



# **Clinical Genomics – Early Stage Cancer Screening**

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    - I. Sun et al.
    - II. Qin et al.
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# Existing Studies

**A population-scale analysis of 36 gut microbiome studies reveals universal species signatures for common diseases.**

Sun, W., Zhang, Y., Guo, R. Et al.  
2024

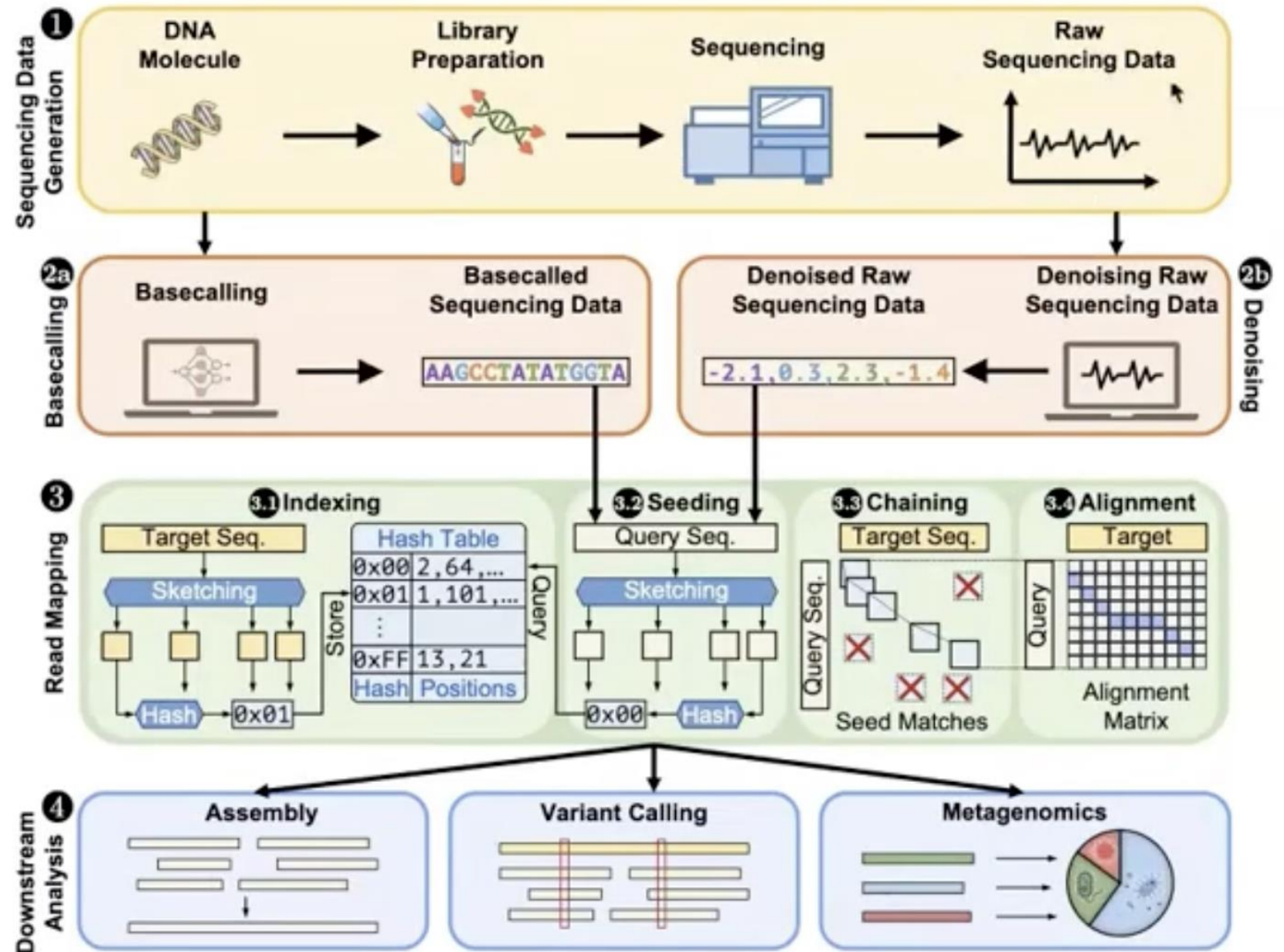
**Consistent signatures in the human gut microbiome of old- and young-onset colorectal cancer.**

