

Inflammaging and the Lung



Elizabeth J. Kovacs, PhD^{a,*}, Devin M. Boe, BA^b, Lisbeth A. Boule, PhD^c,
Brenda J. Curtis, PhD^d

KEYWORDS

- Inflammaging • Elderly • Infection • Host defense • Macrophage • Neutrophils
- Inflammation • Immunosenescence

KEY POINTS

- Age-dependent changes in immune responses cause increased morbidity and mortality in the elderly.
- Inflammaging causes immunosenescence.
- Intestinal permeability in the elderly may be responsible for inflammaging.
- The ability of alveolar macrophages to maintain pulmonary homeostasis following clearance of infection is reduced in the aged.

INTRODUCTION

With advanced age, there are changes in multiple biologic systems¹ including the immune system. Alterations in innate and adaptive immune cells in the aged have been noted.^{2,3} In brief, the age-dependent effects on the innate immune response include diminished pathogen recognition, chemotaxis, and phagocytosis, and in adaptive

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^a Division of GI, Trauma and Endocrine Surgery, Department of Surgery, Mucosal Inflammation Program, GILIP (GI, Liver and Innate Immunity Program), Graduate Program in Immunology, IMAGE (Investigations in Metabolism, Aging, Gender and Exercise), University of Colorado Denver, Anschutz Medical Campus, 12700 East 19th Avenue, Research Complex 2, Mailstop #8620, Aurora, CO 80045, USA; ^b Division of GI, Trauma and Endocrine Surgery, Department of Surgery, Mucosal Inflammation Program, Graduate Program in Immunology, University of Colorado Denver, Anschutz Medical Campus, 12700 East 19th Avenue, Research Complex 2, Room 6460, Aurora, CO 80045, USA; ^c Division of GI, Trauma and Endocrine Surgery, Department of Surgery, Mucosal Inflammation Program, IMAGE, University of Colorado Denver, Anschutz Medical Campus, 12700 East 19th Avenue, Research Complex 2, Room 6460, Aurora, CO 80045, USA; ^d Division of GI, Trauma and Endocrine Surgery, Department of Surgery, Mucosal Inflammation Program, IMAGE, University of Colorado Denver, Anschutz Medical Campus, 12700 East 19th Avenue, Research Complex 2, Room 6018, Aurora, CO 80045, USA

* Corresponding author.

E-mail address: elizabeth.kovacs@ucdenver.edu

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immunity, declining numbers of naive T lymphocytes and reduced cytotoxicity and antibody quality and quantity.² Vaccine efficacy is reduced in the elderly, as are increases in autoimmunity and cancer.² Overall, these immune defects, referred to collectively as immunosenescence, render the host less able to withstand injury or infection relative to younger individuals.

Among the hallmarks of the aging immune system is the persistent low-grade proinflammatory state characterized by heightened basal levels of proinflammatory mediators in the blood.⁴ Because of this association of advanced age and inflammation, Claudio Franceschi coined the term “inflammaging” in 2000.⁴ Franceschi and coworkers⁴ have reported that, even in healthy aged subjects without confirmed ailments, there is an elevated basal level of proinflammatory mediators, including interleukin (IL)-1 β , IL-6, and tumor necrosis factor- α . The elevated levels of these and other proinflammatory factors in the aged can have local and systemic consequences, none of which are ultimately beneficial to the host. This rise in circulating levels of proinflammatory cytokines and other factors is thought by some to be a driving factor in the development and maintenance of immunosenescence^{4,5} and contribute to chronic diseases of the lung and other organs.^{6–8} In this review, the focus is on inflammaging, immunosenescence, and the lung, but it should be noted that many of the age-dependent changes are neither limited to nor likely to be caused by changes in the aging lung itself, and most of these changes are not observed unless the host is challenged by some form of stressor, such as an injury or infection.

CHANGES IN THE LUNG WITH ADVANCED AGE

A wide range of pulmonary parameters that influence lung immunity are altered with advanced age as described in [Table 1](#).

INNATE IMMUNE CELLS OF THE LUNG AND CHANGES WITH ADVANCED AGE
Macrophages

The primary resident innate immune cell in the airway is the alveolar macrophage. This multifaceted cell serves as the first line of defense against invading pathogens and

Table 1 Aging of the lung	
Lung Functions That Are Changed with Age	Reference
↓ Mucociliary escalator: reduced ability to clear microbes and debris from the airway	9,10
↑ Expression of proteins associated with bacteria attachment and infiltration in the pulmonary epithelial cells, including polymeric immunoglobulin receptor and platelet-activating factor receptor	11,12
↑ Expression of markers of cellular senescence	6,13
↓ Epithelial expression of antimicrobial peptides	14,15
↑ Levels of complement and surfactant proteins	15
↑ Proteostasis (and the loss of ability of cells from the aged to properly control protein abundance, proper folding, and degradation)	16,17
↑ Susceptibility to pulmonary infections	2,11,12,18–22
Dysbiosis (or the imbalance) of the pulmonary microbiome in the absence of infection and after infection	23–28

plays a critical role in lung immunologic homeostasis. Macrophages are capable of initiating and resolving an inflammatory response.^{29–31} This ability to play divergent roles is caused by macrophage plasticity. Macrophages can adapt and even change phenotype in response to environmental cues, enabling them to adapt to varying conditions and perform a plethora of diverse functions.^{32–35} Historically, this stimulus-induced shift in macrophage phenotype was referred to as M1 and M2 phenotypes with M1 being proinflammatory and M2 anti-inflammatory.^{36,37} However, because of poor definition and inconsistencies in the cell surface markers defining these two phenotypes, a group of expert macrophage research investigators recently redefined macrophage classification terminology so that they are more narrowly classified based on the source of the macrophages and activation stimuli, and the specific group of markers associated with the particular activation phenotype.³⁸ Regardless of nomenclature, under resting conditions, alveolar macrophages maintain an anti-inflammatory profile to keep the pulmonary airway in check and are capable of rapidly springing into action, becoming strongly proinflammatory when alerted by the presence of foreign material (**Fig. 1**). After pathogen clearance, the ability of alveolar macrophages to promote resolution and return to an anti-inflammatory resting phenotype is equally important for maintenance of lung homeostasis.

