

## **Tutorial: Running ANTx on machines without graphic support “working without graphical user interfaces, GUIs”**

**This tutorial shows how to perform some basic steps without GUIs, for instance when running on a HPC-machine.**

### **CONTENTS**

- 1) **OPTIONAL:** How to set the paths of ELASTIX in UNIX (LINUX)-systems:
- 2) **OPTIONAL:** Open an interactive session on HPC and start Matlab
- 3) **BASICS**
  - ADD ANTx-PATHS
  - GO TO STUDY-FOLDER
  - UPDATE ANT-TOOLBOX
  - CREATE A PROJECT-FILE:
  - LOAD AN ANTx-PROJECT-FILE ("proj.m")
  - CHECK WHETHER THE PROJECT-FILE IS LOADED
- 4) **IMPORT BRUKER RAW-DATA**
- 5) **VISUALIZE FILES AND FOLDERS**
- 6) **SELECTION OF ANIMALS**
- 7) **RENAME FILES**
- 8) **REGISTER “t2.nii” TO TEMPLATE SPACE (STANDARD SPACE)**
- 9) **EXTRACT THE FIRST 3D-VOLUME FROM THE 4D-VOLUME 'dti\_b100.nii'**
- 10) **COREGISTER 'dti\_b100\_1stIMG.nii' TO ‘t2.nii’**
- 11) **TRANSFORM ANOTHER IMAGE TO STANDARD-SPACE**
- 12) **TRANSFORM ANOTHER IMAGE TO NATIVE-SPACE**
- 13) **CHECK REGISTRATION in STANDARD-SPACE - CREATE HTML-FILE**
- 14) **CHECK REGISTRATION in NATIVE-SPACE - CREATE HTML-FILE**
- 15) **REGIONWISE PARAMETER-EXTRACTION**

## 1) OPTIONAL: How to set the paths of ELASTIX in UNIX (LINUX)-systems:

### MAKE ELASTIX RUNNING ON UNIX/LINUX-Systems

#### 1) SET PATH OF ELASTIX IN .bashrc-FILE

##### FOR INFORMATION:

see: Elastix-manual (section 3.2)

see: <https://usermanual.wiki/Document/elastix490manual.1389615963/html#pf17>

Linux: Add the following lines to your .bashrc file:

```
export PATH=folder/bin:$PATH
export LD_LIBRARY_PATH=folder/lib:$LD_LIBRARY_PATH
```

...where "folder" is the path to the linux-Elastix-folder (which is located in the antx2-folder): YOUR-DRIVE\antx2\mritools\elastix\elastix linux64 v4.7

##### EXAMPLE:

My linux-elastix-folder is "/sc-projects/sc-proj-agtiermrt/Daten-2/ressources/antx2/mritools/elastix/elastix linux64 v4.7/". Thus, my bashrc-file is modified as follows:

```
#-----[ELASTIX-PATH in bashrc]-----
export PATH=/sc-projects/sc-proj-agtiermrt/Daten-2/ressources/antx2/mritools/elastix/elastix_linux64_v4.7/bin:$PATH
export LD_LIBRARY_PATH=/sc-projects/sc-proj-agtiermrt/Daten-2/ressources/antx2/mritools/elastix/elastix linux64 v4.7/lib:$LD_LIBRARY_PATH
#-----
```

#### 2) RELOAD .bashrc-FILE & TEST ELASTIX

- save .bashrc-file, exit editor, then type the following to reload the .bashrc-file again:  
**source .bashrc**

- check installation, by typing:

**elastix**

- if successful, a message is displayed:

Use "elastix --help" for information about elastix-usage.

## 2) OPTIONAL: Open interactive session on HPC and start Matlab

### OPEN INTERACTIVE JOB on HPC (optional)

```
srun --time 7-00 --mem=64G --ntasks=8 --pty bash -i
```

-Please check the parameters, here for 7 days, 64 Gb Ram, 8 cores

### LOAD MATLAB-MODULE AND START MATLAB (optional)

```
module load scientific/matlab/R2021b
```

```
matlab
```

-to access Matlab might be different on another machine!

## 3) BASICS

### FROM NOW ON type in the MATLAB CMD-WINDOW...

#### ADD ANTx-PATHS

Go to the ANTx-patd and link all necessary paths using "antlink"-command:

```
cd /sc-projects/sc-proj-agtiermrt/Daten-2/ressources/antx2/
antlink
```

#### GO TO STUDY-FOLDER

Create an empty study folder (here "groeschel"). The study-folder is the folder where the registration of several animals of a study is performed.

```
cd /sc-projects/sc-proj-agtiermrt/Daten-2/mri/projects/groeschel/
```

## UPDATE ANT-TOOLBOX

This is not mandatory... just type **updateantx(2)** to update the toolbox, i.e. obtain the latest version from GitHub. For more info type help updateantx.

```
updateantx(2)
```

## CREATE AN ANT<sub>x</sub>-PROJECT-FILE:

```
makeproject('projectname',fullfile(pwd,'proj.m'),'voxsize',[.07 .07 .07],'wa_refpath','/sc-projects/sc-proj-agtiermrt/Daten-2/ressources/anttemplates/mouse_Allen2017HikishimaLR','wa_species','mouse');
```

-here the project-file "**proj.m**" is created using a target voxel size of 0.07 x 0.07 x 0.07 mm, the **animal template** is "**mouse Allen2017HikishimaLR**", with the species '**mouse**'.

