



# biobeam Documentation

*Release 0.1.0*

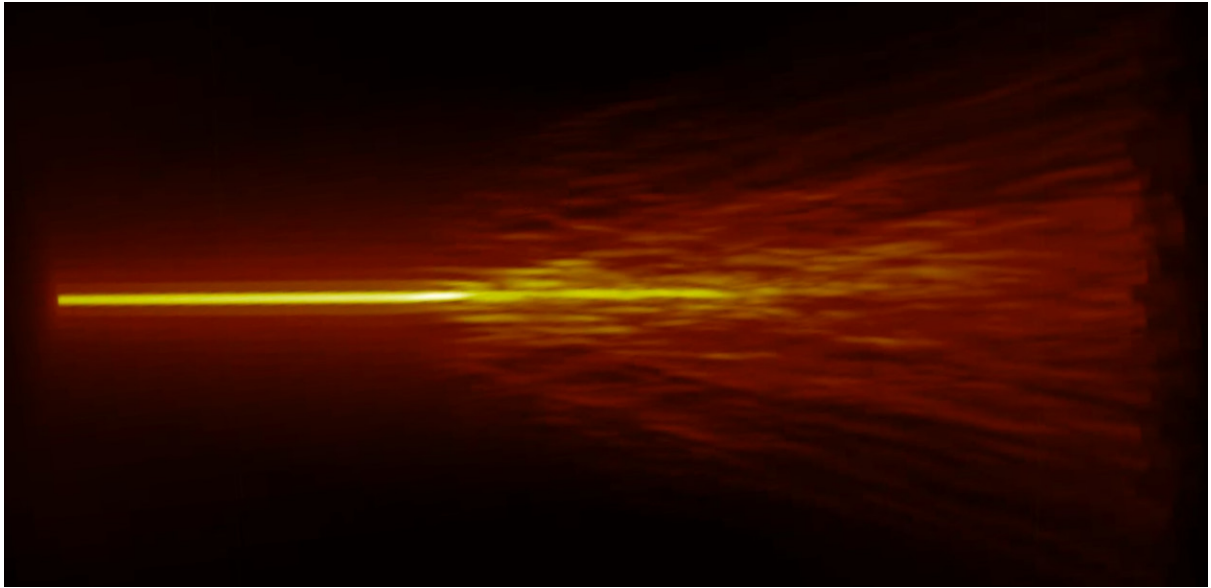
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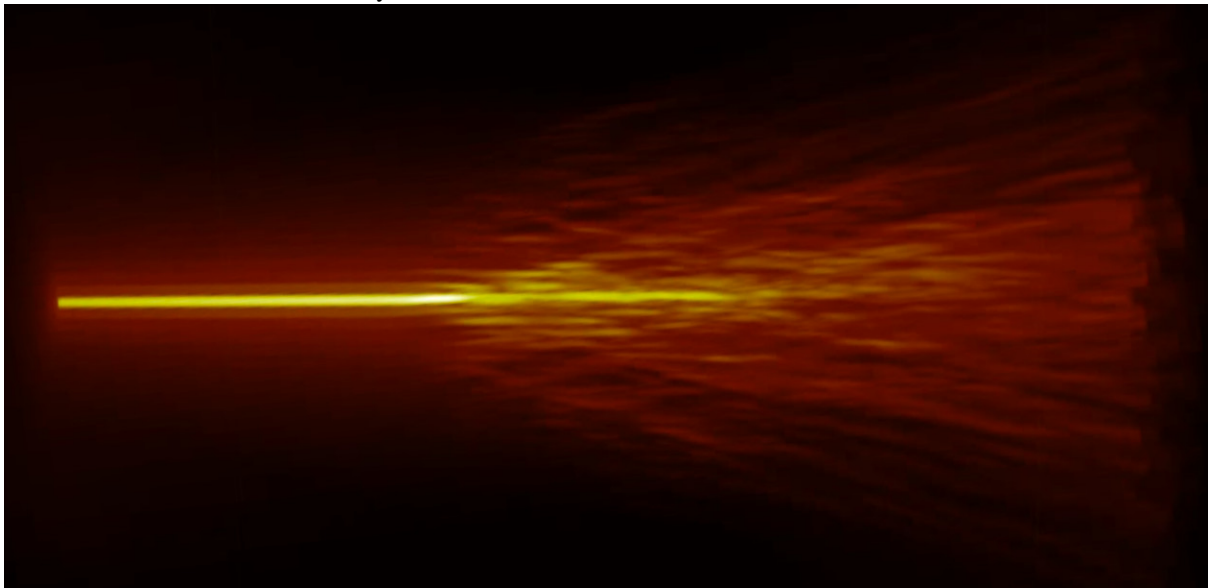
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## 1 Introduction

Biobeams is awesome. And so are you!



