## CARL MCCOMBE

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## **Brief profile**

A current Ph.D. candidate in the Research School of Biology at The Australian National University. I am a molecular biologist/biochemist investigating the functions of proteins involved in plant-microbe interactions.

Education	
The Australian National University, Australia Ph.D. candidate in the Research School of Biology	<u> 2020 - Present</u>
The Australian National University, Australia B.S. with First Class Honours in Biology. GPA: 7.00/7.00 Awarded University Medal	<u>2019</u>
Flinders University, Australia B.S (Biotechnology) GPA: 6.95/7.00	<u> 2016 - 2018</u>

## **Preprints & Publications**

- CL McCombe, A Wegner, CS Zamora, F Casanova, S Aditya, JR Greenwood, L Wirtz, S Paula, E England, S Shang, DJ Ericsson, E Oliveira-Garcia, SJ Williams, U Schaffrath (2023). "Plant pathogenic fungi hijack phosphate starvation signaling with conserved enzymatic effectors" bioRxiv https://doi.org/10.1101/2023.11.14.566975
- CL McCombe, AM Catanzariti, JR Greenwood, AM Desai, MA Outram, DS Yu, DJ Ericsson, SE Brenner, PN Dodds, B Kobe, DA Jones, SJ Williams (2023). "A rust-fungus Nudix hydrolase effector decaps mRNA *in vitro* and interferes with plant immune pathways" *New Phytologist https://doi.org/10.1111/nph.18727*
- DS Yu, MA Outram, A Smith, **CL McCombe**, PB Khambalkar, SA Rima, X Sun, L Ma, DJ Ericsson, DA Jones, SJ Williams (2023). "The structural repertoire of *Fusarium oxysporum f. sp. lycopersici* effectors revealed by experimental and computational studies" *eLife* <a href="https://doi.org/10.7554/eLife.89280.1">https://doi.org/10.7554/eLife.89280.1</a>
- CL McCombe, JR Greenwood, PS Solomon, SJ Williams (2022). "Molecular plant immunity against biotrophic, hemibiotrophic, and necrotrophic fungi." Essays in Biochemistry <a href="https://doi.org/10.1042/EBC20210073">https://doi.org/10.1042/EBC20210073</a>

Awards	
Hirota Naora award - Best presentation at the RSB HDR conference	<u>2022</u>
CPG award presentation at Combio	<u>2022</u>
Runner-up best student presentation at East Coast Protein Meeting	<u>2022</u>
Postgraduate research award - Australian Institute of Nuclear Science and Engineering	<u>2020</u>
Australian Government Research Training Program stipend	<u>2020</u>
University medal – The Australian National University	<u>2019</u>
RSB director's prize in Honours – The Australian National University	<u>2019</u>
Honours scholarship - Australian Institute of Nuclear Science and Engineering	<u>2019</u>
Chancellor's letter of commendation – Flinders University	2016/2017/2018

## Research experience

# The Australian National University PhD candidate

2020 - Present

- Designed and executed experiments to investigate the enzymatic activity of uncharacterized proteins involved in plant-microbe interactions both *in vitro* and *in planta*
- Assisted in the training of new students, including a primary supervisor role for three semester-long undergraduate student projects
- Disseminated experimental results in research publications and conference presentations
- Helped manage ordering for lab consumables. Assisted in routine lab maintenance and organization

#### The Australian National University

#### Technical assistant - COVID-19 genomic sequencing

2021

• I was a member of the team responsible for COVID-19 genomic surveillance in the Australian Capital Territory during 2021. I received cDNA samples and prepared multiplexed libraries for Nanopore sequencing.

## The Australian National University

**Honours student** 

<u>2019</u>

• The Honours project aimed to determine the function of AvrM14, a protein involved in the pathogenesis of flax rust. This project had a focus on structural biology and linking protein structure to biochemical function.

## **Flinders University**

Research assistant

2018

• Worked both independently and in a team completing various general molecular biology techniques to aid in research projects

#### Research skills

A non-exhaustive list of the research techniques/tools used during my research career.

- Recombinant protein expression (E. coli) and purification
- Fast protein liquid chromatography (FPLC) using AKTA systems
- X-ray crystallography and structural biology
- Agroinfiltration of *N. benthamiana* for transient gene expression
- Various enzyme assays (especially for Nudix hydrolases)
- Plant RNA extraction, purification and RT-qPCR
- RNA-sequencing data analysis
- RNA synthesis, 5' capping, and radiolabeling
- Various general molecular biology techniques (e.g., western blotting, molecular cloning etc.)
- Nanopore library preparation for COVID-19 genome sequencing
- Python programming language (especially for organizing large datasets and data visualization)

## **Teaching experience**

## The Australian National University

2022

## General microbiology (BIOL2142) laboratory demonstrating

- Teaching and supervising students in microbiology lab classes
- Marking of scientific reports

### The Australian National University

2021 and 2022

#### Molecular gene technology (BIOL2162) workshop tutor

• Providing guidance to, and teaching, students who were completing online zoom workshops covering a variety of molecular biology research techniques

## The Australian National University

2021 - Present

## Biochemistry and human nutrition (BIOL2171) laboratory demonstrating

- Teaching and supervising students in biochemistry lab classes
- Marking of laboratory reports

#### The Australian National University

2020, 2022/23

#### Advanced studies course (SCNC2101) research project design and lab supervision

• Designing and supervising semester-long undergraduate research projects for second- and third-year students

#### Peer review

Molecular Plant Pathology Journal of Experimental Botany 2023

2021 & 2022

References available on request