

Comparative functional enrichment analysis between subciliary locations for all proteins and cell types combined

Konstantin Kahnert

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Load libraries

```
library(tidyverse)
library(clusterProfiler)
library(enrichplot)
library(org.Hs.eg.db)
library(Cairo)
library(svglite)
library(VennDiagram)
library(grid)
library(simplifyEnrichment)
```

Define function

```
# map_cluster_number:
# - x: enrichResult or compareClusterResult (from clusterProfiler)
# - df: simplifyGO result data.frame with columns ID (term IDs) and Cluster_num
# - comp: if TRUE, use x@compareClusterResult, else x@result
# Returns a data.frame of all enrich columns + term_size + Cluster_num
map_cluster_number <- function(x,
                               df,
                               comp = FALSE) {
  ## 1. Standardize the input DF's column names
  colnames(df) <- c("ID", "Cluster_num")

  ## 2. Select the correct slot from the clusterProfiler object
  if (comp) {
    # from compareClusterResult
    results <- x@compareClusterResult
  } else {
    # from a single enrichResult
    results <- x@result
  }

  ## 3. Get term_size (the numerator of the "BgRatio" string)
```

```

## e.g. "12/21273" → 12
results$term_size <- as.numeric(
  sapply(strsplit(results$BgRatio, "/"),
    function(parts) as.numeric(parts[1]))
)

## 4. Merge the enrichment results with the cluster assignments
merged_results <- merge(
  results,
  df,
  by = "ID",
  all.x = TRUE
)

## 5. Return the processed data frame
return(merged_results)
}

# Function to map Ensembl IDs to gene symbols for a single row
map_geneID_to_symbol <- function(geneID_str) {
  geneIDs <- unlist(strsplit(geneID_str, "/"))
  mapped_ids <- bitr(geneIDs,
    fromType = "ENSEMBL",
    toType = "SYMBOL",
    OrgDb = org.Hs.eg.db)

  # drop duplicates in ensembl column and keep first occurrence
  mapped_ids <- mapped_ids[!duplicated(mapped_ids$ENSEMBL),]

  gene_symbols <- mapped_ids$SYMBOL
  return(paste(gene_symbols, collapse = "/"))
}

# Function to map Ensembl IDs to gene symbols for a single row
map_geneID_to_name <- function(geneIDs) {
  mapped_ids <- bitr(geneIDs,
    fromType = "ENSEMBL",
    toType = c("GENENAME", "SYMBOL"),
    OrgDb = org.Hs.eg.db)

  # drop duplicates in ensembl column and keep first occurrence
  mapped_ids <- mapped_ids[!duplicated(mapped_ids$ENSEMBL),]

  return(mapped_ids)
}

```

Data loading and preprocessing

Set input and output paths

```
# set input and output paths
in_path <- "/mnt/Data/Projects/Cilia/revision/NonRestricted/data/"
out_path <- "/mnt/Data/Projects/Cilia/revision/NonRestricted/analysis/GO_BP/"
```

Load data

```
# Load the data
df_all <- read.delim(paste0(in_path, "All_files_combined_as_cytoscape_input.csv"), sep = "\t", header =
head(df_all))
```

```
##                               BasalBody_ BasalBody_ASC52telo
## ENSG000000001497                True                False
## ENSG000000002330                False                False
## ENSG000000002549                False                False
## ENSG000000003249                False                False
## ENSG000000003756                False                False
## ENSG000000004766                True                 False
##                               BasalBody_hTERT_RPE1_serum_starved BasalBody_RPTEC_TERT1
## ENSG000000001497                                           True                False
## ENSG000000002330                                           False                False
## ENSG000000002549                                           False                False
## ENSG000000003249                                           False                False
## ENSG000000003756                                           False                False
## ENSG000000004766                                           True                 True
##                               PrimaryCilia_ PrimaryCilia_ASC52telo
## ENSG000000001497                True                False
## ENSG000000002330                False                False
## ENSG000000002549                False                False
## ENSG000000003249                False                False
## ENSG000000003756                False                False
## ENSG000000004766                True                 True
##                               PrimaryCilia_hTERT_RPE1_serum_starved PrimaryCilia_RPTEC_TERT1
## ENSG000000001497                                           True                False
## ENSG000000002330                                           False                False
## ENSG000000002549                                           False                False
## ENSG000000003249                                           False                False
## ENSG000000003756                                           False                False
## ENSG000000004766                                           True                 True
##                               PrimaryCiliaTip_ PrimaryCiliaTip_ASC52telo
## ENSG000000001497                False                False
## ENSG000000002330                False                False
## ENSG000000002549                False                False
## ENSG000000003249                False                False
## ENSG000000003756                False                False
## ENSG000000004766                False                False
##                               PrimaryCiliaTip_hTERT_RPE1_serum_starved
```

##	ENSG00000001497		False			
##	ENSG00000002330		False			
##	ENSG00000002549		False			
##	ENSG00000003249		False			
##	ENSG00000003756		False			
##	ENSG00000004766		False			
##		PrimaryCiliaTip_RPTEC_TERT1	PrimaryCiliaTZ_			
##	ENSG00000001497	False	True			
##	ENSG00000002330	False	False			
##	ENSG00000002549	False	False			
##	ENSG00000003249	False	False			
##	ENSG00000003756	False	False			
##	ENSG00000004766	False	False			
##		PrimaryCiliaTZ_ASC52telo				
##	ENSG00000001497	False				
##	ENSG00000002330	False				
##	ENSG00000002549	False				
##	ENSG00000003249	False				
##	ENSG00000003756	False				
##	ENSG00000004766	False				
##		PrimaryCiliaTZ_hTERT_RPE1_serum_starved				
##	ENSG00000001497	True				
##	ENSG00000002330	False				
##	ENSG00000002549	False				
##	ENSG00000003249	False				
##	ENSG00000003756	False				
##	ENSG00000004766	False				
##		PrimaryCiliaTZ_RPTEC_TERT1	Nucleus	Mitotic Membrane Cytoplasm		
##	ENSG00000001497	False	True	False	False	True
##	ENSG00000002330	False	False	False	False	True
##	ENSG00000002549	False	False	False	False	True
##	ENSG00000003249	False	True	False	False	False
##	ENSG00000003756	False	True	False	True	True
##	ENSG00000004766	False	False	False	True	True
##		BasalBody_num	PrimaryCilia_num	PrimaryCiliaTip_num		
##	ENSG00000001497	1	1	0		
##	ENSG00000002330	0	0	0		
##	ENSG00000002549	0	0	0		
##	ENSG00000003249	0	0	0		
##	ENSG00000003756	0	0	0		
##	ENSG00000004766	2	3	0		
##		PrimaryCiliaTZ_num	ASC52telo	hTERT_RPE1_serum_starved		
##	ENSG00000001497	1	False	True		
##	ENSG00000002330	0	False	False		
##	ENSG00000002549	0	False	False		
##	ENSG00000003249	0	False	False		
##	ENSG00000003756	0	False	False		
##	ENSG00000004766	0	True	True		
##		RPTEC_TERT1				
##	ENSG00000001497	False				
##	ENSG00000002330	False				
##	ENSG00000002549	False				
##	ENSG00000003249	False				
##	ENSG00000003756	False				

```
## ENSG00000004766      True
```

Map gene names and symbols

```
# Identify columns that contain "num" in their names
num_columns <- grep("num", names(df_all), value = TRUE)

# Convert all other columns from string to logical
df_all <- df_all %>%
  mutate(across(-all_of(num_columns), ~ as.logical(.)))

# map gene IDs to gene names
mapped_ids <- map_geneID_to_name(rownames(df_all))

# index by Ensembl
rownames(mapped_ids) <- mapped_ids$ENSEMBL

# pull out exactly one entry per row of df_all
df_all$GeneSymbol <- mapped_ids[ rownames(df_all), "SYMBOL" ]
df_all$GeneName   <- mapped_ids[ rownames(df_all), "GENENAME" ]

# add rownames as column and reset index
df_all$Ensembl_ID <- rownames(df_all)
rownames(df_all) <- NULL

# reorder columns to have Ensembl_ID first, Symbol and Gene name first and then all other columns
df_all <- df_all %>% dplyr::select(Ensembl_ID, GeneSymbol, GeneName, everything())
```

Split data by location

```
# Perform filtering again
df_bb <- df_all %>% filter(BasalBody_ == TRUE)
df_pc <- df_all %>% filter(PrimaryCilia_ == TRUE)
df_tip <- df_all %>% filter(PrimaryCiliaTip_ == TRUE)
df_tz <- df_all %>% filter(PrimaryCiliaTZ_ == TRUE)
df_nucleus <- df_all %>% filter(Nucleus == TRUE)
df_mitotic <- df_all %>% filter(Mitotic == TRUE)
df_membrane <- df_all %>% filter(Membrane == TRUE)
df_cytoplasm <- df_all %>% filter(Cytoplasm == TRUE)

# save input data as csv file
write.csv(df_bb, file = paste0(out_path, "NonRestricted_comparison_locations_input_bb.csv"), row.names = FALSE)
write.csv(df_pc, file = paste0(out_path, "NonRestricted_comparison_locations_input_pc.csv"), row.names = FALSE)
write.csv(df_tip, file = paste0(out_path, "NonRestricted_comparison_locations_input_tip.csv"), row.names = FALSE)
write.csv(df_tz, file = paste0(out_path, "NonRestricted_comparison_locations_input_tz.csv"), row.names = FALSE)
write.csv(df_nucleus, file = paste0(out_path, "NonRestricted_comparison_locations_input_nucleus.csv"), row.names = FALSE)
write.csv(df_mitotic, file = paste0(out_path, "NonRestricted_comparison_locations_input_mitotic.csv"), row.names = FALSE)
write.csv(df_membrane, file = paste0(out_path, "NonRestricted_comparison_locations_input_membrane.csv"), row.names = FALSE)
write.csv(df_cytoplasm, file = paste0(out_path, "NonRestricted_comparison_locations_input_cytoplasm.csv"), row.names = FALSE)
```

```

# filter gene_id by location
gene_id_all <- df_all$Ensembl_ID
gene_id_bb <- df_bb$Ensembl_ID
gene_id_pc <- df_pc$Ensembl_ID
gene_id_tip <- df_tip$Ensembl_ID
gene_id_tz <- df_tz$Ensembl_ID
gene_id_nucleus <- df_nucleus$Ensembl_ID
gene_id_mitotic <- df_mitotic$Ensembl_ID
gene_id_membrane <- df_membrane$Ensembl_ID
gene_id_cytoplasm <- df_cytoplasm$Ensembl_ID