- a suitable template has to be downloaded from google-drive :

<https://drive.google.com/drive/folders/1q5XOOVLvUYLqYsQJLqNRF7OK8fNwYhI9>

The template should be unzipped and stored where it could be reached (do not save the downloaded template in the current project-folder!). Creating a project-file has to be done only once!

## LOAD THE PROJECT-FILE "proj.m"

Here we load the created project-file "**proj.m**":

```
loadconfig(fullfile(pwd,'proj.m'));
```

## CHECK WHETHER THE PROJECT-FILE IS LOADED

```
global an;  
an
```

The global variable "an" (struct) contains the basic information for this study. In the CMD-window you should the following:

```
project: 'NEW PROJECT'  
datpath: '/sc-projects/sc-proj-agtiermrt/Daten-2/mri/projects/groeschel/dat'  
voxsize: [0.0700 0.0700 0.0700]  
wa: [1x1 struct]
```

## 4) IMPORT BRUKER-DATA

Before doing this step, I just created the 'raw'-folder within the study's folder and copied the datasets of two animals into the 'raw'-folder. First we just read the file-information of the Bruker raw-data from the "raw"-folder (fullfile(pwd,'raw')) which is located in the current study folder.

The file-information will be stored in the resulting w-struct.

```
w=xbrucker2nifti(fullfile(pwd,'raw'),0,[],[],'gui',0,'show',1); % first read all data and show it
```

The w-struct contains the file-information table "d" with header "hd". This table is listed in the CMD-window when running this command. You could save & reload the struct and import some data later on (advantage: loading time is reduced). To show the table again just type:

```
w.showtable(w) ; %to show the table in CMD-window
```

If graphic is supported you could also visualize the table in an extra window via w.showtable2(w) .

Here, the raw-data folder contains two data-sets (i.e. the data from two animals):

