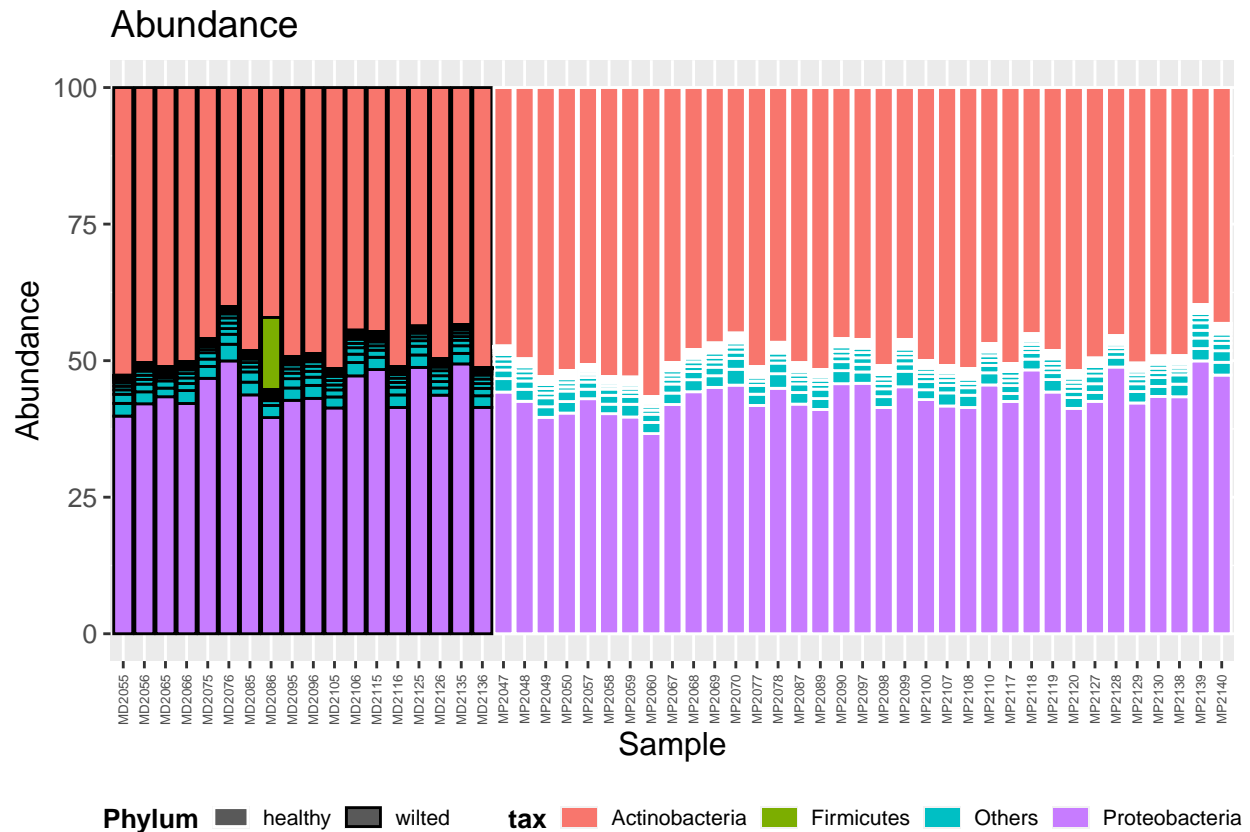
```
Barras_Species <- Abundance_barras(merge_Bacteria, 'Phylum', 'Treatment', 10.0)
Barras_Species[1]
```

```
## [[1]]
```



Barras_Species[2]

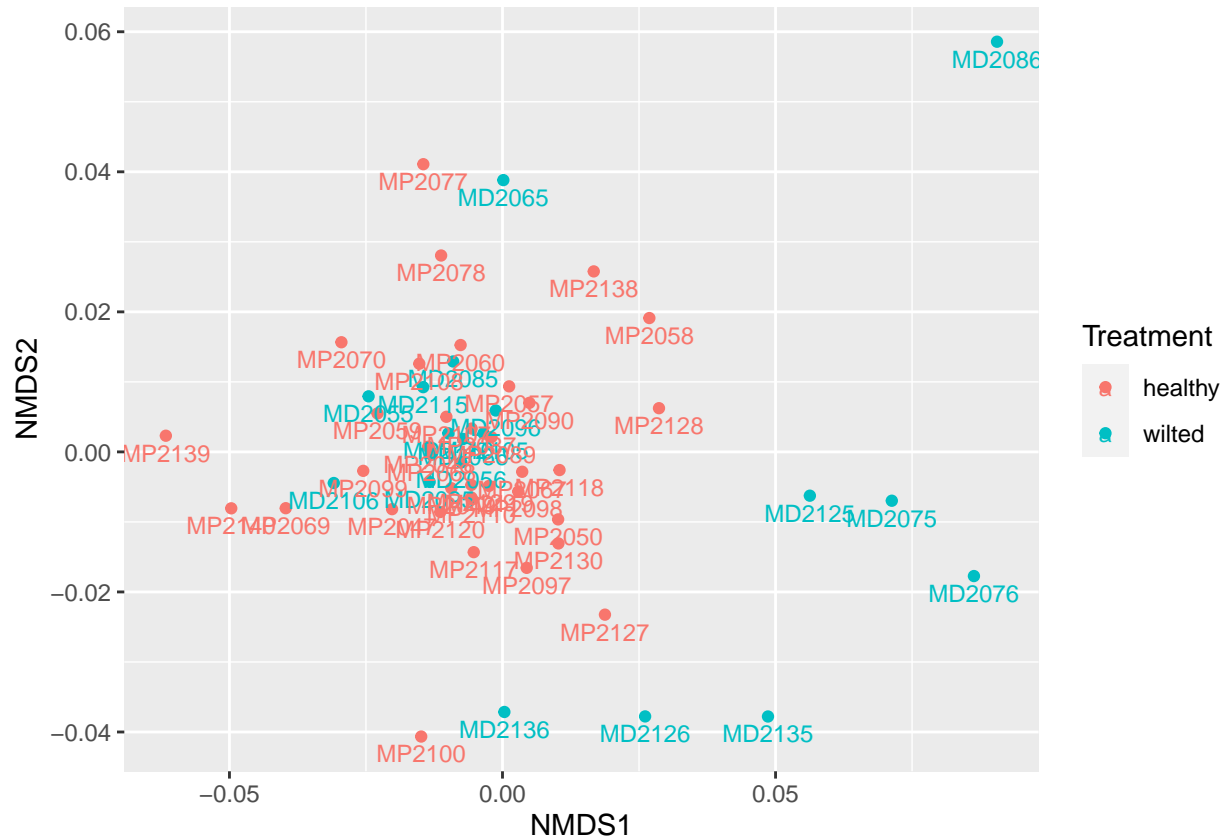
[[1]]



```
Beta_diversity(merge_Bacteria , 'Phylum' , 'Treatment', 'bray')
```