```

Combine bb & tz and pc & tip

```

# combine bb and tz
df_bb_tz <- rbind(df_bb, df_tz)

# drop duplicates based on Ensembl_ID column
df_bb_tz <- df_bb_tz[!duplicated(df_bb_tz$Ensembl_ID), ]
gene_id_bb_tz <- df_bb_tz$Ensembl_ID

# combine pc and tip
df_pc_tip <- rbind(df_pc, df_tip)

# drop duplicates based on Ensembl_ID column
df_pc_tip <- df_pc_tip[!duplicated(df_pc_tip$Ensembl_ID), ]
gene_id_pc_tip <- df_pc_tip$Ensembl_ID

# combine pc and tip and tz
df_pc_tip_tz <- rbind(df_pc, df_tip, df_tz)

# drop duplicates based on Ensembl_ID column
df_pc_tip_tz <- df_pc_tip_tz[!duplicated(df_pc_tip_tz$Ensembl_ID), ]
gene_id_pc_tip_tz <- df_pc_tip_tz$Ensembl_ID

# get intersection of all genes
gene_id_all_cilia <- Reduce(union, list(gene_id_bb_tz, gene_id_pc_tip))
print(paste("Number of genes in all cilia locations:", length(gene_id_all_cilia)))

```

```
## [1] "Number of genes in all cilia locations: 715"
```

All proteins

GO BP enrichment analysis

```

ego <- enrichGO(gene          = gene_id_all,
                 OrgDb         = org.Hs.eg.db,

```

```

        keyType      = 'ENSEMBL',
        ont           = "BP",
        pAdjustMethod = "BH",
        pvalueCutoff  = 0.01,
        qvalueCutoff  = 0.01,
        readable      = TRUE)

ego@result <- ego@result[ego@result$p.adjust < 0.01,]
ego

## #
## # over-representation test
## #
## #...@organism      Homo sapiens
## #...@ontology      BP
## #...@keytype       ENSEMBL
## #...@gene          chr [1:1360] "ENSG000000001497" "ENSG000000002330" "ENSG000000002549" ...
## #...pvalues adjusted by 'BH' with cutoff <0.01
## #...1019 enriched terms found
## 'data.frame':    1019 obs. of  12 variables:
## $ ID              : chr  "G0:0044782" "G0:0060271" "G0:0099111" "G0:0003341" ...
## $ Description      : chr  "cilium organization" "cilium assembly" "microtubule-based transport" "cilium
## $ GeneRatio        : chr  "252/1265" "238/1265" "95/1265" "94/1265" ...
## $ BgRatio          : chr  "441/21273" "411/21273" "235/21273" "272/21273" ...
## $ RichFactor       : num  0.571 0.579 0.404 0.346 0.684 ...
## $ FoldEnrichment   : num  9.61 9.74 6.8 5.81 11.51 ...
## $ zScore           : num  45.9 45 22.5 20.1 23.1 ...
## $ pvalue           : num  2.66e-195 6.63e-186 1.69e-54 6.69e-47 6.33e-46 ...
## $ p.adjust         : num  1.50e-191 1.87e-182 3.19e-51 9.45e-44 7.15e-43 ...
## $ qvalue           : num  1.01e-191 1.26e-182 2.14e-51 6.34e-44 4.80e-43 ...
## $ geneID           : chr  "ZMYND10/MARK4/FUZ/MKS1/EHD3/NUDCD3/EHD2/IFT88/CLXN/OFD1/MAP4/LIMA1/MPHOSPH9
## $ Count            : int  252 238 95 94 52 96 50 40 68 64 ...
## #...Citation
## S Xu, E Hu, Y Cai, Z Xie, X Luo, L Zhan, W Tang, Q Wang, B Liu, R Wang, W Xie, T Wu, L Xie, G Yu. Us

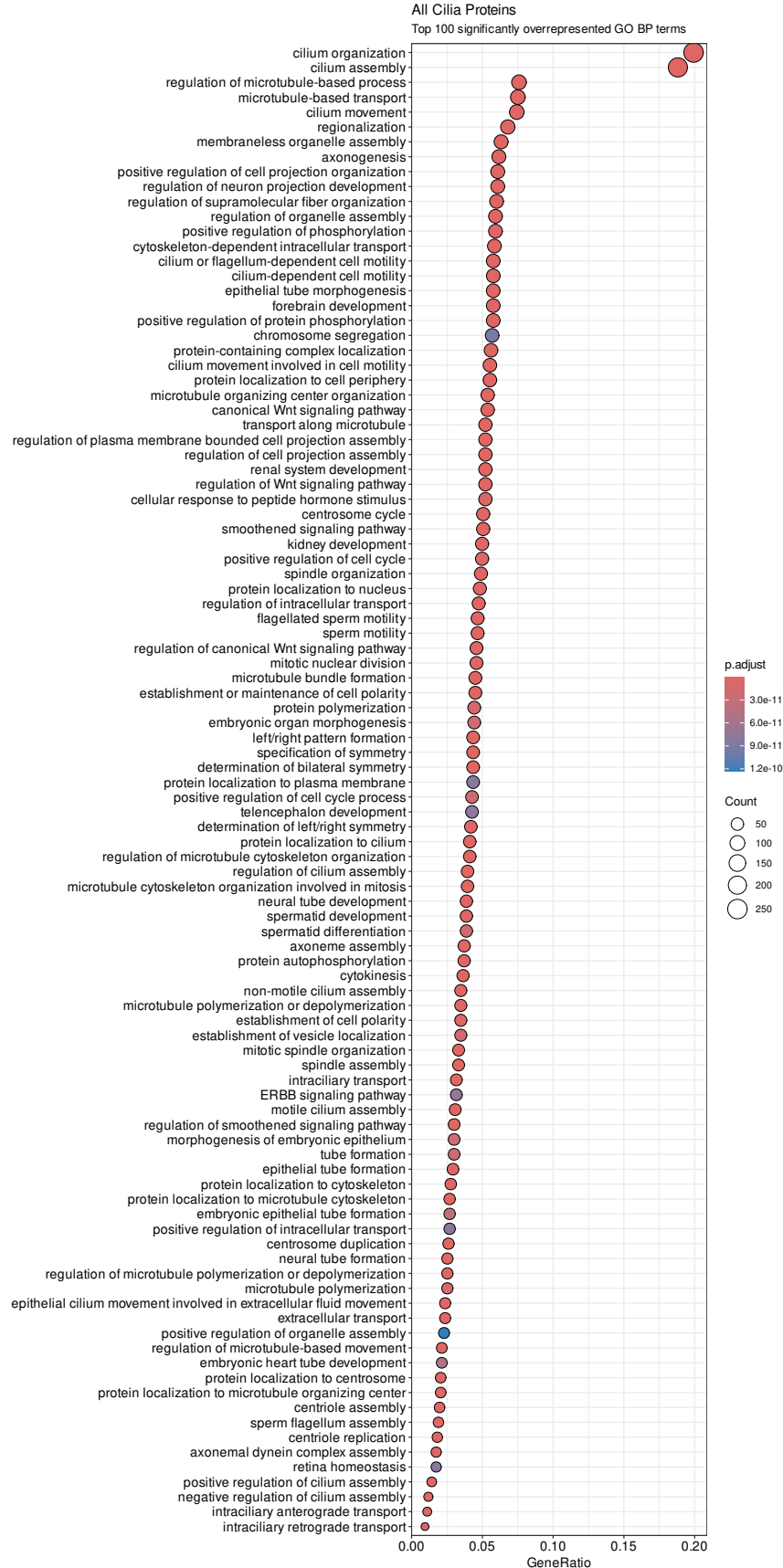
```

Dot plot of top 100 enriched proteins

```

dotplot(ego, showCategory=100, label_format = 50) + ggtitle("All Cilia Proteins", subtitle = "Top 100 s
scale_y_discrete(labels = function(x) str_wrap(x, width = 70))

```




```
# save dotplot as svg file
ggsave(paste0(out_path, "NonRestricted_all_proteins_enrich_GO_BP_dotplot.svg"), plot = last_plot(), dev = "svg")
```

Compare biological themes for different locations (cilia only, PC+tip vs TZ+BB)

```
# prepare input list
input_genes <- list(
  PC_tip = gene_id_pc_tip,
  BB_TZ = gene_id_bb_tz
)
```

GO BP enrichment analysis

```
# Perform the compareCluster analysis
comp <- compareCluster(geneCluster = input_genes,
  fun = "enrichGO",
  OrgDb = org.Hs.eg.db,
  keyType = 'ENSEMBL',
  ont = "BP",
  pAdjustMethod = "BH",
  pvalueCutoff = 0.01,
  qvalueCutoff = 0.01)
```

Dot plot of all enriched terms

```
dotplot(comp, showCategory = NULL) + ggtitle("GO BP Enrichment Comparison", subtitle = "qvalue < 0.01") +
  scale_y_discrete(labels = function(x) str_wrap(x, width = 100))
```



```
# save dotplot as svg file
ggsave(paste0(out_path, "NonRestricted_comparison_locations_cilia_only_combined_GO_BP_dotplot.svg"), pl
```