BRUKER DATA									
set	SubjectId	StudNo	ExpNo	PrcNo	MRseq	protocol	sizeMB	date	file
1	20200925MG_LAERMRT_MGR000025	1	1	1	FLASH	01_1_localizer_CRP	0.393216	20-Oct-2020 15:07:20	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	10	1	SINGLEPULSE	02_6_freqAdj_SINGLEPULSE	0.004096	20-Oct-2020 15:05:32	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	11	1	DTIEpi	02_7_DTI_EPI_seg_b2500_37dir	112.0666	20-Oct-2020 15:06:12	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	11	2	DTIEpi	nan	129.7613	20-Oct-2020 15:06:59	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	13	1	STEAM	04_2_localized_shim_IC_single	0.004096	20-Oct-2020 15:07:11	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	16	1	STEAM	04_3_localized_shim_MGB	0.004096	20-Oct-2020 15:02:38	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	17	1	STEAM	04_3_STEAM_1H_MGB	0.004096	20-Oct-2020 15:05:13	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	19	1	STEAM	04_2_localized_shim_IC_single	0.004096	20-Oct-2020 15:08:29	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	2	1	FLASH	01_2_localizer_multi_slice	1.10592	20-Oct-2020 15:09:05	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	20	1	STEAM	04_2_localized_shim_IC_single	0.004096	20-Oct-2020 15:05:33	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	21	1	STEAM	04_2_localized_shim_IC_single	0.004096	20-Oct-2020 15:04:55	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	22	1	FieldMap	B0Map-ADJ_B0MAP	1.048576	20-Oct-2020 15:05:31	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	23	1	STEAM	04_2_localized_shim_IC_single	0.004096	20-Oct-2020 15:05:32	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	24	1	FLASH	01_2_localizer_multi_slice	1.10592	20-Oct-2020 15:03:47	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	25	1	RARE	03_T2_TurboRARE_CRP_MapShim	2.94912	20-Oct-2020 15:08:28	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	26	1	FieldMap	B0Map-ADJ_B0MAP	1.048576	20-Oct-2020 15:03:51	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	27	1	STEAM	04_2_localized_shim_IC_single	0.004096	20-Oct-2020 15:04:48	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	28	1	STEAM	04_2_STEAM_1H_IC_single	0.004096	20-Oct-2020 15:05:25	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	29	1	STEAM	04_3_localized_shim_MGB	0.004096	20-Oct-2020 15:03:52	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	3	1	RARE	03_T2_TurboRARE_CRP	6.5536	20-Oct-2020 15:05:38	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	31	1	STEAM	04_2_localized_shim_IC_single	0.004096	20-Oct-2020 15:05:39	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	33	1	STEAM	04_2_localized_shim_IC_single	0.004096	20-Oct-2020 15:08:23	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	35	1	STEAM	04_2_STEAM_1H_IC_single	0.004096	20-Oct-2020 15:07:33	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	4	1	DTIEpi	02_1_DTI_EPI_seg_b1000_6dir	20.64384	20-Oct-2020 15:08:11	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	4	2	DTIEpi	nan	129.7613	20-Oct-2020 15:08:24	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	5	1	FieldMap	B0Map-ADJ_B0MAP	1.048576	20-Oct-2020 15:03:53	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	6	1	SINGLEPULSE	02_2_freqAdj_SINGLEPULSE	0.004096	20-Oct-2020 15:05:00	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1	20200925MG_LAERMRT_MGR000025	1	7	1	DTIEpi	02_3_DTI_EPI_seg_b9000_13dir	41.28768	20-Oct-2020 15:08:40	/sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje

```

1 20200925MG_LAERMRT_MGR000025 1 7 2 DTIEpi nan 129.7613 20-Oct-2020 15:09:00 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1 20200925MG_LAERMRT_MGR000025 1 8 1 SINGLEPULSE 02_4_freqAdj_SINGLEPULSE 0.004096 20-Oct-2020 15:08:25 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1 20200925MG_LAERMRT_MGR000025 1 9 1 DTIEpi 02_5_DTI_EPI_seg_b100_25dir 76.67712 20-Oct-2020 15:04:13 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
1 20200925MG_LAERMRT_MGR000025 1 9 2 DTIEpi nan 129.7613 20-Oct-2020 15:04:41 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 1 1 FLASH 01_1_Localizer_CRP 0.393216 20-Oct-2020 14:57:50 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 10 1 DTIEpi 02_7_DTI_EPI_seg_b2500_37dir 112.0666 20-Oct-2020 14:56:04 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 10 2 DTIEpi nan 129.7613 20-Oct-2020 14:56:29 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 11 1 STEAM 04_2_Localized_shim_IC_single 0.004096 20-Oct-2020 14:57:36 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 12 1 STEAM 04_2_STEAM_IH_IC_single 0.004096 20-Oct-2020 14:54:36 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 13 1 STEAM 04_3_Localized_shim_MGB 0.004096 20-Oct-2020 14:57:43 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 14 1 STEAM 04_3_STEAM_IH_MGB 0.004096 20-Oct-2020 14:57:16 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 15 1 FieldMap B0Map-ADJ_B0Map 1.048576 20-Oct-2020 14:59:45 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 16 1 STEAM 04_2_Localized_shim_IC_single 0.004096 20-Oct-2020 14:54:32 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 17 1 STEAM 04_2_Localized_shim_IC_single 0.004096 20-Oct-2020 14:55:28 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 18 1 STEAM 04_2_Localized_shim_IC_single 0.004096 20-Oct-2020 14:56:48 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 19 1 STEAM 04_2_Localized_shim_IC_single 0.004096 20-Oct-2020 14:59:15 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 2 1 FLASH 01_2_Localizer_multi_slice 1.10592 20-Oct-2020 14:59:33 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 20 1 STEAM 04_2_Localized_shim_IC_single 0.004096 20-Oct-2020 14:56:38 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 21 1 STEAM 04_2_Localized_shim_IC_single 0.004096 20-Oct-2020 14:54:36 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 23 1 STEAM 04_2_Localized_shim_IC_single 0.004096 20-Oct-2020 14:55:32 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 3 1 RARE 03_T2_TurboRARE_CRP 6.5536 20-Oct-2020 14:57:06 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 4 1 DTIEpi 02_1_DTI_EPI_seg_b100_6dir 20.64384 20-Oct-2020 14:58:07 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 4 2 DTIEpi nan 129.7613 20-Oct-2020 14:58:30 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 5 1 SINGLEPULSE 02_2_freqAdj_SINGLEPULSE 0.004096 20-Oct-2020 14:54:33 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 6 1 DTIEpi 02_3_DTI_EPI_seg_b900_13dir 41.28768 20-Oct-2020 14:54:49 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 6 2 DTIEpi nan 129.7613 20-Oct-2020 14:55:18 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 7 1 SINGLEPULSE 02_4_freqAdj_SINGLEPULSE 0.004096 20-Oct-2020 14:59:22 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 8 1 DTIEpi 02_5_DTI_EPI_seg_b1600_25dir 76.67712 20-Oct-2020 14:58:47 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 8 2 DTIEpi nan 129.7613 20-Oct-2020 14:59:07 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje
2 20200925MG_LAERMRT_MGR000027 1 9 1 SINGLEPULSE 02_6_freqAdj_SINGLEPULSE 0.004096 20-Oct-2020 14:54:33 /sc-projects/sc-proj-agtiermrt/Daten-2/mri/proje

```

Here we want to import the turboRARE-image (“**03 T2 TurboRARE CRP.nii**”) and the image “**DTI EPI seg b100 6dir**”.