This is something I want to say that is not in the docstring.

## 2 Installation

To nicely render the 3d output it is advisable to install *Spimagine*, an OpenCL accelerated renderer [Spimagine](#)

```
pip install spimagine
```

After that you should be able to simply do

```
pip install biobeams
```

## 3 Basic Usage

### 3.1 Beam propagation

`class biobeam.Bpm3d` (*size=None, shape=None, units=None, dn=None, lam=0.5, n0=1.0, simul\_xy=None, simul\_z=1, n\_volumes=1, enforce\_subsampling=False, fftplan\_kwargs={}*)

the main class for gpu accelerated bpm propagation

`__init__` (*size=None, shape=None, units=None, dn=None, lam=0.5, n0=1.0, simul\_xy=None, simul\_z=1, n\_volumes=1, enforce\_subsampling=False, fftplan\_kwargs={}*)

#### Parameters

- **size** (*(Sx, Sy, Sz)*) – the size of the geometry in microns (Sx,Sy,Sz)
- **shape** (*(Nx, Ny, Nz)*) – the shape of the geometry in pixels (Nx,Ny,Nz)
- **units** (*(dx, dy, dz)*) – the voxelsizes in microns (dx,dy,dz)
- **dn** (*ndarray (float32|complex64)*) – refractive index distribution, `dn.shape != (Nz,Ny,Nx)`
- **lam** (*float*) – the wavelength in microns
- **n0** (*float*) – the refractive index of the surrounding media
- **simul\_xy** (*(Nx, Ny, Nz), optional*) – the shape of the 2d computational geometry in pixels (Nx,Ny) (e.g. subsampling in xy)
- **simul\_z** (*int, optional*) – the subsampling factor along z
- **n\_volumes** (*int*) – splits the domain into chunks if GPU memory is not large enough (will be set automatically)

#### Example

```
>>> m = Bpm3d(size = (10,10,10), shape = (256,256,256), units = (0.1,0.1,0.1), lam = 0.488, n0 = 1.0)
```

**aberr\_at** (*NA=0.4, center=(0, 0, 0), n\_zern=20, n\_integration\_steps=200*)

*c = (cx,cy,cz)* in relative pixel coordinates wrt the center

returns phi, zern

**aberr\_field\_grid** (*NA, cxs, cys, cz, n\_zern=20, n\_integration\_steps=200*)

*cxs, cys* are equally spaced 1d arrays defining the grid

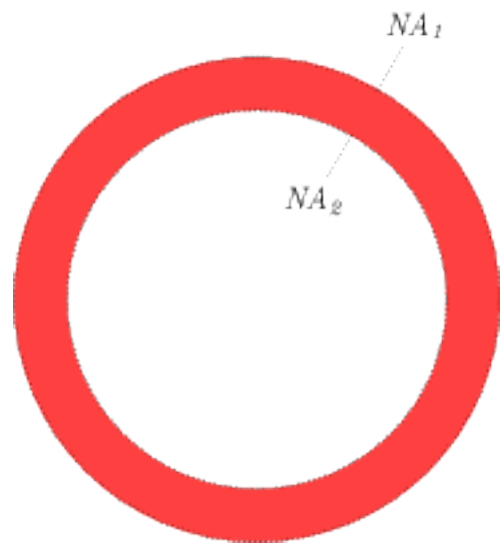
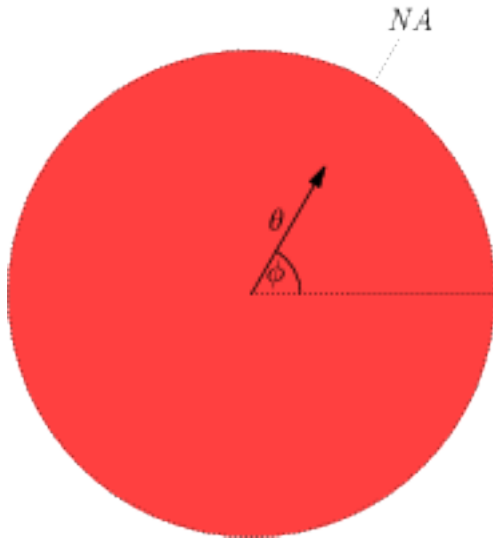
**propagate** (*u0=None, offset=0, return\_comp='field', return\_shape='full', free\_prop=False, dn\_mean\_method='none', \*\*kwargs*)

**kwargs:** `return_comp` in ["field", "intens"] `return_shape` in ["last", "full"] `free_prop` = False | True  
`dn_mean_method` = "none", "global", "local"