Visualize overlap of enriched terms as Venn diagram

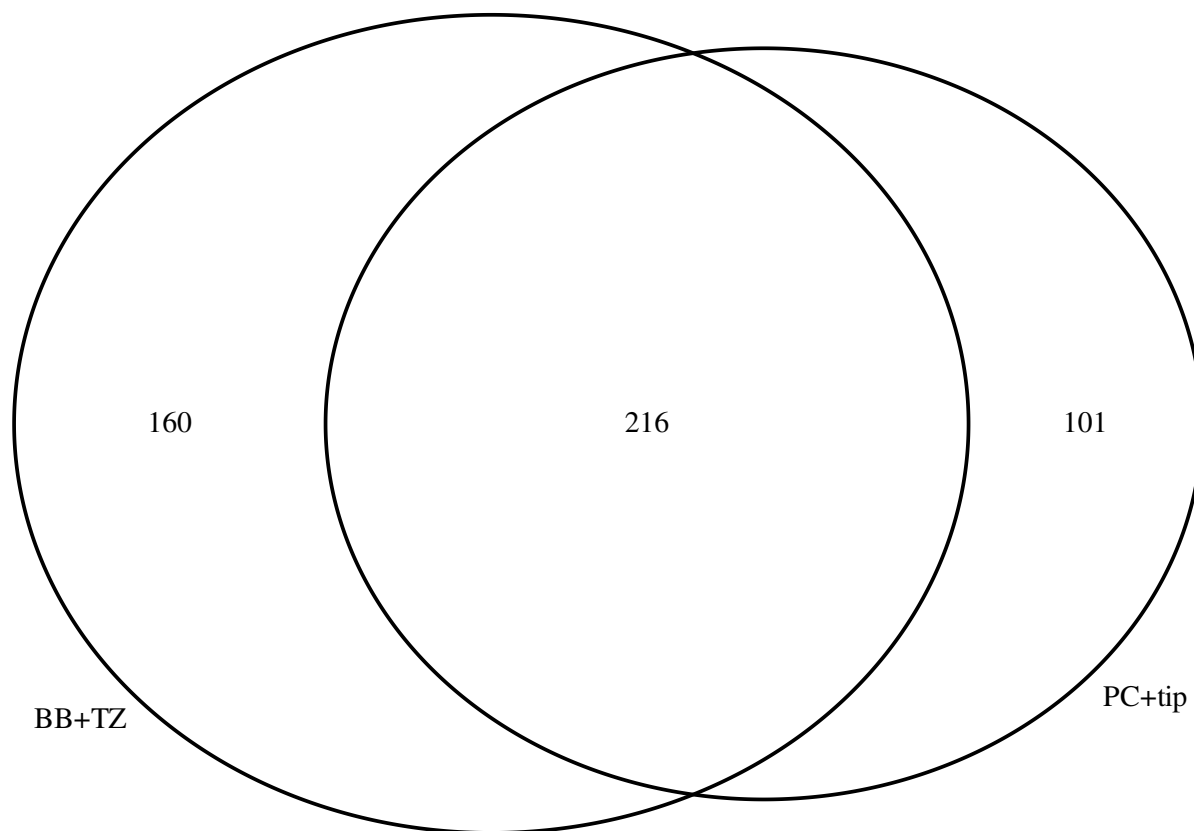
```
# extract results
results <- comp@compareClusterResult

# split by location
pc_tip <- results[results$Cluster == "PC_tip",]
bb_tz <- results[results$Cluster == "BB_TZ",]

# Create a list of the four sets
go_lists <- list(
  PC_tip = pc_tip$ID,
  BB_TZ = bb_tz$ID
)

# Plot the Venn diagram
venn.plot <- venn.diagram(
  x = go_lists,
  category.names = c("PC+tip", "BB+TZ"),
  filename = NULL,
  output = TRUE
)

grid.newpage()
grid.draw(venn.plot)
```



```
# Save the captured plot as an SVG file
svglite(paste0(out_path, "NonRestricted_comparison_locations_cilia_only_combined_GO_BP_venn.svg"), width=1000, height=1000)
grid.draw(venn.plot)
dev.off()
```

```
## cairo_pdf
##      2
```

Determine uniquely enriched and unspecifically enriched terms

```
# calculate intersection of the two
unspecific_terms <- intersect(pc_tip$ID, bb_tz$ID)

# remove all unspecific terms
specific_terms <- results %>% filter(!ID %in% unspecific_terms)

# create a copy of comp
```

```
comp_filtered <- comp  
  
# update results in comp  
comp_filtered@compareClusterResult <- specific_terms
```

Dot plot of uniquely enriched terms

```
# plot(xx, type="dot", caption="GO Enrichment Comparison")  
dotplot(comp_filtered, showCategory = NULL) + ggtitle("GO BP Enrichment Comparison", subtitle = "qvalue  
scale_y_discrete(labels = function(x) str_wrap(x, width = 100))
```



```
# save dotplot as svg file
ggsave(paste0(out_path, "NonRestricted_comparison_locations_cilia_only_combined_GO_BP_dotplot_specific_
```

Prepare data for clustering of enriched terms

```
# Split by location
pc_tip <- comp_filtered@compareClusterResult[comp_filtered@compareClusterResult$Cluster == "PC_tip", ]
bb_tz <- comp_filtered@compareClusterResult[comp_filtered@compareClusterResult$Cluster == "BB_TZ", ]

# Subset for pc_tip
pc_tip <- comp_filtered@compareClusterResult[
  comp_filtered@compareClusterResult$Cluster == "PC_tip",
]

# Subset for bb_tz
bb_tz <- comp_filtered@compareClusterResult[
  comp_filtered@compareClusterResult$Cluster == "BB_TZ",
]

# Create new compareClusterResult objects for each subset
comp_filtered_pc_tip <- comp_filtered
comp_filtered_bb_tz <- comp_filtered

comp_filtered_pc_tip@compareClusterResult <- pc_tip
comp_filtered_bb_tz@compareClusterResult <- bb_tz
```

Cluster results - PC&tip

```
go_id = pc_tip$ID
mat = GO_similarity(go_id,
  ont = 'BP',
  db = 'org.Hs.eg.db',
  measure = "Sim_Relevance_2006"
)
```

Plot cluster heatmap

```
# Capture the plot
heatmap_plot <- grid.grabExpr({
  df <- simplifyGO(mat,
    method = 'binary_cut',
    plot = TRUE,
    column_title = "GO BP terms only significant in primary cilia incl. tip",
    use_raster = FALSE,
    order_by_size = TRUE,
    fontsize_range = c(18, 36),
    max_words = 6,
```

```

        word_cloud_grob_param = list(col = 'black',
                                      max_width = unit(200, "mm"))
    })

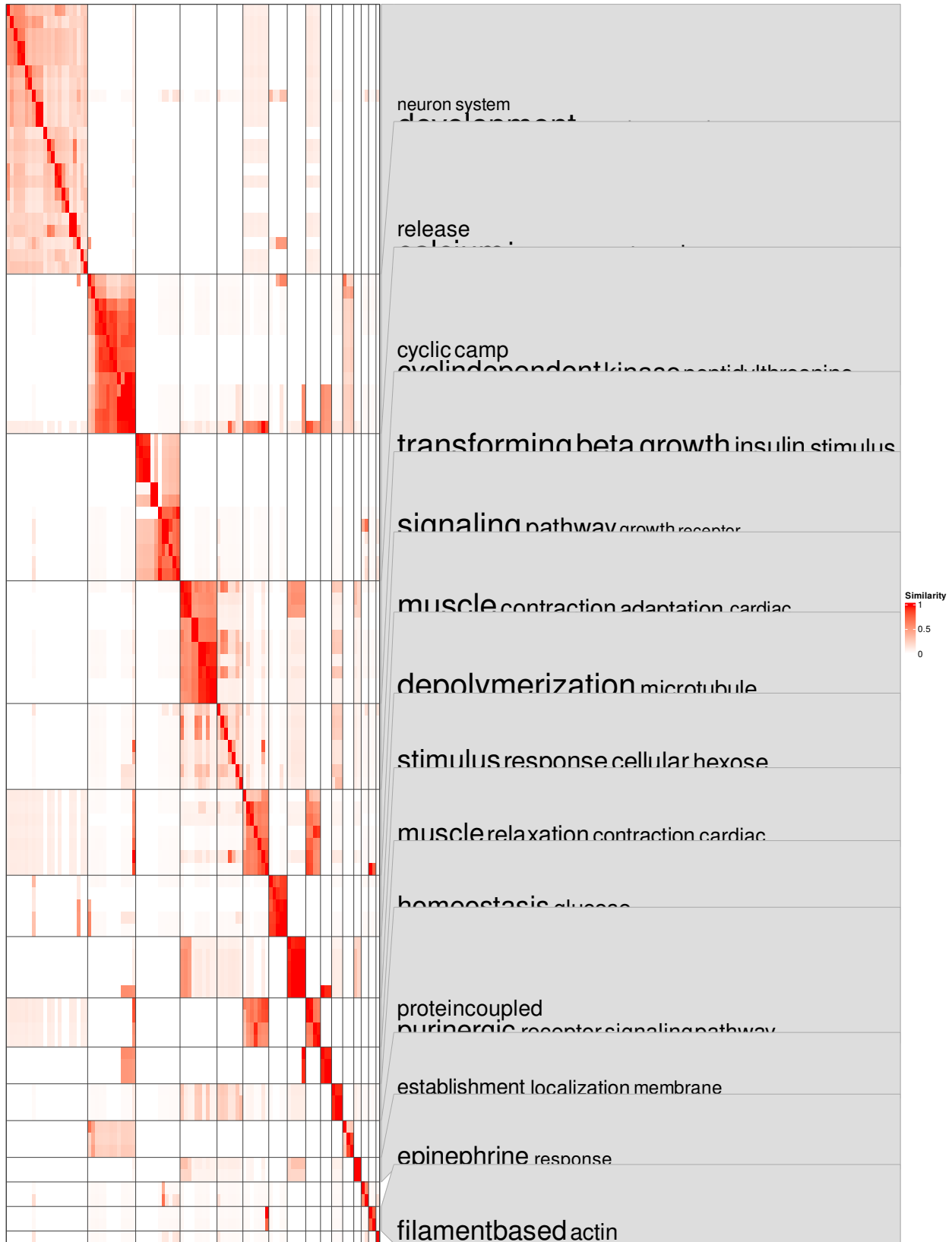
    # Save the captured plot as an SVG file
    svglite(paste0(out_path, "NonRestricted_comparison_locations_cilia_only_combined_GO_BP_dotplot_specific"),
            grid.draw(heatmap_plot)
    dev.off()

## cairo_pdf
##      2

grid.newpage()
grid.draw(heatmap_plot)