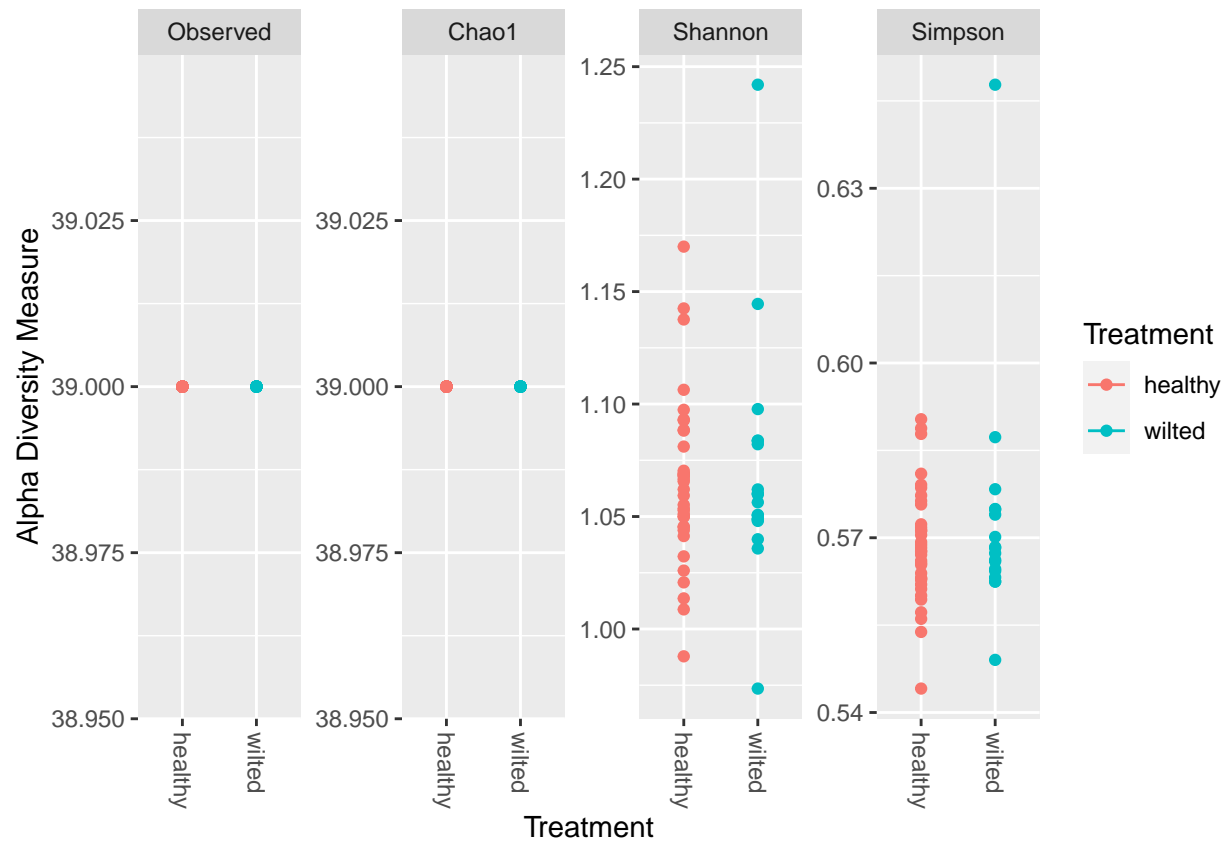
```
## Square root transformation
## Wisconsin double standardization
## Run 0 stress 0.1520004
## Run 1 stress 0.1541159
## Run 2 stress 0.1719744
## Run 3 stress 0.1665614
## Run 4 stress 0.1676962
## Run 5 stress 0.1591786
## Run 6 stress 0.151391
## ... New best solution
## ... Procrustes: rmse 0.008729679 max resid 0.0517727
## Run 7 stress 0.1707262
## Run 8 stress 0.1602944
## Run 9 stress 0.1580849
## Run 10 stress 0.1711338
## Run 11 stress 0.152401
## Run 12 stress 0.1614016
## Run 13 stress 0.1601939
## Run 14 stress 0.1554219
## Run 15 stress 0.1648666
## Run 16 stress 0.1554203
## Run 17 stress 0.1696002
## Run 18 stress 0.1624861
```

```
## Run 19 stress 0.1554219
## Run 20 stress 0.1641734
## *** Best solution was not repeated -- monoMDS stopping criteria:
##      20: stress ratio > sratmax
```



```
Alpha_diversity(merge_Bacteria , 'Phylum' , 'Treatment')
```

```
## Warning in estimate_richness(physeq, split = TRUE, measures = measures): The data you have provided
## any singletons. This is highly suspicious. Results of richness
## estimates (for example) are probably unreliable, or wrong, if you have already
## trimmed low-abundance taxa from the data.
##
## We recommended that you find the un-trimmed data and retry.
```

— Bacteria by Familia

```
Barras_Species <- Abundance_barras(merge_Bacteria, 'Family', 'Treatment', 10.0)
Barras_Species[1]
```