Multiple factors are involved in the resolution of inflammation in the lung. These include but are not limited to (1) clearance of the pathogen or debris; (2) reduced production of neutrophil chemokines; and (3) removal of apoptotic cells, including effete neutrophils. All of these processes are orchestrated by alveolar macrophages.³⁹ It should be noted that the inability of macrophages to perform these functions can result in prolonged inflammation, which if left unchecked can result in damage to

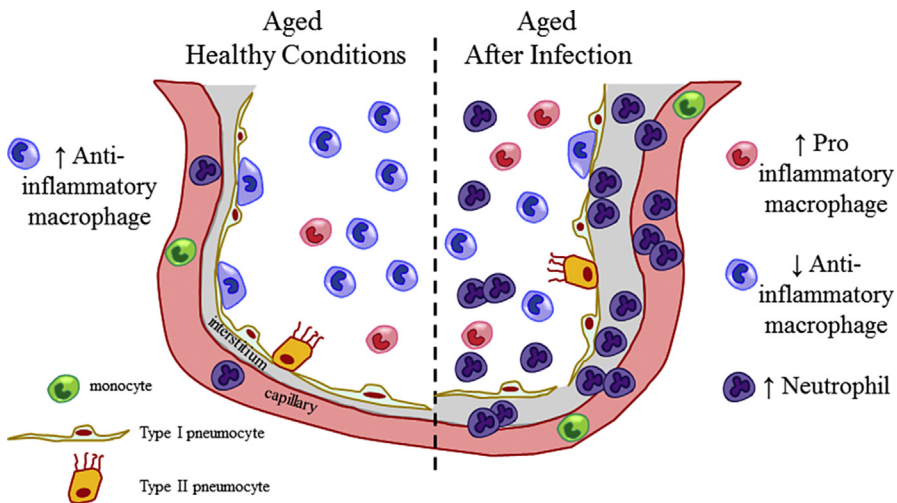


Fig. 1. Innate immune phenotype of the aged lung. Regardless of age, under healthy conditions, the major leukocyte of the distal lung is the alveolar macrophage. These multifaceted cells exist in an anti-inflammatory state to limit inflammation and maintain pulmonary homeostasis. A variety of pathogenic conditions alter alveolar macrophage function. In the young, alveolar macrophages can rapidly respond to external stimuli, such as bacteria; clear infections; and return to their anti-inflammatory state. In contrast, in the elderly, alveolar macrophages fail to mount an adequate response to infectious insult, are slow at recruiting neutrophils to help combat the respiratory pathogens, and are unable to return to their anti-inflammatory phenotype, thus leaving the lung in a compromised state.

lung tissue.⁴⁰ Central to the restoration of pulmonary homeostasis is the removal of neutrophils, which is associated with a shift in alveolar macrophages phenotype to an anti-inflammatory profile.³⁹

With advanced age, it is clear that the ability of macrophages to perform their normal functions is impaired and that inflammaging plays a role in this altered response despite the lack of change in macrophage number. A comprehensive review of macrophage function and aging is available.³⁶ In brief, in vivo and in vitro studies conducted in humans and in various animal models suggest that many but not all of the functions of macrophages are slowed or diminished in magnitude in the aged, leaving the host unable to shift between phenotypes when needed.^{2,33,36} Some of the better documented age-dependent changes in macrophage function are highlighted in [Table 2](#).

Neutrophils

The neutrophil is a key innate immune cell that is often the first cell type to be recruited to sites of injury and infection. Neutrophils are capable of performing a variety of anti-microbial functions that play a critical role in removing pathogens from tissues during the early stages of lung infections. Within minutes after recognition of foreign material, macrophages become activated and initiate a cascade of events that includes the release of chemoattractant cytokines that recruit neutrophils. Working together, macrophages and neutrophils join forces to remove and destroy infectious organisms.^{58,59} Neutrophil functions that are altered with advanced age are shown in [Table 3](#).