```


GO BP terms only significant in primary cilia incl. tip



Process and save results

```
# add cluster number from GO term clustering
results <- map_cluster_number(comp_filtered_pc_tip,
                             df = df,
                             comp = TRUE
)

# Apply the function to each row of the DataFrame
results$GeneSymbol <- sapply(results$geneID, map_geneID_to_symbol)

# save results as csv file
write.csv(results, file = paste0(out_path, "NonRestricted_comparison_locations_cilia_only_combined_GO_BP_dotplot_specific.csv"))
```

Cluster results - BB&TZ

```
go_id = bb_tz$ID
mat = GO_similarity(go_id,
                   ont = 'BP',
                   db = 'org.Hs.eg.db',
                   measure = "Sim_Relevance_2006"
)
```

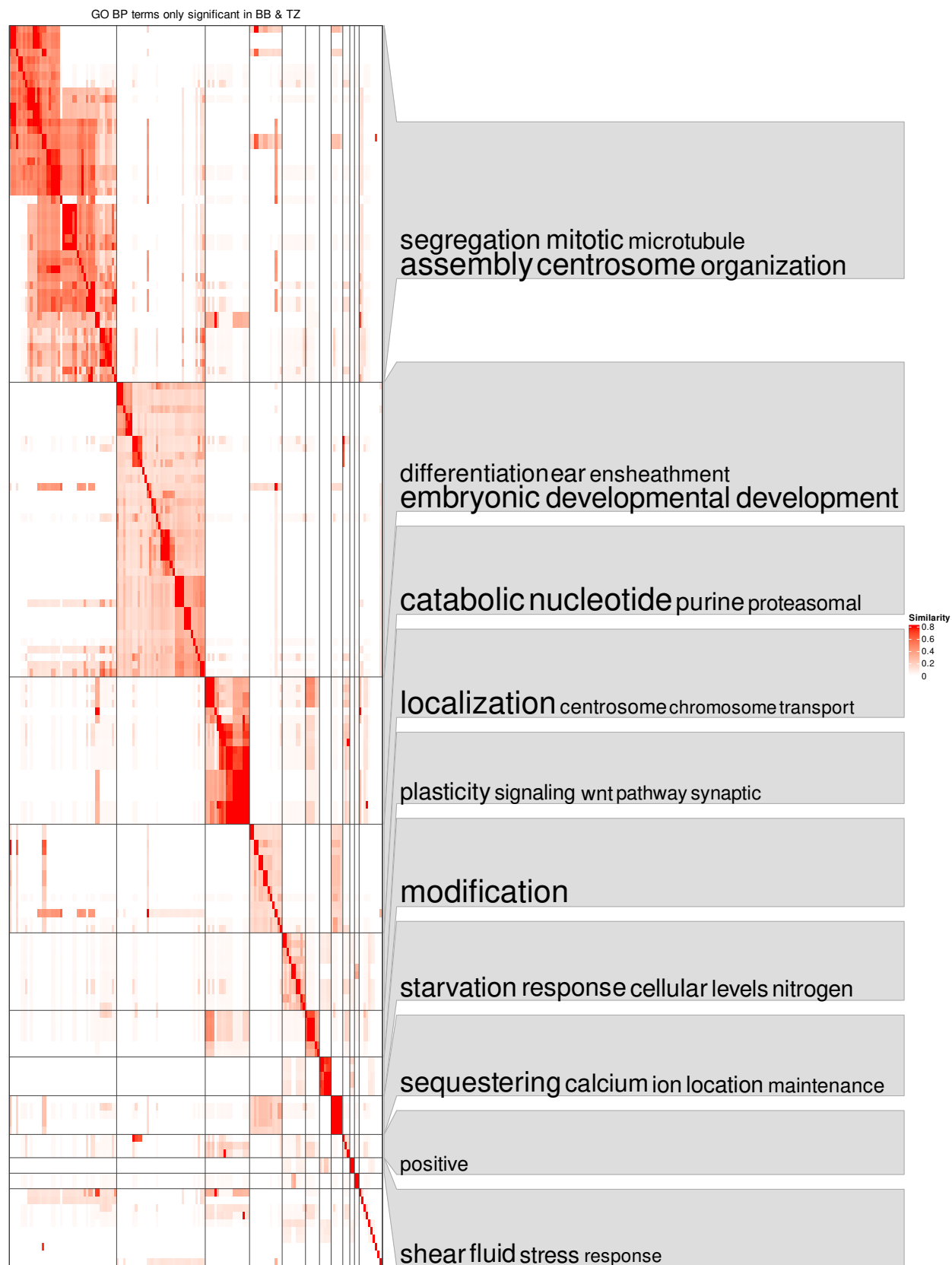
Plot cluster heatmap

```
# Capture the plot
heatmap_plot <- grid.grabExpr({
  df <- simplifyGO(mat,
                  method = 'binary_cut',
                  plot = TRUE,
                  column_title = "GO BP terms only significant in BB & TZ",
                  use_raster = FALSE,
                  order_by_size = TRUE,
                  fontsize_range = c(18, 36),
                  max_words = 6,
                  word_cloud_grob_param = list(col = 'black',
                                                max_width = unit(200, "mm")))
})

# Save the captured plot as an SVG file
svglite(paste0(out_path, "NonRestricted_comparison_locations_cilia_only_combined_GO_BP_dotplot_specific.svg"),
        grid.draw(heatmap_plot)
dev.off()

## cairo_pdf
##      2
```

```
grid.newpage()  
grid.draw(heatmap_plot)
```



Process and save results

```
# add cluster number from GO term clustering
results <- map_cluster_number(comp_filtered_bb_tz,
                             df = df,
                             comp = TRUE
)

# Apply the function to each row of the DataFrame
results$GeneSymbol <- sapply(results$geneID, map_geneID_to_symbol)

# save results as csv file
write.csv(results, file = paste0(out_path, "NonRestricted_comparison_locations_cilia_only_combined_GO_BP_"))
```

Filter for terms enriched in both locations

```
# get results as data frame
comp_results <- comp@compareClusterResult

# get all terms significant in each location
pc_tip <- comp_results %>% filter(Cluster == "PC_tip")
bb_tz <- comp_results %>% filter(Cluster == "BB_TZ")

# calculate intersection of the two
unspecific_terms <- intersect(pc_tip$ID, bb_tz$ID)

# remove all unspecific terms
unspecific_terms <- comp_results %>% filter(ID %in% unspecific_terms)

# create a copy of comp
comp_filtered <- comp

# update results in comp
comp_filtered@compareClusterResult <- unspecific_terms
```

Dot plot of shared terms

```
# plot dotplot
dotplot(comp_filtered, showCategory = NULL) + ggtitle("GO BP Enrichment Comparison", subtitle = "qvalue") +
  scale_y_discrete(labels = function(x) str_wrap(x, width = 100))
```



```
# save dotplot as svg file
ggsave(paste0(out_path, "NonRestricted_comparison_locations_cilia_only_combined_GO_BP_dotplot_shared_terms.svg"))
```

Prepare data for clustering of shared enriched terms

```
# Split by location
pc_tip <- comp_filtered@compareClusterResult[comp_filtered@compareClusterResult$Cluster == "PC_tip", ]
bb_tz <- comp_filtered@compareClusterResult[comp_filtered@compareClusterResult$Cluster == "BB_TZ", ]

# Create new compareClusterResult objects for each subset
comp_filtered_pc_tip <- comp_filtered
comp_filtered_bb_tz <- comp_filtered

comp_filtered_pc_tip@compareClusterResult <- pc_tip
comp_filtered_bb_tz@compareClusterResult <- bb_tz
```

Cluster results - Both locations

```
go_id = pc_tip$ID
mat = GO_similarity(go_id,
  ont = 'BP',
  db = 'org.Hs.eg.db',
  measure = "Sim_Relevance_2006"
)
```