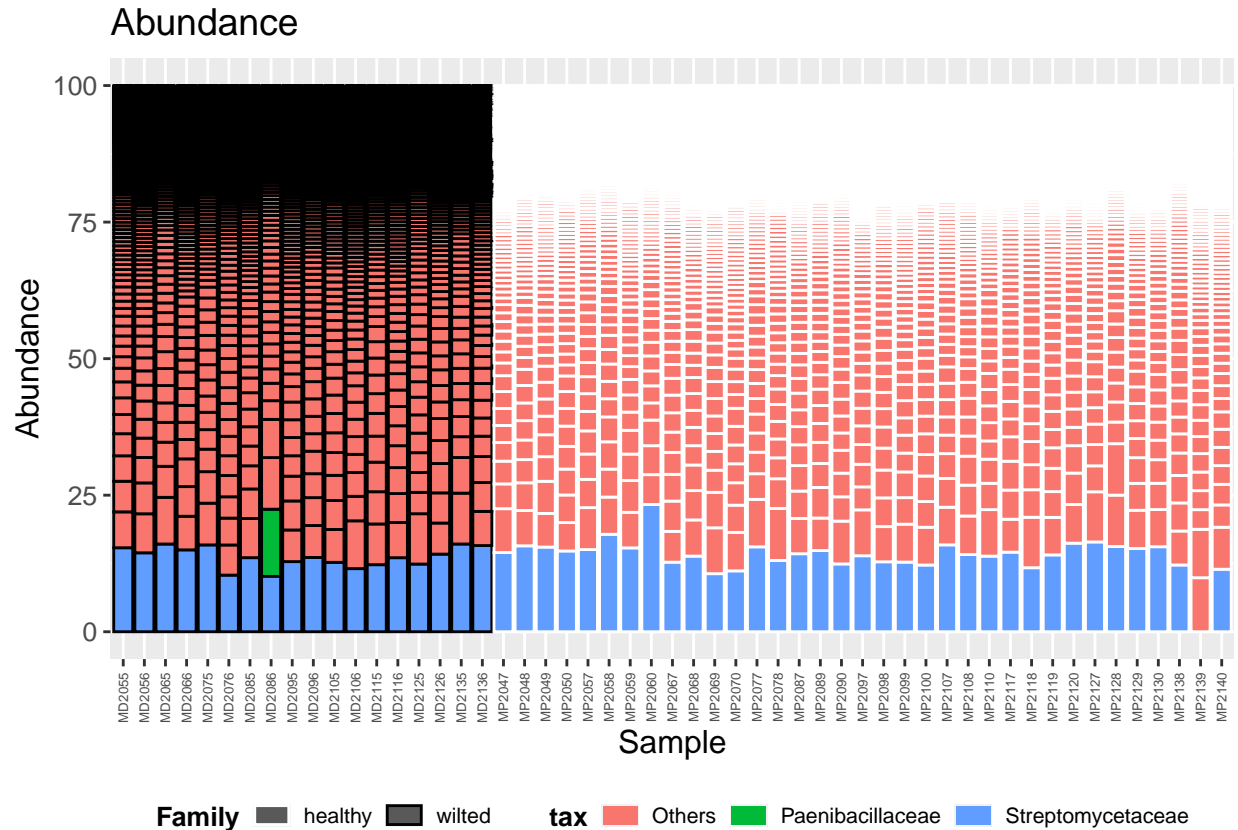
[[1]]

Family

Actinomycesaceae	Chloromonadaceae	Halobacterioviraceae	Nocardiaceae	St
Actinopolymorphaceae	Clostridiaceae	Halobacteroidaceae	Nocardioidaceae	St
Actinopolysporaceae	Clostridiales Family XVI. Incertae Sedis	Halomonadaceae	Nocardiopsaceae	St
Aerococcaceae	Clostridiales Family XVII. Incertae Sedis	Halothiobacillaceae	Nostocaceae	St
Aeromonadaceae	Cohaesibacteraceae	Hapalosiphonaceae	Oceanospirillaceae	St
Akkermansiaceae	Coleofasciculaceae	Helicobacteraceae	Oculatellaceae	St
Alcaligenaceae	Colwelliaceae	Heliothecaceae	Odoribacteraceae	St
Alcanivoracaceae	Comamonadaceae	Holospiraceae	Oleiphilaceae	St
Alicyclobacillaceae	Conexibacteraceae	Hydrogenimonaceae	Opiritaceae	St
Alteromonadaceae	Coprothermobacteraceae	Hydrogenophilaceae	Orbaceae	St
Amoebophilaceae	Coriobacteriaceae	Hydrogenothermaceae	Ornithinimicrobiaceae	Su
Amorphaceae	Corynebacteriaceae	Hyellaceae	Oscillatoriaceae	Su
Anaerohalosphaeraceae	Coxiellaceae	Hymenobacteraceae	Oscillospiraceae	Su
Anaerolineaceae	Crocinitomicaceae	Hyphomicrobiaceae	Oxalobacteraceae	Su
Anaeromyxobacteraceae	Cyclobacteriaceae	Hyphomonadaceae	Paenibacillaceae	Su
Anaplasmataceae	Cytophagaceae	Iamiaceae	Paludibacteraceae	Sy
Aphanizomenonaceae	Deferribacteraceae	Ichthyobacteriaceae	Parachiamydiaceae	Sy
Aphanothecaceae	Dehalococcoidaceae	Idiomarinaceae	Parvibaculaceae	Sy
Aquificaceae	Deinococcaceae	Ignavibacteriaceae	Parvularculaceae	Sy
Archangiaceae	Demeguinaceae	Ilumatobacteraceae	Pasteurellaceae	Sy
Arcobacteraceae	Dermabacteraceae	Immundisolibacteraceae	Pectobacteriaceae	Sy
Ardenticatenaceae	Dermacoccaceae	Intrasporangiaceae	Pelagibacteraceae	Sy
Atopobiaceae	Dermatophilaceae	Isosphaeraceae	Peptococcaceae	Ta
Atribacteraceae	Dermocarpellaceae	Jatrophihabitantaceae	Peptoniphilaceae	Te
Aurantimonadaceae	Desulfallaceae	Jiangellaceae	Peptostreptococcaceae	Te
Azonexaceae	Desulfarculaceae	Jonesiaceae	Petrotoxiaceae	Te
Azospirillaceae	Desulfatobacteriaceae	Kaistiaceae	Phototrophicaceae	Te
Bacillaceae	Desulfobacteriaceae	Kangiellaceae	Phreatobacteraceae	Te
Bacterioviraceae	Desulfobulbaceae	Kineosporiaceae	Phycisphaeraceae	Tr
Bacteroidaceae	Desulfocapsaceae	Kiritimatiellaceae	Phyllobacteriaceae	Tr
Baekduiaceae	Desulfohalobiaceae	Koferiaceae	Pirellulaceae	Tr
Barnesiellaceae	Desulfomicrobiaceae	Koleobacteraceae	Piscirickettsiaceae	Tr
Bartonellaceae	Desulfosudaceae	Kordiimonadaceae	Planctomycetaceae	Tr
Bdellovibrionaceae	Desulfotomaculaceae	Kosmotogaceae	Planococcaceae	Tr
Beijerinckiaceae	Desulfotomaculaceae	Kribbellaceae	Pleomorphomonadaceae	Tr
Bernardetiaceae	Desulfurellaceae	Ktedonosporobacteraceae	Polyangiaceae	Tr
Beutenbergiaceae	Desulfurobacteriaceae	Kytococcaceae	Porphyrimonadaceae	Tr
Bifidobacteriaceae	Desulfuromonadaceae	Labilitrachaceae	Prevotellaceae	Tr
Blastochloridaceae	Devosiaceae	Lachnospiraceae	Prochlorococcaceae	Tr
Blattabacteriaceae	Dictyoglomaceae	Lacipirellulaceae	Prochlorotrichaceae	Tr
Bogoriellaceae	Dietziaceae	Lactobacillaceae	Prolixibacteraceae	Tr
Borrelliaceae	Dissulfurispiraceae	Lawsonellaceae	Promicromonosporaceae	Tr
Boseaceae	Dysgonomonadaceae	Legionellaceae	Propionibacteriaceae	Tr
Brachyspiraceae	Ectothiorhodospiraceae	Leptolyngbyaceae	Proteinivoraceae	Tr
Bradymonadaceae	Eggerthellaceae	Leptospiaceae	Pseudanabaenaceae	Tr
Bradyrhizobiaceae	Egibacteraceae	Leptotrichiaceae	Pseudoalteromonadaceae	Tr
Breoghanaceae	Egicoccaceae	Lichenihabitantaceae	Pseudomonadaceae	Tr
Brevibacteriaceae	Elioraeaceae	Limnochordaceae	Pseudonocardaceae	Tr
Brucellaceae	Elusimicrobiaceae	Listeriaceae	Psychromonadaceae	Tr
Bruguierivoracaceae	Emcibacteraceae	Litoricolaceae	Punicococcaceae	Tr
Bryobacteraceae	Endomicrobiaceae	Magnetococcaceae	Rhabdochlamydiaceae	Tr
Budviciaceae	Endozoicomonadaceae	Mangrovivirgaceae	Rhizobiaceae	Tr
Burkholderiaceae	Enterobacteriaceae	Maricaulaceae	Rhodanobacteraceae	Ti
Caedimonadaceae	Enterococcaceae	Marinifiliaceae	Rhodobacteraceae	Ti
Caldilineaceae	Entomoplasmataceae	Marinifiliaceae	Rhodocyclaceae	Tc
Caldiseriaceae	Erwinaceae	Marinobacteraceae	Rhodospirillaceae	Tr
Calditrichaceae	Erysipelotrichaceae	Mariprofundaceae	Rhodothermaceae	Tr
Calothrixaceae	Erythrobacteraceae	Marinimaceae	Rickettsiaceae	Tr