A PARADOX: AGING CAUSES HIGHER CYTOKINE LEVELS IN VIVO, YET REDUCED PRODUCTION BY INFLAMMATORY CELLS IN VITRO

The cellular sources of the mediators responsible for inflammaging remain unknown. Interestingly, there is a disconnect between the in vivo and in vitro effects of stimulation on the inflammatory response in young adult and older subjects and in cells isolated from those subjects. From human and rodent studies in which an inflammatory stimulus, such as lipopolysaccharide, is given in vivo, it is clear that the inflammatory response is of greater magnitude and duration in older subjects relative to younger.^{88–90} In contrast, in vitro stimulation of certain cell subsets, including blood monocytes, lung, or peritoneal macrophages from aged subjects, yields lower levels of cytokines relative to cells from younger individuals,^{44–46,91,92} suggesting either that monocyte/macrophages are not a major source of these mediators in vivo or that there are additional factors responsible for this discrepancy. The effects of aging on monocyte/macrophage functions were comprehensively reviewed elsewhere.³⁶

Table 2 Aging and macrophages	
Alterations in Macrophage Function with Advanced Age	Reference
↓ Toll-like receptor expression (mRNA and protein) and downstream signaling (in most but not all studies)	23,41–46
↓ Production of proinflammatory and immunomodulatory cytokines, including tumor necrosis factor- α , IL-6, IL-1 β , and CCL2 (monocyte chemoattractant protein-1) after stimulation by various agonists	44–50
↓ Telomere length	51
↑ Regulators of immune signaling, such as A20, a deubiquitinase that, in turn, inhibits toll-like receptor signaling and nuclear factor- κ B activation	11,12
↓ Phagocytosis and pathogen clearance	7,49,52–57

Table 3
Aging and neutrophils

Alterations in Neutrophil Function with Advanced Age	Reference
↓ Chemotaxis	60–67
No change in chemokinesis	60
↓ Phagocytosis	62,64,68–75
↓ Production of reactive oxygen species	61,62,64,70,71,76–78
↓ Generation of neutrophil extracellular traps	79–81
↓ Production of proinflammatory cytokines and mediators, including IL-6, IL-8, myeloperoxidase, elastase, and ↑ production of anti-inflammatory cytokines, IL-10	76,82,83
No increase in lifespan following stimulation	83–87

WHAT CAUSES INFLAMMAGING?

There are multiple theories about the origin and perpetuation of inflammaging. Ones that have gained press over time include classical ideas about increased oxidative stress, DNA damage, and telomere shortening.^{1,2} In brief, it is believed that with advanced age there is an increase in posttranslational modification of macromolecules including DNA, proteins, and lipids that stimulate leukocytes and other cells to secrete proinflammatory cytokines; and senescence of immune and nonimmune cells leading to an increased release of inflammatory mediators via a senescence-associated secretory phenotype.^{1,2} Additionally, a complementary and newer theory about the initiation of inflammaging is emerging and gaining support in the literature. This theory revolves around changes in intestinal permeability that allows bacteria and bacterial products (eg, endotoxin and peptidoglycan) to translocate into the lymphatic system and ultimately the bloodstream where they can trigger the low systemic inflammation in the elderly.

In brief, changes in aged intestine include dysbiosis of intestinal microbiota in animal models of aging and in elderly humans,^{93–96} and decreased integrity of the intestinal epithelial cell barrier in mice and humans.^{97–102}

AGING, DYSBIOSIS OF INTESTINAL MICROBIOME, AND THE GUT-LIVER-LUNG AXIS

Extensive clinical and experimental evidence reveals that the intestinal barrier integrity plays a role in inflammaging that in turn alters pulmonary inflammation. The gut hypothesis states that heightened intestinal permeability, along with changes in immune function of the gut, results in increased translocation of bacteria and bacterial products.^{103–105}

Like the lung, the intestine is an organ that is exposed to the outside environment with a large surface area. Although the lung and intestine provide different biologic functions, they share in common the feature of needing to maintain compartmental barriers that must remain intact to permit normal organ function to occur and to protect the host from invading pathogens. Those barriers are created by the epithelium lining the lumen of the respiratory and gastrointestinal tract. The integrity of tight junctions between adjacent epithelial cells is an essential part of these barriers. In young and the aged, this barrier is maintained in part by the complex interactions between the multiple proteins making up tight junctions, including occludins and claudins, along with multiple adaptor and scaffolding proteins. Under normal conditions, the

epithelium maintains a semipermeable barrier permitting passage of smaller molecules while preventing the movement of other materials to its underlying mucosal tissue. Regardless of the organ, breach of the epithelial barrier allows inappropriate access of microbial organisms and debris to the underlying mucosa, which can cause inflammation and tissue damage.^{106–109} The integrity of this barrier is perturbed in a plethora of disease states, such as reflux esophagitis, cancer, and inflammatory bowel disease (discussed elsewhere).^{108,109}

One mechanism of altering the epithelial status quo is mediated by the enzyme myosin light chain kinase (MLCK), the long 210-kDa form that remains inactive in the cytoplasm of epithelial (and endothelial) cells. When activated, MLCK phosphorylates myosin regulatory light-chain (MLC) at serine 19, allowing it to interact with actin. The interaction between actin and MLC causes cytoskeletal sliding, which disrupts tight junctions and creates a gap in the epithelial barrier,^{110,111} thus permitting the uncontrolled flow of fluid, bacteria, bacterial products, and other materials across the epithelial lining.^{112,113} Of interest to research on the elderly, the same set of proinflammatory mediators that are elevated in the circulation of the aged and serve as hallmarks of inflammaging, namely IL-1 β , IL-6, and tumor necrosis factor- α , can trigger the activation of MLCK. Additionally, in the lung, when MLCK is activated in the capillary lining endothelial cells, it results in paracellular permeability, which can lead to pulmonary edema.¹¹⁰ One of the consequences of the leakiness of the intestinal epithelium is the translocation of bacteria from the intestinal lumen to the underlying mucosal tissue and to regional lymph nodes. Subsequently, these products can traffic to the liver where they can stimulate production of proinflammatory cytokines (Fig. 2). If not appropriately contained by the aging immune system, the dissemination of