## 4 Input Beam patterns

### 4.1 Gaussian/Bessel beams

Gaussian/Bessel beams



`biobeam.focus_field_beam(shape=(128, 128, 128), units=(0.1, 0.1, 0.1), lam=0.5, NA=0.6, n0=1.0, return_all_fields=False, n_integration_steps=200)`

calculates the focus field for a perfect, aberration free optical system for x polarized illumination via the vectorial debye diffraction integral (see <sup>1</sup>). The pupil function is given by numerical aperture(s) NA (that can be a list to model bessell beams, see further below)

#### Parameters

- **shape** ( $N_x, N_y, N_z$ ) – the shape of the geometry
- **units** ( $dx, dy, dz$ ) – the pixel sizes in microns
- **lam** (*float*) – the wavelength of light used in microns
- **NA** (*float/list*) – the numerical aperture(s) of the illumination objective that is either a single number (for gaussian beams) or an even length list of NAs (for bessell beams), e.g.  $NA = [0.5, 0.55]$  lets light through the annulus  $0.5 < 0.55$  (making a bessell beam ) or  $NA = [0.1, 0.2, 0.5, 0.6]$  lets light through the annulus  $0.1 < 0.2$  and  $0.5 < 0.6$  making a beating double bessell beam...
- **n0** (*float*) – the refractive index of the medium
- **n\_integration\_steps** (*int*) – number of integration steps to perform
- **return\_all\_fields** (*boolean*) – if True returns also the complex vectorial field components

<sup>1</sup> Matthew R. Foreman, Peter Toeroek, *Computational methods in vectorial imaging*, Journal of Modern Optics, 2011, 58, 5-6, 339

### Returns

- **u** (*ndarray*) – the intensity of the focus field
- **(u,ex,ey,ez)** (*list(ndarray)*) – the intensity of the focus field and the complex field components (if `return_all_fields` is `True`)

### Example

```
>>> u = focus_field_beam((128,128,128), (0.1,0.1,.1), lam=.5, NA = .4)
```

### References

`biobeam.focus_field_beam_plane(shape=(128, 128), units=(0.1, 0.1), z=0.0, lam=0.5, NA=0.6, n0=1.0, ex_g=None, n_integration_steps=200)`  
calculates the complex 2d input field at position -z of a perfect, aberration free optical system

### Parameters

- **shape** (*Nx, Ny*) – the 2d shape of the geometry
- **units** (*dx, dy*) – the pixel sizes in microns
- **z** (*float*) – defocus position in microns, such that the beam would focus at z e.g. an input field with  $z = 10$ . would hav its focus spot after 10 microns
- **lam** (*float*) – the wavelength of light used in microns
- **NA** (*float/list*) – the numerical aperture(s) of the illumination objective that is either a single number (for gaussian beams) or an even length list of NAs (for bessell beams), e.g.  $NA = [0.5, 0.55]$  lets light through the annulus  $0.5 < 0.55$  (making a bessell beam ) or  $NA = [0.1, 0.2, 0.5, 0.6]$  lets light through the annulus  $0.1 < 0.2$  and  $0.5 < 0.6$  making a beating double bessell beam...
- **n0** (*float*) – the refractive index of the medium
- **n\_integration\_steps** (*int*) – number of integration steps to perform

**Returns** **ex** – the complex field

**Return type** `ndarray`

### Example

```
>>> # the input pattern of a bessell beam that will focus after 4 microns
>>> ex = focus_field_beam_plane((256,256), (0.1,0.1), z = 4, lam=.5, NA = (.4,.5))
```

See also:

`biobeam.focus_field_beam()` the 3d function

## 4.2 Cylindrical Lens



```
biobeam.focus_field_cylindrical (shape=(128, 128, 128), units=(0.1, 0.1, 0.1),  
                                lam=0.5, NA=0.3, n0=1.0, return_all_fields=False,  
                                n_integration_steps=100)
```

calculates the focus field for a perfect, aberration free cylindrical lens after x polarized illumination via the vectorial debye diffraction integral (see <sup>2</sup>). The pupil function is given by the numerical aperture NA

#### Parameters

- **shape** (*Nx, Ny, Nz*) – the shape of the geometry
- **units** (*dx, dy, dz*) – the pixel sizes in microns
- **lam** (*float*) – the wavelength of light used in microns
- **NA** (*float*) – the numerical aperture of the lens
- **n0** (*float*) – the refractive index of the medium
- **return\_all\_fields** (*boolean*) – if True, returns u,ex,ey,ez where ex/ey/ez are the complex field components
- **n\_integration\_steps** (*int*) – number of integration steps to perform
- **return\_all\_fields** – if True returns also the complex vectorial field components

#### Returns

- **u** (*ndarray*) – the intensity of the focus field
- **(u,ex,ey,ez)** (*list(ndarray)*) – the intensity of the focus field and the complex field components (if return\_all\_fields is True)