Barras_Species [2]

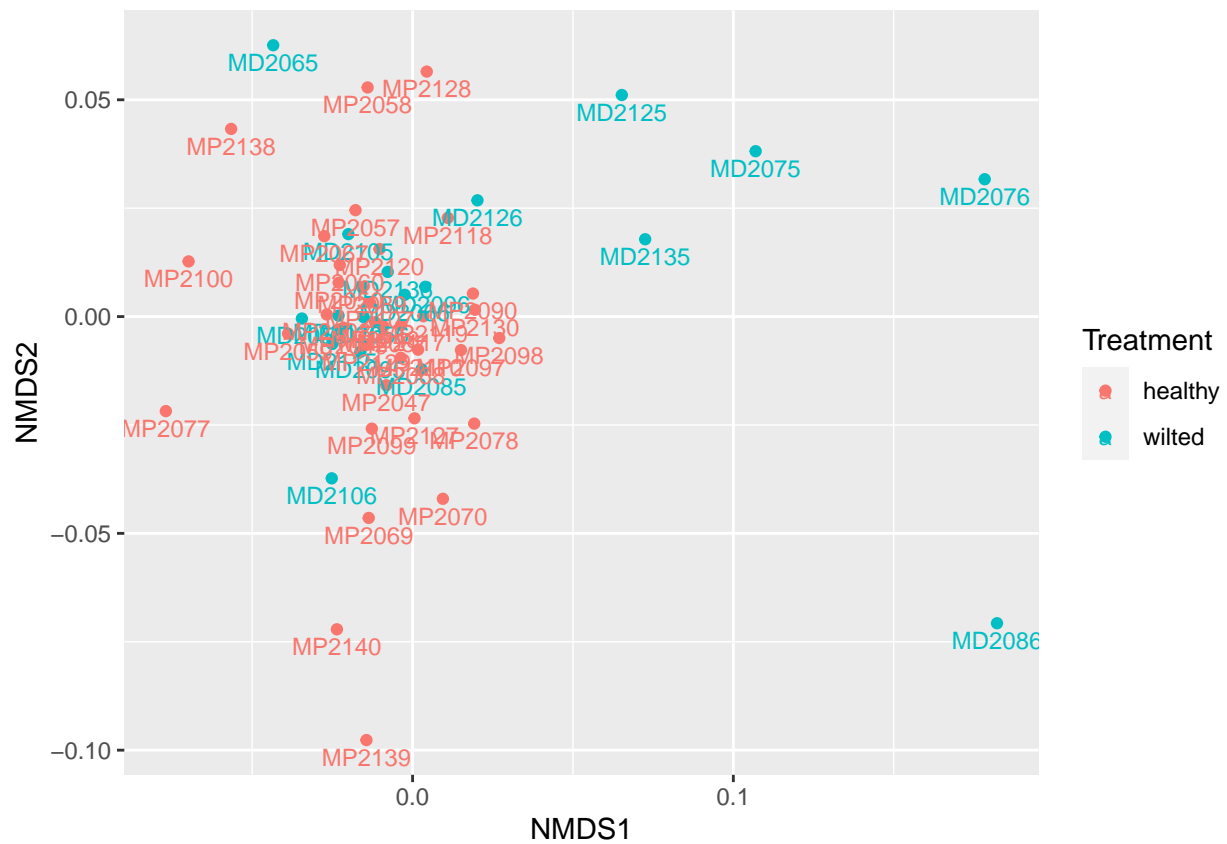
[[1]]