To visualize the filtered table run the following command (note that the w-struct is used as 1<sup>st</sup> argument):

```
w2=xbrucker2nifti(w,0,[],[], 'gui',0,'show',1,'fit',{'pro','03_T2_TurboRARE_CRP|EPI_seg_b100'});
```

the table now looks as follows:

BRUKER DATA									
set	SubjectId	StudNo	ExpNo	PrnNo	MRseq	protocol	sizeMB	date	file
1	20200925MG_LAERMRT_MGR000025	1	3	1	RARE	03_T2_TurboRARE_CRP	6.5536	20-Oct-2020 15:05:38	/sc-projects/sc-
1	20200925MG_LAERMRT_MGR000025	1	4	1	DTIEpi	02_1_DTI_EPI_seg_b100_6dir	20.64384	20-Oct-2020 15:08:11	/sc-projects/sc-
2	20200925MG_LAERMRT_MGR000027	1	3	1	RARE	03_T2_TurboRARE_CRP	6.5536	20-Oct-2020 14:57:06	/sc-projects/sc-
2	20200925MG_LAERMRT_MGR000027	1	4	1	DTIEpi	02_1_DTI_EPI_seg_b100_6dir	20.64384	20-Oct-2020 14:58:07	/sc-projects/sc-

Now let’s import these data. For this just set the ‘show’-parameter to 0:

```
w2=xbrucker2nifti(w,0,[],[], 'gui',0,'show',0,'fit',{'pro','03_T2_TurboRARE_CRP|EPI_seg_b100'});
```

## 5) VISUALIZE FILES AND FOLDERS

-type **dispf**iles to see a file-by-folder matrix in the command-window (see dispfiles for further help):  
dispfiles

The following is displayed in the CMD-window:

FILE x FOLDER			
	20200925MG_LAERMRT_MGR000025	20200925MG_LAERMRT_MGR000027	counts
02_1_DTI_EPI_seg_b100_6dir_1.nii			2/2
03_T2_TurboRARE_CRP_1.nii	+	+	2/2
counts	2/2	2/2	

Here we see that the study’s “**dat**”-folder now contains two animal-folders (“**20200925MG\_LAERMRT\_MGR000025**” and “**20200925MG\_LAERMRT\_MGR000027**”). Each Folder contains the two imported files (“**02 1 DTI EPI seg b100 6dir 1.nii**” and “**03 T2 TurboRARE CRP 1.nii**”).

## 6) SELECTION OF ANIMALS

Before renaming the files let’s first select the animals we want to process.

Here we will select all animals:

```
mdirs=antcb('getallsubjects')
```

Note that the variable “**mdirs**” contains the full paths names of the selected animals:

```
{'/sc-projects/sc-proj-agtiermrt/Daten-2/mri/projects/groeschel/dat/20200925MG_LAERMRT_MGR000025'}
{'/sc-projects/sc-proj-agtiermrt/Daten-2/mri/projects/groeschel/dat/20200925MG_LAERMRT_MGR000027'}
```

Alternative selection of all animals:

```
mdirs =antcb('selectdirs',[1:2 ])
```

```
mdirs=antcb('selectdirs','all')
```

## 7) RENAME FILES

Here we rename the file '**03 T2 TurboRARE CRP 1.nii**' to 't2.nii'. Note that the name convention of “**t2.nii**” is mandatory, because this file is used for registration to standard space. I.e., the registration relies on the name 't2.nii'!

The renaming of the file '**03 T2 TurboRARE CRP 1.nii**' is just because I hate long names.

Let’s rename '03\_T2\_TurboRARE\_CRP\_1.nii' to t2.nii':

```
xrename(0,'03_T2_TurboRARE_CRP_1.nii','t2.nii',';', 'dirs',mdirs);
and rename '02_1_DTI_EPI_seg_b100_6dir_1.nii' to 'dti_b100.nii':
xrename(0,'02_1_DTI_EPI_seg_b100_6dir_1.nii','dti_b100.nii',';', 'dirs',mdirs);
```

Here for safety reasons, we make a copy of the original file and rename the copied file. Note that copying and renaming of the copied version is defined via the colon-symbol (:); the 4<sup>th</sup> input arg). Alternatively, to rename the original files just keep the 4<sup>th</sup> arg empty.