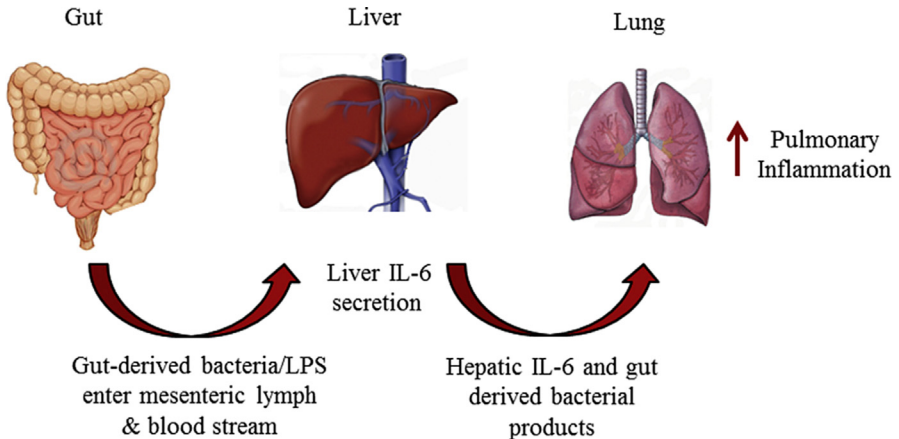


Fig. 2. The gut-liver-lung axis. Under healthy conditions, the epithelial cells lining the intestine maintain tight junctions preventing luminal contents from invading the underlying mucosal tissues. In the aged, it is thought that epithelial cell tight junctions loosen, possibly in response to the presence of the proinflammatory cytokines associated with inflammaging. This loosening of junctional complexes and subsequent increase in paracellular permeability allows gut-derived bacteria, bacterial products, and endotoxins to enter the mesenteric lymph and the bloodstream. Bacteria and their products then trigger Kupffer cells and other cells in the liver to produce and secrete proinflammatory cytokines, including IL-6. Hepatic-derived IL-6, along with the gut-derived bacteria products in the circulation, promotes baseline lung inflammation, which can then be further exacerbated in the aged after injury or infection. LPS, lipopolysaccharide.

bacteria and/or release of bacteria and bacterial products, such as endotoxins, throughout the body can occur, leading to prolonged and exacerbated inflammation in all organs, which likely contributes to increased morbidity and mortality in the aged. Hence, the intestine and its microbial contents can play a critical role in inducing or exacerbating complications in various patient populations^{111,114} and in the aged.^{101,102,115–117}

SUMMARY AND FUTURE DIRECTIONS

Factors or treatments that reduce inflammaging are of interest to basic and clinical researchers because they may be able to dampen the prolonged and heightened inflammation seen in the elderly after attempting to combat an infection. Thoughts about the design of therapeutic interventions to reduce inflammaging can be directed either at cells themselves or the proinflammatory environment in which they reside. Animal studies involving adoptive transfer of subsets of leukocytes are in progress, as are numerous clinical and basic research studies investigating antioxidant and anti-inflammatory agents to attenuate the overexuberant inflammatory response in the aged. Some believe that taking the indirect approach of reducing intestinal inflammation or restoring the intestinal microbiota may have benefit, but this is not without controversy.^{118–120} It would be of interest to determine if patients receiving anti-inflammatory therapies for other conditions have restored intestinal barrier function and if this, in turn, improves systemic responses to the injury or infection in the aged population. Further exploration of these direct and indirect avenues of therapeutic manipulation may be of benefit to the overall health of the aged and with that will likely improve overall lung health of the elderly.

REFERENCES

1. Lopez-Otin C, Blasco MA, Partridge L, et al. The hallmarks of aging. *Cell* 2013; 153(6):1194–217.
2. Frasca D, Blomberg BB. Inflammaging decreases adaptive and innate immune responses in mice and humans. *Biogerontology* 2016;17(1):7–19.
3. Shaw AC, Goldstein DR, Montgomery RR. Age-dependent dysregulation of innate immunity. *Nat Rev Immunol* 2013;13(12):875–87.
4. Franceschi C, Bonafe M, Valensin S, et al. Inflamm-aging. An evolutionary perspective on immunosenescence. *Ann N Y Acad Sci* 2000;908:244–54.
5. Solana R, Tarazona R, Gayoso I, et al. Innate immunosenescence: effect of aging on cells and receptors of the innate immune system in humans. *Semin Immunol* 2012;24(5):331–41.
6. Franceschi C, Campisi J. Chronic inflammation (inflammaging) and its potential contribution to age-associated diseases. *J Gerontol A Biol Sci Med Sci* 2014; 69(Suppl 1):S4–9.
7. Hearps AC, Martin GE, Angelovich TA, et al. Aging is associated with chronic innate immune activation and dysregulation of monocyte phenotype and function. *Aging Cell* 2012;11(5):867–75.
8. Murray MA, Chotirmall SH. The impact of immunosenescence on pulmonary disease. *Mediators Inflamm* 2015;2015:692546.
9. Ho JC, Chan KN, Hu WH, et al. The effect of aging on nasal mucociliary clearance, beat frequency, and ultrastructure of respiratory cilia. *Am J Respir Crit Care Med* 2001;163(4):983–8.
10. Svartengren M, Falk R, Philipson K. Long-term clearance from small airways decreases with age. *Eur Respir J* 2005;26(4):609–15.