Plot cluster heatmap

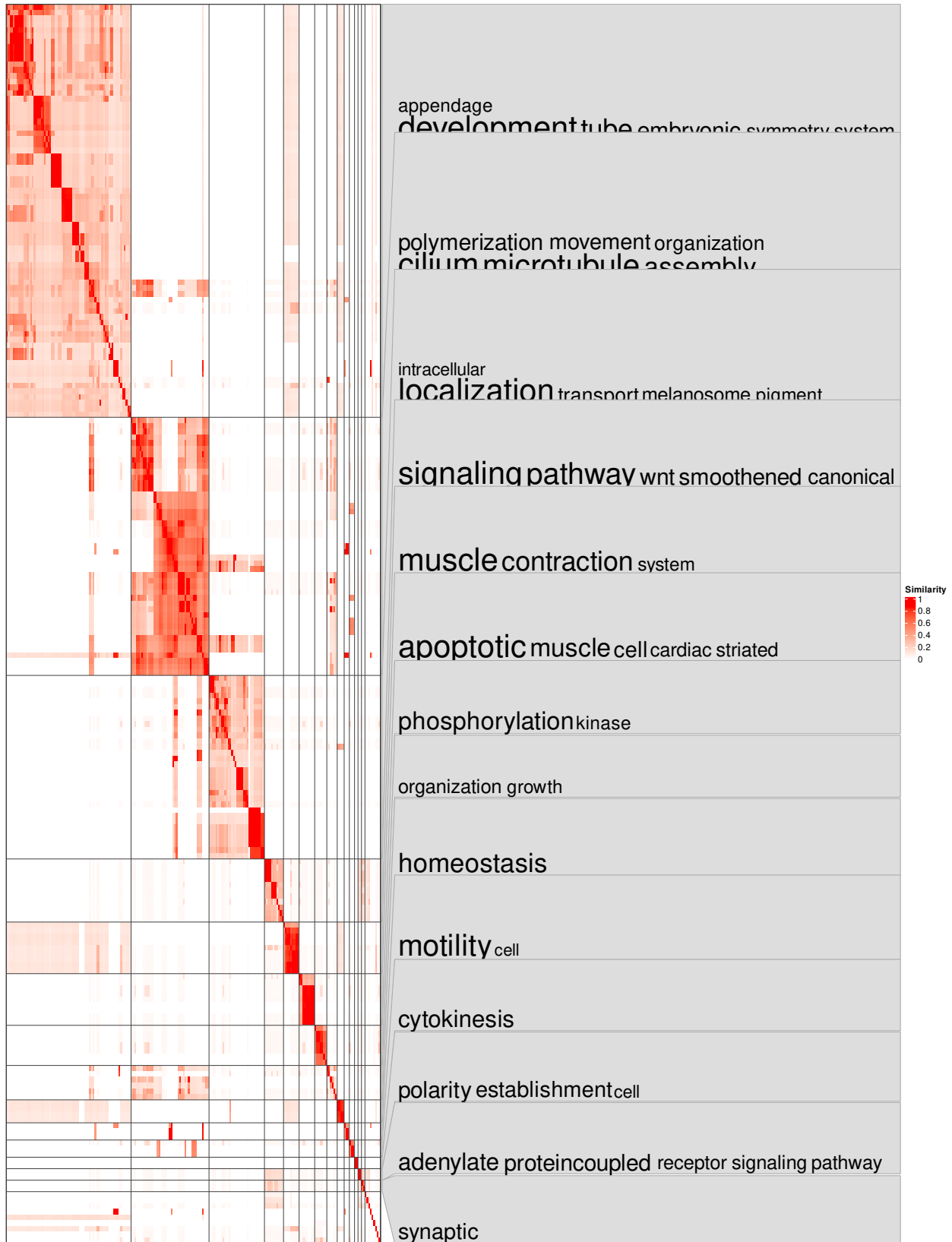
```
# Capture the plot
heatmap_plot <- grid.grabExpr({
  df <- simplifyGO(mat,
    method = 'binary_cut',
    plot = TRUE,
    column_title = "GO BP terms significant in both locations",
    use_raster = FALSE,
    order_by_size = TRUE,
    fontsize_range = c(18, 36),
    max_words = 6,
    word_cloud_grob_param = list(col = 'black',
                                  max_width = unit(200, "mm")))
})

# Save the captured plot as an SVG file
svglite(paste0(out_path, "NonRestricted_comparison_locations_cilia_only_combined_GO_BP_dotplot_specific.svg"))
grid.draw(heatmap_plot)
dev.off()

## cairo_pdf
## 2
```

```
grid.newpage()  
grid.draw(heatmap_plot)
```


GO BP terms significant in both locations



Process and save results

```
# add cluster number from GO term clustering
results <- map_cluster_number(comp_filtered_pc_tip,
                             df = df,
                             comp = TRUE
)

# Apply the function to each row of the DataFrame
results$GeneSymbol <- sapply(results$geneID, map_geneID_to_symbol)

# save results as csv file
write.csv(results, file = paste0(out_path, "NonRestricted_comparison_locations_cilia_only_combined_GO_BI"))

# add cluster number from GO term clustering
results <- map_cluster_number(comp_filtered_bb_tz,
                             df = df,
                             comp = TRUE
)

# Apply the function to each row of the DataFrame
results$GeneSymbol <- sapply(results$geneID, map_geneID_to_symbol)

# save results as csv file
write.csv(results, file = paste0(out_path, "NonRestricted_comparison_locations_cilia_only_combined_GO_BI"))
```

Compare biological themes for different locations (cilia vs other parts of cell)

```
# Split data by location
input_genes <- list(
  PrimaryCiliaTip = gene_id_tip,
  PrimaryCilia = gene_id_pc,
  PrimaryCiliaTZ = gene_id_tz,
  BasalBody = gene_id_bb,
  Cytoplasm = gene_id_cytoplasm,
  Membrane = gene_id_membrane,
  Nucleus = gene_id_nucleus,
  Mitotic = gene_id_mitotic
)
```

GO BP enrichment analysis

```
# Perform the compareCluster analysis
comp <- compareCluster(geneCluster = input_genes,
                      fun = "enrichGO",
                      OrgDb = org.Hs.eg.db,
```

```
keyType      = 'ENSEMBL',  
ont          = "BP",  
pAdjustMethod = "BH",  
pvalueCutoff = 0.01,  
qvalueCutoff = 0.01)
```

```
# get results  
results <- comp@compareClusterResult  
  
# Apply the function to each row of the DataFrame  
results$GeneSymbol <- sapply(results$geneID, map_geneID_to_symbol)  
  
# save results as csv file  
write.csv(results, file = paste0(out_path, "NonRestricted_comparison_all_locations_GO_BP_result.csv"))
```

Dot plot of all enriched terms

```
dotplot(comp, showCategory = NULL) + ggtitle("GO BP Enrichment Comparison", subtitle = "qvalue < 0.01")  
  scale_y_discrete(labels = function(x) str_wrap(x, width = 60))
```

Entity	2017	2018	2019	2020	2021	2022	2023	2024	2025	2026	2027	2028	2029	2030	2031	2032	2033	2034	2035	2036	2037	2038	2039	2040	2041	2042	2043	2044	2045	2046	2047	2048	2049	2050	2051	2052	2053	2054	2055	2056	2057	2058	2059	2060	2061	2062	2063	2064	2065	2066	2067	2068	2069	2070	2071	2072	2073	2074	2075	2076	2077	2078	2079	2080	2081	2082	2083	2084	2085	2086	2087	2088	2089	2090	2091	2092	2093	2094	2095	2096	2097	2098	2099	2100	2101	2102	2103	2104	2105	2106	2107	2108	2109	2110	2111	2112	2113	2114	2115	2116	2117	2118	2119	2120	2121	2122	2123	2124	2125	2126	2127	2128	2129	2130	2131	2132	2133	2134	2135	2136	2137	2138	2139	2140	2141	2142	2143	2144	2145	2146	2147	2148	2149	2150	2151	2152	2153	2154	2155	2156	2157	2158	2159	2160	2161	2162	2163	2164	2165	2166	2167	2168	2169	2170	2171	2172	2173	2174	2175	2176	2177	2178	2179	2180	2181	2182	2183	2184	2185	2186	2187	2188	2189	2190	2191	2192	2193	2194	2195	2196	2197	2198	2199	2200	2201	2202	2203	2204	2205	2206	2207	2208	2209	2210	2211	2212	2213	2214	2215	2216	2217	2218	2219	2220	2221	2222	2223	2224	2225	2226	2227	2228	2229	2230	2231	2232	2233	2234	2235	2236	2237	2238	2239	2240	2241	2242	2243	2244	2245	2246	2247	2248	2249	2250	2251	2252	2253	2254	2255	2256	2257	2258	2259	2260	2261	2262	2263	2264	2265	2266	2267	2268	2269	2270	2271	2272	2273	2274	2275	2276	2277	2278	2279	2280	2281	2282	2283	2284	2285	2286	2287	2288	2289	2290	2291	2292	2293	2294	2295	2296	2297	2298	2299	2300	2301	2302	2303	2304	2305	2306	2307	2308	2309	2310	2311	2312	2313	2314	2315	2316	2317	2318	2319	2320	2321	2322	2323	2324	2325	2326	2327	2328	2329	2330	2331	2332	2333	2334	2335	2336	2337	2338	2339	2340	2341	2342	2343	2344	2345	2346	2347	2348	2349	2350	2351	2352	2353	2354	2355	2356	2357	2358	2359	2360	2361	2362	2363	2364	2365	2366	2367	2368	2369	2370	2371	2372	2373	2374	2375	2376	2377	2378	2379	2380	2381	2382	2383	2384	2385	2386	2387	2388	2389	2390	2391	2392	2393	2394	2395	2396	2397	2398	2399	2400	2401	2402	2403	2404	2405	2406	2407	2408	2409	2410	2411	2412	2413	2414	2415	2416	2417	2418	2419	2420	2421	2422	2423	2424	2425	2426	2427	2428	2429	2430	2431	2432	2433	2434	2435	2436	2437	2438	2439	2440	2441	2442	2443	2444	2445	2446	2447	2448	2449	2450	2451	2452	2453	2454	2455	2456	2457	2458	2459	2460	2461	2462	2463	2464	2465	2466	2467	2468	2469	2470	2471	2472	2473	2474	2475	2476	2477	2478	2479	2480	2481	2482	2483	2484	2485	2486	2487	2488	2489	2490	2491	2492	2493	2494	2495	2496	2497	2498	2499	2500	2501	2502	2503	2504	2505	2506	2507	2508	2509	2510	2511	2512	2513	2514	2515	2516	2517	2518	2519	2520	2521	2522	2523	2524	2525	2526	2527	2528	2529	2530	2531	2532	2533	2534	2535	2536	2537	2538	2539	2540	2541	2542	2543	2544	2545	2546	2547	2548	2549	2550	2551	2552	2553	2554	2555	2556	2557	2558	2559	2560	2561	2562	2563	2564	2565	2566	2567	2568	2569	2570	2571	2572	2573	2574	2575	2576	2577	2578	2579	2580	2581	2582	2583	2584	2585	2586	2587	2588	2589	2590	2591	2592	2593	2594	2595	2596	2597	2598	2599	2600	2601	2602	2603	2604	2605	2606	2607	2608	2609	2610	2611	2612	2613	2614	2615	2616	2617	2618	2619	2620	2621	2622	2623	2624	2625	2626	2627	2628	2629	2630	2631	2632	2633	2634	2635	2636	2637	2638	2639	2640	2641	2642	2643	2644	2645	2646	2647	2648	2649	2650	2651	2652	2653	2654	2655	2656	2657	2658	2659	2660	2661	2662	2663	2664	2665	2666	2667	2668	2669	2670	2671	2672	2673	2674	2675	2676	2677	2678	2679	2680	2681	2682	2683	2684	2685	2686	2687	2688	2689	2690	2691	2692	2693	2694	2695	2696	2697	2698	2699	2700	2701	2702	2703	2704	2705	2706	2707	2708	2709	2710	2711	2712	2713	2714	2715	2716	2717	2718	2719	2720	2721	2722	2723	2724	2725	2726	2727	2728	2729	2730	2731	2732	2733	2734	2735	2736	2737	2738	2739	2740	2741	2742	2743	2744	2745	2746	2747	2748	2749	2750	2751	2752	2753	2754	2755	2756	2757	2758	2759	2760	2761	2762	2763	2764	2765	2766	2767	2768	2769	2770	2771	2772	2773	2774	2775	2776	2777	2778	2779	2780	2781	2782	2783	2784	2785	2786	2787	2788	2789	2790	2791	2792	2793	2794	2795	2796	2797	2798	2799	2800	2801	2802	2803	2804	2805	2806	2807	2808	2809	2810	2811	2812	2813	2814	2815	2816	2817	2818	2819	2820	2821	2822	2823	2824	2825	2826	2827	2828	2829	2830	2831	2832	2833	2834	2835	2836	2837	2838	2839	2840	2841	2842	2843	2844	2845	2846	2847	2848	2849	2850	2851	2852	2853	2854	2855	2856	2857	2858	2859	2860	2861	2862	2863	2864	2865	2866	2867	2868	2869	2870	2871	2872	2873	2874	2875	2876	2877	2878	2879	2880	2881	2882	2883	2884	2885	2886	2887	2888	2889	2890	2891	2892	2893	2894	2895	2896	2897	2898	2899	2900	2901	2902	2903	2904	2905	2906	2907	2908	2909	2910	2911	2912	2913	2914	2915	2916	2917	2918	2919	2920	2921	2922	2923	2924	2925	2926	2927	2928	2929	2930	2931	2932	2933	2934	2935	2936	2937	2938	2939	2940	2941	2942	2943	2944	2945	2946	2947	2948	2949	2950	2951	2952	2953	2954	2955	2956	2957	2958	2959	2960	2961	2962	2963	2964	2965	2966	2967	2968	2969	2970	2971	2972	2973	2974	2975	2976	2977	2978	2979	2980	2981	2982	2983	2984	2985	2986	2987	2988	2989	2990	2991	2992	2993	2994	2995	2996	2997	2998	2999	3000	3001	3002	3003	3004	3005	3006	3007	3008	3009	3010	3011	3012	3013	3014	301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```