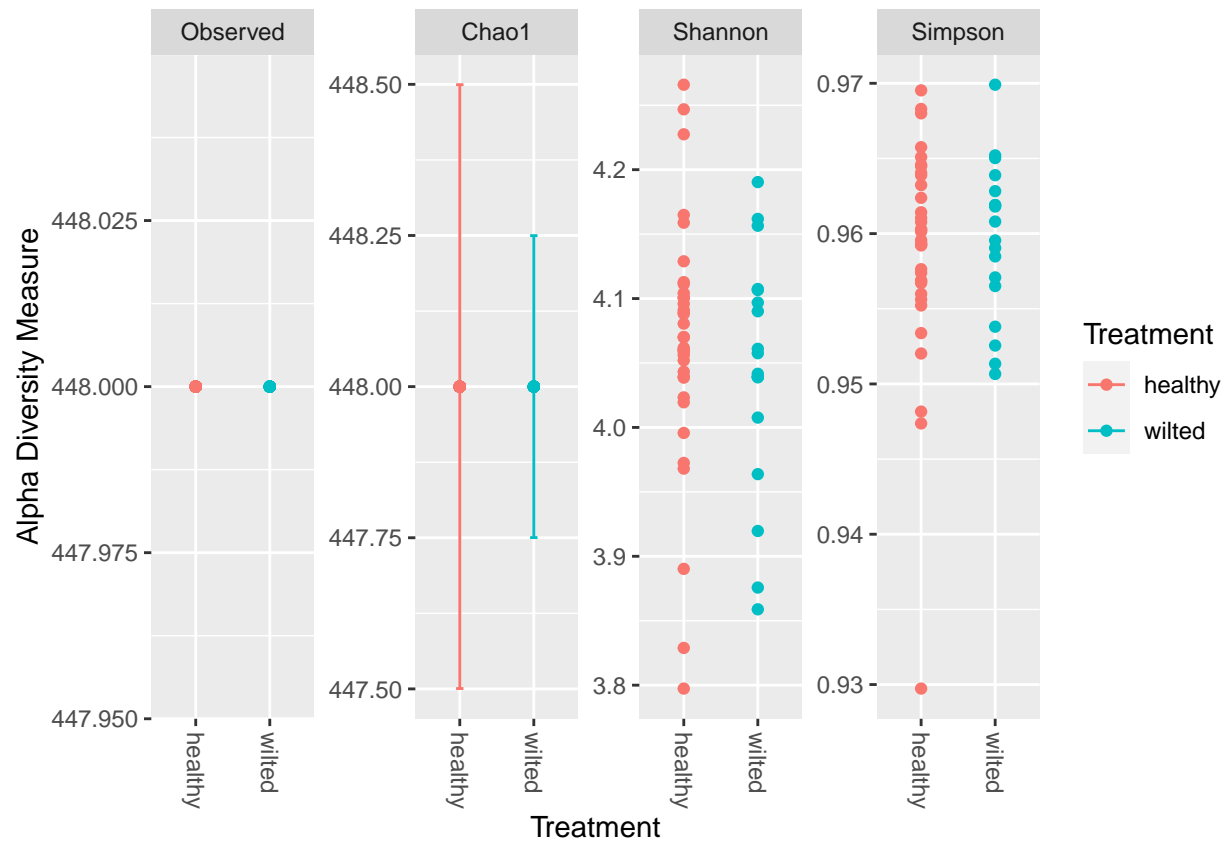
```
Beta_diversity(merge_Bacteria , 'Family' , 'Treatment', 'bray')
```

```
## Wisconsin double standardization
## Run 0 stress 0.1412073
## Run 1 stress 0.1374745
## ... New best solution
## ... Procrustes: rmse 0.06404146 max resid 0.362518
## Run 2 stress 0.1405164
## Run 3 stress 0.1792916
## Run 4 stress 0.1401022
## Run 5 stress 0.1729145
## Run 6 stress 0.1370379
## ... New best solution
## ... Procrustes: rmse 0.01572709 max resid 0.05915357
## Run 7 stress 0.1391838
## Run 8 stress 0.1390465
## Run 9 stress 0.1721853
## Run 10 stress 0.1400264
## Run 11 stress 0.1412067
## Run 12 stress 0.1374452
## ... Procrustes: rmse 0.01063197 max resid 0.04902825
## Run 13 stress 0.1570078
## Run 14 stress 0.1401021
## Run 15 stress 0.14556
## Run 16 stress 0.137194
```

```
## ... Procrustes: rmse 0.01158332  max resid 0.05694715
## Run 17 stress 0.1704098
## Run 18 stress 0.1441743
## Run 19 stress 0.1371941
## ... Procrustes: rmse 0.01162238  max resid 0.0569763
## Run 20 stress 0.1517993
## *** Best solution was not repeated -- monoMDS stopping criteria:
##      20: stress ratio > sratmax
```



```
Alpha_diversity(merge_Bacteria , 'Family' , 'Treatment')
```



```
## ----- Bacteria by Genero
```

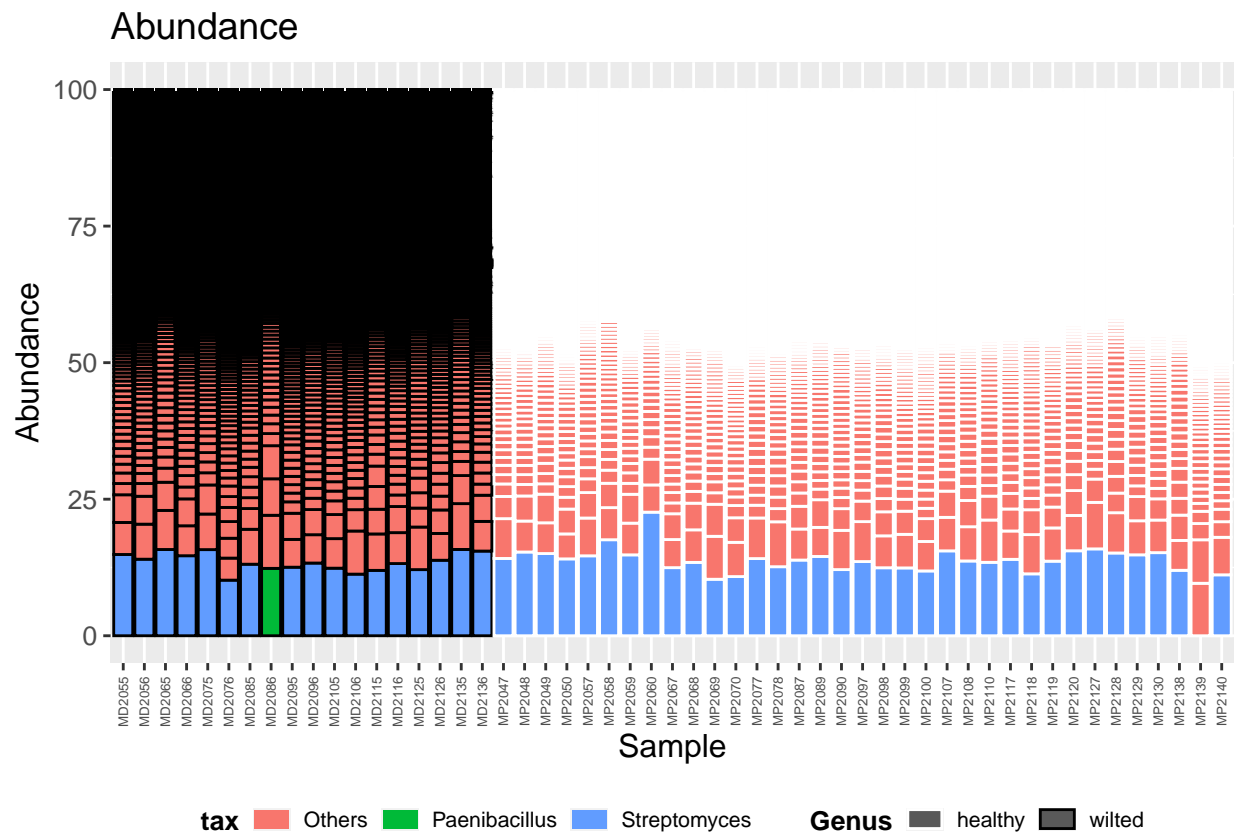
```
Barras_Species <- Abundance_barras(merge_Bacteria, 'Genus', 'Treatment', 10.0)
Barras_Species[1]
```

```
## [[1]]
```