#### Example

```
>>> u, ex, ey, ez = focus_field_cylindrical((128,128,128), (0.1,0.1,.1), lam=.5, NA = .4, ret
```

#### References

```
biobeam.focus_field_cylindrical_plane (shape=(128, 128), units=(0.1, 0.1), z=0.0,  
                                lam=0.5, NA=0.3, n0=1.0, ex_g=None,  
                                n_integration_steps=200)
```

calculates the complex 2d input field at position -z of a perfect, aberration free optical system

#### Parameters

- **shape** (*Nx, Ny*) – the 2d shape of the geometry
- **units** (*dx, dy*) – the pixel sizes in microns
- **z** (*float*) – defocus position in microns, such that the beam would focus at z e.g. an input field with z = 10. would hav its focus spot after 10 microns
- **lam** (*float*) – the wavelength of light used in microns
- **NA** (*float/list*) – the numerical aperture(s) of the illumination objective that is either a single number (for gaussian beams) or an even length list of NAs (for bessell beams), e.g. NA = [0.5,0.55] lets light through the annulus 0.5<0.55 (making a bessell beam ) or NA = [0.1,0.2,0.5,0.6] lets light through the annulus 0.1<0.2 and 0.5<0.6 making a beating double bessell beam...
- **n0** (*float*) – the refractive index of the medium
- **n\_integration\_steps** (*int*) – number of integration steps to perform

#### Returns **ex** – the complex field

<sup>2</sup> Colin J. R. Sheppard: Cylindrical lenses—focusing and imaging: a review, Appl. Opt. 52, 538-545 (2013)

**Return type** ndarray

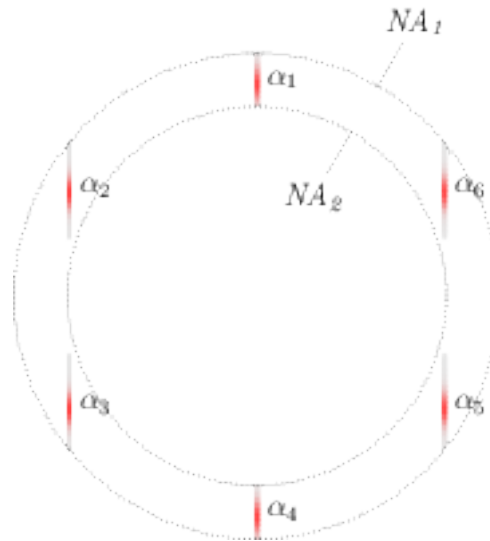
### Example

```
>>> # the input pattern of a bessell beam that will focus after 4 microns
>>> ex = focus_field_cylindrical_plane((256,256), (0.1,0.1), z = 4., lam=.5, NA = (.4,.5))
```

**See also:**

`biobeam.focus_field_cylindrical()` the 3d function

## 4.3 Bessel Lattices



```
biobeam.focus_field_lattice(shape=(128, 128, 128), units=(0.1, 0.1, 0.1), lam=0.5, NA1=0.4,
                             NA2=0.5, sigma=0.1, kpoints=6, return_all_fields=False,
                             n0=1.0, n_integration_steps=100)
```

Calculates the focus field for a bessel lattice. The pupil function consists out of discrete points (kpoints) superimposed on an annulus (NA1<NA2) which are smeared out by a 1d gaussian of given sigma creating an array of bessel beams in the focal plane (see <sup>3</sup>).

### Parameters

- **shape** ( $N_x, N_y, N_z$ ) – the shape of the geometry
- **units** ( $dx, dy, dz$ ) – the pixel sizes in microns
- **lam** (*float*) – the wavelength of light used in microns
- **NA1** (*float/list*) – the numerical aperture of the inner ring
- **NA2** (*float/list*) – the numerical aperture of the outer ring
- **sigma** (*float*) – the standard deviation of the gaussian smear function applied to each point on the aperture (the bigger sigma, the tighter the sheet in y)
- **kpoints** (*int/ (2,N) array*) – defines the set of points on the aperture that create the lattice, can be - a (2,N) ndarray, such that kpoints[:,i] are the coordinates of the ith point - a single int, defining points on a regular polygon (e.g. 4 for a square lattice, 6 for a hex lattice)  $k_i = \arcsin \frac{NA_1 + NA_2}{2n_0} \begin{pmatrix} \cos \phi_i \\ \sin \phi_i \end{pmatrix}$ ,  $\phi_i = \frac{\pi}{2} + \frac{2i}{N}$
- **n0** (*float*) – the refractive index of the medium

<sup>3</sup> Chen et al. Lattice light-sheet microscopy: imaging molecules to embryos at high spatiotemporal resolution. Science 346, (2014).