Now, we check whether the new files exist via dispfiles:

dispfiles

FILE	x	FOLDER			
			20200925MG_LAERMRT_MGR000025	20200925MG_LAERMRT_MGR000027	counts
02_1_DTI_EPI_seg_b100_6dir_1.nii	+				2/2
03_T2_TurboRARE_CRP_1.nii	+				2/2
dti_b100.nii	+				2/2
t2.nii	+				2/2
counts		4/4		4/4	

## 8) REGISTER “t2.nii” TO TEMPLATE SPACE (STANDARD SPACE)

Registration of **“t2.nii”** to standard space is done in 4 steps: [1] initialization, [2] rough rigid registration, [3] segmentation and [4] warping. You can perform these steps (‘task’) isolated & sequentially or combined,. Note however, that task ‘2’ can be only performed when task ‘1’ has been already performed. Here the rough registration is done automatically (autoreg’, 1). We also use parallel processing across animals ('parfor',1); Please check the memory and number of cores when using parallel processing:

```
xwarp3('batch','task',[1:4 ],'autoreg',1,'parfor',1, 'mdirs',mdirs(:) );
```

Now let’s check whether the **“t2.nii”** is transformed to standard-space (new name is: **“x t2.nii”**) and the template (**“AVGT.nii”**) and atlas (**“ANO.nii”**) is back-transformed to native-space (new names: **“ix ANO.nii”** & **“ix AVGT.nii”**) via:

```
dispfiles('flt','^x.*.nii|^ix.*.nii');
```

FILE	x	FOLDER			
			20200925MG_LAERMRT_MGR000025	20200925MG_LAERMRT_MGR000027	counts
ix_ANO.nii	+				2/2
ix_AVGT.nii	+				2/2
x_t2.nii	+				2/2
counts		3/3		3/3	

Here we see that all three files were created for each data set

## 9) Extract the first 3d-volume from the 4D-vlume 'dti\_b100.nii'

Use the following command to extract the 1<sup>st</sup> volume (4<sup>th</sup> input arg: 1) of **‘dti\_b100.nii’** and save it as **‘dti\_b100\_1stIMG.nii’** for all animals (mdirs):

```
xrename(0,'dti_b100.nii','dti_b100_1stIMG.nii','1','dirs', mdirs );
```

Again, check the existence of the new files via:

```
dispfiles('flt','^dt')
```

FILE	x	FOLDER			
			20200925MG_LAERMRT_MGR000025	20200925MG_LAERMRT_MGR000027	counts
dti_b100.nii	+				2/2
dti_b100_1stIMG.nii	+				2/2
counts		2/2		2/2	

## 10) COREGISTER 'dti\_b100\_1stIMG.nii' to ‘t2.nii’

Now let’s coregister the **file ‘dti\_b100\_1stIMG.nii’** (moving image) to the **‘t2.nii’**-image (fixed image):

```
z=[];
z.TASK={ '[2]' }; ;% flag 2 (or ‘2’) signals to perform coregistration via SPM
z.targetImg1={ 't2.nii' }; ;% TARGET-IMAGE
z.sourceImg1={ 'dti_b100_1stIMG.nii' }; ;% SOURCE-IMAGE
z.sourceImgNum1=[1]; ;% IN CASE OF 4D-vol use 1st 3d-volume of SOURCE
z.applyImg1= " ; % HERE, THE TRAFO IS NOT APPLIED TO OTHER IMAGES
z.cost_fun='nmi';
z.sep=[7 2 1 0.5 0.1 0.05];
z.tol=[0.01 0.01 0.01 0.001 0.001 0.001];
```

```

z.fwhm=[4 4];
z.centering=[0];
z.reslicing=[1];                                ;% OUTPUT-IMAGE IS RESLICED TO MATCH WITH TARGET
z.interpOrder='auto';
z.prefix='r3';                                    ;% OUTPUT FILE-PREFIX
z.warping=[0]; %WARPING IS "OFF"
z.isparallel=1;                                  ;% PARALLEL PROCESSING ENABLED
xcoreg(0,z,mdirs);

```

Again, check existence of the registered file:

```
dispfiles('flt','^r3')
```

FILE	x	FOLDER
-----		
20200925MG_LAERMRT_MGR000025 20200925MG_LAERMRT_MGR000027 counts		
=====		
r3c_dti_b100_1stIMG.nii	+	2/2
r3c_t2.nii	+	2/2
counts	2/2	2/2

Here we see that the file 'r3c dti b100 1stIMG.nii' exists for each animal. The file "r3c t2.nii" is just a copy of the unchanged target-file ('t2.nii', fixed image).