11. Hinojosa CA, Akula Suresh Babu R, Rahman MM, et al. Elevated A20 contributes to age-dependent macrophage dysfunction in the lungs. *Exp Gerontol* 2014;54:58–66.
12. Hinojosa E, Boyd AR, Orihuela CJ. Age-associated inflammation and toll-like receptor dysfunction prime the lungs for pneumococcal pneumonia. *J Infect Dis* 2009;200(4):546–54.
13. Shivshankar P, Boyd AR, Le Saux CJ, et al. Cellular senescence increases expression of bacterial ligands in the lungs and is positively correlated with increased susceptibility to pneumococcal pneumonia. *Aging Cell* 2011;10(5):798–806.
14. Simell B, Vuorela A, Ekstrom N, et al. Aging reduces the functionality of anti-pneumococcal antibodies and the killing of *Streptococcus pneumoniae* by neutrophil phagocytosis. *Vaccine* 2011;29(10):1929–34.
15. Moliva JI, Rajaram MV, Sidiki S, et al. Molecular composition of the alveolar lining fluid in the aging lung. *Age* 2014;36(3):9633.
16. Kaushik S, Cuervo AM. Proteostasis and aging. *Nat Med* 2015;21(12):1406–15.
17. Meiners S, Eickelberg O, Konigshoff M. Hallmarks of the ageing lung. *Eur Respir J* 2015;45(3):807–27.
18. Boyd AR, Shivshankar P, Jiang S, et al. Age-related defects in TLR2 signaling diminish the cytokine response by alveolar macrophages during murine pneumococcal pneumonia. *Exp Gerontol* 2012;47(7):507–18.
19. Verschoor CP, Johnstone J, Loeb M, et al. Anti-pneumococcal deficits of monocyte-derived macrophages from the advanced-age, frail elderly and related impairments in PI3K-AKT signaling. *Hum Immunol* 2014;75(12):1192–6.
20. Kline KA, Bowdish DM. Infection in an aging population. *Curr Opin Microbiol* 2016;29:63–7.
21. Chen MM, Palmer JL, Plackett TP, et al. Age-related differences in the neutrophil response to pulmonary pseudomonas infection. *Exp Gerontol* 2014;54:42–6.
22. Jackaman C, Radley-Crabb HG, Soffe Z, et al. Targeting macrophages rescues age-related immune deficiencies in C57BL/6J geriatric mice. *Aging Cell* 2013;12(3):345–57.
23. Stearns JC, Davidson CJ, McKeon S, et al. Culture and molecular-based profiles show shifts in bacterial communities of the upper respiratory tract that occur with age. *ISME J* 2015;9(5):1246–59.
24. de Steenhuijsen Pters WA, Huijskens EG, Wyllie AL, et al. Dysbiosis of upper respiratory tract microbiota in elderly pneumonia patients. *ISME J* 2016;10(1):97–108.
25. Krone CL, Biesbroek G, Trzcinski K, et al. Respiratory microbiota dynamics following *Streptococcus pneumoniae* acquisition in young and elderly mice. *Infect Immun* 2014;82(4):1725–31.
26. Thevaranjan N, Whelan FJ, Puchta A, et al. *Streptococcus pneumoniae* colonization disrupts the microbial community within the upper respiratory tract of aging mice. *Infect Immun* 2016;84(4):906–16.
27. Krone CL, Trzcinski K, Zborowski T, et al. Impaired innate mucosal immunity in aged mice permits prolonged *Streptococcus pneumoniae* colonization. *Infect Immun* 2013;81(12):4615–25.
28. Whelan FJ, Verschoor CP, Stearns JC, et al. The loss of topography in the microbial communities of the upper respiratory tract in the elderly. *Ann Am Thorac Soc* 2014;11(4):513–21.
29. Herold S, Mayer K, Lohmeyer J. Acute lung injury: how macrophages orchestrate resolution of inflammation and tissue repair. *Front Immunol* 2011;2:65.