- **n\_integration\_steps** (*int*) – number of integration steps to perform
- **return\_all\_fields** (*boolean*) – if True, returns u,ex,ey,ez where ex/ey/ez are the complex vector field components

#### Returns

- **u** (*ndarray*) – the intensity of the focus field
- **(u,ex,ey,ez)** (*list(ndarray)*) – the intensity of the focus field and the complex field components (if return\_all\_fields is True)

#### Example

```
>>> u = focus_field_lattice((128,128,128), (0.1,0.1,.1), lam=.5, NA1 = .44, NA2 = .55, kpoint
```

#### References

biobeam.**focus\_field\_lattice\_plane** (*shape=(256, 256), units=(0.1, 0.1), z=0.0, lam=0.5, NA1=0.4, NA2=0.5, sigma=0.1, kpoints=6, n0=1.0, apodization\_bound=10, ex\_g=None, n\_integration\_steps=100*)

calculates the complex 2d input field at position -z of a for a besell lattice beam.

#### Parameters

- **shape** (*Nx, Ny*) – the shape of the geometry
- **units** (*dx, dy*) – the pixel sizes in microns
- **z** (*float*) – defocus position in microns, such that the beam would focus at z e.g. an input field with z = 10. would have its focal spot after 10 microns
- **lam** (*float*) – the wavelength of light used in microns
- **NA1** (*float/list*) – the numerical aperture of the inner ring
- **NA2** (*float/list*) – the numerical aperture of the outer ring
- **sigma** (*float*) – the standard deviation of the gaussian smear function applied to each point on the aperture (the bigger sigma, the tighter the sheet in y)
- **kpoints** (*int/ (2,N) array*) – defines the set of points on the aperture that create the lattice, can be - a (2,N) ndarray, such that kpoints[:,i] are the coordinates of the ith point - a single int, defining points on a regular polygon (e.g. 4 for a square lattice, 6 for a hex lattice)  $k_i = \arcsin \frac{NA_1+NA_2}{2n_0} \begin{pmatrix} \cos \phi_i \\ \sin \phi_i \end{pmatrix}$ ,  $\phi_i = \frac{\pi}{2} + \frac{2i}{N}$
- **n0** (*float*) – the refractive index of the medium
- **apodization\_bound** (*int*) – width of the region where the input field is tapered to zero (with a hamming window) on the +/- x borders
- **n\_integration\_steps** (*int*) – number of integration steps to perform
- **return\_all\_fields** (*boolean*) – if True, returns u,ex,ey,ez where ex/ey/ez are the complex vector field components

**Returns** **u** – the 2d complex field

**Return type** ndarray



### Example

```
>>> u = focus_field_lattice_plane((128,128), (0.1,0.1), z = 2., lam=.5, NA1 = .44, NA2 = .55,
```

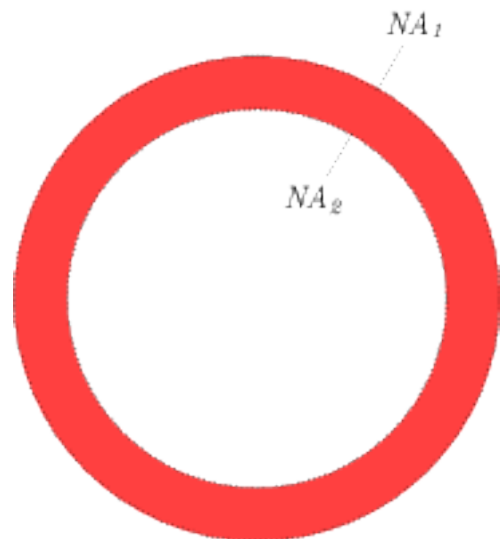
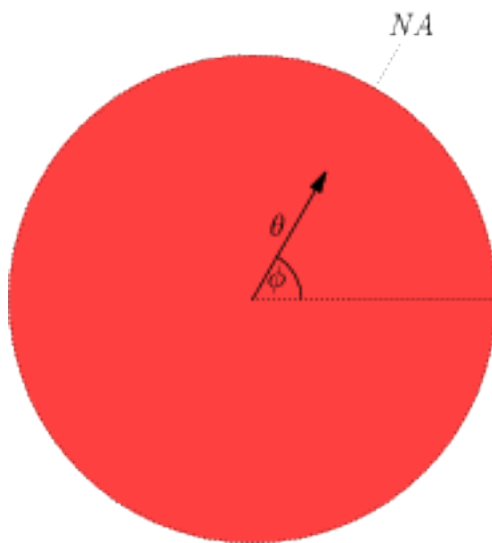
See also:

