

Using the GEOquery package

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January 3, 2008

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1 Overview of GEO

The NCBI Gene Expression Omnibus (GEO) serves as a public repository for a wide range of high-throughput experimental data. These data include single and dual channel microarray-based experiments measuring mRNA, genomic DNA, and protein abundance, as well as non-array techniques such as serial analysis of gene expression (SAGE), and mass spectrometry proteomic data. Currently, nearly 200,000 samples and over 4000 different platforms are represented in GEO!

At the most basic level of organization of GEO, there are four basic entity types. The first three (Sample, Platform, and Series) are supplied by users; the fourth, the dataset, is compiled and curated by GEO staff from the user-submitted data.¹

1.1 Platforms

A Platform record describes the list of elements on the array (e.g., cDNAs, oligonucleotide probesets, ORFs, antibodies) or the list of elements that may be detected and quantified in that experiment (e.g., SAGE tags, peptides). Each Platform record is assigned a unique and stable GEO accession number (GPLxxx). A Platform may reference many Samples that have been submitted by multiple submitters.

1.2 Samples

A Sample record describes the conditions under which an individual Sample was handled, the manipulations it underwent, and the abundance measurement of each element derived from it. Each Sample record is assigned a unique and stable GEO accession number (GSMxxx). A Sample entity must reference only one Platform and may be included in multiple Series.

1.3 Series

A Series record defines a set of related Samples considered to be part of a group, how the Samples are related, and if and how they are ordered. A Series provides a focal point and description of the experiment as a whole. Series records may also contain tables describing extracted data, summary conclusions, or analyses. Each Series record is assigned a unique and stable GEO accession number (GSExxx). Series records are available in a couple of formats which are handled by GEOquery independently. The smaller and new GSEMatrix files are quite fast to parse; a simple flag is used by GEOquery to choose to use GSEMatrix files (see below).

1.4 Datasets

GEO DataSets (GDSxxx) are curated sets of GEO Sample data. A GDS record represents a collection of biologically and statistically comparable GEO Samples and forms the basis

¹See <http://www.ncbi.nih.gov/geo> for more information

of GEO's suite of data display and analysis tools. Samples within a GDS refer to the same Platform, that is, they share a common set of probe elements. Value measurements for each Sample within a GDS are assumed to be calculated in an equivalent manner, that is, considerations such as background processing and normalization are consistent across the dataset. Information reflecting experimental design is provided through GDS subsets.

2 Getting Started using GEOquery

Getting data from GEO is really quite easy. There is only one command that is needed, `getGEO`. This one function interprets its input to determine how to get the data from GEO and then parse the data into useful R data structures. Usage is quite simple:

```
> library(GEOquery)
```

This loads the GEOquery library.

```
> gds <- getGEO("GDS1")
```

File stored at:

```
/tmp/RtmpVSFUaf/GDS1.soft.gz
```

```
parsing geodata
```

```
parsing subsets
```

```
ready to return
```

Now, `gds` contains the R data structure (of class *GDS*) that represents the GDS1 entry from GEO. You'll note that the filename used to store the download was output to the screen (but not saved anywhere) for later use to a call to `getGEO(filename=...)`.

We can do the same with any other GEO accession, such as GSM3, a GEO sample.

```
> gsm <- getGEO("GSM3")
```

File stored at:

```
/tmp/RtmpVSFUaf/GSM3.soft
```

3 GEOquery Data Structures

The GEOquery data structures really come in two forms. The first, comprising *GDS*, *GPL*, and *GSM* all behave similarly and accessors have similar effects on each. The fourth GEOquery data structure, *GSE* is a composite data type made up of a combination of *GSM* and *GPL* objects. I will explain the first three together first.

3.1 The GDS, GSM, and GPL classes

Each of these classes is comprised of a metadata header (taken nearly verbatim from the SOFT format header) and a GEODataTable. The GEODataTable has two simple parts, a Columns part which describes the column headers on the Table part. There is also a *show* method for each class. For example, using the gsm from above:

```
> Meta(gsm)

$channel_count
[1] "1"

$contact_address
[1] "6 Center Drive"

$contact_city
[1] "Bethesda"

$contact_country
[1] "USA"

$contact_department
[1] "LCDB"

$contact_email
[1] "oliver@helix.nih.gov"

$contact_fax
[1] "301-496-5239"

$contact_institute
[1] "NIDDK, NIH"

$contact_name
[1] "Brian,,Oliver"

$contact_phone
[1] "301-496-5495"

$contact_state
[1] "MD"

$contact_web_link
[1] "http://www.niddk.nih.gov/intram/people/boliver.htm"
```