lex	Desulfuricoccus	Labyrinthula	Paramecium	Sulfolobus
flexum	Desulfatibacillum	Labrys	Paroceanicella	Succinivibrio
habitans	Desulfatobacterium	Laceyella	Parolsenella	Suicoccus
luna	Desulfobacca	Lachnoanaerobaculum	Parvibaculum	Sulfidibacter
marina	Desulfobacter	Lachnoclostridium	Parvimonas	Sulfitobacter
ncola	Desulfobacula	Lachnospira	Parvularcula	Sulfuricaulis
rhabdus	Desulfobulbus	Lacibacter	Pasteurella	Sulfuricella
rufa	Desulfocapsa	Lacimicrobium	Paucibacter	Sulfuricurvum
salimonas	Desulfococcus	Lacinutrix	Paucilactobacillus	Sulfuriferula
salinus	Desulfocurvibacter	Lacipirellula	Pauljensenia	Sulfuriflexus
sediminimonas	Desulfofarcimen	Lacrimispora	Pectinatus	Sulfurifustis
sphaera	Desulfoglaeba	Lacticaseibacillus	Pectobacterium	Sulfurihydrogenibium
talea	Desulfoglobium	Lactiplantibacillus	Pediococcus	Sulfurimicrobium
ibacter	Desulfoluna	Lactobacillus	Pedobacter	Sulfurimonas
hidicoccus	Desulfolutivibrio	Lactococcus	Pedococcus	Sulfuriroseicoccus
hnia	Desulfomarina	Lacunisphaera	Pelagerythrobacter	Sulfuritalea
nobacterium	Desulfomicrobium	Lancefieldella	Pelagibacterium	Sulfuritortus
angium	Desulfomonile	Lapidilactobacillus	Pelagovum	Sulfurivermis
bacter	Desulfonema	Laribacter	Pelostegia	Sulfurospirillum
cibacterium	Desulforamulus	Larkinella	Pelobacter	Sulfurovum
ibacter	Desulforapulum	Latilactobacillus	Pelodictyon	Sutcliffeella
imonas	Desulfosarcina	Lautropia	Pelolinea	Sutterella
ratoleum	Desulfoscipio	Lawsonella	Pelosinus	Suttonella
nicrococcus	Desulfosediminicola	Lawsonia	Pengzhengrongella	Swingsia
nophonus	Desulfosporosinus	Leadbetterella	Peptacetobacter	Symbiobacterium
obacter	Desulfosudis	Leclercia	Peptoclostridium	Symmachella
a	Desulfotalea	Lederbergia	Peptoniphilus	Synechococcus
acaulis	Desulfotomaculum	Leeuwenhoekella	Peribacillus	Synechocystis
lassotoga	Desulfovibrio	Legionella	Periweissella	Syntrophobacter
tibacter	Desulfurispirillum	Leifsonia	Permianibacter	Syntrophobotulus
obium	Desulfurivibrio	Leisingera	Persephonella	Syntrophomonas
acter	Desulfurobacterium	Lelliottia	Persicimonas	Syntrophotalea
ntiibacter	Desulfuromonas	Leminorella	Peteryoungia	Syntrophothermus
ntimicrobium	Devosia	Lentibacillus	Petrimonas	Syntrophus
ntimonas	Devriesea	Lentilactobacillus	Petrocella	Tabrizicola
ticoccus	Dialister	Lentilitoribacter	Petrotoga	Tamiana
imonas	Diaminobutyricimonas	Lentzea	Phaeobacter	Tannerella
liella	Diaphorobacter	Leptodesmis	Phascolarctobacterium	Tardibacter
idibacter	Dichelobacter	Leptolyngbya	Phenyllobacterium	Tardiphaga
wickia	Dickeya	Leptosira	Phnomibacter	Tateyamaria
acterium	Dictyoglomus	Leptospirillum	Phocaeicola	Tatlockia
rcus	Dietzia	Leptothermofonsia	Phoenicibacter	Tatumella
hizobium	Dinoroseobacter	Leptothrix	Photobacterium	Tautonia
pira	Dissulfurimicrobium	Leptotrichia	Photorhabdus	Taylorella
pirillum	Dissulfurispira	Leucobacter	Phototrophicus	Telmatocola
obacter	Dokdonella	Leuconostoc	Phreatobacter	Tenacibaculum
llus	Dokdonia	Levilactobacillus	Phycococcus	Tenuifilum
eriplanes	Dolichospermum	Liberibacter	Phycisphaera	Tepidanaerobacter
eriovorax	Dolosigranulum	Lichenicola	Phyllobacterium	Tepidibacter
eroides	Dongshaeta	Lichenihabitans	Phytobacter	Tepidiforma
duia	Dorea	Ligilactobacillus	Phytohabitans	Tepidimonas
esiella	Draconibacterium	Lignipirellula	Pigmentiphaga	Tepiditoga
onella	Duganella	Limihaloglobus	Pikeienueella	Terasakiella
lea	Duncaniella	Limnobacter	Pimelobacter	Teredinibacter
lovibrio	Dyadobacter	Limnobaculum	Pirellula	Terribacillus
giatota	Dyella	Limnochorda	Pirellulimonas	Terricaulis
irickia	Dysgonomonas	Limnoglobus	Piscirickettsia	Terriglobus
	Dysnomobacter	Limnolobus	Piscirickettsia	Terrirhabdus

Barras_Species [2]