`biobeam.focus_field_lattice()` the corresponding 3d function

## 5 Focus field calculations

### 5.1 Gaussian/Bessel beams

Gaussian/Bessel beams



`biobeam.focus_field_beam(shape=(128, 128, 128), units=(0.1, 0.1, 0.1), lam=0.5, NA=0.6, n0=1.0, return_all_fields=False, n_integration_steps=200)`

calculates the focus field for a perfect, aberration free optical system for x polarized illumination via the vectorial debye diffraction integral (see <sup>1</sup>). The pupil function is given by numerical aperture(s) NA (that can be a list to model bessel beams, see further below)

#### Parameters

<sup>1</sup> Matthew R. Foreman, Peter Toeroek, *Computational methods in vectorial imaging*, Journal of Modern Optics, 2011, 58, 5-6, 339

- **shape** ( $N_x, N_y, N_z$ ) – the shape of the geometry
- **units** ( $dx, dy, dz$ ) – the pixel sizes in microns
- **lam** ( $float$ ) – the wavelength of light used in microns
- **NA** ( $float/list$ ) – the numerical aperture(s) of the illumination objective that is either a single number (for gaussian beams) or an even length list of NAs (for bessell beams), e.g.  $NA = [0.5, 0.55]$  lets light through the annulus  $0.5 < 0.55$  (making a bessell beam ) or  $NA = [0.1, 0.2, 0.5, 0.6]$  lets light through the annulus  $0.1 < 0.2$  and  $0.5 < 0.6$  making a beating double bessell beam...
- **n0** ( $float$ ) – the refractive index of the medium
- **n\_integration\_steps** ( $int$ ) – number of integration steps to perform
- **return\_all\_fields** ( $boolean$ ) – if True returns also the complex vectorial field components

### Returns

- **u** ( $ndarray$ ) – the intensity of the focus field
- **(u,ex,ey,ez)** ( $list(ndarray)$ ) – the intensity of the focus field and the complex field components (if return\_all\_fields is True)

### Example

```
>>> u = focus_field_beam((128,128,128), (0.1,0.1,.1), lam=.5, NA = .4)
```

### References

`biobeam.focus_field_beam_plane(shape=(128, 128), units=(0.1, 0.1), z=0.0, lam=0.5, NA=0.6, n0=1.0, ex_g=None, n_integration_steps=200)`  
calculates the complex 2d input field at position -z of a perfect, aberration free optical system

### Parameters

- **shape** ( $N_x, N_y$ ) – the 2d shape of the geometry
- **units** ( $dx, dy$ ) – the pixel sizes in microns
- **z** ( $float$ ) – defocus position in microns, such that the beam would focus at z e.g. an input field with  $z = 10$ . would hav its focus spot after 10 microns
- **lam** ( $float$ ) – the wavelength of light used in microns
- **NA** ( $float/list$ ) – the numerical aperture(s) of the illumination objective that is either a single number (for gaussian beams) or an even length list of NAs (for bessell beams), e.g.  $NA = [0.5, 0.55]$  lets light through the annulus  $0.5 < 0.55$  (making a bessell beam ) or  $NA = [0.1, 0.2, 0.5, 0.6]$  lets light through the annulus  $0.1 < 0.2$  and  $0.5 < 0.6$  making a beating double bessell beam...
- **n0** ( $float$ ) – the refractive index of the medium
- **n\_integration\_steps** ( $int$ ) – number of integration steps to perform

**Returns** **ex** – the complex field

**Return type** `ndarray`

### Example

```
>>> # the input pattern of a bessel beam that will focus after 4 microns
>>> ex = focus_field_beam_plane((256,256), (0.1,0.1), z = 4, lam=.5, NA = (.4,.5))
```

See also:

`biobeam.focus_field_beam()` the 3d function

## 5.2 Cylindrical Lens



```
biobeam.focus_field_cylindrical(shape=(128, 128, 128), units=(0.1, 0.1, 0.1),
                                lam=0.5, NA=0.3, n0=1.0, return_all_fields=False,
                                n_integration_steps=100)
```

calculates the focus field for a perfect, aberration free cylindrical lens after x polarized illumination via the vectorial debye diffraction integral (see <sup>2</sup>). The pupil function is given by the numerical aperture NA

### Parameters

- **shape** ( $N_x, N_y, N_z$ ) – the shape of the geometry
- **units** ( $dx, dy, dz$ ) – the pixel sizes in microns
- **lam** (*float*) – the wavelength of light used in microns
- **NA** (*float*) – the numerical aperture of the lens
- **n0** (*float*) – the refractive index of the medium
- **return\_all\_fields** (*boolean*) – if True, returns  $u, ex, ey, ez$  where  $ex/ey/ez$  are the complex field components
- **n\_integration\_steps** (*int*) – number of integration steps to perform
- **return\_all\_fields** – if True returns also the complex vectorial field components