\$`contact_zip/postal_code`

[1] "20892"

\$data_row_count

[1] "3456"

\$description

[1] "Testis dissected from adult (12-24 hours post-eclosion) *Drosophila melanogaster* of

[2] "Keywords = gonad, male, sex"

\$geo_accession

[1] "GSM3"

\$last_update_date

[1] "May 27 2005"

\$molecule_ch1

[1] "total RNA"

\$organism_ch1

[1] "*Drosophila melanogaster*"

\$platform_id

[1] "GPL5"

\$series_id

[1] "GSE462"

\$source_name_ch1

[1] "y w[67c1]/Y testis"

\$status

[1] "Public on Oct 18 2000"

\$submission_date

[1] "Oct 18 2000"

\$supplementary_file

[1] "NONE"

\$title

```
[1] "testis a"
```

```
$type
```

```
[1] "RNA"
```

```
> Table(gsm)[1:5, ]
```

	ID_REF	SIGNAL_RAW	BKD_FORM	NORM_FORM	BKD_RAW	NORM_VALUE	CONST	VALUE
1	1	138392.6	no	no	101113.8	395070.1	39542	76820.87
2	2	100973.5	no	no	101113.8	395070.1	39542	39401.71
3	3	118994.0	no	no	101113.8	395070.1	39542	57422.25
4	4	108126.1	yes	no	101113.8	395070.1	39542	46554.27
5	5	293362.1	no	no	101113.8	395070.1	39542	231790.33

```
> Columns(gsm)
```

	Column	Description
1	ID_REF	
2	SIGNAL_RAW	raw signal
3	BKD_FORM	
4	NORM_FORM	
5	BKD_RAW	raw background as taken in four quarters of microarray
6	NORM_VALUE	normalization value
7	CONST	constant value
8	VALUE	

The *GPL* behaves exactly as the *GSM* class. However, the *GDS* has a bit more information associated with the *Columns* method:

```
> Columns(gds)
```

	sample	gender	tissue
1	GSM3	male	testis
2	GSM4	male	testis
3	GSM5	male	gonadectomized male
4	GSM6	male	gonadectomized male
5	GSM7	female	ovary
6	GSM8	female	ovary
7	GSM9	female	gonadectomized female
8	GSM10	female	gonadectomized female

	description
1	Value for GSM3: testis a; src: y w[67c1]/Y testis
2	Value for GSM4: testis b; src: y w[67c1]/Y testis
3	Value for GSM5: male a; src: y w[67c1]/Y male

```

4      Value for GSM6: male b; src: y w[67c1]/Y
5      Value for GSM7: ovary a; src: y w[67c1] ovary
6      Value for GSM8: ovary b; src: y w[67c1] ovary
7      Value for GSM9: female a; src: y w[67c1] female
8      Value for GSM10: female b; src: y w[67c1] female

```

3.2 The GSE class

The *GSE* is the most confusing of the GEO entities. A GSE entry can represent an arbitrary number of samples run on an arbitrary number of platforms. The *GSE* has a metadata section, just like the other classes. However, it doesn't have a *GEODDataTable*. Instead, it contains two lists, accessible using *GPLList* and *GSMList*, that are each lists of *GPL* and *GSM* objects. To show an example:

```
> gse <- getGEO("GSE462", GSEMatrix = FALSE)
```

```
File stored at:
```

```
/tmp/RtmpVSFUaf/GSE462.soft.gz
```

```
Parsing....
```

```
^PLATFORM = GPL5
```

```
^SAMPLE = GSM3
```

```
^SAMPLE = GSM4
```

```
^SAMPLE = GSM5
```

```
^SAMPLE = GSM6
```

```
^SAMPLE = GSM7
```

```
^SAMPLE = GSM8
```

```
^SAMPLE = GSM9
```

```
> Meta(gse)
```

```
$contact_address
```

```
[1] "6 Center Drive"
```

```
$contact_city
```

```
[1] "Bethesda"
```

```
$contact_country
```

```
[1] "USA"
```

```
$contact_department
```

```
[1] "LCDB"
```

```
$contact_email
```

```
[1] "oliver@helix.nih.gov"
```

```

$contact_fax
[1] "301-496-5239"

$contact_institute
[1] "NIDDK, NIH"

$contact_name
[1] "Brian,,Oliver"

$contact_phone
[1] "301-496-5495"

$contact_state
[1] "MD"

$contact_web_link
[1] "http://www.niddk.nih.gov/intram/people/boliver.htm"

$`contact_zip/postal_code`
[1] "20892"

$contributor
[1] "Justen,,Andrews"      "Gerard,G,Bouffard" "Chris,,Cheadle"
[4] "Jining,,LÃij"         "Kevin,G,Becker"    "Brian,,Oliver"

$geo_accession
[1] "GSE462"

$last_update_date
[1] "Oct 28 2005"

$platform_id
[1] "GPL5"

$pubmed_id
[1] "11116097"

$sample_id
[1] "GSM10" "GSM3"  "GSM4"  "GSM5"  "GSM6"  "GSM7"  "GSM8"  "GSM9"

$status

