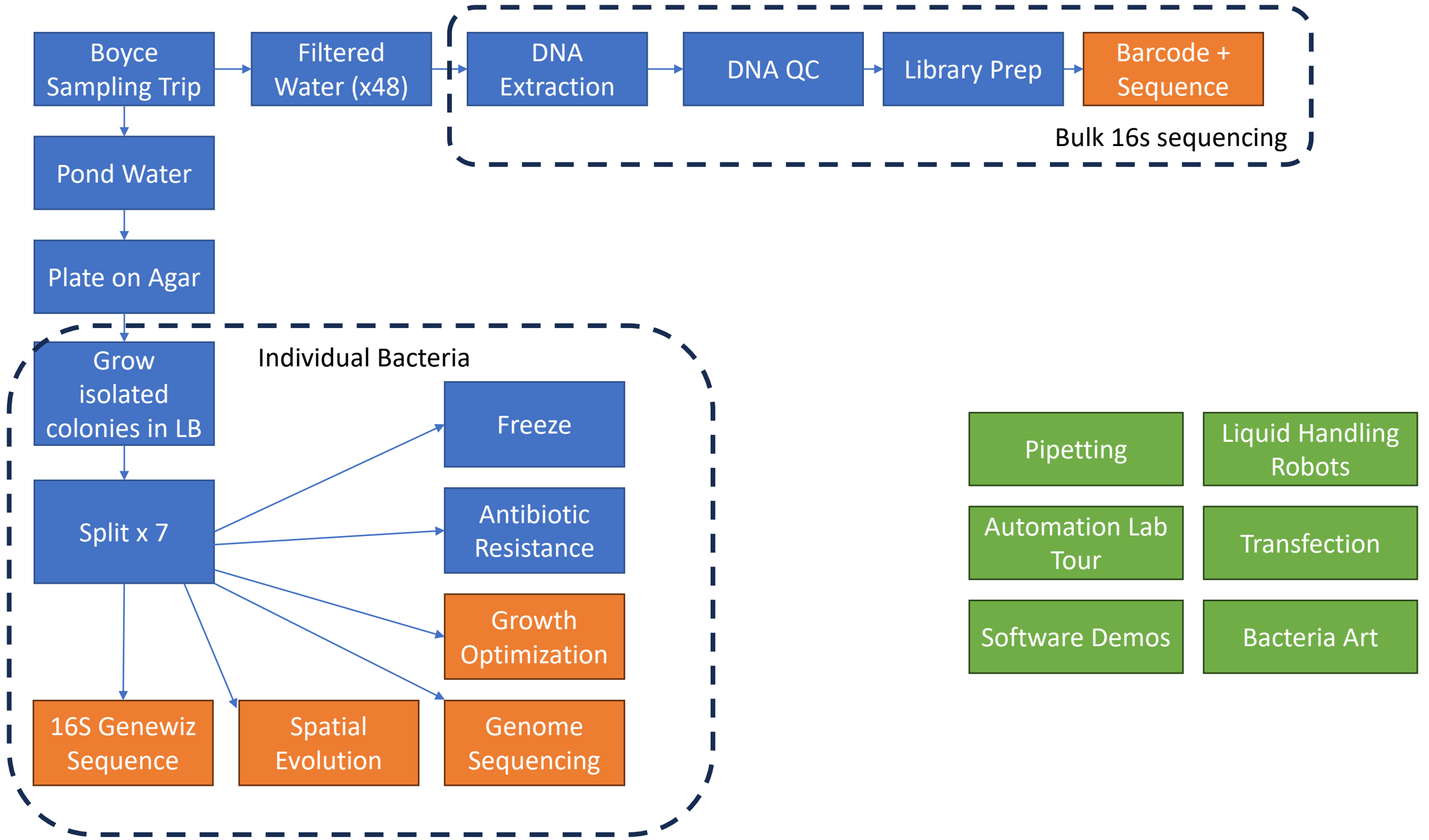
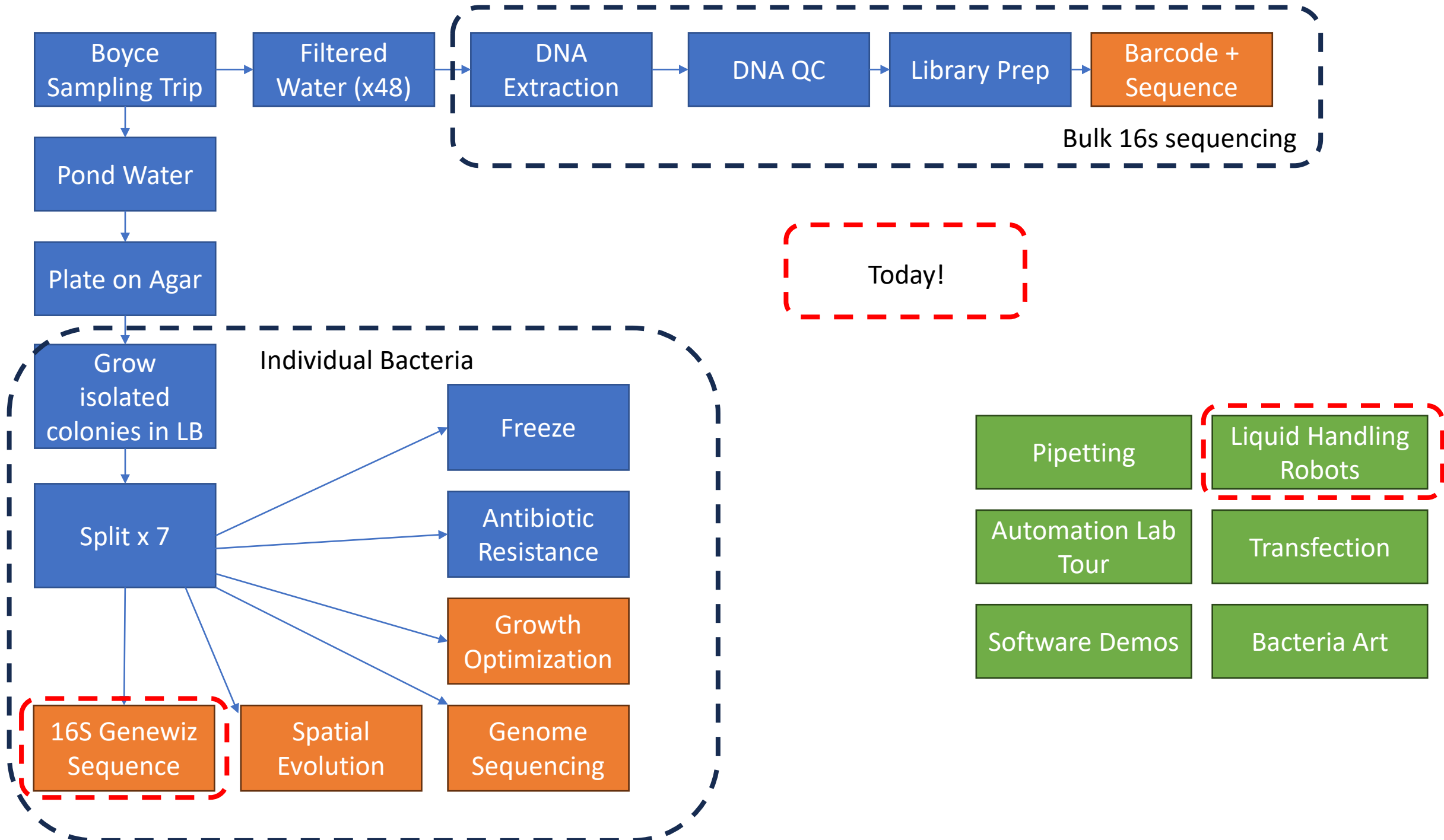
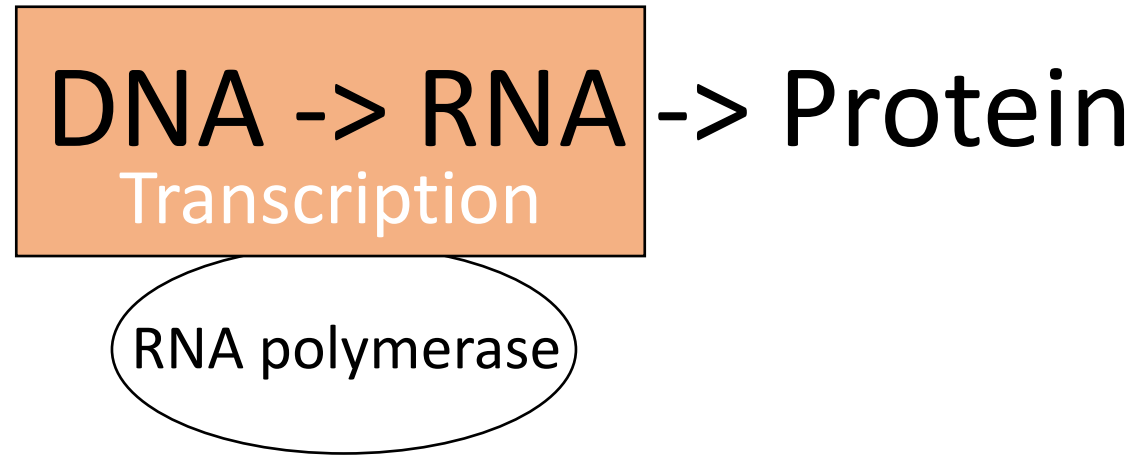


# CompBio Pre-College Lab





# Molecular Biology - Central Dogma



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## article

*Nature* **227**, 561 - 563 (08 August 1970); doi:10.1038/227561a0

## Central Dogma of Molecular Biology

FRANCIS CRICK

MRC Laboratory of Molecular Biology, Hills Road, Cambridge CB2 2QH

**The central dogma of molecular biology deals with the detailed residue-by-residue transfer of sequential information. It states that such information cannot be transferred from protein to either protein or nucleic acid.**

---

# Molecular Biology - Central Dogma

DNA -> RNA -> Protein  
Translation

Ribosome

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## article

*Nature* **227**, 561 - 563 (08 August 1970); doi:10.1038/227561a0

## Central Dogma of Molecular Biology

FRANCIS CRICK

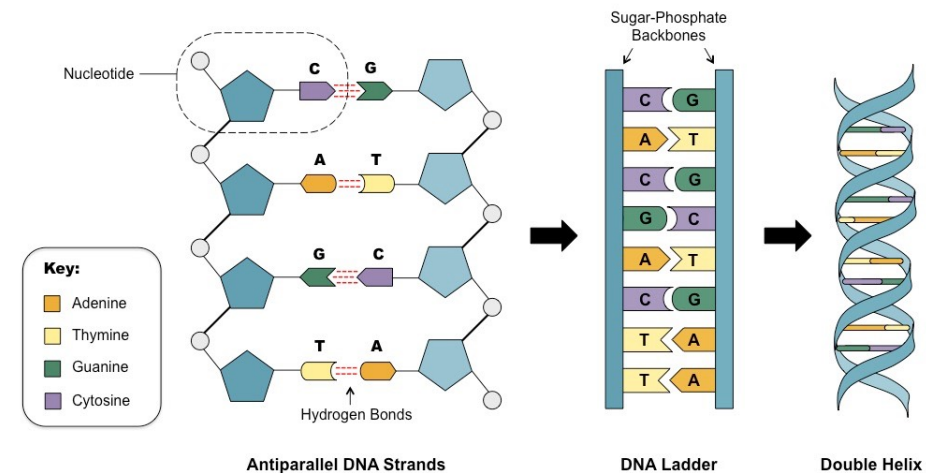
MRC Laboratory of Molecular Biology, Hills Road, Cambridge CB2 2QH

**The central dogma of molecular biology deals with the detailed residue-by-residue transfer of sequential information. It states that such information cannot be transferred from protein to either protein or nucleic acid.**