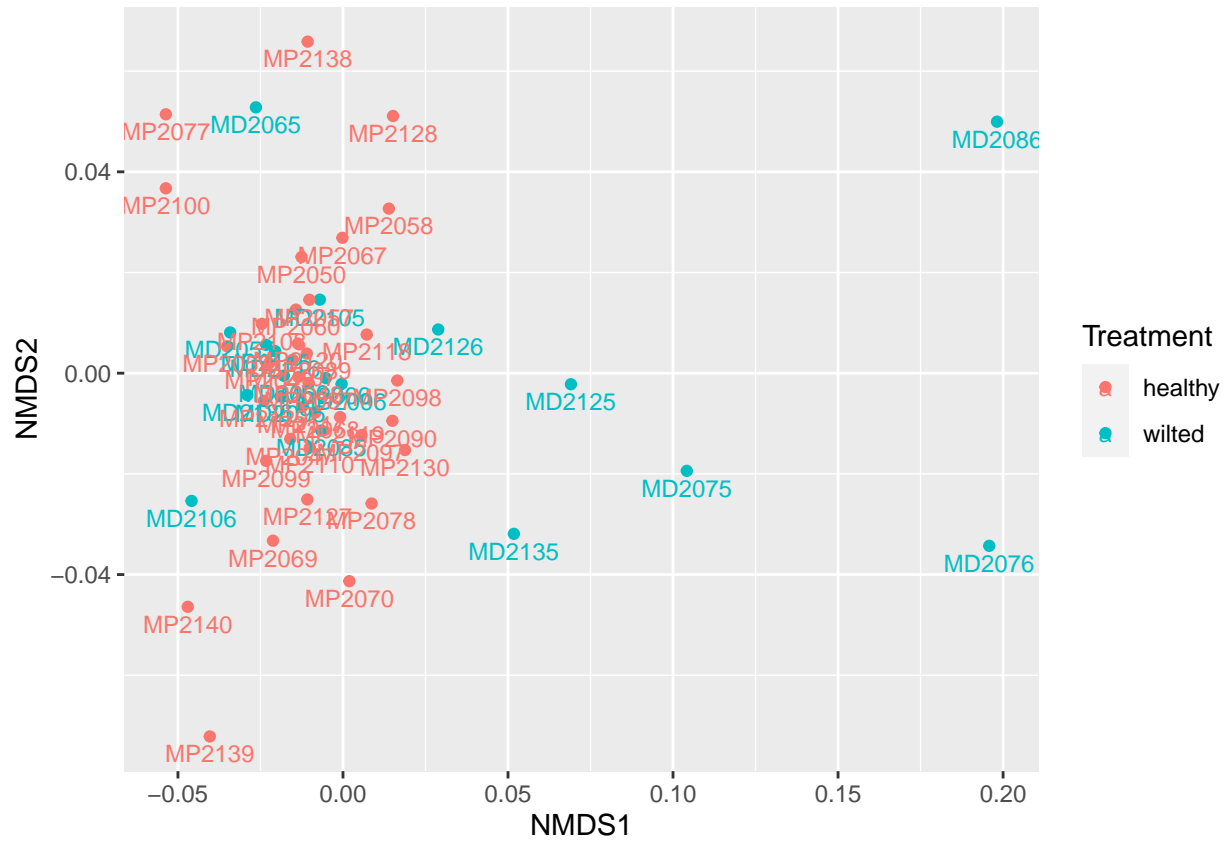
[[1]]



```
Beta_diversity(merge_Bacteria , 'Genus' , 'Treatment', 'bray')
```

```
## Wisconsin double standardization
## Run 0 stress 0.1467945
## Run 1 stress 0.1432835
## ... New best solution
## ... Procrustes: rmse 0.08892711 max resid 0.3292679
## Run 2 stress 0.1624125
## Run 3 stress 0.1452004
## Run 4 stress 0.1476409
## Run 5 stress 0.1490739
## Run 6 stress 0.1438418
## Run 7 stress 0.1481202
## Run 8 stress 0.1437861
## Run 9 stress 0.1480106
## Run 10 stress 0.1490668
## Run 11 stress 0.1476747
## Run 12 stress 0.1511661
## Run 13 stress 0.1432842
## ... Procrustes: rmse 0.0008987919 max resid 0.003811659
## ... Similar to previous best
## Run 14 stress 0.1638381
## Run 15 stress 0.1519965
## Run 16 stress 0.1476188
## Run 17 stress 0.1469869
```

```
## Run 18 stress 0.1578524
## Run 19 stress 0.1547472
## Run 20 stress 0.1500188
## *** Best solution repeated 1 times
```

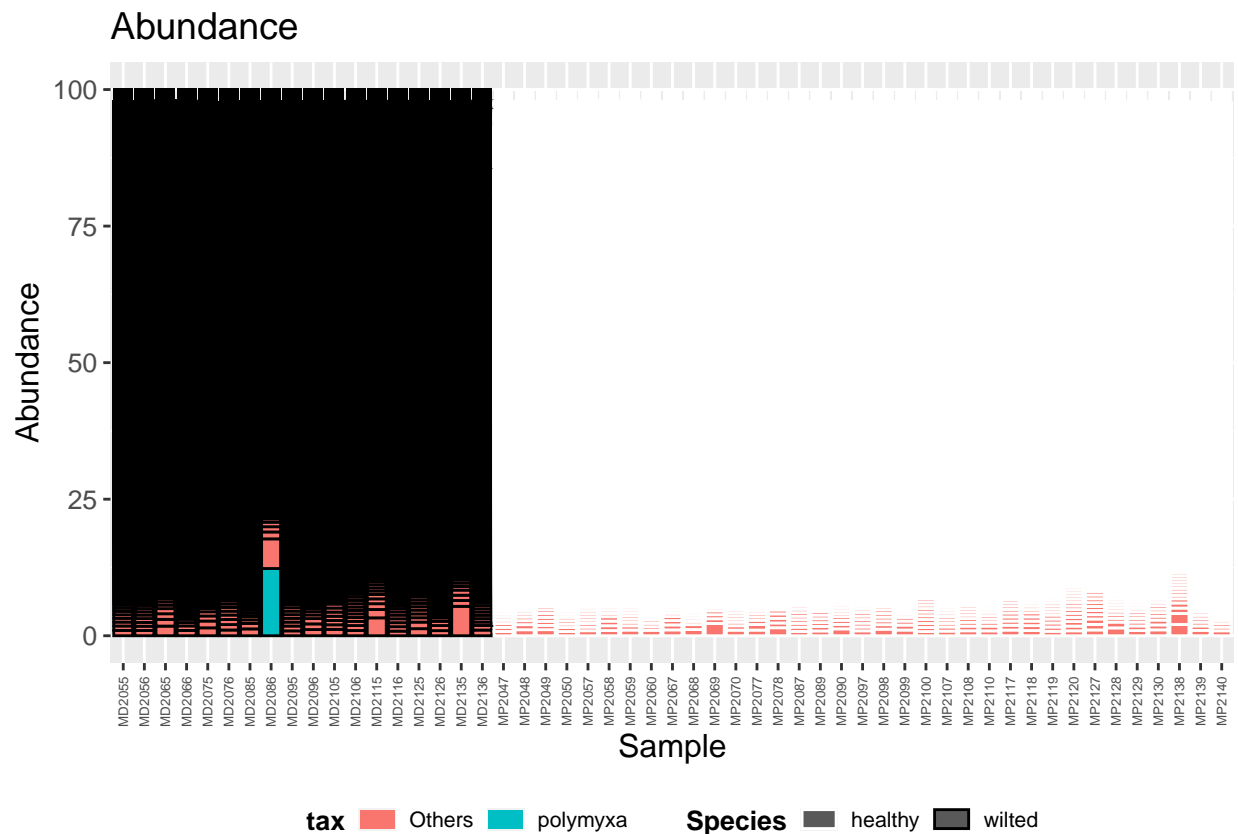


```
Alpha_diversity(merge_Bacteria , 'Genus' , 'Treatment')
```