30. Aggarwal NR, King LS, D'Alessio FR. Diverse macrophage populations mediate acute lung inflammation and resolution. *Am J Physiol Lung Cell Mol Physiol* 2014;306(8):L709–25.
31. Porcheray F, Viaud S, Rimaniol AC, et al. Macrophage activation switching: an asset for the resolution of inflammation. *Clin Exp Immunol* 2005;142(3):481–9.
32. Glezeva N, Horgan S, Baugh JA. Monocyte and macrophage subsets along the continuum to heart failure: misguided heroes or targetable villains? *J Mol Cell Cardiol* 2015;89(Pt B):136–45.
33. Malyshev I, Malyshev Y. Current concept and update of the macrophage plasticity concept: intracellular mechanisms of reprogramming and M3 macrophage “switch” Phenotype. *Biomed Res Int* 2015;2015:341308.
34. Das A, Sinha M, Datta S, et al. Monocyte and macrophage plasticity in tissue repair and regeneration. *Am J Pathol* 2015;185(10):2596–606.
35. Mantovani A, Biswas SK, Galdiero MR, et al. Macrophage plasticity and polarization in tissue repair and remodelling. *J Pathol* 2013;229(2):176–85.
36. Albright JM, Dunn RC, Shults JA, et al. Advanced age alters monocyte and macrophage responses. *Antioxid Redox Signal* 2016;25(15):805–15.
37. Sica A, Mantovani A. Macrophage plasticity and polarization: in vivo veritas. *J Clin Invest* 2012;122(3):787–95.
38. Murray PJ, Allen JE, Biswas SK, et al. Macrophage activation and polarization: nomenclature and experimental guidelines. *Immunity* 2014;41(1):14–20.
39. Ariel A, Maridonneau-Parini I, Rovere-Querini P, et al. Macrophages in inflammation and its resolution. *Front Immunol* 2012;3:324.
40. Sindrilaru A, Peters T, Wieschalka S, et al. An unrestrained proinflammatory M1 macrophage population induced by iron impairs wound healing in humans and mice. *J Clin Invest* 2011;121(3):985–97.
41. Shaw AC, Panda A, Joshi SR, et al. Dysregulation of human toll-like receptor function in aging. *Ageing Res Rev* 2011;10(3):346–53.
42. De Nardo D. Toll-like receptors: activation, signalling and transcriptional modulation. *Cytokine* 2015;74(2):181–9.
43. Kaparakis M, Philpott DJ, Ferrero RL. Mammalian NLR proteins; discriminating foe from friend. *Immunol Cell Biol* 2007;85(6):495–502.
44. Renshaw M, Rockwell J, Engleman C, et al. Cutting edge: impaired toll-like receptor expression and function in aging. *J Immunol* 2002;169(9):4697–701.
45. Boehmer ED, Goral J, Faunce DE, et al. Age-dependent decrease in toll-like receptor 4-mediated proinflammatory cytokine production and mitogen-activated protein kinase expression. *J Leukoc Biol* 2004;75(2):342–9.
46. Boehmer ED, Meehan MJ, Cutro BT, et al. Aging negatively skews macrophage TLR2- and TLR4-mediated pro-inflammatory responses without affecting the IL-2-stimulated pathway. *Mech Ageing Dev* 2005;126(12):1305–13.
47. Chelvarajan RL, Collins SM, Van Willigen JM, et al. The unresponsiveness of aged mice to polysaccharide antigens is a result of a defect in macrophage function. *J Leukoc Biol* 2005;77(4):503–12.
48. Chelvarajan RL, Liu Y, Popa D, et al. Molecular basis of age-associated cytokine dysregulation in LPS-stimulated macrophages. *J Leukoc Biol* 2006;79(6):1314–27.
49. Liang S, Domon H, Hosur KB, et al. Age-related alterations in innate immune receptor expression and ability of macrophages to respond to pathogen challenge in vitro. *Mech Ageing Dev* 2009;130(8):538–46.
50. Mahbub S, Deburghgraeve CR, Kovacs EJ. Advanced age impairs macrophage polarization. *J Interferon Cytokine Res* 2012;32(1):18–26.

51. Arai Y, Martin-Ruiz CM, Takayama M, et al. Inflammation, but not telomere length, predicts successful ageing at extreme old age: a longitudinal study of semi-supercentenarians. *EBioMedicine* 2015;2(10):1549–58.
52. Swift ME, Kleinman HK, DiPietro LA. Impaired wound repair and delayed angiogenesis in aged mice. *Lab Invest* 1999;79(12):1479–87.
53. Albright JF, Albright JW. Senescence of natural/innate resistance to infection. Totowa (NY): Humana Press, Inc; 2003.
54. Lynch AM, Murphy KJ, Deighan BF, et al. The impact of glial activation in the aging brain. *Aging Dis* 2010;1(3):262–78.
55. Linehan E, Dombrowski Y, Snoddy R, et al. Aging impairs peritoneal but not bone marrow-derived macrophage phagocytosis. *Aging Cell* 2014;13(4):699–708.
56. Aprahamian T, Takemura Y, Goukassian D, et al. Ageing is associated with diminished apoptotic cell clearance in vivo. *Clin Exp Immunol* 2008;152(3):448–55.
57. Arnardottir HH, Dalli J, Colas RA, et al. Aging delays resolution of acute inflammation in mice: reprogramming the host response with novel nano-proresolving medicines. *J Immunol* 2014;193(8):4235–44.
58. Silva MT. When two is better than one: macrophages and neutrophils work in concert in innate immunity as complementary and cooperative partners of a myeloid phagocyte system. *J Leukoc Biol* 2010;87(1):93–106.
59. Silva MT, Correia-Neves M. Neutrophils and macrophages: the main partners of phagocyte cell systems. *Front Immunol* 2012;3:174.
60. Sapey E, Greenwood H, Walton G, et al. Phosphoinositide 3-kinase inhibition restores neutrophil accuracy in the elderly: toward targeted treatments for immunosenescence. *Blood* 2014;123(2):239–48.
61. Di Lorenzo G, Balistreri CR, Candore G, et al. Granulocyte and natural killer activity in the elderly. *Mech Ageing Dev* 1999;108(1):25–38.
62. Polignano A, Tortorella C, Venezia A, et al. Age-associated changes of neutrophil responsiveness in a human healthy elderly population. *Cytobios* 1994;80(322):145–53.
63. McLaughlin B, O'Malley K, Cotter TG. Age-related differences in granulocyte chemotaxis and degranulation. *Clin Sci* 1986;70(1):59–62.
64. Antonaci S, Jirillo E, Ventura MT, et al. Non-specific immunity in aging: deficiency of monocyte and polymorphonuclear cell-mediated functions. *Mech Ageing Dev* 1984;24(3):367–75.
65. Nomellini V, Brubaker AL, Mahbub S, et al. Dysregulation of neutrophil CXCR2 and pulmonary endothelial ICAM-1 promotes age-related pulmonary inflammation. *Aging Dis* 2012;3(3):234–47.
66. Nomellini V, Faunce DE, Gomez CR, et al. An age-associated increase in pulmonary inflammation after burn injury is abrogated by CXCR2 inhibition. *J Leukoc Biol* 2008;83(6):1493–501.
67. Brubaker AL, Rendon JL, Ramirez L, et al. Reduced neutrophil chemotaxis and infiltration contributes to delayed resolution of cutaneous wound infection with advanced age. *J Immunol* 2013;190(4):1746–57.
68. Butcher SK, Chahal H, Nayak L, et al. Senescence in innate immune responses: reduced neutrophil phagocytic capacity and CD16 expression in elderly humans. *J Leukoc Biol* 2001;70(6):881–6.
69. Butcher SK, Killampalli V, Chahal H, et al. Effect of age on susceptibility to post-traumatic infection in the elderly. *Biochem Soc Trans* 2003;31(2):449–51.