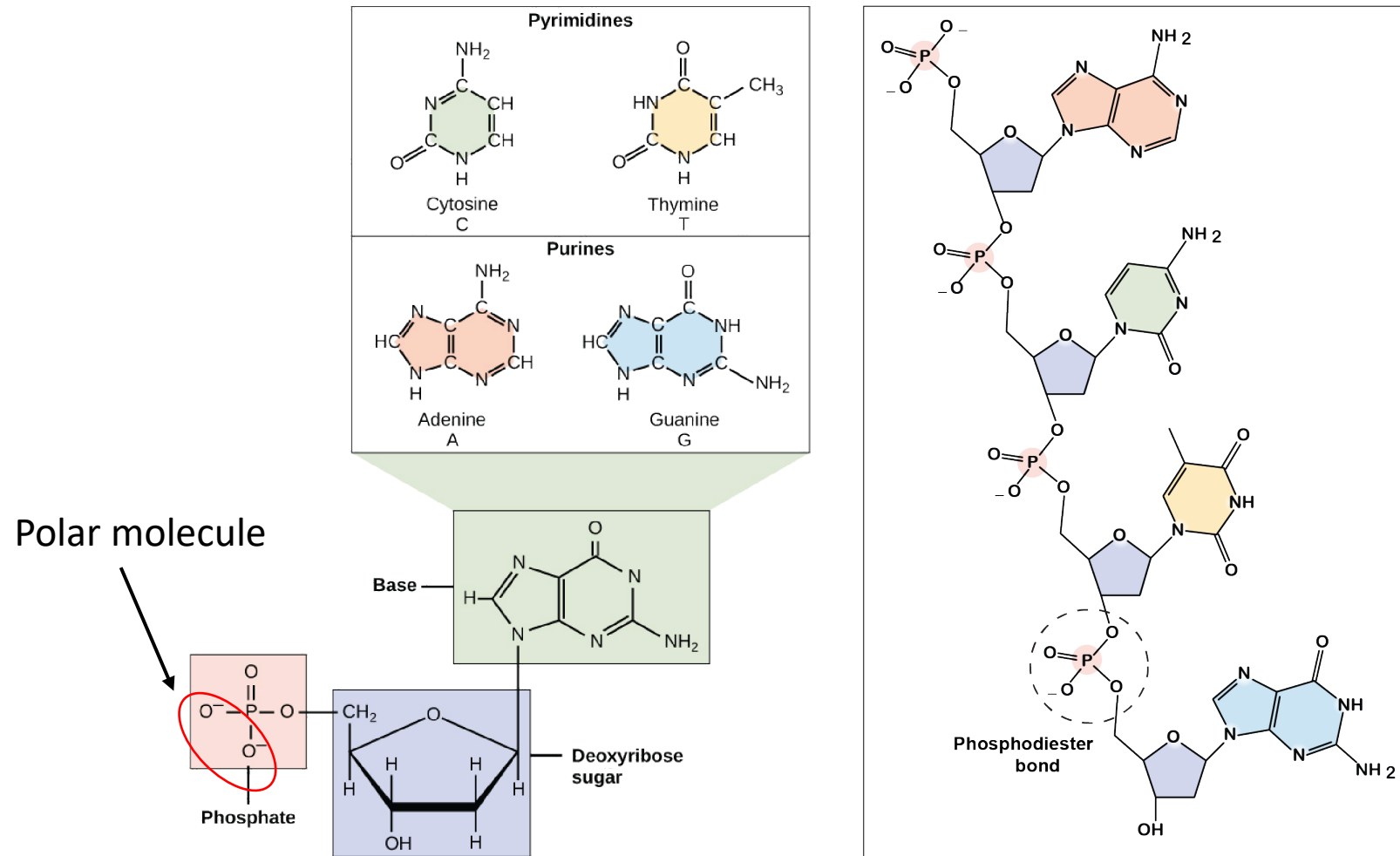
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# DNA Structure

- Molecule carrying genetic information to allow a cell to generate proteins and function.
- Each strand is composed of nucleotides
- Nucleotides:
  - nitrogen containing base (adenine, guanine, cytosine, thymine) (A,G,C,T)
  - Deoxyribose
  - Phosphate group
- Double Helix Structure
  - A  $\leftrightarrow$  T
  - G  $\leftrightarrow$  C



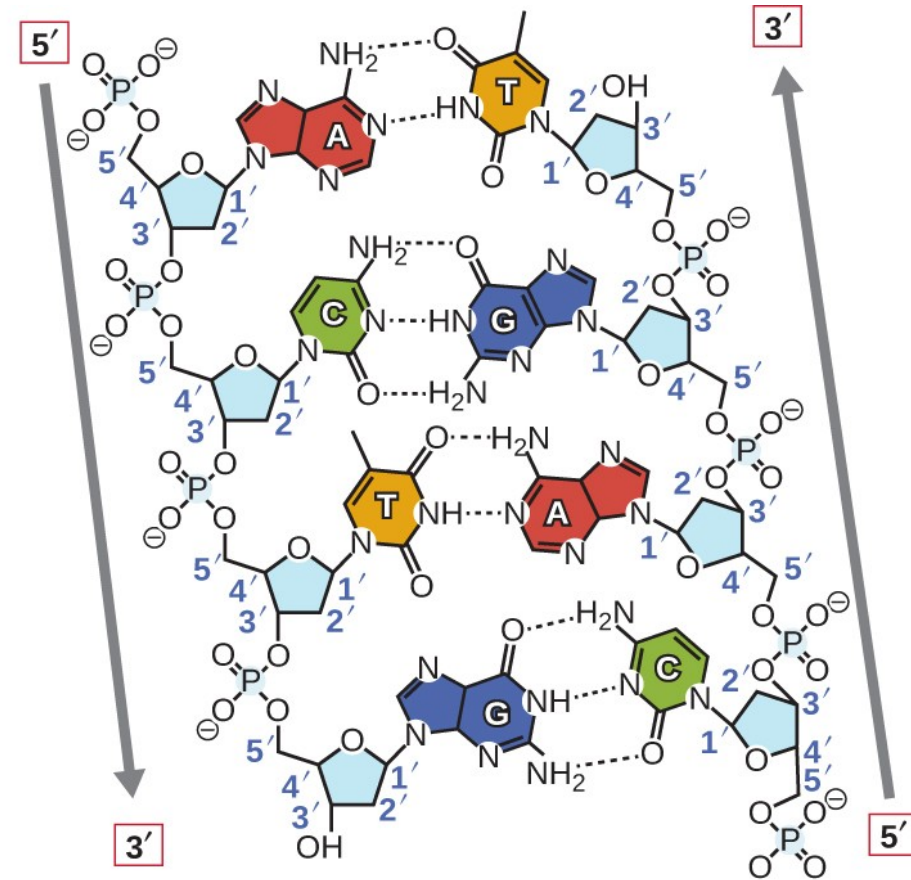
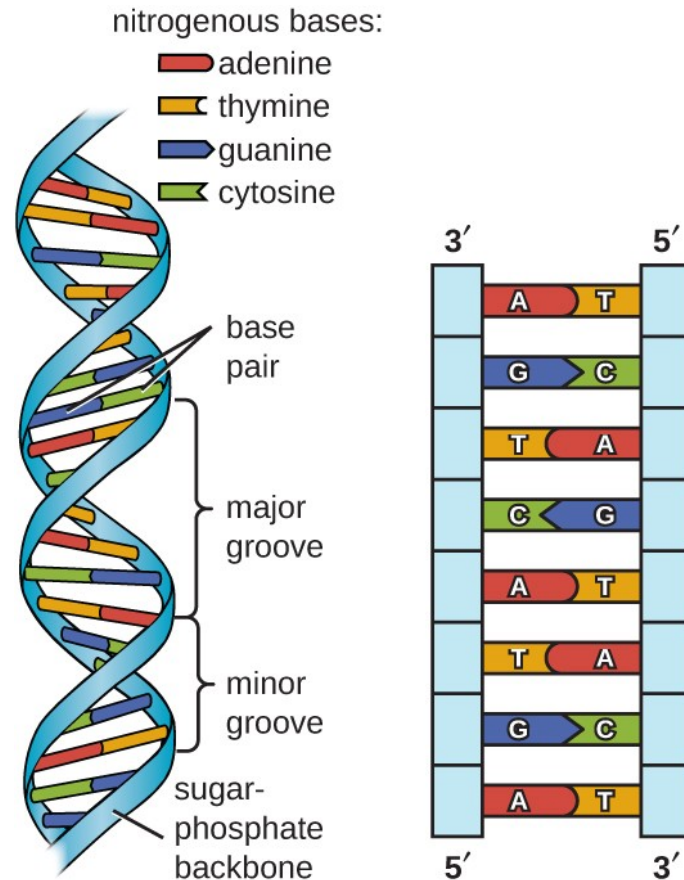
# DNA structure



Dissolves in water (polar)

Precipitate in Ethanol (non-polar)

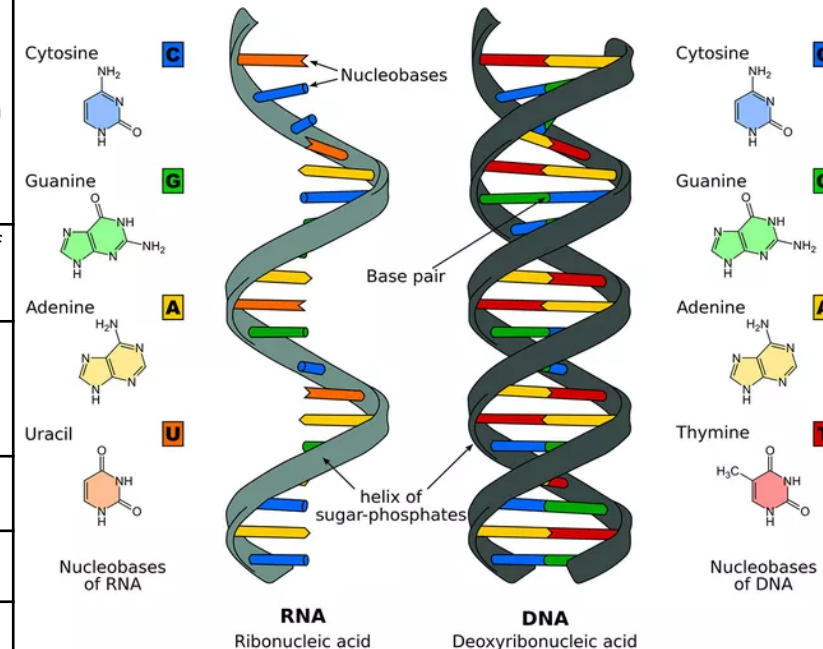
# DNA primary and secondary structures





# RNA structure

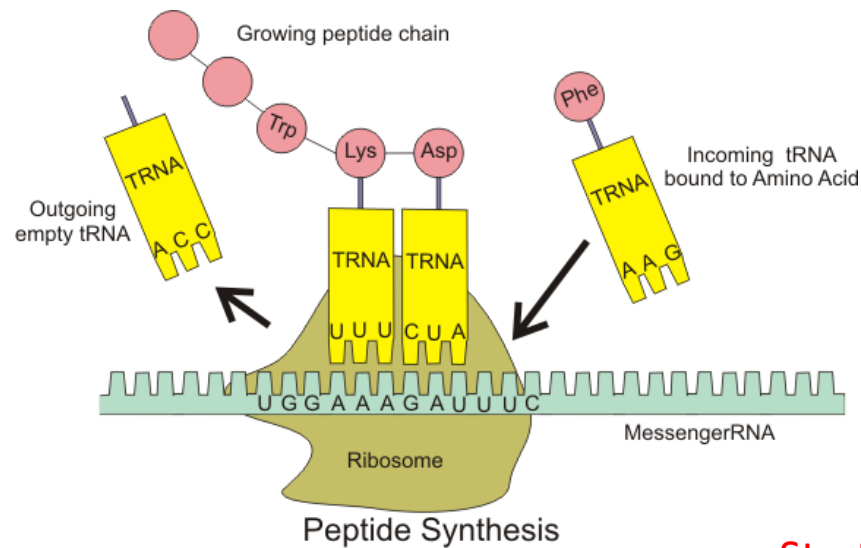
Comparison	DNA	RNA
Name	DeoxyriboNucleic Acid	RiboNucleic Acid
Function	Long-term storage of genetic information; transmission of genetic information to make other cells and new organisms.	Used to transfer the genetic code from the nucleus to the ribosomes to make proteins. RNA is used to transmit genetic information in some organisms and may have been the molecule used to store genetic blueprints in primitive organisms.
Structural Features	B-form double helix.	A-form helix. shorter chains of nucleotides.
Composition of Bases and Sugars	deoxyribose sugar phosphate backbone adenine, guanine, cytosine, thymine bases	ribose sugar phosphate backbone adenine, guanine, cytosine, uracil bases
Propagation	self-replicating.	synthesized from DNA on an as-needed basis.
Base Pairing	A <-> T   G <-> C	A <-> U G <-> C
Reactivity	The C-H bonds make it fairly stable, The small grooves in the helix also serve as protection.	The O-H bond in the ribose of RNA makes the molecule more reactive. The large grooves in the molecule make it susceptible to enzyme attack. Constantly produced, used, degraded, and recycled.
Ultraviolet Damage	susceptible to UV damage.	relatively resistant to UV damage.



# Types of RNA

- Messenger RNA (mRNA) – conveys genetic information that directs the synthesis of specific proteins.
- Transfer RNA (tRNA) – delivers amino acids to the ribosome.
- Ribosomal RNA (rRNA) – links amino acids together to create coded proteins.
- Micro RNA (miRNA) – small non-coding RNA that post transcriptionally regulate gene expression.