```



```
[1] "Public on Jul 16 2003"
```

```
$submission_date
```

```
[1] "Jun 25 2003"
```

```
$summary
```

```
[1] "Identification and annotation of all the genes in the sequenced Drosophila genome i
```

```
$title
```

```
[1] "Analysis of transcription in the Drosophila melanogaster testis"
```

```
$type
```

```
[1] "other"
```

```
> names(GSMList(gse))
```

```
[1] "GSM10" "GSM3"  "GSM4"  "GSM5"  "GSM6"  "GSM7"  "GSM8"  "GSM9"
```

```
> GSMList(gse)[[1]]
```

```
An object of class "GSM"
```

```
channel_count
```

```
[1] "1"
```

```
contact_address
```

```
[1] "6 Center Drive"
```

```
contact_city
```

```
[1] "Bethesda"
```

```
contact_country
```

```
[1] "USA"
```

```
contact_department
```

```
[1] "LCDB"
```

```
contact_email
```

```
[1] "oliver@helix.nih.gov"
```

```
contact_fax
```

```
[1] "301-496-5239"
```

```
contact_institute
```

```
[1] "NIDDK, NIH"
```

```
contact_name
```

```
[1] "Brian,,Oliver"
```

```
contact_phone
```

```
[1] "301-496-5495"
```

```
contact_state
```

```
[1] "MD"
```

```

contact_web_link
[1] "http://www.niddk.nih.gov/intram/people/boliver.htm"
contact_zip/postal_code
[1] "20892"
data_row_count
[1] "3456"
description
[1] "Whole adult male minus (12-24 hours post-eclosion) Drosophila melanogaster of the g
geo_accession
[1] "GSM10"
last_update_date
[1] "Mar 09 2006"
molecule_ch1
[1] "total RNA"
organism_ch1
[1] "Drosophila melanogaster"
platform_id
[1] "GPL5"
series_id
[1] "GSE462"
source_name_ch1
[1] "y w[67c1] female"
status
[1] "Public on Oct 18 2000"
submission_date
[1] "Oct 18 2000"
title
[1] "female b"
type
[1] "RNA"
An object of class "GEODataTable"
***** Column Descriptions *****
      Column      Description
1      ID_REF
2 SIGNAL_RAW      raw signal
3      BKD_FORM
4      NORM_FORM
5      BKD_RAW      raw background
6 NORM_VALUE normalization value
7      CONST      constant value
8      VALUE
***** Data Table *****

```

	ID_REF	SIGNAL_RAW	BKD_FORM	NORM_FORM	BKD_RAW	NORM_VALUE	CONST	VALUE
1	1	4486.49	0	0	3379.579	23337.54	39542	55845.45
2	2	3482.51	0	0	3379.579	23337.54	39542	41058.05
3	3	3812.39	0	0	3379.579	23337.54	39542	45916.78
4	4	3257.56	1	0	3379.579	23337.54	39542	37744.81
5	5	5436.91	0	0	3379.579	23337.54	39542	69843.97

3451 more rows ...

```
> names(GPLList(gse))
```

```
[1] "GPL5"
```

See below for an additional, preferred method of obtaining GSE information.

4 Converting to BioConductor ExpressionSets and limma MALists

GEO datasets are (unlike some of the other GEO entities), quite similar to the *limma* data structure *MAList* and to the *Biobase* data structure *ExpressionSet*. Therefore, there are two functions, `GDS2MA` and `GDS2eSet` that accomplish that task.