70. Fulop T Jr, Foris G, Worum I, et al. Age-dependent alterations of Fc gamma receptor-mediated effector functions of human polymorphonuclear leucocytes. *Clin Exp Immunol* 1985;61(2):425–32.
71. Wenisch C, Patruta S, Daxbock F, et al. Effect of age on human neutrophil function. *J Leukoc Biol* 2000;67(1):40–5.
72. Amaya RA, Baker CJ, Keitel WA, et al. Healthy elderly people lack neutrophil-mediated functional activity to type V group B *Streptococcus*. *J Am Geriatr Soc* 2004;52(1):46–50.
73. Butcher S, Chahel H, Lord JM. Review article: ageing and the neutrophil: no appetite for killing? *Immunology* 2000;100(4):411–6.
74. Alonso-Fernandez P, Puerto M, Mate I, et al. Neutrophils of centenarians show function levels similar to those of young adults. *J Am Geriatr Soc* 2008;56(12):2244–51.
75. Esparza B, Sanchez H, Ruiz M, et al. Neutrophil function in elderly persons assessed by flow cytometry. *Immunol Invest* 1996;25(3):185–90.
76. Dalboni TM, Abe AE, de Oliveira CE, et al. Activation profile of CXCL8-stimulated neutrophils and aging. *Cytokine* 2013;61(3):716–9.
77. Tortorella C, Ottolenghi A, Pugliese P, et al. Relationship between respiratory burst and adhesiveness capacity in elderly polymorphonuclear cells. *Mech Ageing Dev* 1993;69(1–2):53–63.
78. Fu YK, Arkins S, Li YM, et al. Reduction in superoxide anion secretion and bactericidal activity of neutrophils from aged rats: reversal by the combination of gamma interferon and growth hormone. *Infect Immun* 1994;62(1):1–8.
79. Kruger P, Saffarzadeh M, Weber AN, et al. Neutrophils: between host defense, immune modulation, and tissue injury. *PLoS Pathog* 2015;11(3):e1004651.
80. Tseng CW, Kyme PA, Arruda A, et al. Innate immune dysfunctions in aged mice facilitate the systemic dissemination of methicillin-resistant *S. aureus*. *PLoS One* 2012;7(7):e41454.
81. Hazeldine J, Harris P, Chapple IL, et al. Impaired neutrophil extracellular trap formation: a novel defect in the innate immune system of aged individuals. *Aging Cell* 2014;13(4):690–8.
82. Qian F, Guo X, Wang X, et al. Reduced bioenergetics and toll-like receptor 1 function in human polymorphonuclear leukocytes in aging. *Aging (Albany NY)* 2014;6(2):131–9.
83. Schroder AK, von der Ohe M, Kolling U, et al. Polymorphonuclear leucocytes selectively produce anti-inflammatory interleukin-1 receptor antagonist and chemokines, but fail to produce pro-inflammatory mediators. *Immunology* 2006;119(3):317–27.
84. Fulop T Jr, Fouquet C, Allaire P, et al. Changes in apoptosis of human polymorphonuclear granulocytes with aging. *Mech Ageing Dev* 1997;96(1–3):15–34.
85. Tortorella C, Simone O, Piazzolla G, et al. Role of phosphoinositide 3-kinase and extracellular signal-regulated kinase pathways in granulocyte macrophage-colony-stimulating factor failure to delay fas-induced neutrophil apoptosis in elderly humans. *J Gerontol A Biol Sci Med Sci* 2006;61(11):1111–8.
86. Fortin CF, Lesur O, Fulop T Jr. Effects of aging on triggering receptor expressed on myeloid cells (TREM)-1-induced PMN functions. *FEBS Lett* 2007;581(6):1173–8.
87. Wessels I, Jansen J, Rink L, et al. Immunosenescence of polymorphonuclear neutrophils. *ScientificWorldJournal* 2010;10:145–60.
88. Gomez CR, Goral J, Ramirez L, et al. Aberrant acute-phase response in aged interleukin-6 knockout mice. *Shock* 2006;25(6):581–5.