# Protein synthesis - 64 different codons produce 20 different amino acids



		Second Letter				
		U	C	A	G	
1st letter	U	UUU   Phe UUC   UUA   Leu UUG	UCU   Ser UCC   UCA   UCG	UAU   Tyr UAC   UAA   Stop UAG   Stop	UGU   Cys UGC   UGA   Stop UGG   Trp	3rd letter
	C	CUU   Leu CUC   CUA   CUG	CCU   Pro CCC   CCA   CCG	CAU   His CAC   CAA   Gln CAG	CGU   Arg CGC   CGA   CGG	
	A	AUU   Ile AUC   AUA   AUG   Met	ACU   Thr ACC   ACA   ACG	AAU   Asn AAC   AAA   Lys AAG	AGU   Ser AGC   AGA   Arg AGG	
	G	GUU   Val GUC   GUA   GUG	GCU   Ala GCC   GCA   GCG	GAU   Asp GAC   GAA   Glu GAG	GGU   Gly GGC   GGA   GGG	

Start codon

No amino acids

Stretch between AUG and stop codon called Open reading frame (ORF).



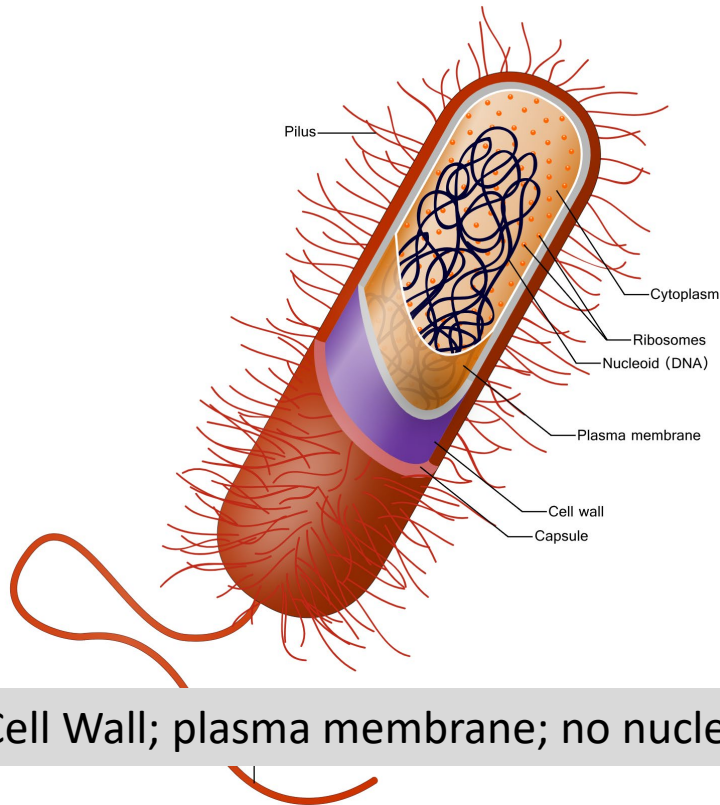
<https://youtu.be/gG7uCskUOrA>

# Common Operations on DNA

1. Collect it (**DNA Extraction**)
2. Quantify total (fluorescent tagging, light absorbance)
3. Estimate size distribution (gel electrophoresis)
4. Make copies
5. Read it
6. Cut it
7. Connect it
8. Move it
9. Change it

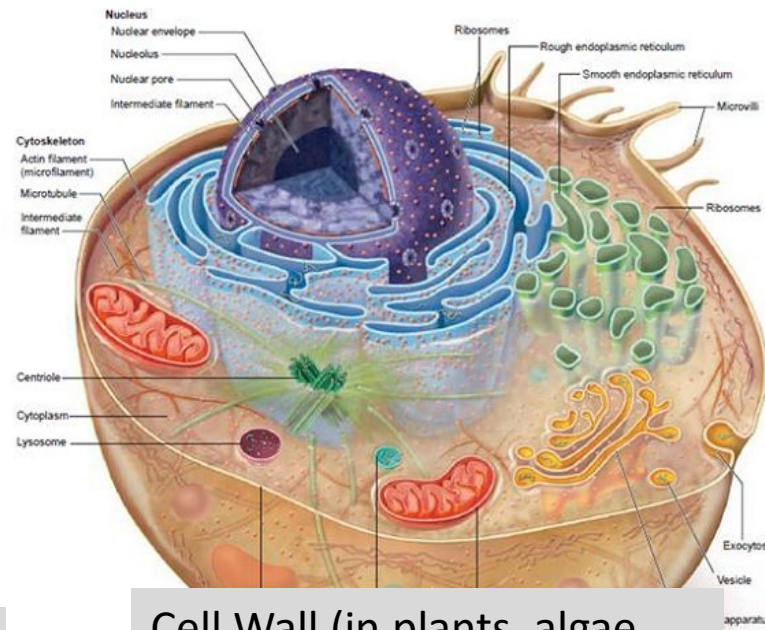
# Cellular Organization affects how we extract DNA

## Prokaryotes (bacteria)



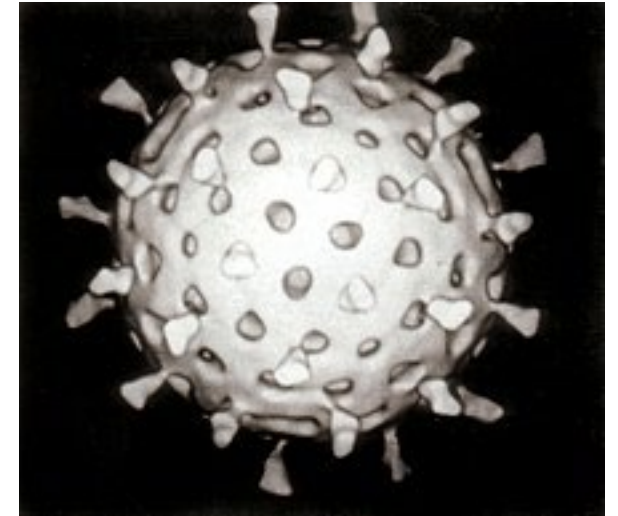
Cell Wall; plasma membrane; no nucleus

## Eukaryotes (algae, plants, animals, etc.)



Cell Wall (in plants, algae only); plasma membrane; DNA in nucleus

## Viruses

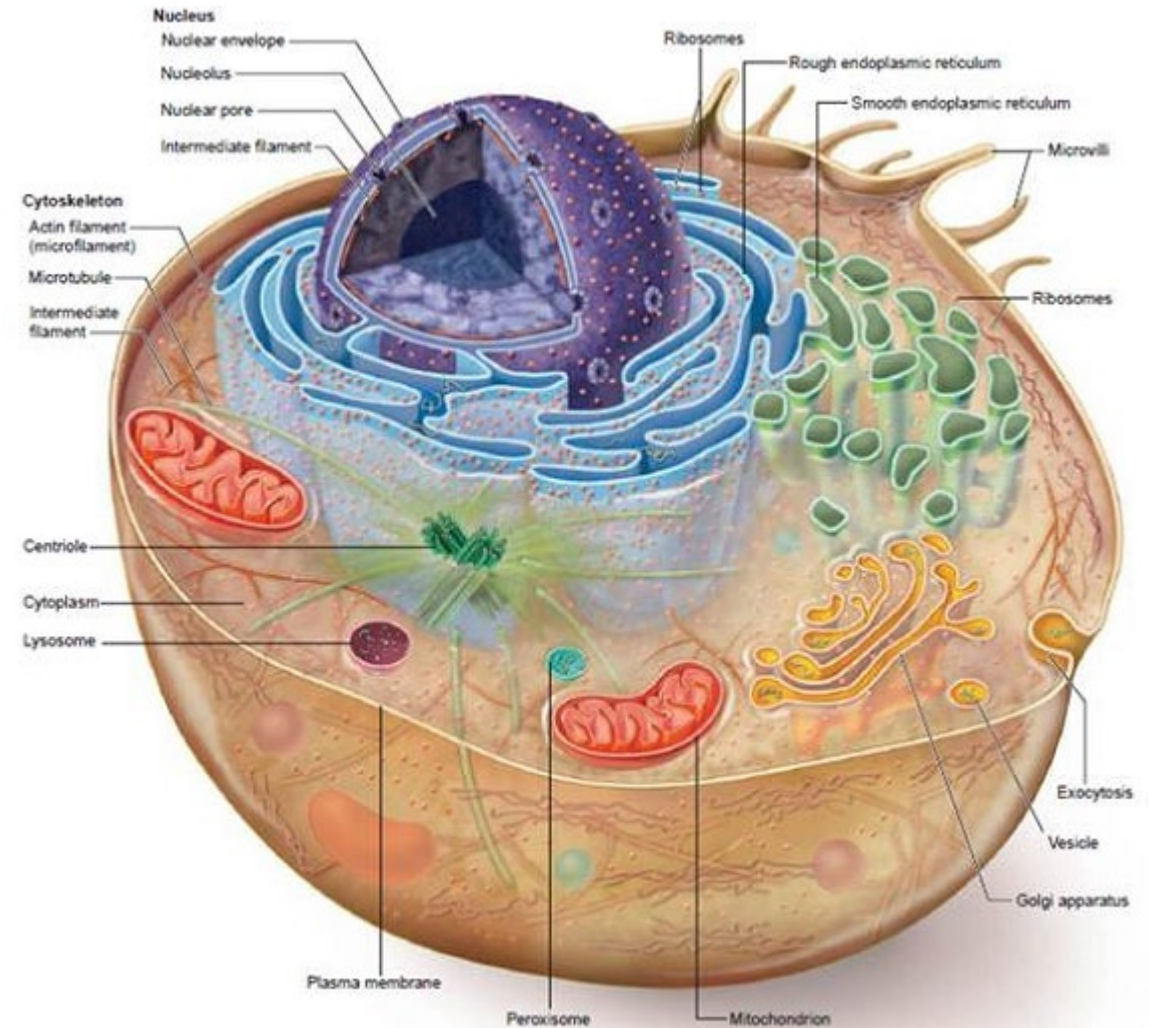


Capsid; plasma membrane (sometimes); RNA or DNA



# DNA Extraction

- Collect cells/tissue sample
- Lyse the cells
- Break down lipids
- Break down proteins
- Remove debris
- Assess quality of extraction



# Common Operations on DNA

1. Collect it (**DNA Extraction**)
2. Quantify total (**fluorescent tagging, light absorbance**)
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# Fluorescence Spectroscopy

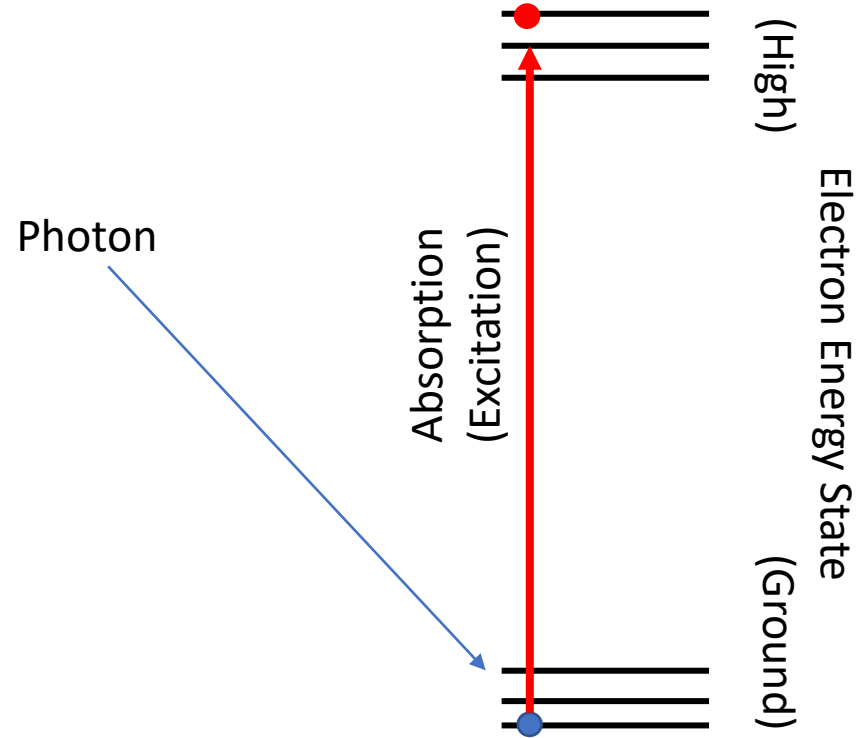
Generate signal by modifying and controlling wavelength of light reaching the sensor.