e		lactamica	sp. 'Soap Lake #7'	sp. KSB-15	sp. X
		lactarius	sp. (Blaberus giganteus)	sp. KSM-R2A25	sp. X
		lactatifermentans	sp. (Blatta orientalis)	sp. KSM-R2A30	sp. X
		lactatiformans	sp. (Blattella germanica)	sp. KT 15	sp. X
		lactis	sp. (Cryptocercus kyebangensis)	sp. KT25b	sp. X
		lactucaae	sp. (Cryptocercus punctulatus) str. Cpu	sp. KTR9	sp. X
		lacunae	sp. (ex Adelges kitamiensis)	sp. KUDC0405	sp. X
		lacus	sp. (ex Biomphalaria glabrata)	sp. KUDC0406	sp. X
		lacuslunae	sp. (Mastotermes darwiniensis)	sp. KUDC1026	sp. X
		lacustris	sp. (Nauphoeta cinerea)	sp. KUDC1714	sp. X
		laguerreae	sp. (Periplaneta americana)	sp. KUIN-1	sp. X
		laidlawii	sp. 001	sp. Kuro-4	sp. X
		laixinhei	sp. 007	sp. KW1	sp. X
		laikuanensis	sp. 008	sp. KX20019	sp. X
		laminatus	sp. 02C 26	sp. KY-GH-1	sp. X
		lanienae	sp. 09C 129	sp. KY-YJ-3	sp. X
		lanii	sp. 09RB8471	sp. KY3	sp. Y
		lansingensis	sp. 09RB8910	sp. KY5	sp. Y
		lanthieri	sp. 1_2014MBL_MicDiv	sp. KY70	sp. Y
		lapagei	sp. 1_2015MBL_MicDiv	sp. KY75	sp. Y
		lapidicaptus	sp. 1-1C	sp. L-07	sp. Y
		lapsinanis	sp. 1.5R	sp. L-2-11	sp. Y
		lari	sp. 1(2017)	sp. L-8-10	sp. Y
		lariciata	sp. 103DPR2	sp. L-8-3	sp. Y
		larrymorei	sp. 1063	sp. L-A4	sp. Y
		larvae	sp. 107-1	sp. L12M9	sp. Y
		lata	sp. 1070	sp. L1A13	sp. Y
		latens	sp. 10FS3-1	sp. L1A34	sp. Y
		laterosporus	sp. 10M-3C3	sp. L1A9	sp. Y
		laumondii	sp. 10RB9215	sp. L1139	sp. Y
		lautus	sp. 11-1-2	sp. L1SW	sp. Y
		lavamentivorans	sp. 11-B-312	sp. L2A1	sp. Y
		lavendulae	sp. 113-1-2	sp. L2A11	sp. Y
		leachii	sp. 113-3-3	sp. L3-i22	sp. Y
		leadbetteri	sp. 113-3-9	sp. L3-i23	sp. Y
		leeuwenhoekii	sp. 1137	sp. L3A3	sp. Y
		leguminosarum	sp. 113P3	sp. L3A6	sp. Y
		lehensis	sp. 114	sp. L3A8	sp. Y
		lekithochrous	sp. 11515TR	sp. L5	sp. Y
		lemovicicum	sp. 116-D4	sp. L51/94	sp. Y
		lemurum	sp. 119287	sp. L5B5	sp. Y
		lenghuensis	sp. 11B	sp. L6-1	sp. Y
		lenta	sp. 11kri321	sp. L9-4	sp. Y
		lentiflavum	sp. 12200R-103	sp. LA-2-3-30-S1-D2	sp. Y
		lentis	sp. 13-15	sp. LA112445	sp. Y
		lentocellum	sp. 131	sp. LA31	sp. Y
		lentus	sp. 131-2-1	sp. LAB-08	sp. Y
		leopoldii	sp. 131-2-5	sp. LADL05-105	sp. Y
		leprae	sp. 131-3-5	sp. LAS2	sp. Y
		lepromatosis	sp. 13159349	sp. LB1	sp. Y
		leptomitiformis	sp. 135	sp. LBG001	sp. Y
		lettingae	sp. 140616W15	sp. LBUM 1475	sp. Y
andiella		liangguodongii	sp. 14171R-50	sp. LBUM 1480	sp. Y
ibbosus		liangshanensis	sp. 14181154	sp. LBUM 1482	sp. Y
ifera		liaowanqingii	sp. 143-21	sp. LBUM920	sp. Y
		libanensis	sp. 144S4	sp. LC2018020214	sp. Y
		licheniformis	sp. 15-184	sp. LC6	sp. Y

Barras_Species[2]

[[1]]



```
Beta_diversity(merge_Bacteria , 'Species' , 'Treatment' , 'bray')
```

```
## Wisconsin double standardization
## Run 0 stress 0.1597841
## Run 1 stress 0.1603297
## Run 2 stress 0.1601333
## ... Procrustes: rmse 0.05751254 max resid 0.2742671
## Run 3 stress 0.1746425
## Run 4 stress 0.1682795
## Run 5 stress 0.1745846
## Run 6 stress 0.1595746
## ... New best solution
## ... Procrustes: rmse 0.06179215 max resid 0.2864908
## Run 7 stress 0.1667257
## Run 8 stress 0.1622331
## Run 9 stress 0.1853643
## Run 10 stress 0.1892493
## Run 11 stress 0.1632423
## Run 12 stress 0.1899387
## Run 13 stress 0.1618986
## Run 14 stress 0.1826662
## Run 15 stress 0.1604222
## Run 16 stress 0.1734581
## Run 17 stress 0.1590693
## ... New best solution
```

```
Alpha_diversity(merge_Bacteria , 'Species' , 'Treatment')
```