### Returns

- **u** (*ndarray*) – the intensity of the focus field
- **(u,ex,ey,ez)** (*list(ndarray)*) – the intensity of the focus field and the complex field components (if `return_all_fields` is True)

### Example

```
>>> u, ex, ey, ez = focus_field_cylindrical((128,128,128), (0.1,0.1,.1), lam=.5, NA = .4, ret
```

### References

```
biobeam.focus_field_cylindrical_plane(shape=(128, 128), units=(0.1, 0.1), z=0.0,
                                        lam=0.5, NA=0.3, n0=1.0, ex_g=None,
                                        n_integration_steps=200)
```

calculates the complex 2d input field at position -z of a perfect, aberration free optical system

### Parameters

<sup>2</sup> Colin J. R. Sheppard: Cylindrical lenses—focusing and imaging: a review, Appl. Opt. 52, 538-545 (2013)

- **shape** ( $N_x, N_y$ ) – the 2d shape of the geometry
- **units** ( $dx, dy$ ) – the pixel sizes in microns
- **z** (*float*) – defocus position in microns, such that the beam would focus at z e.g. an input field with  $z = 10$ . would hav its focus spot after 10 microns
- **lam** (*float*) – the wavelength of light used in microns
- **NA** (*float/list*) – the numerical aperture(s) of the illumination objective that is either a single number (for gaussian beams) or an even length list of NAs (for bessell beams), e.g.  $NA = [0.5, 0.55]$  lets light through the annulus  $0.5 < 0.55$  (making a bessell beam ) or  $NA = [0.1, 0.2, 0.5, 0.6]$  lets light through the annulus  $0.1 < 0.2$  and  $0.5 < 0.6$  making a beating double bessell beam...
- **n0** (*float*) – the refractive index of the medium
- **n\_integration\_steps** (*int*) – number of integration steps to perform

**Returns** **ex** – the complex field

**Return type** ndarray

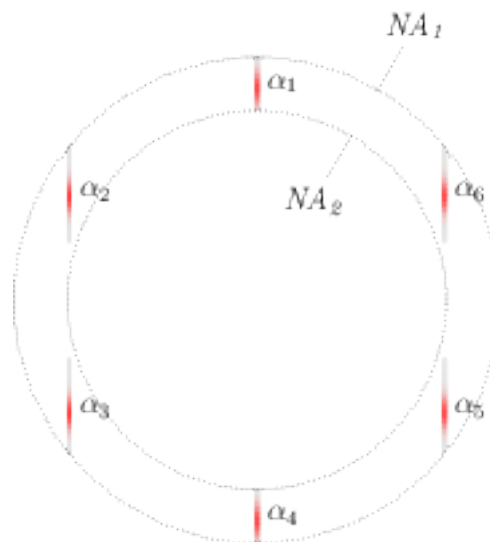
### Example

```
>>> # the input pattern of a bessell beam that will focus after 4 microns
>>> ex = focus_field_cylindrical_plane((256,256), (0.1,0.1), z = 4., lam=.5, NA = (.4,.5))
```

See also:

`biobeam.focus_field_cylindrical()` the 3d function

## 5.3 Bessel Lattices



```
biobeam.focus_field_lattice(shape=(128, 128, 128), units=(0.1, 0.1, 0.1), lam=0.5, NA1=0.4,
                             NA2=0.5, sigma=0.1, kpoints=6, return_all_fields=False,
                             n0=1.0, n_integration_steps=100)
```

Calculates the focus field for a bessell lattice. The pupil function consists out of discrete points ( $kpoints$ ) superimposed on an annulus ( $NA1 < NA2$ ) which are smeared out by a 1d gaussian of given sigma creating an array of bessell beams in the focal plane (see <sup>3</sup>).

### Parameters

<sup>3</sup> Chen et al. Lattice light-sheet microscopy: imaging molecules to embryos at high spatiotemporal resolution. Science 346, (2014).