1. Attach fluorescent tag to protein/target
2. Excite tag using light
3. Measure emission from excited tag

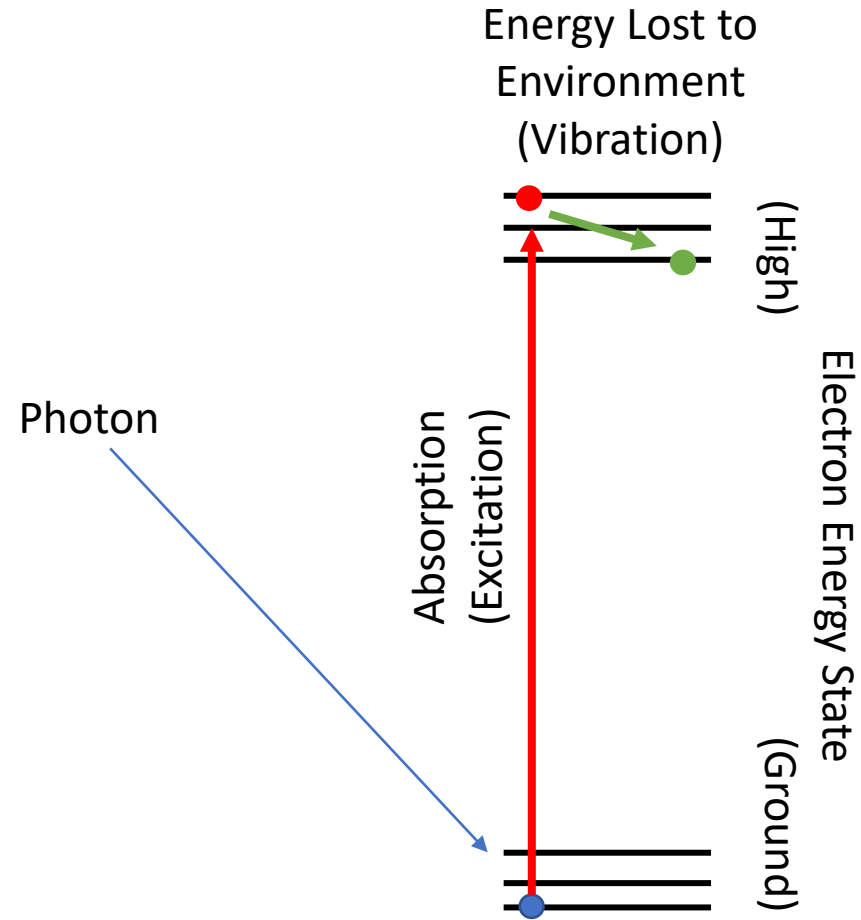
# Excitation and Emission (very important for modern biological research!)



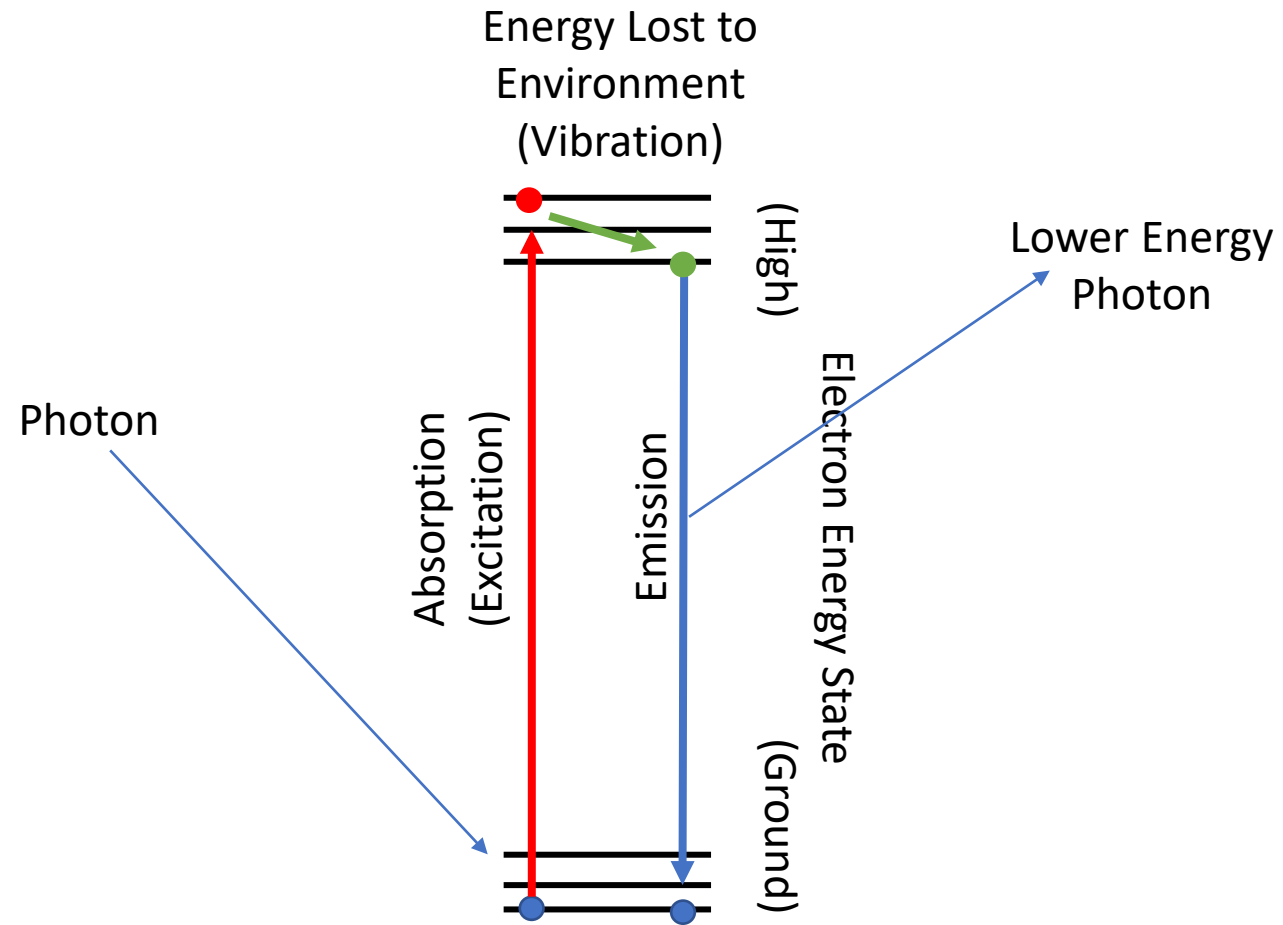
# Excitation and Emission



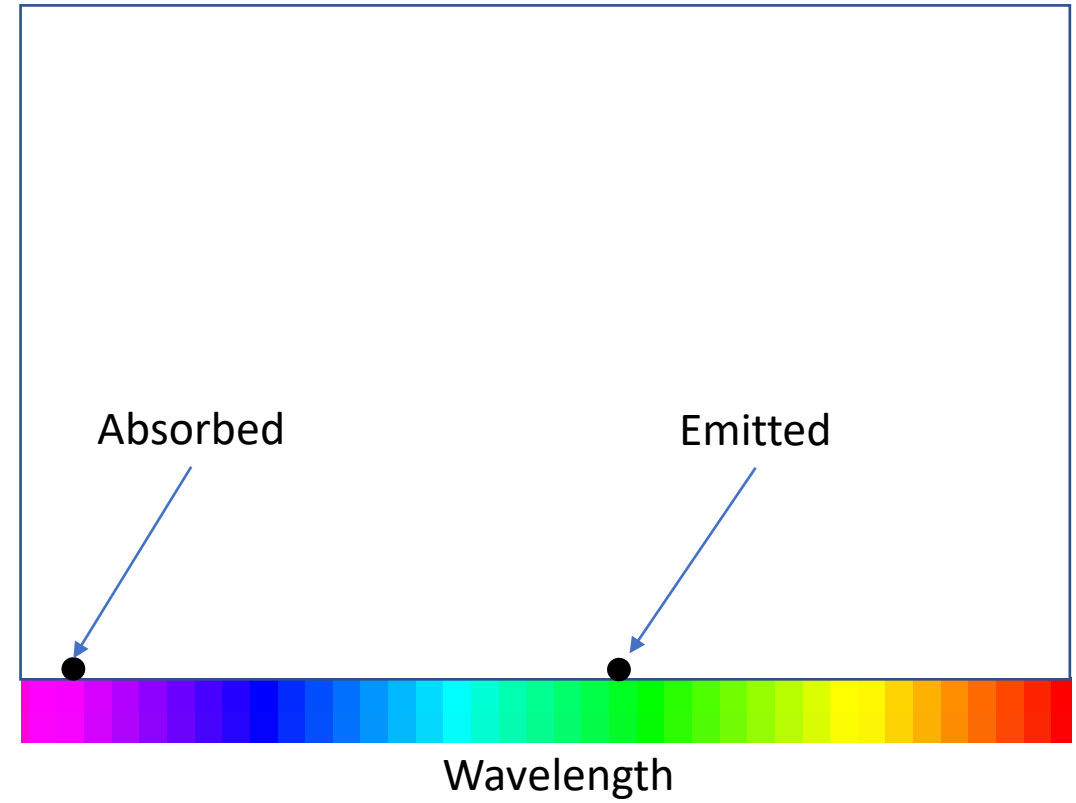
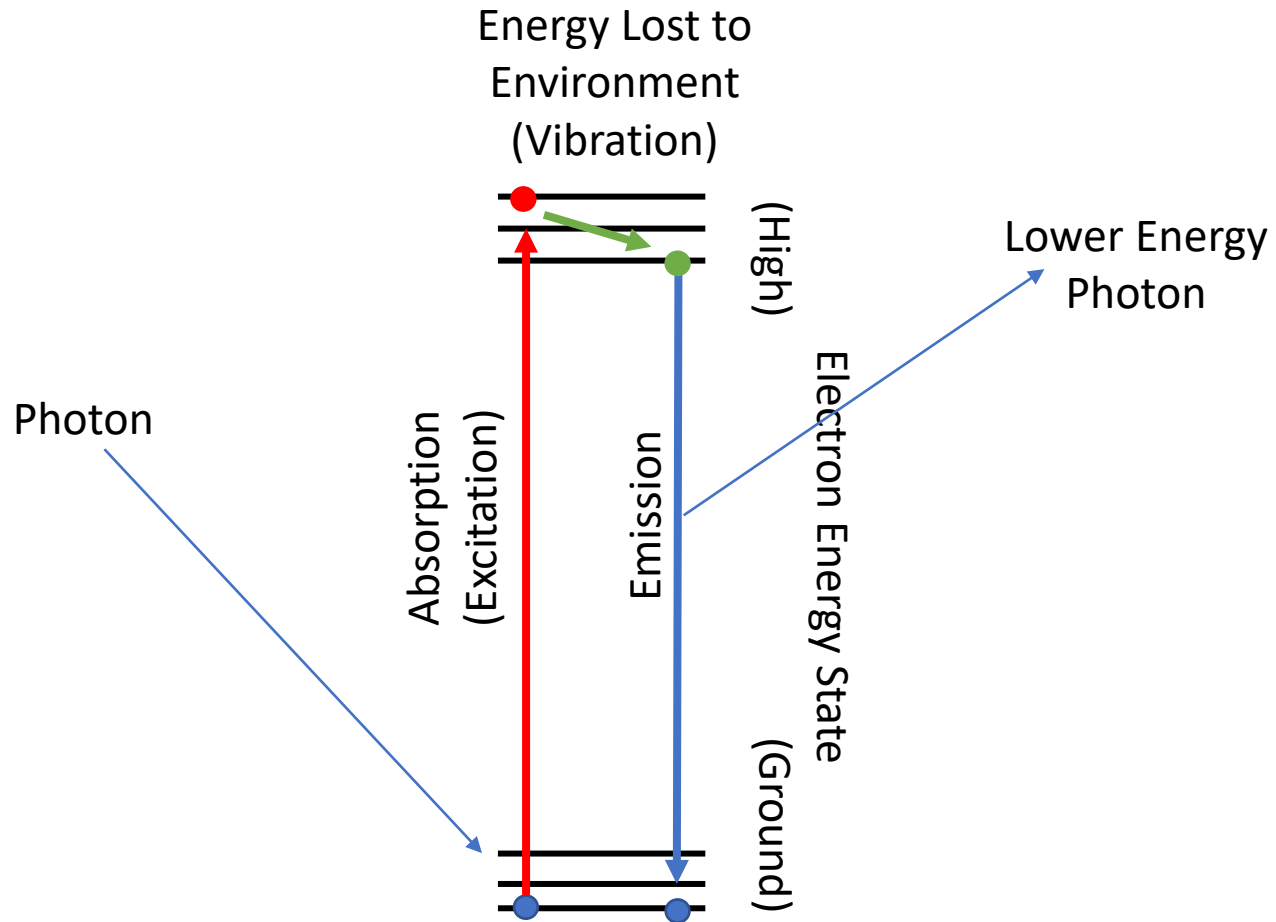
# Excitation and Emission