# save dotplot as svg file
ggsave(paste0(out_path, "NonRestricted_comparison_all_locations_GO_BP_dotplot.svg"), plot = last_plot())

# extract results
results <- comp@compareClusterResult$ID

# get unique terms
results <- unique(results)

# filter for p.adjust < 0.01 and qvalue < 0.01
results_signif <- comp@compareClusterResult %>% filter(p.adjust < 0.01 & qvalue < 0.01)
results_signif <- unique(results_signif$ID)

# print number of unique terms
print(paste("Number of unique terms in comp:", length(results)))

## [1] "Number of unique terms in comp: 941"

print(paste("Number of unique terms in comp:", length(results_signif)))

## [1] "Number of unique terms in comp: 941"

```

Filter for cilia terms

Here we filter for terms that are enriched in any of the four cilia locations, so basically cutting of the long dotplot above to only show the terms of the cilia locations.

```

# get results as data frame
comp_results <- comp@compareClusterResult

# get all terms that are significant in any of the 4 non cilia locations
non_cilia_terms <- comp_results %>% filter(Cluster == "Cytoplasm" | Cluster == "Membrane" | Cluster == "Nucleus" | Cluster == "PlasmaMembrane")

# get all the terms that are significant in any of the cilia locations
cilium_terms <- comp_results %>% filter(Cluster == "PrimaryCilia" | Cluster == "PrimaryCiliaTip" | Cluster == "SecondaryCilia")

# remove all cilia terms from non_cilia_terms
non_cilia_only_terms <- non_cilia_terms %>% filter(!ID %in% cilium_terms$ID)

# remove all non cilia only terms from cilium_terms
cilium_only_terms <- cilium_terms %>% filter(!ID %in% non_cilia_only_terms$ID)

# filter comp results for terms in cilium_only_terms
comp_results_filtered <- comp_results %>% filter(ID %in% cilium_only_terms$ID)

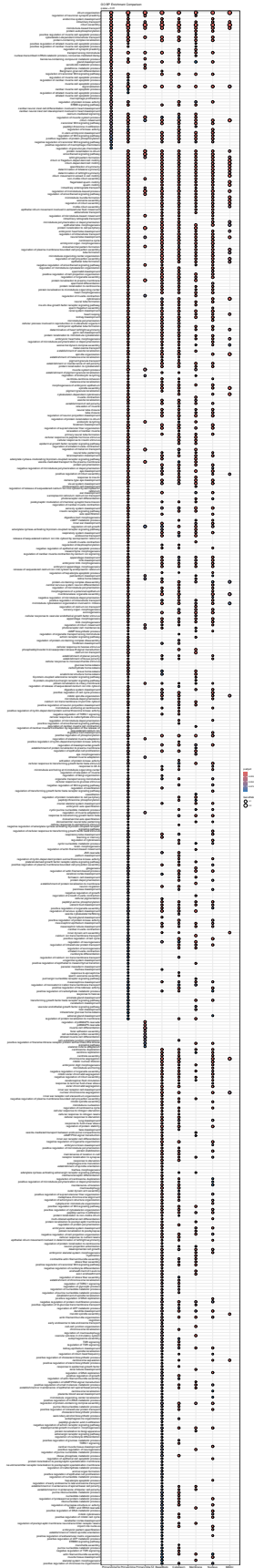
# create a copy of comp
comp_filtered <- comp

# update results in comp
comp_filtered@compareClusterResult <- comp_results_filtered

```

Dot plot of terms enriched in ciliary locations

```
# plot dotplot  
dotplot(comp_filtered, showCategory = NULL) + ggtitle("GO BP Enrichment Comparison", subtitle = "qvalue  
scale_y_discrete(labels = function(x) str_wrap(x, width = 80))
```



```
# save dotplot as svg file
ggsave(paste0(out_path, "NonRestricted_comparison_all_comparison_locations_GO_BP_dotplot_filtered_cilia
```

Filter for cilia only terms

Here we filter for terms that are only enriched in any of the four cilia locations and not in any of the other four non cilia locations. Note: This removes the non cilia locations from the plot as there is no enrichment of any of the terms in any non cilia location.

```
# get results as data frame
comp_results <- comp@compareClusterResult

# get all terms that are significant in any of the 4 non cilia locations
non_cilia_terms <- comp_results %>% filter(Cluster == "Cytoplasm" | Cluster == "Membrane" | Cluster ==

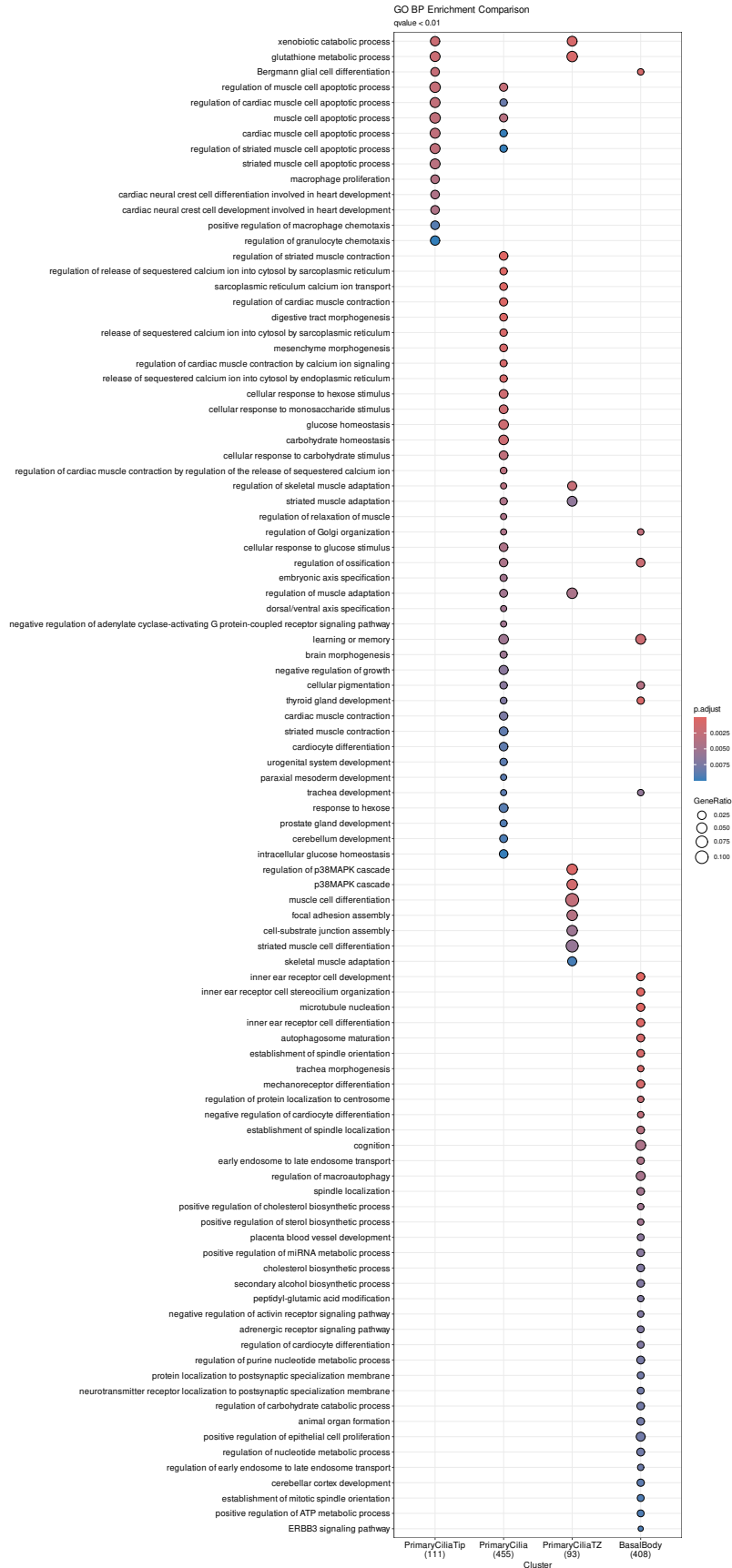
# remove all non cilia terms from results
cilia_only_terms <- comp_results %>% filter(!ID %in% non_cilia_terms$ID)

