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Condensate quantification - mRNA molecules within condensates (How to use)

Before start

- Download the repository or the content of this folder.
- For Github downloads: example smFISH image (large file) is provided as 'tiftxt'. A link to download the image is within the text file.
 - https://doi.org/10.17632/8jvrnztdvc.1 (location: smFISH images quantification > example_image).
 - https://doi.org/10.17632/8jvrnztdvc.2 (location: condensate quantification > example-data).
 - This guide uses the smiFISH image from the folder: ./code-image-analysis-c-elegans/smFISH-images-quantification/example-image.
- Important: this script uses results from smFISH images quantification (./code-image-analysis-c-elegans/smFISH-images-quantification).
- Important: an average PSF of experimental single mRNA molecules is required.

Requirements and set up

Requirements:

- Tested in MATLAB v R2018b or higher
 - Required toolboxes:
 Image Processing Toolbox
 Statistics and Machine Learning Toolbox
- Additional requirements:

FISH-quant (1,2,#)

https://bitbucket.org/muellerflorian/fish_quant/src/master/

Set up:

- Copy the scripts provided (./condensate-quantification-scripts) to the MATLAB folder (usually located in Documents directory).
- Install FISH-quant:
 - Go to https://bitbucket.org/muellerflorian/fish_quant/src/master/ (1,2).
 - Click Downloads and then download the repository (1,2).
 - Unzip the file in the MATLAB folder (Documents directory) (1,2).
 - Go to ./Documents/MATLAB/FISH_quant/Documentation (1,2).
 - Open FISH-QUANT__Tutorials.pdf (1,2).
 - Follow the section 1.1 Install FISH-quant for Matlab (1,2).

This script performs the following analysis:

- Inputs:
 - MATLAB workspace with condensate coordinates: .mat file obtained after smFISH images quantification.
 - **smFISH image**: smFISH image used to run **smFISH images quantification**.
 - Average PSF of experimental single mRNA molecules: it can be obtained as described previously (1,2).
- Analysis:
 - Estimation of the number of mRNA molecules within condensates segmented in **smFISH images quantification**. Estimates are computed by integrated intensity and cumulative intensity (3).
- Outputs
 - CSV table file with columns:

```
"imageID": image name
"Cell": oocyte
"condensate": condensate index
"SigmaXY": estimated sigma in x and y of condensate
"SigmaZ": estimated sigma in z of condensate
"Amplitud" estimated amplitude of condensate
"BGD": estimated background of condensate
"MaxI": maximum intensity of condensate
"volume_dil": volume of condensate after morphological dilation
"volume": volume of condensate
"mean_intensity": average intensity of condensate
"cumulative_intensity": cumulative intensity of condensate
"median_intensity": median intensity of condensate
```

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```
"n_molecules_intg": number of mRNA molecules within condensate, calculated by integrated intensity
"n_molecules_cum": number of mRNA molecules within condensate, calculated by cumulative intensity
"n_molecules_cum_ctrl": number of mRNA molecules within condensate, calculated by cumulative intensity
(computing control)
```

User guide

Data provided

Location: ./example-data

- MATLAB workspace with condensate coordinates: ./example-data/Dil_Coor_FISH_GFP_w4_Spn4.mat.
- smFISH image: ./example-data/w4 Spn4.tif.
- Average PSF of experimental single mRNA molecules: ./example-data/experimental_single_mol_PSF_8x6_mRNA_AVG_ns.tif.

Analysis (Procedure):

Pre-analysis

- 1. Make a folder with the *inputs* (as in _/example-data).
 - Dil_Coor_FISH_GFP_w4_Spn4.mat
 - *w4_Spn4*.tif
 - experimental_single_mol_PSF_8x6_mRNA_AVG_ns.tif
- 2. The script will look for the key words in **bold** to identify each input. This can be modified as shown below.

Analysis

- 3. Open MATLAB.
- 4. In the command window type:

```
condensates_quantification_script
```

and then press enter.

5. Select the folder with the *inputs* (as in ./example-data).

The default parameters of the script include:

```
%===== file ID indicators
files = struct; % do not modify
files.matlab_ws = '*Coor*'; % unique file identifier for MATLAB workspace
files.FISH_img = '*Spn4'; % unique file identifier for smFISH image
files.mRNA_img = 'experimental_single_mol*'; % unique file identifier for mRNA image
%====== modify below only if you need to change the microscope parameters
define_microscope_parameters = 0; % yes = 1, no = 0
...
```

if define_microscope_parameters = 1, the script will ask for microscope parameters in the command window.

To change this:

- Type in the command window:
 - edit condensates_quantification_script
- Press enter.
- Then, default values can be manually modified (lines # 9 to # 15).

Outputs:

Provided output examples:

The quantification results, for the example data provided in this tutorial, are included (location: ./quantification-example-data).

References

- 1. Mueller, F., Senecal, A., Tantale, K. et al. FISH-quant: automatic counting of transcripts in 3D FISH images. Nat Methods 10, 277-278 (2013).
- 2. Tsanov, N., Samacoits, A., Chouaib, R. et al. smiFISH and FISH-quant a flexible single RNA detection approach with super-resolution capability, Nucleic Acids Research 44 (22), e165 (2016).

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3. Cardona et al., Self-demixing of mRNA copies buffers mRNA:mRNA and mRNA:regulator stoichiometries, Cell (2023), https://doi.org/10.1016/j.cell.2023.08.018