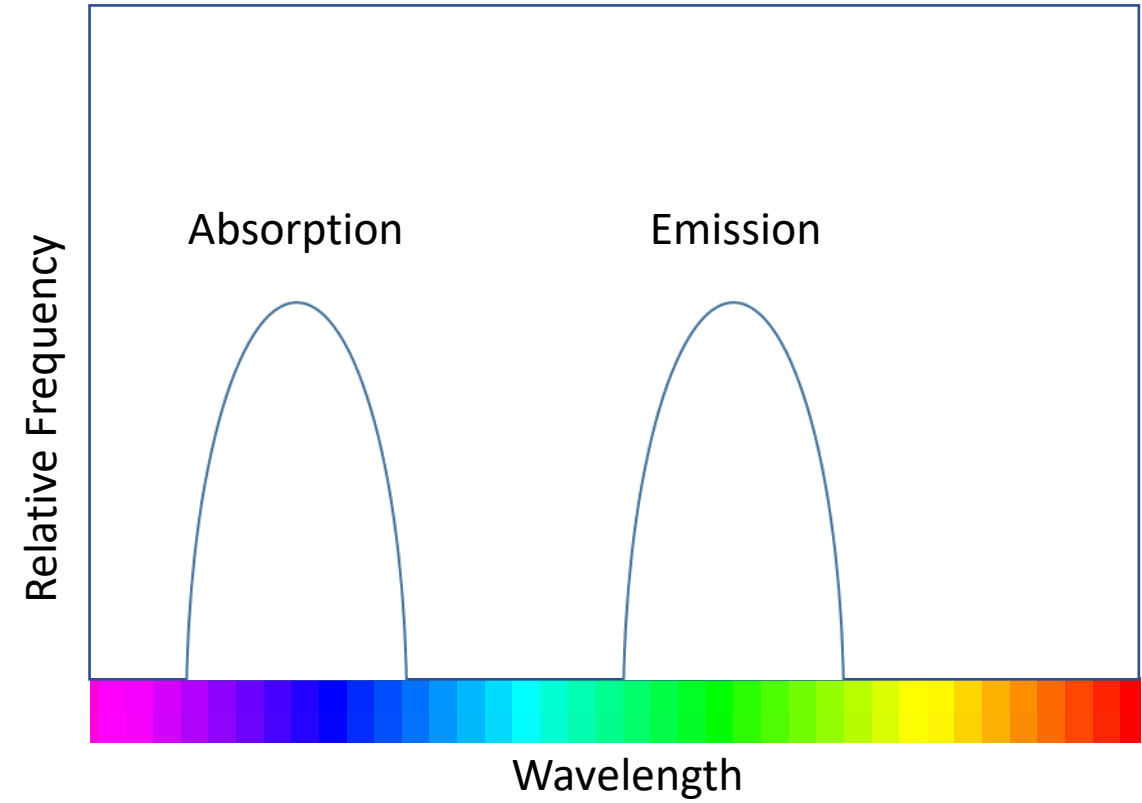
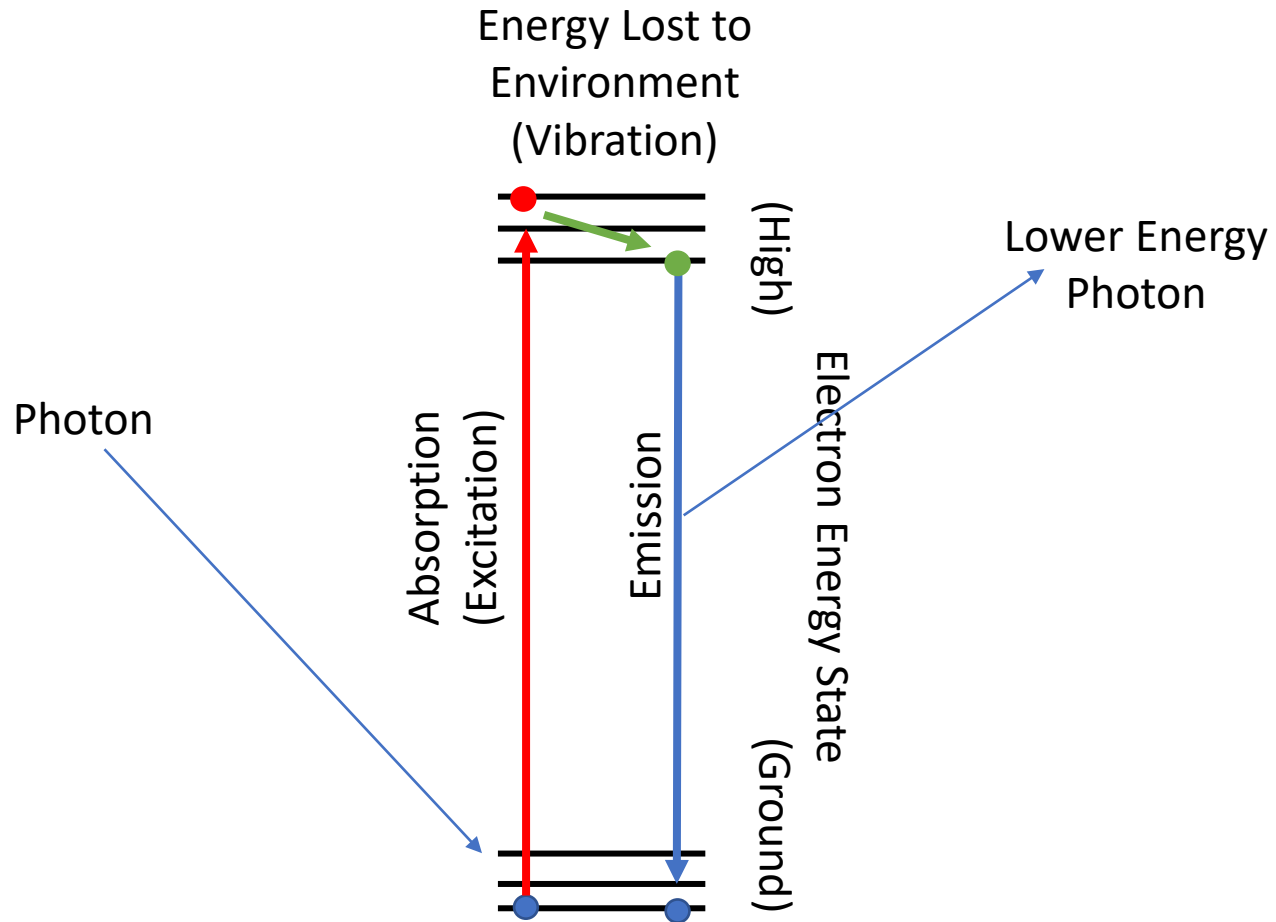
# Excitation and Emission



# Excitation and Emission



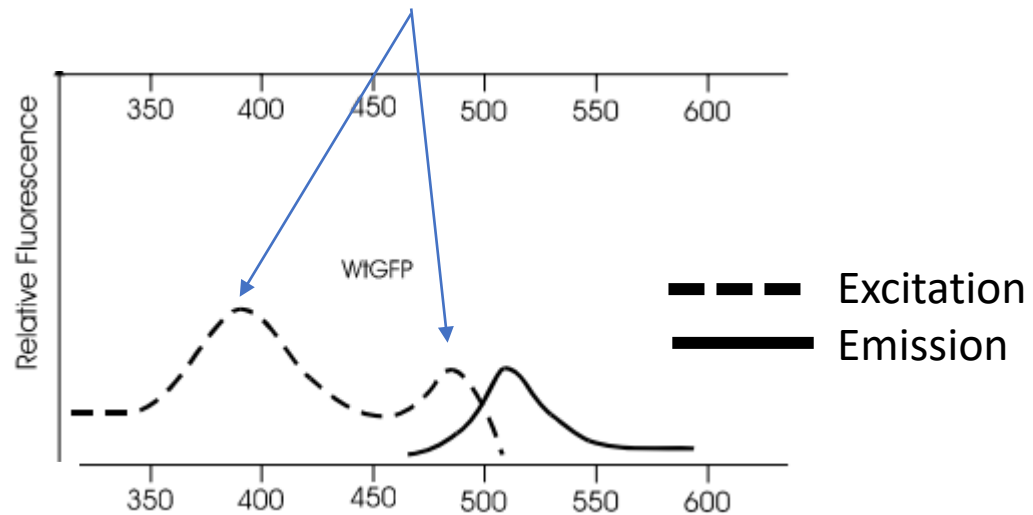
# Excitation and Emission



# Green Fluorescent Protein

- Derived from *Aequorea Victoria*
- Required  $\text{Ca}^{2+}$  to fluoresce

Two peaks are not ideal.



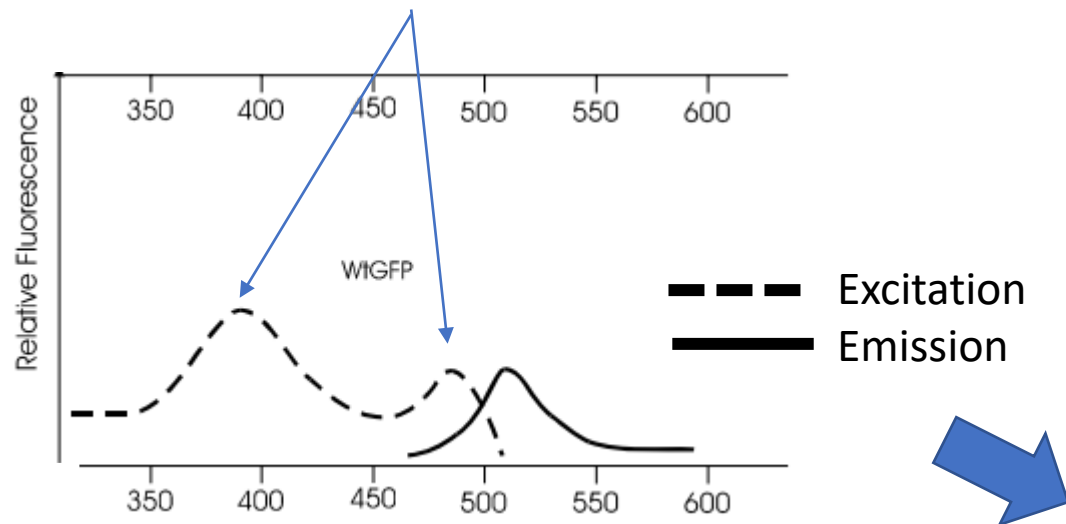
By Mnolf (Photo taken in the Monterey Bay Aquarium, CA, USA) [GFDL (<http://www.gnu.org/copyleft/fdl.html>), CC-BY-SA-3.0 (<http://creativecommons.org/licenses/by-sa/3.0/>) or CC BY-SA 2.0 (<http://creativecommons.org/licenses/by-sa/2.0/>)], via Wikimedia Commons



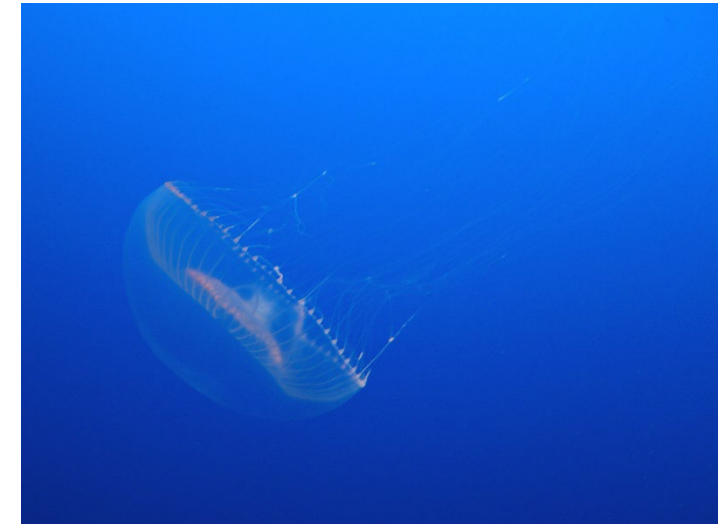
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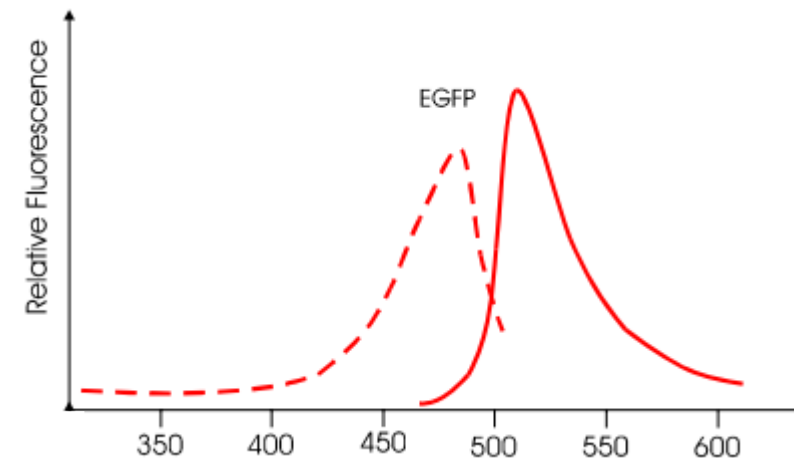
Two peaks are not ideal.