# create a copy of comp
comp_filtered <- comp

# update results in comp
comp_filtered@compareClusterResult <- cilia_only_terms
```

Dot plot of terms only enriched in ciliary locations

```
# plot dotplot
dotplot(comp_filtered, showCategory = NULL) + ggtitle("GO BP Enrichment Comparison", subtitle = "qvalue
  scale_y_discrete(labels = function(x) str_wrap(x, width = 100))
```

```

# save dotplot as svg file
ggsave(paste0(out_path, "NonRestricted_comparison_all_comparison_locations_GO_BP_dotplot_filtered_cilia

# extract results
results <- comp@compareClusterResult$ID

# get unique terms
results <- unique(results)

# print number of unique terms
print(paste("Number of unique terms in comp:", length(results)))

```

```
## [1] "Number of unique terms in comp: 941"
```

Simplify results

```

comp_simplified <- simplify(comp, cutoff=0.6, by="p.adjust", select_fun=min)

# print number of unique terms
print(paste("Number of significant terms:", nrow(comp)))

```

```
## [1] "Number of significant terms: 2172"
```

```
print(paste("Number of significant terms after simplifying:", nrow(comp_simplified)))
```

```
## [1] "Number of significant terms after simplifying: 462"
```

```

# extract results
results <- comp_simplified@compareClusterResult$ID

# get unique terms
results <- unique(results)

# print number of unique terms
print(paste("Number of unique terms in comp_simplified:", length(results)))

```

```
## [1] "Number of unique terms in comp_simplified: 256"
```

```

# get results
results <- comp_simplified@compareClusterResult

# Apply the function to each row of the DataFrame
results$GeneSymbol <- sapply(results$geneID, map_geneID_to_symbol)

# save results as csv file
write.csv(results, file = paste0(out_path, "NonRestricted_comparison_all_locations_GO_BP_result_simplif

```

Dot plot of all enriched terms (simplified)

```
# plot dotplot  
dotplot(comp_simplified, showCategory = NULL) + ggtitle("GO BP Enrichment Comparison", subtitle = "qval")  
  scale_y_discrete(labels = function(x) str_wrap(x, width = 80))
```


Filter for cilia terms

Here we filter for terms that are enriched in any of the four cilia locations, so basically cutting of the long dotplot above to only show the terms of the cilia locations. But this time using the simplified results.

```
# get results as data frame
comp_results <- comp_simplified@compareClusterResult

# get all terms that are significant in any of the 4 non cilia locations
non_cilia_terms <- comp_results %>% filter(Cluster == "Cytoplasm" | Cluster == "Membrane" | Cluster == "SecondaryCilia" | Cluster == "SecondaryCiliaTip")

# get all the terms that are significant in any of the cilia locations
cilia_terms <- comp_results %>% filter(Cluster == "PrimaryCilia" | Cluster == "PrimaryCiliaTip" | Cluster == "SecondaryCilia" | Cluster == "SecondaryCiliaTip")

# remove all cilia terms from non_cilia_terms
non_cilia_only_terms <- non_cilia_terms %>% filter(!ID %in% cilia_terms$ID)

# remove all non cilia only terms from cilia terms
cilia_only_terms <- cilia_terms %>% filter(!ID %in% non_cilia_only_terms$ID)

# filter comp results for terms in cilia_only_terms
comp_results_filtered <- comp_results %>% filter(ID %in% cilia_only_terms$ID)

# create a copy of comp
comp_simplified_filtered <- comp_simplified

# update results in comp
comp_simplified_filtered@compareClusterResult <- comp_results_filtered
```

Dot plot of terms enriched in ciliary locations (simplified)

```
# plot dotplot
dotplot(comp_simplified_filtered, showCategory = NULL) + ggtitle("GO BP Enrichment Comparison", subtitle = "Cilia Locations") +
  scale_y_discrete(labels = function(x) str_wrap(x, width = 80))
```



```
# save dotplot as svg file
ggsave(paste0(out_path, "NonRestricted_comparison_all_comparison_locations_GO_BP_dotplot_simplified_06_06_2020.svg"))
```

Filter for cilia only terms

Here we filter again for terms that are only enriched in any of the four cilia locations and not in any of the other four non cilia locations. But this time using the simplified results. Note: This removes the non cilia locations from the plot as there is no enrichment of any of the terms in any non cilia location.

```
# get results as data frame
comp_results <- comp_simplified@compareClusterResult

# get all terms that are significant in any of the 4 non cilia locations
non_cilia_terms <- comp_results %>% filter(Cluster == "Cytoplasm" | Cluster == "Membrane" | Cluster == "Nucleus" | Cluster == "Mitochondrion")

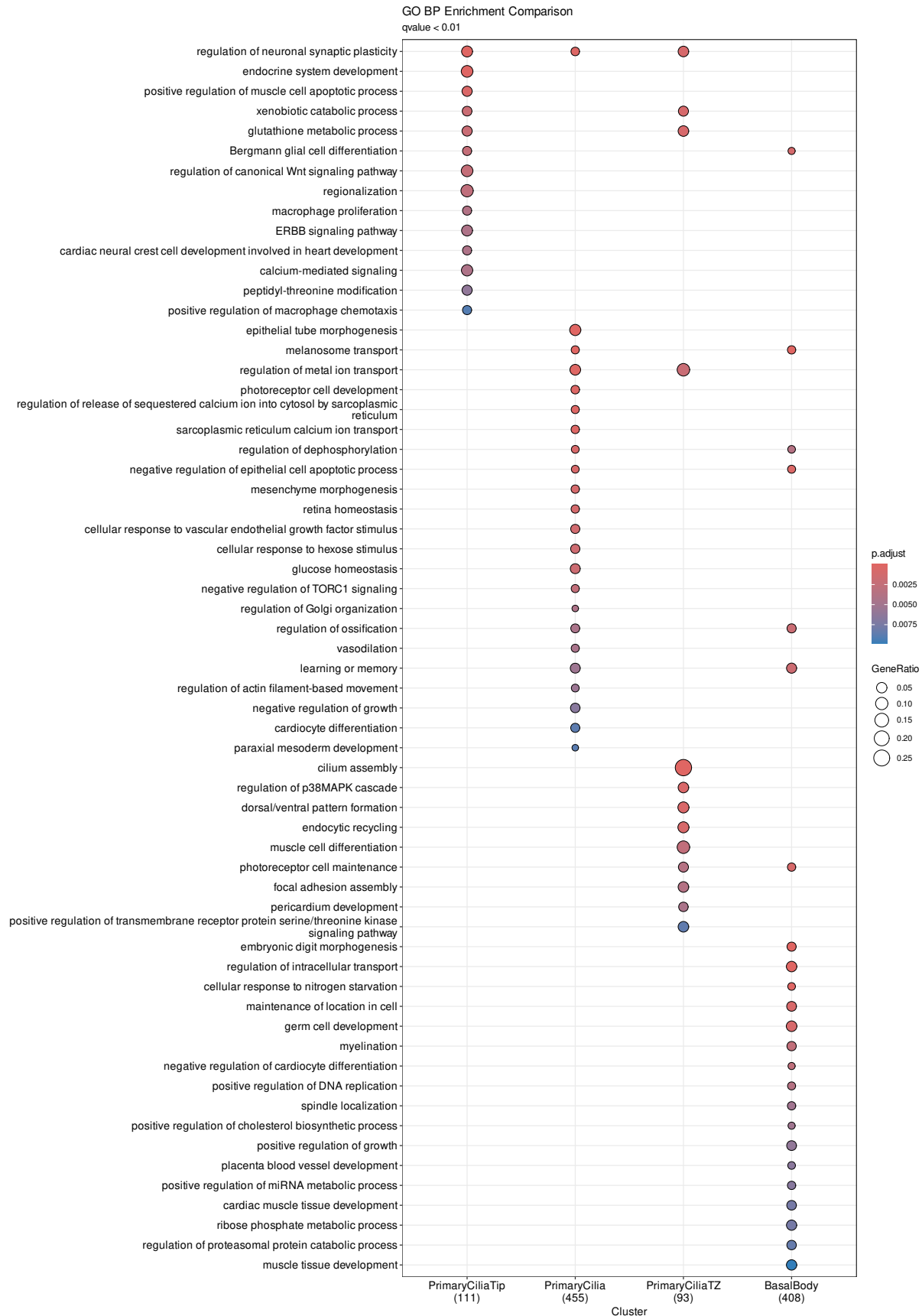
# remove all non cilia terms from results
cilium_only_terms <- comp_results %>% filter(!ID %in% non_cilia_terms$ID)

# create a copy of comp
comp_simplified_filtered <- comp_simplified

# update results in comp
comp_simplified_filtered@compareClusterResult <- cilium_only_terms
```

Dot plot of terms only enriched in ciliary locations(simplified)

```
# plot dotplot
dotplot(comp_simplified_filtered, showCategory = NULL) + ggtitle("GO BP Enrichment Comparison", subtitle = "Cilia only terms") +
  scale_y_discrete(labels = function(x) str_wrap(x, width = 80))
```