- **shape** ( $N_x, N_y, N_z$ ) – the shape of the geometry
- **units** ( $dx, dy, dz$ ) – the pixel sizes in microns
- **lam** ( $float$ ) – the wavelength of light used in microns
- **NA1** ( $float/list$ ) – the numerical aperture of the inner ring
- **NA2** ( $float/list$ ) – the numerical aperture of the outer ring
- **sigma** ( $float$ ) – the standard deviation of the gaussian smear function applied to each point on the aperture (the bigger sigma, the tighter the sheet in y)
- **kpoints** ( $int/ (2, N) \text{ array}$ ) – defines the set of points on the aperture that create the lattice, can be - a (2,N) ndarray, such that kpoints[:,i] are the coordinates of the ith point - a single int, defining points on a regular polygon (e.g. 4 for a square lattice, 6 for a hex lattice)  $k_i = \arcsin \frac{NA_1 + NA_2}{2n_0} \begin{pmatrix} \cos \phi_i \\ \sin \phi_i \end{pmatrix}$ ,  $\phi_i = \frac{\pi}{2} + \frac{2i}{N}$
- **n0** ( $float$ ) – the refractive index of the medium
- **n\_integration\_steps** ( $int$ ) – number of integration steps to perform
- **return\_all\_fields** ( $boolean$ ) – if True, returns u,ex,ey,ez where ex/ey/ez are the complex vector field components

### Returns

- **u** ( $ndarray$ ) – the intensity of the focus field
- **(u,ex,ey,ez)** ( $list(ndarray)$ ) – the intensity of the focus field and the complex field components (if return\_all\_fields is True)

### Example

```
>>> u = focus_field_lattice((128,128,128), (0.1,0.1,.1), lam=.5, NA1 = .44, NA2 = .55, kpoint
```

### References

biobeam.**focus\_field\_lattice\_plane** ( $shape=(256, 256)$ ,  $units=(0.1, 0.1)$ ,  $z=0.0$ ,  $lam=0.5$ ,  $NA1=0.4$ ,  $NA2=0.5$ ,  $sigma=0.1$ ,  $kpoints=6$ ,  $n0=1.0$ ,  $apodization_bound=10$ ,  $ex_g=None$ ,  $n_integration_steps=100$ )

calculates the complex 2d input field at position -z of a for a besell lattice beam.

### Parameters

- **shape** ( $N_x, N_y$ ) – the shape of the geometry
- **units** ( $dx, dy$ ) – the pixel sizes in microns
- **z** ( $float$ ) – defocus position in microns, such that the beam would focus at z e.g. an input field with  $z = 10$ . would have its focal spot after 10 microns
- **lam** ( $float$ ) – the wavelength of light used in microns
- **NA1** ( $float/list$ ) – the numerical aperture of the inner ring
- **NA2** ( $float/list$ ) – the numerical aperture of the outer ring
- **sigma** ( $float$ ) – the standard deviation of the gaussian smear function applied to each point on the aperture (the bigger sigma, the tighter the sheet in y)
- **kpoints** ( $int/ (2, N) \text{ array}$ ) – defines the set of points on the aperture that create the lattice, can be - a (2,N) ndarray, such that kpoints[:,i] are the coordinates of

the  $i$ th point - a single int, defining points on a regular polygon (e.g. 4 for a square lattice, 6 for a hex lattice)  $k_i = \arcsin \frac{NA_1 + NA_2}{2n_0} \begin{pmatrix} \cos \phi_i \\ \sin \phi_i \end{pmatrix}$ ,  $\phi_i = \frac{\pi}{2} + \frac{2i}{N}$

- **n0** (*float*) – the refractive index of the medium
- **apodization\_bound** (*int*) – width of the region where the input field is tapered to zero (with a hamming window) on the +/- x borders
- **n\_integration\_steps** (*int*) – number of integration steps to perform
- **return\_all\_fields** (*boolean*) – if True, returns u,ex,ey,ez where ex/ey/ez are the complex vector field components

**Returns** **u** – the 2d complex field

**Return type** ndarray

### Example

```
>>> u = focus_field_lattice_plane((128,128), (0.1,0.1), z = 2., lam=.5, NA1 = .44, NA2 = .55,
```

**See also:**

`biobeam.focus_field_lattice()` the corresponding 3d function

## 6 Simulating light sheet microscopy

## 7 Aberrations

## 8 Examples

### 8.1 Plane wave scattered by sphere

```
# create the refractive index difference
x = 0.1 * np.arange(-128,128)
Z, Y, X = np.meshgrid(x,x,x,indexing = "ij")
R = np.sqrt(X**1+Y**2+Z**2)
dn = 0.05*(R<2.)

# create the computational geometry
m = Bpm3d(dn = dn, units = (0.1,0.1,0.1), lam = 0.5)

# propagate a plane wave and return the intensity
u = m._propagate()

# vizualize
import matplotlib.pyplot as plt
plt.subplot(1,2,1)
plt.imshow(u[...,128], cmap = "hot")
plt.title("zy slice")
plt.subplot(1,2,2)
plt.imshow(u[128,...], cmap = "hot")
plt.title("xy slice")
```

## 8.2 Light sheet through cell phantom

## 8.3 Computing the psf inside a cell phantom

## 8.4 Aberration from sphere

# 9 Some Examples

or not?

```
print "huhu"
```

```
huhu
```

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