Nobel Prize in Chemistry – 2008

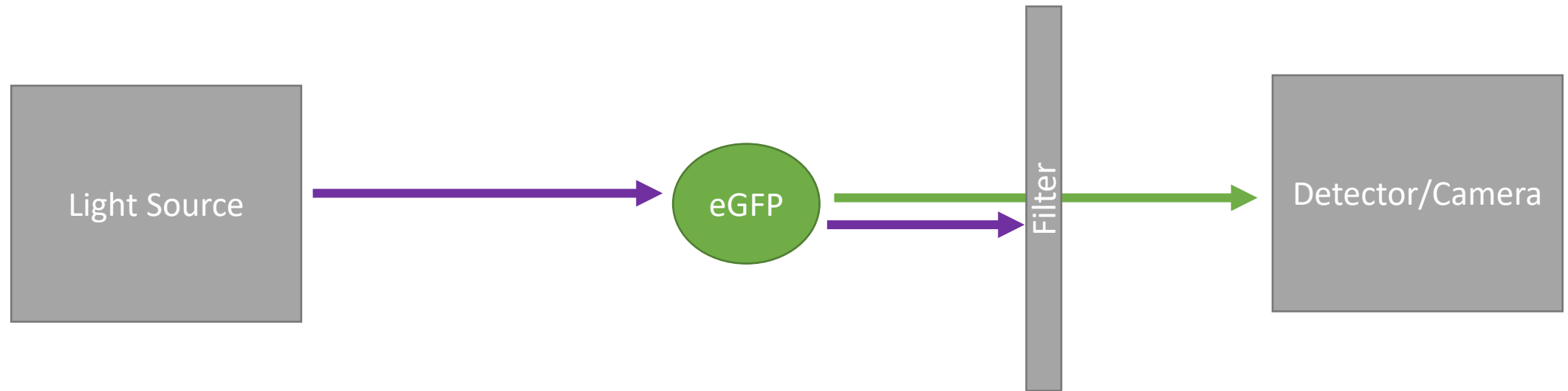


By Mnolf (Photo taken in the Monterey Bay Aquarium, CA, USA) [GFDL (<http://www.gnu.org/copyleft/fdl.html>), CC-BY-SA-3.0 (<http://creativecommons.org/licenses/by-sa/3.0/>) or CC BY-SA 2.0 (<http://creativecommons.org/licenses/by-sa/2.0/>)], via Wikimedia Commons



# Hardware Approaches

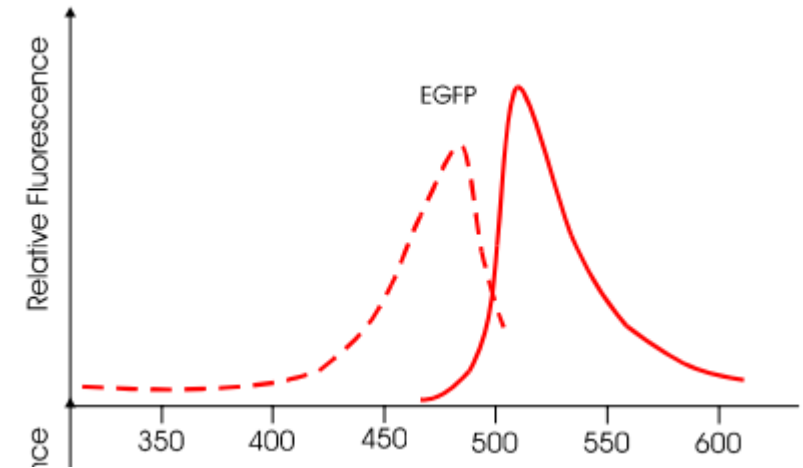
- Excite EGFP with UV light and detect emitted green light



# Why is fluorescence a big deal?

## Specificity!

- Location within cells
  - Modify genes to include fluorescent marker
  - Chemical control
- Fluorescent activation



This gives researchers a lot of flexibility to control fluorescence in their experiments.

# Common Operations on DNA

1. Collect it (**DNA Extraction**)
2. Quantify total (**fluorescent tagging, light absorbance**)
3. Estimate size distribution (**gel electrophoresis**)
4. Make copies
5. Read it
6. Cut it
7. Connect it
8. Move it
9. Change it

# Gel Electrophoresis

Overall Goal: sort substances (nucleic acid strands or proteins) based on size/charge

- Construct Gel
- Prepare samples
- Add experimental samples (as well as ladder and control) to gel
- Run current through gel
- Observe change in gel

# Gel Electrophoresis – Step 1 – The Gel

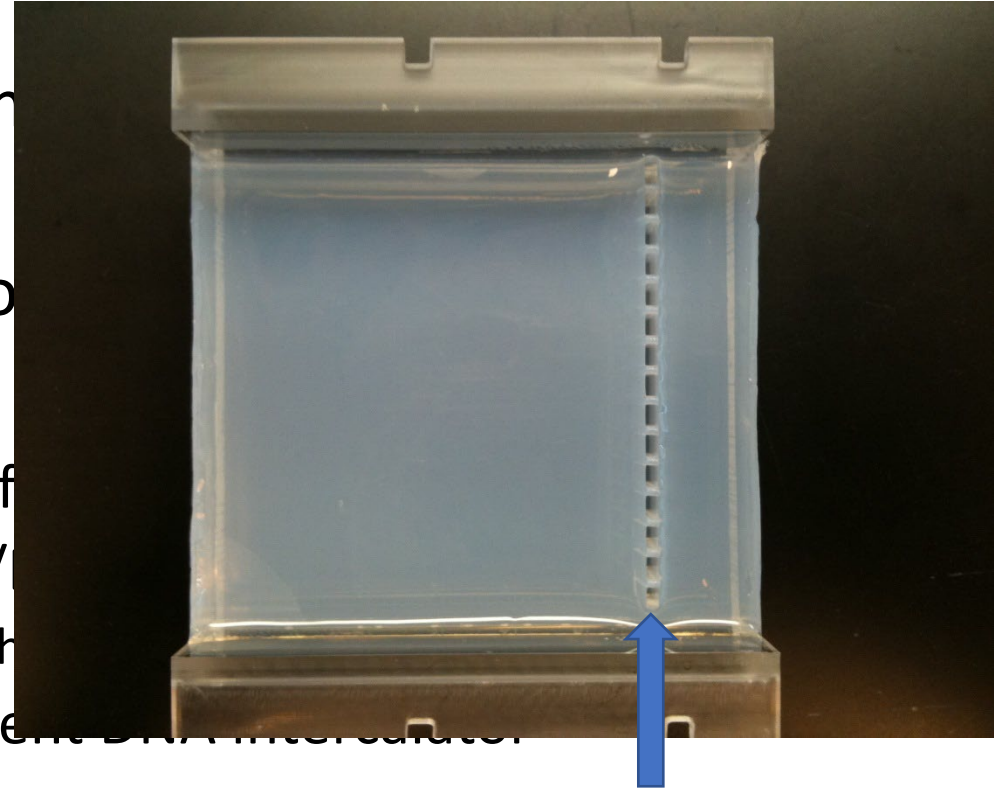
- Goal: generate a mesh through which sample will flow slowly in an organized fashion
- Lanes – lanes are added using a comb to generate wells in the gel
- DNA/RNA:
  - Gel - Agarose gel 0.2-5% weight/ buffer volume – thermal set
  - Buffer – Tris/(Borate or Acetic acid)/EDTA – TBE or TAE
    - Prevent/reduces enzymatic activities that might degrade the DNA
  - Stain – Ethidium bromide – fluorescent DNA intercalator

# Gel Electrophoresis – Step 1 – The Gel

- Goal: generate a mesh through which sample will flow slowly in an organized fashion
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# Gel Electrophoresis – Step 1 – The Gel

- Goal: generate a mesh through which molecules can move in an organized fashion
- Lanes – lanes are added using a comb
- DNA/RNA:
  - Gel - Agarose gel 0.2-5% weight/ buffer
  - Buffer – Tris/(Borate or Acetic acid)/EDTA
    - Prevent/reduces enzymatic activities that can damage DNA/RNA
  - Stain – Ethidium bromide – fluorescent DNA intercalator



Lanes  
(Runs Horizontally)



# Gel Electrophoresis – Step 2 – Sample Prep

1. Denature DNA/RNA
2. Add loading dye, stain

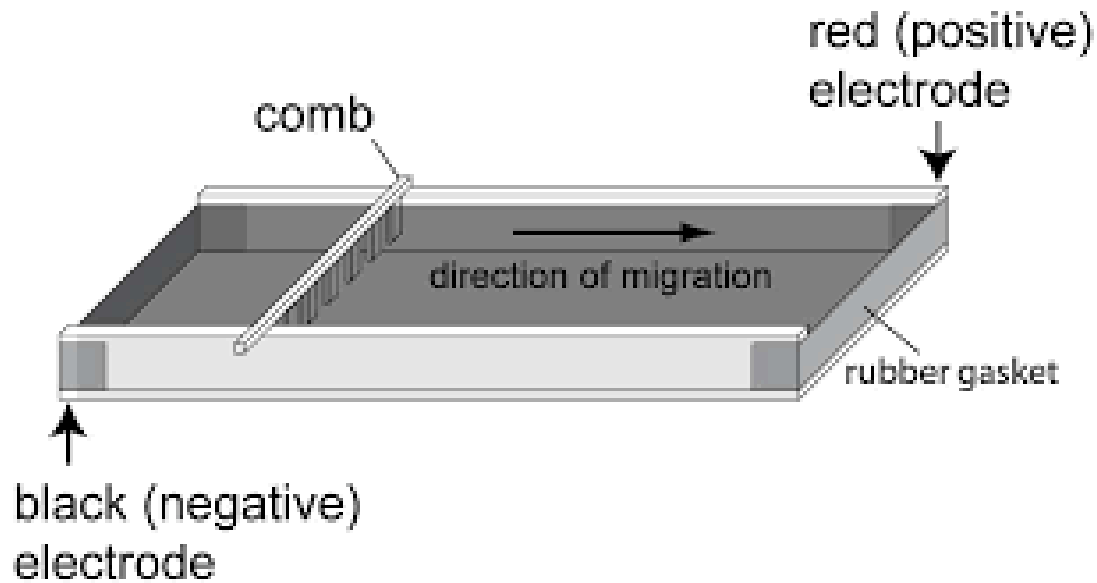
# Gel Electrophoresis – Step 3 – Load Sample

5-20 uL total volume per sample

1. Ladder with known sizes (base pairs for DNA/RNA)
2. Controls
3. Experimental samples

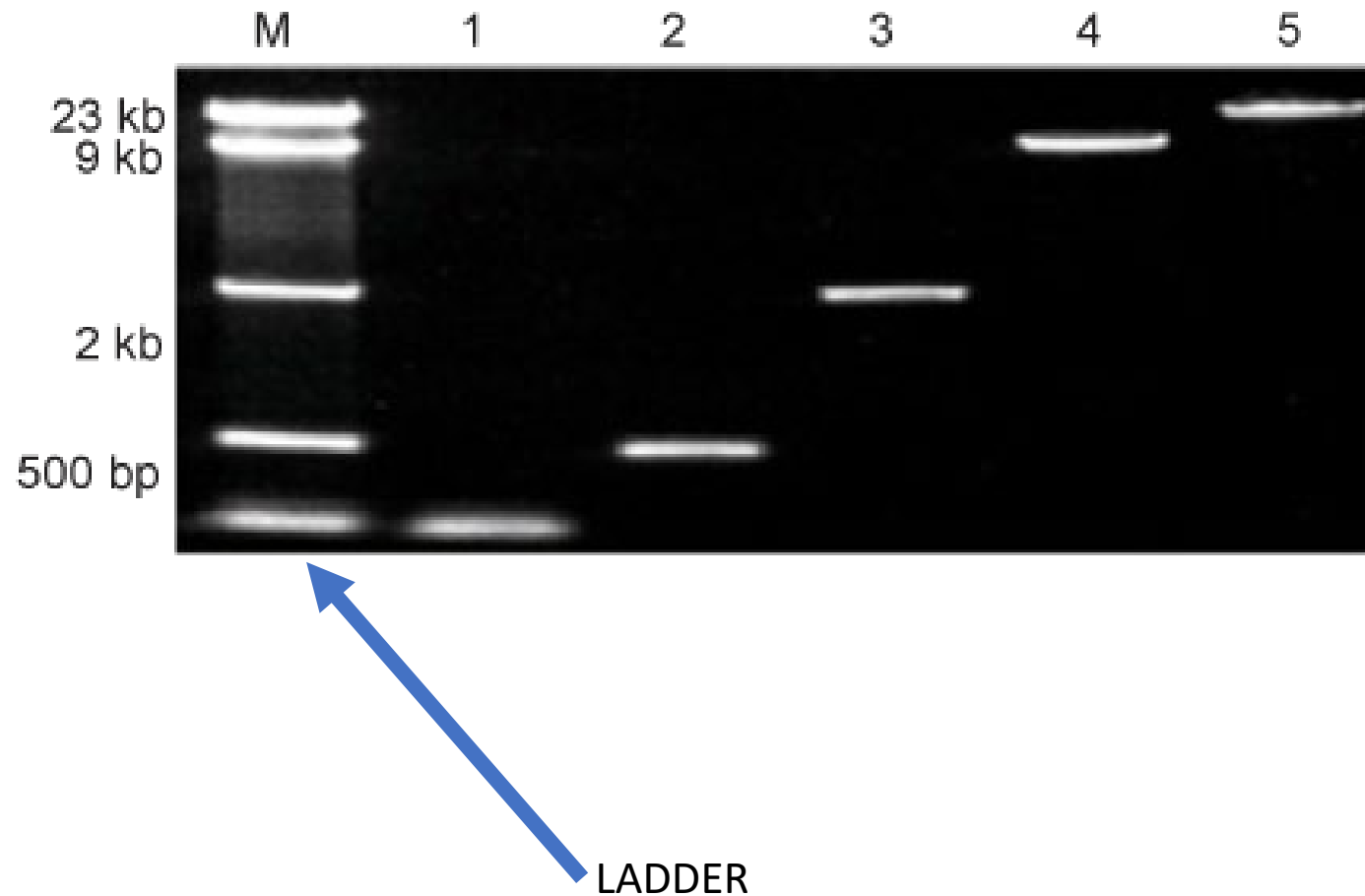
# Gel Electrophoresis – Step 4 – Run the Gel