## 11) TRANSFORM ANOTHER IMAGE TO STANDARD-SPACE

Now, we want to transform the image "r3c dti b100 1stIMG.nii" to standard-space (1<sup>st</sup> arg: 1) for all selected animals (mdirs), using b-spline interpolation (4<sup>th</sup> arg: 4), using the local reorientation information estimated from the rough rigid registration step (5<sup>th</sup> arg: 'local') and indicate that the input-file is located in the animal folder (6<sup>th</sup> arg: struct('source','intern')). This will create the file "x r3c dti b100 1stIMG.nii" in standard-space:

```
fis=doelastix(1,mdirs,{ 'r3c_dti_b100_1stIMG.nii'},4,'local',struct('source','intern'));
```

Let's check the existence of the file "x r3c dti b100 1stIMG.nii":

```
dispfiles('flt','x_.*')
```

FILE	x	FOLDER
-----		
20200925MG_LAERMRT_MGR000025 20200925MG_LAERMRT_MGR000027 counts		
=====		
ix_ANO.nii	+	2/2
ix_AVGT.nii	+	2/2
ix_AVGThemi.nii	+	2/2
x_r3c_dti_b100_1stIMG.nii	+	2/2
x_t2.nii	+	2/2
counts	5/5	5/5

## 12) TRANSFORM ANOTHER IMAGE TO NATIVE-SPACE

In the same way we could also transform an image from standard-space to the template space (for instance the Atlas). Here we transform the template's hemispheric mask ('AVGThemi.nii') to native space (new name: 'ix\_AVGThemi.nii'). For transformation to native space the 1<sup>st</sup> arg is -1, we use NN-interpolation to preserve numbers/hemispheric-IDs (4<sup>th</sup> arg: 0):

```
fis2=doelastix(-1,mdirs,{ 'AVGThemi.nii'},0,'local',struct('source','intern'));
```

Let's check the existence of the file 'ix\_AVGThemi.nii':

```
dispfiles('flt','ix_.*')
```

FILE	x	FOLDER
-----		
20200925MG_LAERMRT_MGR000025 20200925MG_LAERMRT_MGR000027 counts		
=====		
ix_ANO.nii	+	2/2
ix_AVGT.nii	+	2/2
ix_AVGThemi.nii	+	2/2
counts	3/3	3/3

## 13) CHECK REGISTRATION in STANDARD-SPACE - CREATE HTML-FILE

Next, we will create an HTML-file to visualize the overlay of the template "AVGT.nii" and "x t2.nii" (animal in standard-space). The 3<sup>rd</sup> arg defines the output-folder, the 4<sup>th</sup> input, defines the size of the images and the number of slices to visualize:

```
checkreghtml(mdirs,{ 'AVGT.nii','x_t2.nii'},fullfile(pwd,'checks'),struct('size',300,'slices','n20'))
```

Now you can visualize the registration with your Web-browser. The 1<sup>st</sup> image is an animated gif, showing the overlay of the 'AVGT.nii' and 'x t2.nii' images. Click onto the image to toggle between

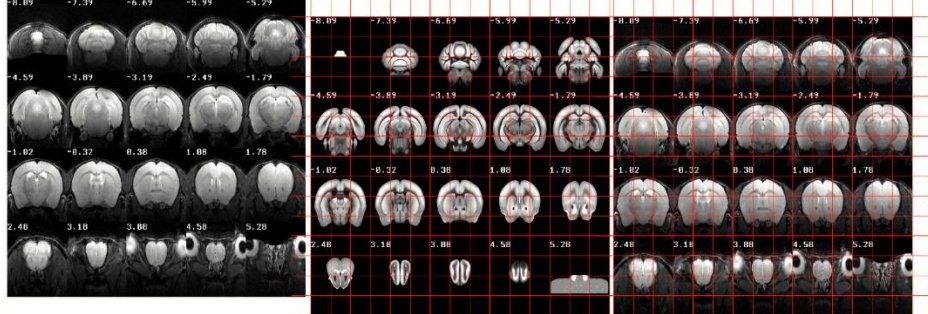


the two images, or hit 'start animation'-button to see the animated gif. The 2<sup>nd</sup> ('AVGT.nii') and 3<sup>rd</sup> image ('x\_t2.nii') shows the two images side-by-side.

[AVGT.nii-x\_t2.nii]

1] 20200925MG\_LAERMRT\_MGR000025

start animation stop animation -zoom +zoom click image to toggle images

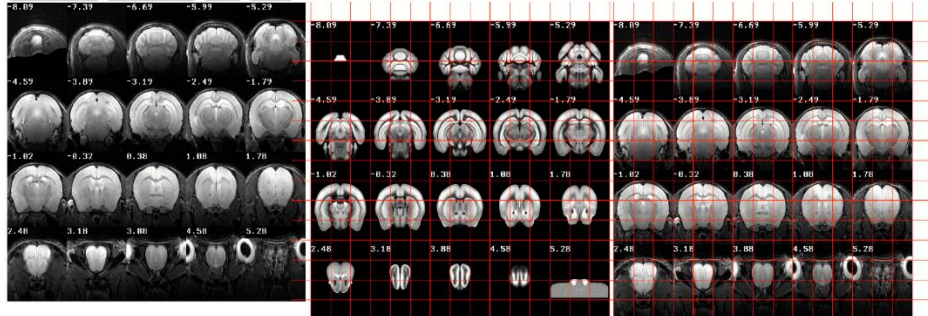


/sc-projects/sc-proj-agtlermrt/Daten-2/mri/projects/groeschel/dat/20200925MG\_LAERMRT\_MGR000025/AVGT.nii  
/sc-projects/sc-proj-agtlermrt/Daten-2/mri/projects/groeschel/dat/20200925MG\_LAERMRT\_MGR000025/x\_t2.nii