Qin, Y., Tong, X., Mei, WJ. et al.  
2024

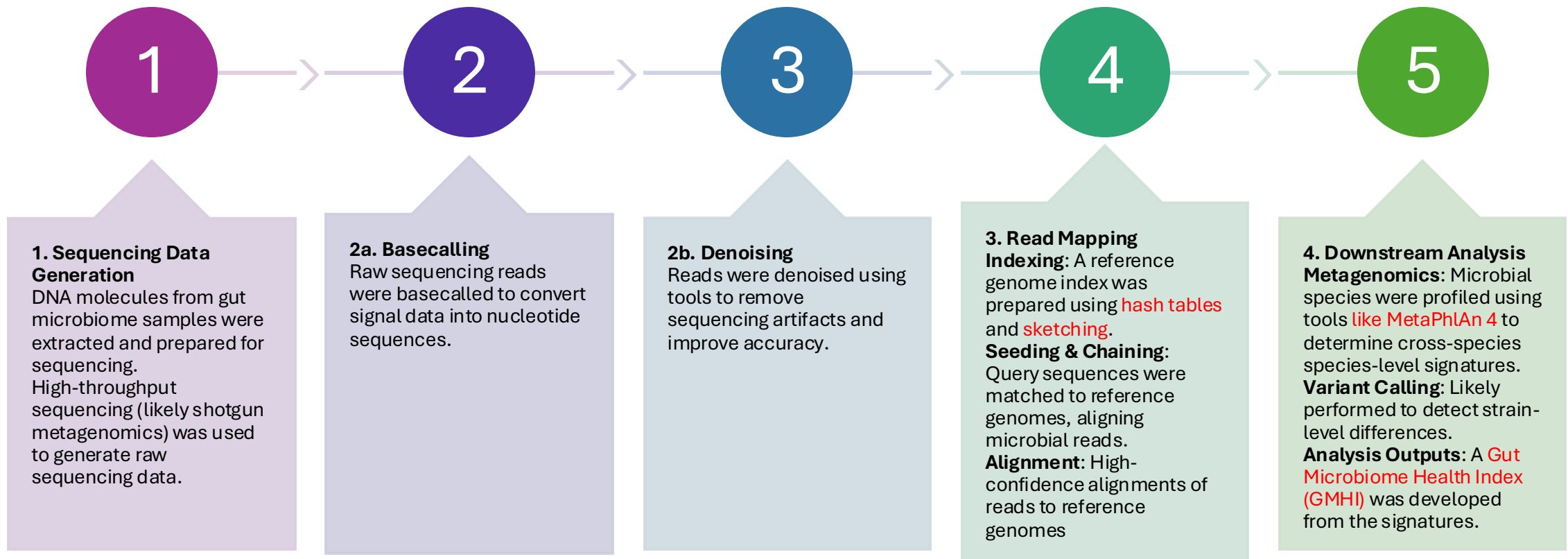
**Microbiome confounders and quantitative profiling challenge predicted microbial targets in colorectal cancer development.**

Tito, R.Y., Verbandt, S., Aguirre Vazquez, M. et al.

# Genome Analysis Pipeline



# Study 1 – Gut Microbiome and Common Diseases



# Study 2 – Old and Young Onset CRC



## 1. Sequencing Data Generation

DNA was extracted from stool samples of CRC patients and controls.

Shotgun sequencing was used to capture microbial communities.



## 2a. Basecalling

Raw sequencing reads were basecalled to nucleotide sequences.



## 2b. Denoising

Reads were denoised to filter noise and technical artifacts.



## 3. Read Mapping

**Indexing:** Reference genomes were indexed using microbial species and strain databases.

**Seeding & Chaining:** Query reads were matched to microbial strains.

**Alignment:** Reads were aligned to genomes for strain-level comparisons using tools like StrainPhlAn3.



## 4. Downstream Analysis

**Metagenomics:** Microbial strain-level profiling was conducted to detect differences between CRC onset groups.

**Assembly:** Possibly used to reconstruct microbial genomes for further strain analysis.

**Variant Calling:** Identified variations associated with disease progression.



# Study 3 – CRC Microbiome Confounders and Predictions

## 1. Sequencing Data Generation

16S **rRNA sequencing** was performed on CRC patient and control samples to capture bacterial taxa.

## 2a. Basecalling

Basecalling converted raw signal data to nucleotide sequences for 16S regions.

## 2b. Denoising

Reads were denoised and processed to remove PCR and sequencing artifacts, ensuring reliable taxonomic assignment.

## 3. Read Mapping

**Indexing:** **Reference databases** for 16S rRNA regions were prepared.

**Seeding & Chaining:** Sequence reads were matched to the reference 16S sequences.

**Alignment:** **High-confidence alignment matrices** were generated to identify bacterial taxa.

## 4. Downstream Analysis

**Metagenomics:** Quantitative profiling was used to **detect species- and genus-level biomarkers**.

**Confounder Analysis:** Tools like **GLMs** were applied to control for confounders such as transit time and inflammation.

**Variant Calling:** Likely omitted due to the focus on 16S data rather than whole genomes.

# Working Trajectory

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Data Acquisition

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Data Understanding and Preprocessing

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Implementing and Understanding Preprocessing Steps (study inspired)

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Initial ML Exploration using R Scripts from Original Studies



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**Training XGBoost in Python**

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**Evaluation and Performance Metrics**

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Mitigating Bias and Expanding Scope

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Final Testing and Documentation



# **GitHub Presentation**



# Reflection

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- Results achieved in time we had are unfortunately inconclusive
- Potentially the issue lies in the metadata and labelling of the data from the study we used
- Outlook and Improvements:
  - Attempt running different algorithms on the data and compare and contrast the results
  - Try running XGboost on data from several different studies to compare/contrast/troubleshoot
  - Take data that only checks for CRC, not other diseases (at least for primary training)
  - Preferably we would have taken more time to access the data from Belgian study as it seemed to have most controls and had clearer labeling