- Run current through the gel
- Negatively charged molecules flow from negative to positive



Agarose (DNA/RNA)

# Gel Electrophoresis – Step 5 - Observe



# Gel Electrophoresis - Uses

- Measure size of molecules
- Isolate molecules of specific size
- Quantify samples

# Common Operations on DNA

1. Collect it (**DNA Extraction**)
2. Quantify total (**fluorescent tagging, light absorbance**)
3. Estimate size distribution (**gel electrophoresis**)
4. Make copies (**polymerase chain reaction**)
5. Read it
6. Cut it
7. Connect it
8. Move it
9. Change it

# DNA Notation (primary and secondary structure)

ssDNA = 5' -ACTGCGATAGACGATGTCCGGATGACA-3'

← Shows sequence

dsDNA = 5' -ACTGCGATAGACGATGTCCGGATGACA-3'  
3' -TGACGCTATCTGCTACAGGCCTACTGT-5'

← Shows sequence  
and pairing

dsDNA =  
5' -  -3'  
3' -  -5'

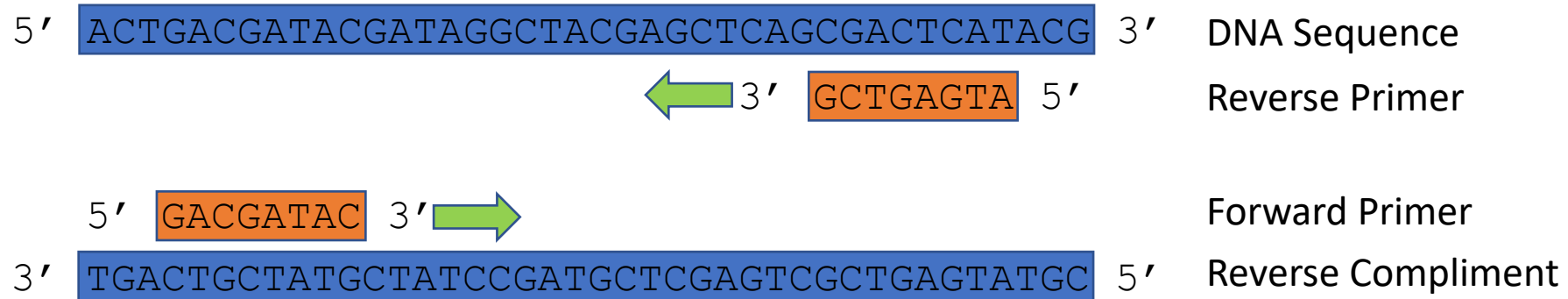
← Shows pairing

# Polymerase Chain Reaction

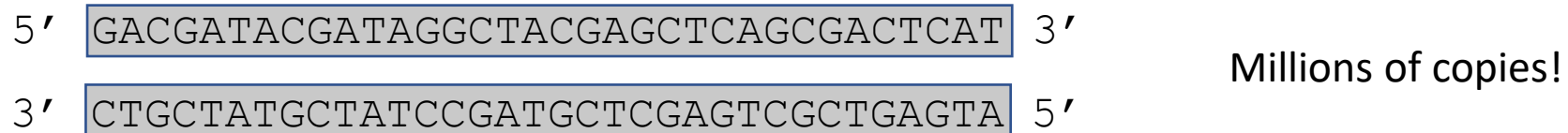
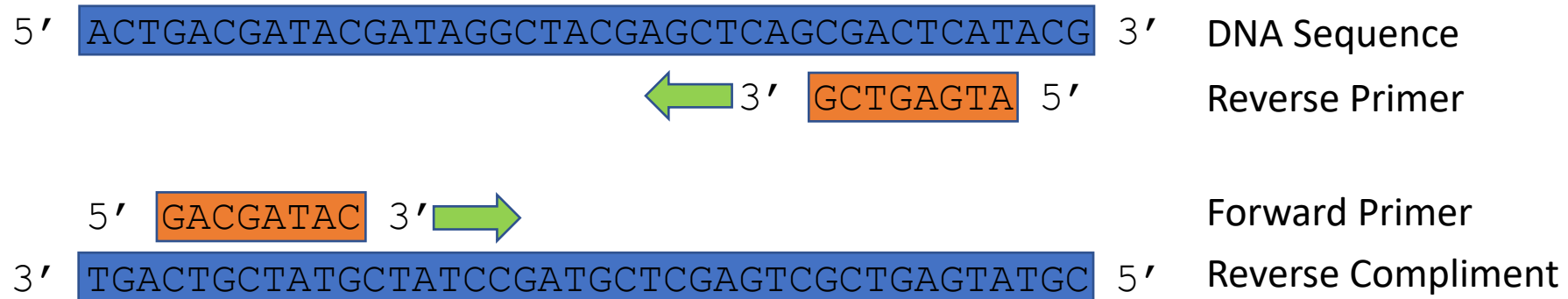
- Purpose : generate many copies of small segments of DNA amounts of DNA
- Requirements:
  - Polymerase
  - Free nucleotides (dNTP)
  - DNA to be copied
  - Primers – very short segments of beginning/end of DNA to be copied
  - Thermocycler
- Polymerase – DNA repair/extension enzyme



# Polymerase Chain Reaction



# Polymerase Chain Reaction



95

72

55

°C



Cycle 1

[www.dnalc.org](http://www.dnalc.org)

# Common Operations on DNA

1. Collect it (**DNA Extraction**)
2. Quantify total (**fluorescent tagging, light absorbance**)
3. Estimate size distribution (**gel electrophoresis**)
4. Make copies (**polymerase chain reaction**)
5. Read it (**sequencing**)
6. Cut it (restriction endonuclease enzymes)
7. Connect it
8. Move it
9. Change it

# Standard Polymerase Chain Reaction

- Purpose : generate many copies of small segments of DNA amounts of DNA
- Requirements:
  - Polymerase
  - Free nucleotides (dNTP)
  - DNA to be copied
  - Primers – very short segments of beginning/end of DNA to be copied
  - Thermocycler
- Polymerase – DNA repair/extension enzyme

Multiple Cycles Product: Millions+ copies of desired DNA

# Polymerase Chain Reaction (for Sanger Sequencing)

- Purpose : generate many copies of small segments of DNA amounts of DNA
  - Requirements:
    - Polymerase
    - Free nucleotides (dNTP) + **fluorescent terminating nucleotides**
    - DNA to be copied
    - ~~• Primers – very short segments of beginning/end of DNA to be copied~~
    - Single primer at beginning of region to be sequenced
    - Thermocycler
  - Polymerase – DNA repair/extension enzyme
- One cycle** product: Millions+ copies of **truncated** desired DNA

# Sanger Sequencing

1. Amplify region of interest using standard PCR
2. Purify product (remove primers and undesired PCR product)
3. Add one primer for side of strand you wish to sequence
4. Replace ***small*** fraction of dNTPs with terminating dNTPs  
Terminating dNTPs: dNTPs which stop the polymerase reaction  
Each terminating nucleotide (G,A,T,C) may be uniquely fluorescently tagged
5. Run one cycle of PCR  
Note: If fluorescent, only one reaction is needed. If not, four reactions are required (one each for G,A,T,C)
6. Run reaction(s) in gel

# Sanger Sequencing

1. Amplify region of interest using standard PCR
2. Purify product (remove primers and undesired PCR product)
3. Add one primer for side of strand you wish to sequence
4. Replace ***small*** fraction of dNTPs with terminating dNTPs

Terminating nucleotides terminate polymerase strand extension

Terminating dNTPs: dNTPs which stop the polymerase reaction

Each terminating nucleotide (G,A,T,C) may be uniquely fluorescently tagged

5. Run one cycle of PCR

Note: If fluorescent, only one reaction is needed. If not, four reactions are required (one each for G,A,T,C)

6. Run reaction(s) in gel



# Terminating Nucleotides Examples

- Nucleotides which terminate polymerase strand extension

ACTGTCTGACATGTGCGCGT

ACTG →

TGACAGACTGTACACGCGCA

	A	C	T	G
Normal	100%	100%	100%	100%
Terminating	0%	0%	0%	0%

Product(s)?

# Terminating Nucleotides

- Nucleotides which terminate polymerase strand extension

ACTGTCTGACATGTGCGCGT

ACTG →

TGACAGACTGTACACGCGCA

	A	C	T	G
Normal	100%	100%	100%	100%
Terminating	0%	0%	0%	0%

Product(s)? ACTGTCTGACATGTGCGCGT

# Terminating Nucleotides

- Nucleotides which terminate polymerase strand extension

ACTGTCTGACATGTGCGCGT

ACTG →

TGACAGACTGTACACGCGCA

	A	C	T	G
Normal	0%	100%	100%	100%
Terminating	100%	0%	0%	0%

Product(s)?

# Terminating Nucleotides

- Nucleotides which terminate polymerase strand extension

ACTGTCTGACATGTGCGCGT

ACTG →

TGACAGACTGTACACGCGCA

	A	C	T	G
Normal	0%	100%	100%	100%
Terminating	100%	0%	0%	0%

Product(s)? ACTGTCTGA

# Terminating Nucleotides

- Nucleotides which terminate polymerase strand extension

ACTGTCTGACATGTGCGCGT

ACTG →

TGACAGACTGTACACGCGCA

	A	C	T	G
Normal	50%	100%	100%	100%
Terminating	50%	0%	0%	0%

Product(s)?

# Terminating Nucleotides

- Nucleotides which terminate polymerase strand extension

ACTGTCTGACATGTGCGCGT

ACTG →

TGACAGACTGTACACGCGCA

	A	C	T	G
Normal	50%	100%	100%	100%
Terminating	50%	0%	0%	0%

Product(s)? ACTGTCTGA; ACTGTCTGACA; ACTGTCTGACATGTGCGCGT

# Multiple Reactions

ACTGTCTGACATGTGCGCGT

## Products

	A	C	T	G	[ ACTGTCTGA ACTGTCTGACA	
Normal	50%	100%	100%	100%		
Terminating	50%	0%	0%	0%		
Normal	100%	50%	100%	100%	[ ACTGTC ACTGTCTGAC ACTGTCTGACATGTGC ACTGTCTGACATGTGCGC	
Terminating	0%	50%	0%	0%		
Normal	100%	100%	50%	100%	[ ACTGT ACTGTCT ACTGTCTGACAT ACTGTCTGACATGT	
Terminating	0%	0%	50%	0%		
Normal	100%	100%	100%	50%	[ ACTGTCTG ACTGTCTGACATG ACTGTCTGACATGTG ACTGTCTGACATGTGCG ACTGTCTGACATGTGCGCG	
Terminating	0%	0%	0%	50%		

# Multiple Reactions

ACTGTCTGACATGTGCGCGT

## Products

	A	C	T	G
Normal	50%	100%	100%	100%
Terminating	50%	0%	0%	0%

ACTGTCTGA  
ACTGTCTGACA

Normal	100%	50%	100%	100%
Terminating	0%	50%	0%	0%

ACTGTC  
ACTGTCTGAC  
ACTGTCTGACATGTGC  
ACTGTCTGACATCTCCCC

Normal	100%	100%	50%	100%
Terminating	0%	0%	50%	0%

ACTGT  
ACTGTCT  
ACTGTCTGAC  
ACTGTCTGAC

Normal	100%	100%	100%	50%
Terminating	0%	0%	0%	50%

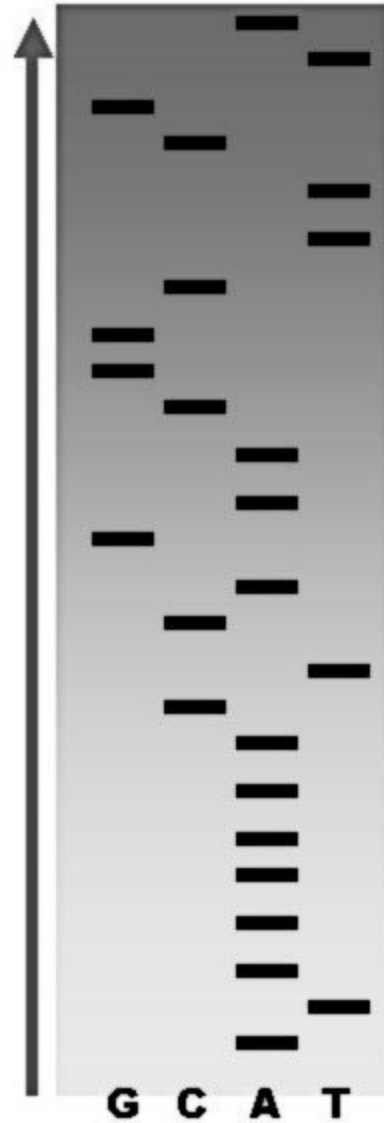
ACTGTCTG  
ACTGTCTGACATG  
ACTGTCTGACATGTG  
ACTGTCTGACATGTGCG  
ACTGTCTGACATGTGCGCG

What is the relative frequency of products ending in G?



