Class 10: Structural Bioinformatics part 1

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What is in the PDB database

The main repository of biomolecular structure info is in the PDB < www.rcsb.org> Lets see what this database contains:

```
stats <- read.csv("pdb_stats.csv", row.names = 1)
stats</pre>
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	161,663	12,592	12,337	200	74	32
Protein/Oligosaccharide	9,348	2,167	34	8	2	0
Protein/NA	8,404	3,924	286	7	0	0
Nucleic acid (only)	2,758	125	1,477	14	3	1
Other	164	9	33	0	0	0
Oligosaccharide (only)	11	0	6	1	0	4
	Total					
Protein (only)	186,898					
Protein/Oligosaccharide	11,559					
Protein/NA	12,621					
Nucleic acid (only)	4,378					
Other	206					
Oligosaccharide (only)	22					

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
as.numeric(stats$X.ray)
```

Warning: NAs introduced by coercion

[1] NA NA NA NA 164 11

We got to get rid of the commas. Can you find a function to get rid of the commas?

```
x <- stats$X.ray
sum( as.numeric(gsub(",", "", x)) )</pre>
```

[1] 182348

I am going to turn this into a function and then use apply() to work on the entire table of data

```
sumcomma <- function (x) {
  sum( as.numeric(gsub(",", "", x)) )
}
sumcomma(stats$X.ray)</pre>
```

[1] 182348

```
n.total <- sumcomma(stats$Total)
n.total</pre>
```

[1] 215684

```
sumcomma(stats$EM)
```

[1] 18817

```
apply(stats, 2, sumcomma)
```

X.ray	EM	NMR	Multiple.methods
182348	18817	14173	230
Neutron	Other	Total	
79	37	215684	

```
apply(stats, 2, sumcomma) / sumcomma(stats$Total)
```

```
X.ray EM NMR Multiple.methods
0.8454405519 0.0872433746 0.0657118748 0.0010663749
Neutron Other Total
0.0003662766 0.0001715473 1.0000000000
```

Q2: What proportion of structures in the PDB are protein?

248,805,733 - 186,898

```
186898/248805733*100
```

[1] 0.07511804

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

Visualizing the HIV-1 protease structure

Mol* ("mol-star") viewer is now everywhere. The homepage os here: https://molstar.org/viewer/ I want to insert my image from Mol* here

Working with bio3d

```
library(bio3d)

pdb <- read.pdb("1hsg")

Note: Accessing on-line PDB file

pdb</pre>
```



Figure 1: My first molecular image



Figure 2: My second molecular image

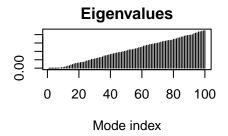
```
Call: read.pdb(file = "1hsg")
  Total Models#: 1
    Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
    Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
    Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
    Non-protein/nucleic Atoms#: 172 (residues: 128)
    Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
  Protein sequence:
     PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
     QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
     ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
     VNIIGRNLLTQIGCTLNF
+ attr: atom, xyz, seqres, helix, sheet,
       calpha, remark, call
  head(pdb$atom)
  type eleno elety alt resid chain resno insert
                                                          У
1 ATOM
          1
                N < NA >
                         PRO
                                Α
                                      1
                                          <NA> 29.361 39.686 5.862 1 38.10
2 ATOM
          2
                         PRO
                                      1 <NA> 30.307 38.663 5.319 1 40.62
               CA <NA>
                               Α
                        PRO
                                     1 <NA> 29.760 38.071 4.022 1 42.64
3 ATOM
               C <NA>
                               Α
4 ATOM
          4
               O <NA>
                        PRO
                                     1 <NA> 28.600 38.302 3.676 1 43.40
                               Α
5 ATOM
          5
               CB <NA>
                        PRO
                                     1 <NA> 30.508 37.541 6.342 1 37.87
                                      1 <NA> 29.296 37.591 7.162 1 38.40
6 ATOM
          6
               CG <NA>
                        PRO
                                Α
  segid elesy charge
1 <NA>
           N
               <NA>
2 <NA>
           C <NA>
3 <NA>
           C <NA>
4 <NA>
           O <NA>
5 <NA>
           C <NA>
6 <NA>
           C
               <NA>
```

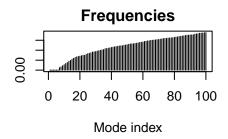
pdbseq(pdb)[25]

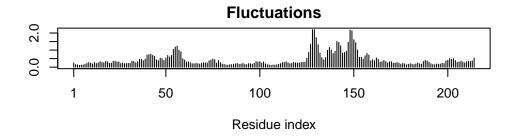
Predicting functional motions of a single structure

We can do bioinformatics prediction of functional motions (ie. flexibility/dynamics):

```
pdb <- read.pdb("6s36")</pre>
 Note: Accessing on-line PDB file
  PDB has ALT records, taking A only, rm.alt=TRUE
  pdb
       read.pdb(file = "6s36")
  Total Models#: 1
     Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)
    Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
    Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
     Non-protein/nucleic Atoms#: 244 (residues: 244)
    Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]
  Protein sequence:
     MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIMDAGKLVT
     DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDKI
     VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
     YYSKEAEAGNTKYAKVDGTKPVAEVRADLEKILG
+ attr: atom, xyz, seqres, helix, sheet,
       calpha, remark, call
  m <- nma(pdb)
Building Hessian...
                           Done in 0.015 seconds.
Diagonalizing Hessian...
                           Done in 0.275 seconds.
```







mktrj(m, file="adk_m7.pdb")