89. Gomez CR, Hirano S, Cutro BT, et al. Advanced age exacerbates the pulmonary inflammatory response after lipopolysaccharide exposure. *Crit Care Med* 2007; 35(1):246–51.
90. Gomez CR, Nomellini V, Baila H, et al. Comparison of the effects of aging and IL-6 on the hepatic inflammatory response in two models of systemic injury: scald injury versus i.p. LPS administration. *Shock* 2009;31(2):178–84.
91. Plowden J, Renshaw-Hoelscher M, Engleman C, et al. Innate immunity in aging: impact on macrophage function. *Aging Cell* 2004;3(4):161–7.
92. Gomez CR, Karavitis J, Palmer JL, et al. Interleukin-6 contributes to age-related alteration of cytokine production by macrophages. *Mediators Inflamm* 2010; 2010:475139.
93. Kim KA, Jeong JJ, Yoo SY, et al. Gut microbiota lipopolysaccharide accelerates inflamm-aging in mice. *BMC Microbiol* 2016;16(1):9.
94. Claesson MJ, Cusack S, O'Sullivan O, et al. Composition, variability, and temporal stability of the intestinal microbiota of the elderly. *Proc Natl Acad Sci U S A* 2011;108(Suppl 1):4586–91.
95. Claesson MJ, Jeffery IB, Conde S, et al. Gut microbiota composition correlates with diet and health in the elderly. *Nature* 2012;488(7410):178–84.
96. Langille MG, Meehan CJ, Koenig JE, et al. Microbial shifts in the aging mouse gut. *Microbiome* 2014;2(1):50.
97. Man AL, Bertelli E, Rentini S, et al. Age-associated modifications of intestinal permeability and innate immunity in human small intestine. *Clin Sci* 2015; 129(7):515–27.
98. Cesar Machado MC, da Silva FP. Intestinal barrier dysfunction in human pathology and aging. *Curr Pharm Des* 2016;22(30):4645–50.
99. Pasternak JA, Kent-Dennis C, Van Kessel AG, et al. Claudin-4 undergoes age-dependent change in cellular localization on pig jejunal villous epithelial cells, independent of bacterial colonization. *Mediators Inflamm* 2015;2015:263629.
100. Valentini L, Ramminger S, Haas V, et al. Small intestinal permeability in older adults. *Physiol Rep* 2014;2(4):e00281.
101. Mabbott NA. A breakdown in communication? Understanding the effects of aging on the human small intestine epithelium. *Clin Sci* 2015;129(7):529–31.
102. Mabbott NA, Kobayashi A, Sehgal A, et al. Aging and the mucosal immune system in the intestine. *Biogerontology* 2015;16(2):133–45.
103. Magnotti LJ, Deitch EA. Burns, bacterial translocation, gut barrier function, and failure. *J Burn Care Rehabil* 2005;26(5):383–91.
104. Deitch EA. Bacterial translocation or lymphatic drainage of toxic products from the gut: what is important in human beings? *Surgery* 2002;131(3):241–4.
105. Deitch EA. Gut-origin sepsis: evolution of a concept. *Surgeon* 2012;10(6):350–6.
106. Ivanov AI. Structure and regulation of intestinal epithelial tight junctions: current concepts and unanswered questions. *Adv Exp Med Biol* 2012;763:132–48.
107. Ulluwishewa D, Anderson RC, McNabb WC, et al. Regulation of tight junction permeability by intestinal bacteria and dietary components. *J Nutr* 2011; 141(5):769–76.
108. Hallstrand TS, Hackett TL, Altemeier WA, et al. Airway epithelial regulation of pulmonary immune homeostasis and inflammation. *Clin Immunol* 2014;151(1): 1–15.
109. Oshima T, Miwa H. Gastrointestinal mucosal barrier function and diseases. *J Gastroenterol* 2016;51(8):768–78.
110. Dudek SM, Garcia JG. Cytoskeletal regulation of pulmonary vascular permeability. *J Appl Physiol* (1985) 2001;91(4):1487–500.

111. Odenwald MA, Turner JR. Intestinal permeability defects: is it time to treat? *Clin Gastroenterol Hepatol* 2013;11(9):1075–83.
112. Turner JR. Molecular basis of epithelial barrier regulation: from basic mechanisms to clinical application. *Am J Pathol* 2006;169(6):1901–9.
113. Turner JR, Rill BK, Carlson SL, et al. Physiological regulation of epithelial tight junctions is associated with myosin light-chain phosphorylation. *Am J Physiol* 1997;273(4 Pt 1):C1378–85.
114. Mittal R, Coopersmith CM. Redefining the gut as the motor of critical illness. *Trends Mol Med* 2014;20(4):214–23.
115. Man AL, Gicheva N, Nicoletti C. The impact of ageing on the intestinal epithelial barrier and immune system. *Cell Immunol* 2014;289(1–2):112–8.
116. Nicoletti C. Age-associated changes of the intestinal epithelial barrier: local and systemic implications. *Expert Rev Gastroenterol Hepatol* 2015;9(12):1467–9.
117. Schiffrin EJ, Morley JE, Donnet-Hughes A, et al. The inflammatory status of the elderly: the intestinal contribution. *Mutat Res* 2010;690(1–2):50–6.
118. Arbolea S, Watkins C, Stanton C, et al. Gut bifidobacteria populations in human health and aging. *Front Microbiol* 2016;7:1204.
119. Salazar N, Valdes-Varela L, Gonzalez S, et al. Nutrition and the gut microbiome in the elderly. *Gut Microbes* 2016;8(2):1–16.
120. van Beek AA, Sovran B, Hugenholtz F, et al. Supplementation with *Lactobacillus plantarum* WCFS1 prevents decline of mucus barrier in colon of accelerated aging Ercc1-Delta7 mice. *Front Immunol* 2016;7:408.