# Sanger Sequencing Gel

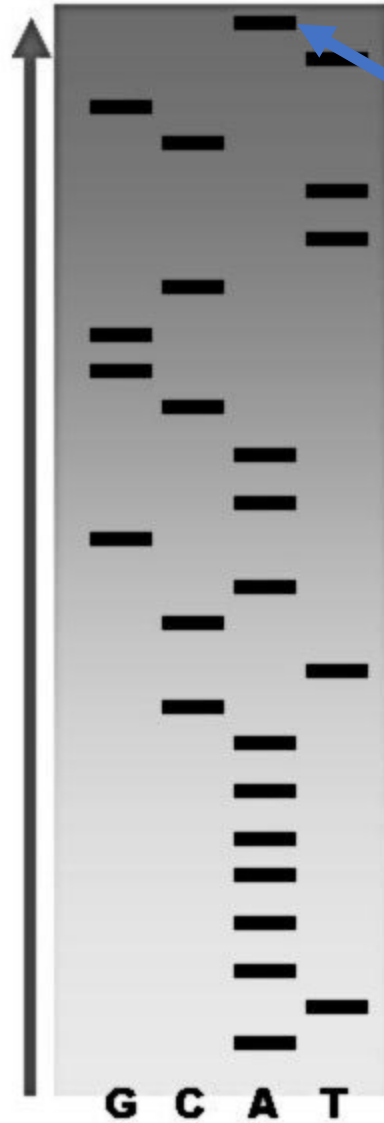
[https://commons.wikimedia.org/wiki/File:Radioactive\\_Fluorescent\\_Seq.jpg](https://commons.wikimedia.org/wiki/File:Radioactive_Fluorescent_Seq.jpg)



Where are the smaller PCR products?

# Sanger Sequencing Gel

[https://commons.wikimedia.org/wiki/File:Radioactive\\_Fluorescent\\_Seq.jpg](https://commons.wikimedia.org/wiki/File:Radioactive_Fluorescent_Seq.jpg)

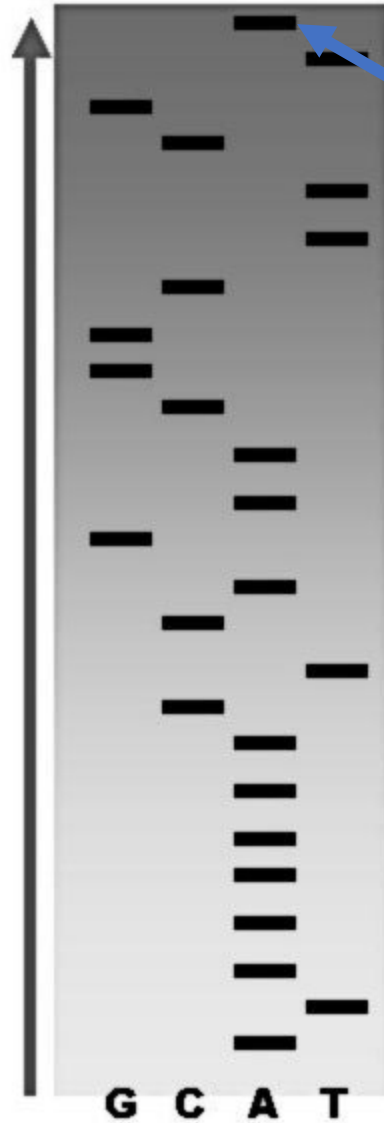


Where are the smaller PCR products?

What is the sequence of product here?

# Sanger Sequencing Gel

[https://commons.wikimedia.org/wiki/File:Radioactive\\_Fluorescent\\_Seq.jpg](https://commons.wikimedia.org/wiki/File:Radioactive_Fluorescent_Seq.jpg)

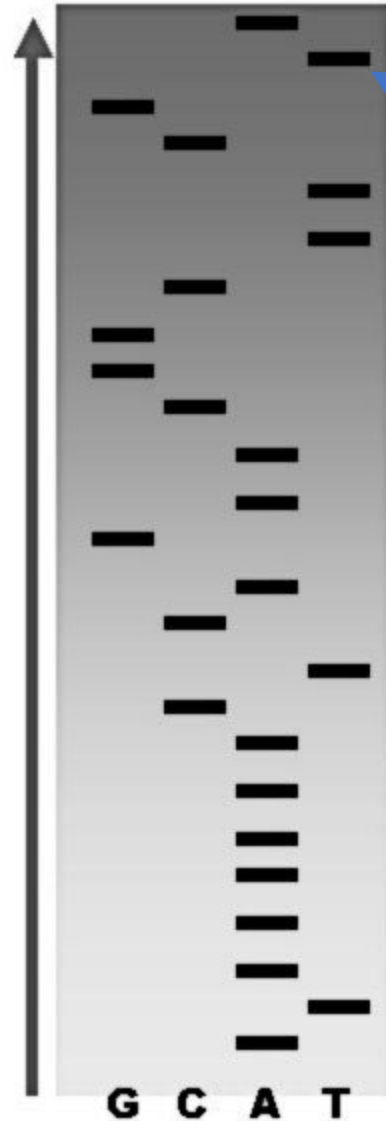


Where are the smaller  
PCR products?

What is the sequence  
of product here?  
[Primer] + A

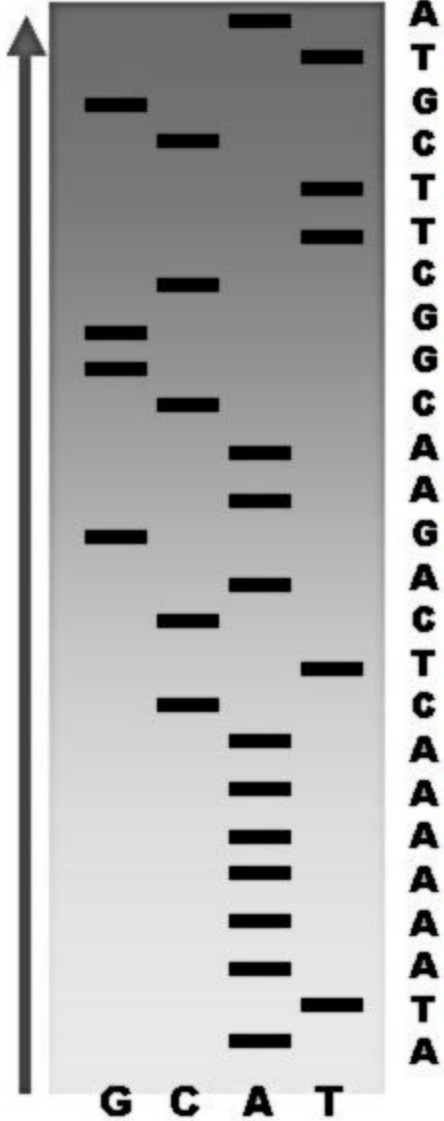
# Sanger Sequencing Gel

[https://commons.wikimedia.org/wiki/File:Radioactive\\_Fluorescent\\_Seq.jpg](https://commons.wikimedia.org/wiki/File:Radioactive_Fluorescent_Seq.jpg)



# Sanger Sequencing Gel

[https://commons.wikimedia.org/wiki/File:Radioactive\\_Fluorescent\\_Seq.jpg](https://commons.wikimedia.org/wiki/File:Radioactive_Fluorescent_Seq.jpg)



# Single Reaction with Fluorescent Terminating Nucleotides

ACTGTCTGACATGTGCGCGT

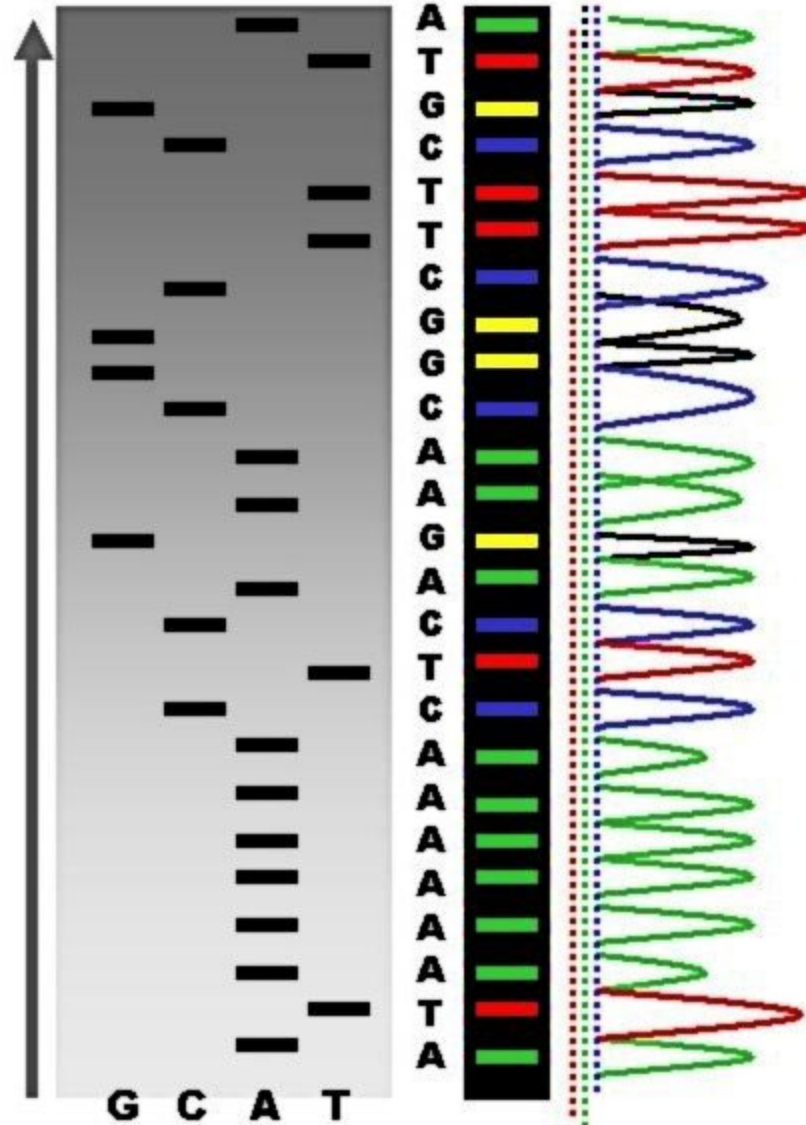
## Products

	A	C	T	G
Normal	50%	50%	50%	50%
Terminating	50%	50%	50%	50%

ACTG**T**  
ACTGT**C**  
ACTGTC**T**  
ACTGTCT**G**  
ACTGTCTG**A**  
ACTGTCTGAC**C**  
ACTGTCTGAC**A**  
ACTGTCTGACA**T**  
ACTGTCTGACAT**G**  
ACTGTCTGACATG**T**  
ACTGTCTGACATGT**G**  
ACTGTCTGACATGTG**C**  
ACTGTCTGACATGTGC**G**  
ACTGTCTGACATGTGCG**C**  
ACTGTCTGACATGTGCGC**G**  
ACTGTCTGACATGTGCGCG**T**

# Sanger Sequencing Gel

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# Sequencing Traces

- Snapgene Viewer (free)
- Sequence file on Canvas



# Today

- Protocol 7 for remaining bacteria
  - Skip WGS prep
  - Only make 2 extra tubes per sample
  - Blank labels provided...