AVGT.nii; DIM: [164 212 158]  
0.07000 0 0 -5.745  
0 0.07000 0 -8.864  
0 0 0.07000 -8.520

2] 20200925MG\_LAERMRT\_MGR000027

start animation stop animation -zoom +zoom click image to toggle images



/sc-projects/sc-proj-agtlermrt/Daten-2/mri/projects/groeschel/dat/20200925MG\_LAERMRT\_MGR000027/AVGT.nii  
/sc-projects/sc-proj-agtlermrt/Daten-2/mri/projects/groeschel/dat/20200925MG\_LAERMRT\_MGR000027/x\_t2.nii

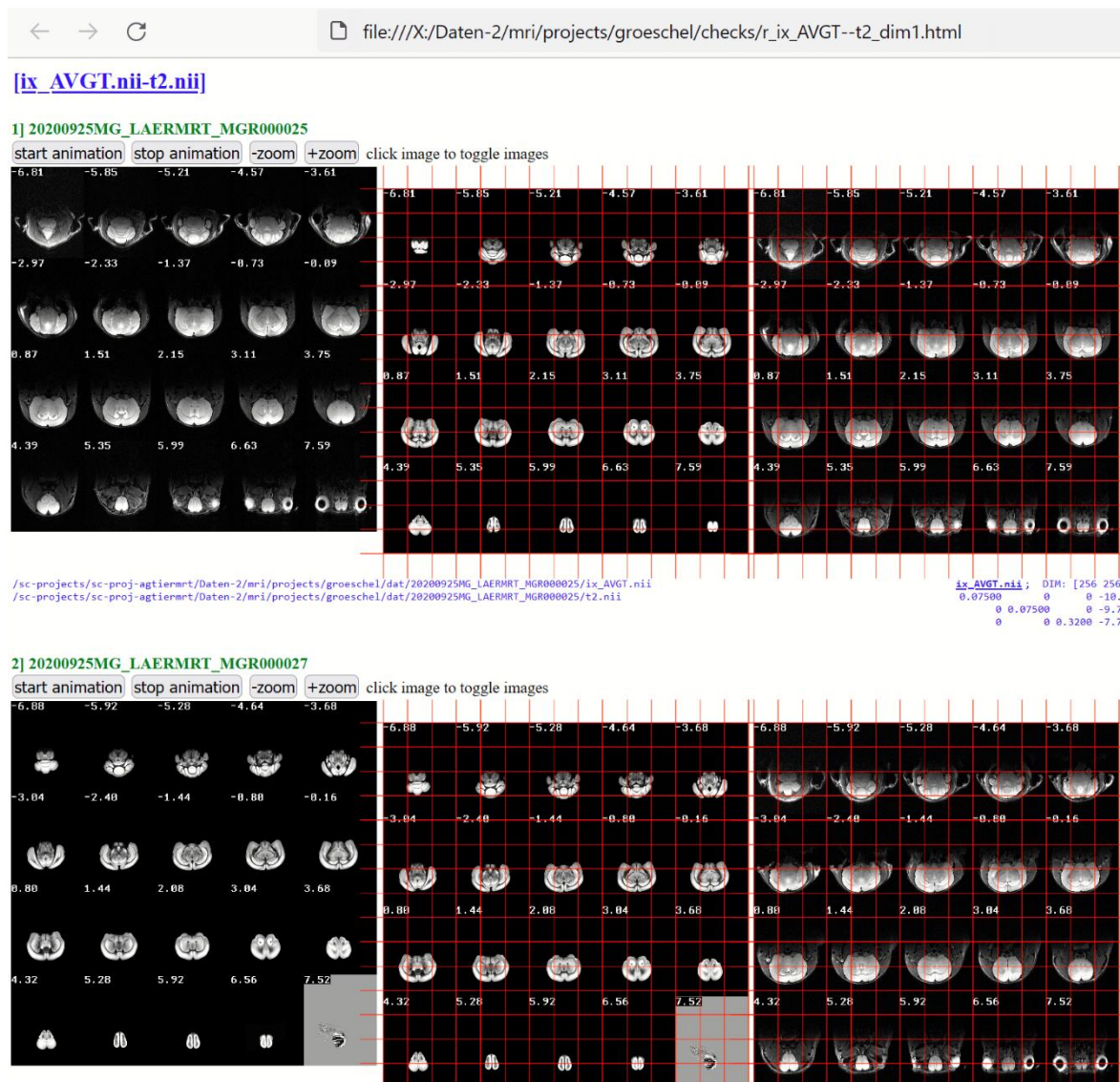
AVGT.nii; DIM: [164 212 158]  
0.07000 0 0 -5.745  
0 0.07000 0 -8.864  
0 0 0.07000 -8.520