```
# save dotplot as svg file
ggsave(paste0(out_path, "NonRestricted_comparison_all_comparison_locations_GO_BP_dotplot_simplified_06_06_2019.svg"))
```

```
# calculate overlap between comp and comp_simplified results by plotting a venn diagramm
# extract results
results <- comp@compareClusterResult$ID
results_simplified <- comp_simplified@compareClusterResult$ID

# print number of unique terms
print(paste("Number of terms in comp:", length(results)))
```

Check overlap between significant terms and simplified terms

```
## [1] "Number of terms in comp: 2172"
```

```
print(paste("Number of terms in comp_simplified:", length(results_simplified)))
```

```
## [1] "Number of terms in comp_simplified: 462"
```

```
# get unique terms
results <- unique(results)
results_simplified <- unique(results_simplified)

# print number of unique terms
print(paste("Number of unique terms in comp:", length(results)))
```

```
## [1] "Number of unique terms in comp: 941"
```

```
print(paste("Number of unique terms in comp_simplified:", length(results_simplified)))
```

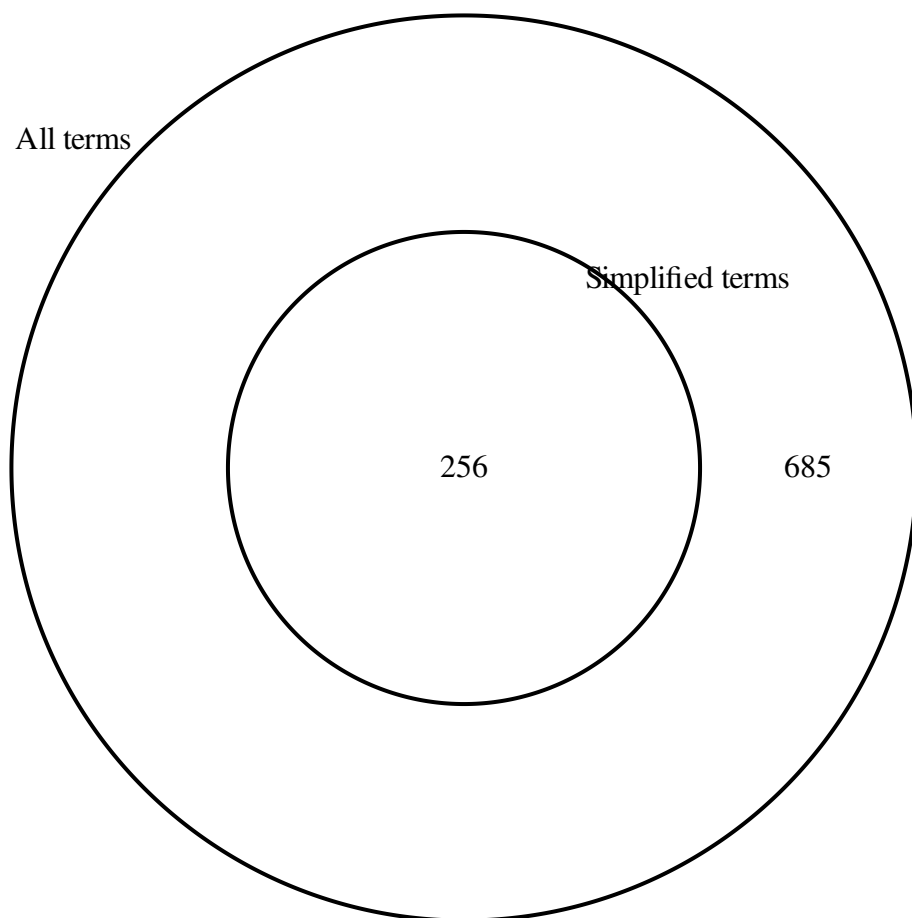
```
## [1] "Number of unique terms in comp_simplified: 256"
```

```
intersection <- intersect(results, results_simplified)
print(paste("Number of terms in intersection:", length(intersection)))
```

```
## [1] "Number of terms in intersection: 256"
```

```
# plot venn diagram
venn.plot <- venn.diagram(
  x = list(results = results, results_simplified = results_simplified),
  category.names = c("All terms", "Simplified terms"),
  filename = NULL,
  output = FALSE
)

grid.newpage()
grid.draw(venn.plot)
```



Session info

```
sessionInfo()
```

```
## R version 4.5.0 (2025-04-11)
## Platform: x86_64-pc-linux-gnu
## Running under: Ubuntu 24.04.2 LTS
##
## Matrix products: default
## BLAS:   /usr/lib/x86_64-linux-gnu/blas/libblas.so.3.12.0
## LAPACK: /usr/lib/x86_64-linux-gnu/lapack/liblapack.so.3.12.0  LAPACK version 3.12.0
##
## locale:
##  [1] LC_CTYPE=en_US.UTF-8      LC_NUMERIC=C
##  [3] LC_TIME=en_US.UTF-8      LC_COLLATE=en_US.UTF-8
##  [5] LC_MONETARY=en_US.UTF-8  LC_MESSAGES=en_US.UTF-8
##  [7] LC_PAPER=en_US.UTF-8     LC_NAME=C
##  [9] LC_ADDRESS=C             LC_TELEPHONE=C
## [11] LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C
##
```

```

## time zone: America/Los_Angeles
## tzcode source: system (glibc)
##
## attached base packages:
## [1] grid      stats4    stats      graphics  grDevices  utils      datasets
## [8] methods   base
##
## other attached packages:
## [1] simplifyEnrichment_2.2.0 VennDiagram_1.7.3      futile.logger_1.4.3
## [4] svglite_2.1.3           Cairo_1.6-2            org.Hs.eg.db_3.21.0
## [7] AnnotationDbi_1.70.0    IRanges_2.42.0         S4Vectors_0.46.0
## [10] Biobase_2.68.0          BiocGenerics_0.54.0    generics_0.1.3
## [13] enrichplot_1.28.2       clusterProfiler_4.16.0 lubridate_1.9.4
## [16] forcats_1.0.0           stringr_1.5.1          dplyr_1.1.4
## [19] purrr_1.0.4             readr_2.1.5            tidyr_1.3.1
## [22] tibble_3.2.1            ggplot2_3.5.2          tidyverse_2.0.0
##
## loaded via a namespace (and not attached):
## [1] RColorBrewer_1.1-3      rstudioapi_0.17.1      jsonlite_2.0.0
## [4] shape_1.4.6.1           magrittr_2.0.3         modeltools_0.2-24
## [7] ggtangle_0.0.6          farver_2.1.2           rmarkdown_2.29
## [10] ragg_1.4.0              GlobalOptions_0.1.2    fs_1.6.6
## [13] vctrs_0.6.5            memoise_2.0.1          ggtree_3.16.0
## [16] htmltools_0.5.8.1       lambda.r_1.2.4         gridGraphics_0.5-1
## [19] plyr_1.8.9             futile.options_1.0.1   cachem_1.1.0
## [22] igraph_2.1.4           mime_0.13              lifecycle_1.0.4
## [25] iterators_1.0.14       pkgconfig_2.0.3        Matrix_1.7-3
## [28] R6_2.6.1               fastmap_1.2.0          gson_0.1.0
## [31] GenomeInfoDbData_1.2.14 shiny_1.10.0           clue_0.3-66
## [34] digest_0.6.37          aplot_0.2.5            colorspace_2.1-1
## [37] patchwork_1.3.0        textshaping_1.0.1      RSQLite_2.3.11
## [40] labeling_0.4.3         timechange_0.3.0       httr_1.4.7
## [43] compiler_4.5.0         bit64_4.6.0-1          withr_3.0.2
## [46] doParallel_1.0.17      BiocParallel_1.42.0    DBI_1.2.3
## [49] R.utils_2.13.0         scatterplot3d_0.3-44   rjson_0.2.23
## [52] tools_4.5.0            ape_5.8-1              flexclust_1.5.0
## [55] httpuv_1.6.16          R.oo_1.27.1            glue_1.8.0
## [58] promises_1.3.2         nlme_3.1-168           GOSemSim_2.34.0
## [61] cluster_2.1.8.1        reshape2_1.4.4         fgsea_1.34.0
## [64] gtable_0.3.6           tzdb_0.5.0             class_7.3-23
## [67] R.methodsS3_1.8.2      data.table_1.17.0      hms_1.1.3
## [70] xml2_1.3.8             XVector_0.48.0         ggrepel_0.9.6
## [73] foreach_1.5.2          pillar_1.10.2          yulab.utils_0.2.0
## [76] later_1.4.2            circlize_0.4.16        splines_4.5.0
## [79] treeio_1.32.0          lattice_0.22-5         bit_4.6.0
## [82] tidyselect_1.2.1       GO.db_3.21.0           ComplexHeatmap_2.24.0
## [85] tm_0.7-16             Biostrings_2.76.0      knitr_1.50
## [88] NLP_0.3-2             xfun_0.52              matrixStats_1.5.0
## [91] stringi_1.8.7          UCSC.utils_1.4.0       lazyeval_0.2.2
## [94] ggfun_0.1.8            yaml_2.3.10            evaluate_1.0.3
## [97] codetools_0.2-20       qvalue_2.40.0          Polychrome_1.5.4
## [100] ggplotify_0.1.2        cli_3.6.5              xtable_1.8-4
## [103] systemfonts_1.2.3      Rcpp_1.0.14            GenomeInfoDb_1.44.0
## [106] png_0.1-8              parallel_4.5.0         simona_1.6.0

```

## [109]	blob_1.2.4	DOSE_4.2.0	slam_0.1-55
## [112]	tidytree_0.4.6	scales_1.4.0	crayon_1.5.3
## [115]	GetoptLong_1.0.5	rlang_1.1.6	cowplot_1.1.3
## [118]	fastmatch_1.1-6	KEGGREST_1.48.0	formatR_1.14