## 14) CHECK REGISTRATION in NATIVE-SPACE - CREATE HTML-FILE

In the same, we can create an overlay of images in native-space. Here we compare the images "t2.nii" and "ix\_AVGT.nii" for all animals and save the output as HTML-file.

Note that the slicing is now done from the 1<sup>st</sup>-dimension ('dim': 1)

```
checkreghtml(mdirs,{ 'ix_AVGT.nii', 't2.nii' },fullfile(pwd,'checks'),struct('size',300,'slices','n20','dim',1))
```



## 15) Regionwise parameter-extraction

We now extract parameters such as volume, mean, median etc. for each region using the image "t2.nii" and the standard atlas (z.atlas= 'ANO.nii'). Here, from the image in native space (z.space= 'native') the parameters are aggregated over the left and right hemisphere (z.hemisphere= 'both'). The resulting Excelfile 'regwise\_t2' is stored in the 'results'-folder (subfolder of the study-folder).

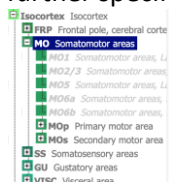
```
z=[];
z.files      = 't2.nii';      % file used for parameter extraction
z.atlas      = 'ANO.nii';     % selected atlas name, atlas has to be the standard space atlas name
z.space      = 'native';     % use images from "native" space
z.hemisphere = 'both';       % hemisphere used: [left,right or both]
z.fileNameOut = 'regwise_t2'; % <optional> name of the output-file.
xgetlabels4(0,z,mdirs);
```

The resulting excel-file contains a sheet for each parameter. Here see the volume-sheet ("vol") with region-wise volume in mm3 for each region (rows) and each animal (columns). See "info"-sheet & xgetlabels4.m for more information:



1	region	20200925MG_LAERMRT_MGR000025	20200925MG_LAERMRT_MGR000027
2	root	494.8288316	483.2800356
3	Basic cell groups and regions	439.7092504	430.7632534
4	Cerebrum	261.3599111	260.6777113
5	Cerebral cortex	209.5541287	207.7019293
6	Cortical plate	200.5649318	198.6353324
7	Isocortex	112.9841616	113.9579612
8	Frontal pole cerebral cortex	0.883799699	0.854999709
9	Frontal pole layer 1	0.131399955	0.17639994
10	Frontal pole layer 2/3	0.363599876	0.280799904
11	Frontal pole layer 5	0.313199893	0.320399891
12	Frontal pole layer 6a	0.075599974	0.077399974
13	Frontal pole layer 6b	0	0
14	Somatomotor areas	23.08859214	22.43699237
15	Somatomotor areas Layer 1	0	0
16	Somatomotor areas Layer 2/3	0	0
17	Somatomotor areas Layer 5	0	0
18	Somatomotor areas Layer 6a	0	0
19	Somatomotor areas Layer 6b	0	0
20	Primary motor area	10.50299643	10.2779965
21	Primary motor area Layer 1	1.349999541	1.274399566
22	Primary motor area Layer 2/3	3.648598759	3.639598762

Note: Zero volume entries as for the “somatomotor areas” is because the region-IDs are not defined/exist in the Allen brain Atlas (see also: <http://atlas.brain-map.org/atlas#atlas=1&plate=100960428&structure=500&x=5245.5&y=3833.5&zoom=-4&resolution=33.45&z=6>) Here we see that the layers of “somatomotor areas” are grayed, i.e. not further specified



Of course you could extract parameters from images in standard space as well:

Example: Just change the following from the above example:

z.files = 'x\_t2.nii'; i.e. the t2w-image in standard space is use  
z.space = 'standard'; this indicated that the image is in standard space  
z.fileNameOut = 'regwise\_x\_t2' just another name for the resulting Excelfile:

#### EXAMPLE: PARAMTER EXTRACTION FROM STANDARD SPACE

```
z=[];
z.files = 'x_t2.nii'; % file used for parameter extraction
z.atlas = 'ANO.nii'; % selected atlas name, atlas has to be the standard space atlas name
z.space = 'standard'; % use images from "standard" space
z.hemisphere = 'both'; % hemisphere used: [left,right or both]
z.fileNameOut = 'regwise_x_t2' ; % <optional> name of the output-file.
xgetlabels4(0,z,mdirs);
```

...End of this tutorial. More progress is hopefully coming soon.