4.1 Getting GSE Series Matrix files as an ExpressionSet

GEO Series are collections of related experiments. In addition to being available as SOFT format files, which are quite large, NCBI GEO has prepared a simpler format file based on tab-delimited text. The `getGEO` function can handle this format and will parse very large GSEs quite quickly. The data structure returned from this parsing is a list of *ExpressionSets*. As an example, we download and parse GSE2553. Unfortunately, *GSEMatrix* parsing is not currently available on windows.

```
> if (.Platform$OS.type != "windows") {
+   gse2553 <- getGEO("GSE2553", GSEMatrix = TRUE)
+   show(gse2553)
+   show(pData(phenoData(gse2553[[1]]))[1:5, c(1, 6, 8)])
+ }
```

```
Found 1 file(s)
GSE2553_series_matrix.txt.gz
File stored at:
/tmp/RtmpVSFUaf/GPL1977.soft
$GSE2553_series_matrix.txt.gz
ExpressionSet (storageMode: lockedEnvironment)
assayData: 12600 features, 181 samples
```

```

element names: exprs
phenoData
  sampleNames: GSM48681, GSM48682, ..., GSM48861 (181 total)
  varLabels and varMetadata description:
    title: NA
    geo_accession: NA
    ....: ...
    data_row_count: NA
    (27 total)
featureData
  featureNames: 1, 2, ..., 12600 (12600 total)
  fvarLabels and fvarMetadata description:
    ID: NA
    PenAt: NA
    ....: ...
    Chimeric_Cluster_IDs: NA
    (13 total)
  additional fvarMetadata: Description, labelDescription
experimentData: use 'experimentData(object)'
Annotation: GPL1977

```

		title	type
GSM48681	Patient sample ST18, Dermatofibrosarcoma	RNA	
GSM48682	Patient sample ST410, Ewing Sarcoma	RNA	
GSM48683	Patient sample ST130, Sarcoma, NOS	RNA	
GSM48684	Patient sample ST293, Malignant Peripheral Nerve Sheath Tumor	RNA	
GSM48685	Patient sample ST367, Liposarcoma	RNA	
	source_name_ch1		
GSM48681	Dermatofibrosarcoma		
GSM48682	Ewing Sarcoma		
GSM48683	Sarcoma, NOS		
GSM48684	Malignant Peripheral Nerve Sheath Tumor		
GSM48685	Liposarcoma		

4.2 Converting GDS to an ExpressionSet

Taking our `gds` object from above, we can simply do:

```
> eset <- GDS2eSet(gds, do.log2 = TRUE)
```

File stored at:

```
/tmp/RtmpVSFUaf/GPL5.soft
```

Now, `eset` is an *ExpressionSet* that contains the same information as in the GEO dataset, including the sample information, which we can see here:

```
> eset
```

```
ExpressionSet (storageMode: lockedEnvironment)
assayData: 3456 features, 8 samples
  element names: exprs
phenoData
  sampleNames: GSM3, GSM4, ..., GSM10 (8 total)
  varLabels and varMetadata description:
    sample: NA
    gender: NA
    tissue: NA
    description: NA
featureData
  featureNames: 1, 2, ..., 3456 (3456 total)
  fvarLabels and fvarMetadata description:
    ID:
      GB_ACC: GenBank accession number LINK_PRE:"http://www.ncbi.nlm.nih.gov/entre
z/query.fcgi?cmd=Search&db=Nucleotide&term="
      ...: ...
      SPOT_QC: spot quality control
      (13 total)
    additional fvarMetadata: labelDescription
experimentData: use 'experimentData(object)'
pubMedIds: 11116097
Annotation:
```

```
> pData(eset)
```

	sample	gender	tissue	
GSM3	GSM3	male	testis	
GSM4	GSM4	male	testis	
GSM5	GSM5	male	gonadectomized male	
GSM6	GSM6	male	gonadectomized male	
GSM7	GSM7	female	ovary	
GSM8	GSM8	female	ovary	
GSM9	GSM9	female	gonadectomized female	
GSM10	GSM10	female	gonadectomized female	
				description
GSM3	Value for GSM3: testis a; src: y w[67c1]/Y testis			
GSM4	Value for GSM4: testis b; src: y w[67c1]/Y testis			

```

GSM5      Value for GSM5: male a; src: y w[67c1]/Y male
GSM6      Value for GSM6: male b; src: y w[67c1]/Y
GSM7      Value for GSM7: ovary a; src: y w[67c1] ovary
GSM8      Value for GSM8: ovary b; src: y w[67c1] ovary
GSM9      Value for GSM9: female a; src: y w[67c1] female
GSM10     Value for GSM10: female b; src: y w[67c1] female

```

4.3 Converting GDS to an MAList

No annotation information (called platform information by GEO) was retrieved from because *ExpressionSet* does not contain slots for gene information, typically. However, it is easy to obtain this information. First, we need to know what platform this GDS used. Then, another call to `getGEO` will get us what we need.

```
> Meta(gds)$platform
```

```
[1] "GPL5"
```

```
> gpl <- getGEO("GPL5")
```

File stored at:

```
/tmp/RtmpVSFUaf/GPL5.soft
```

So, `gpl` now contains the information for GPL5 from GEO. Unlike *ExpressionSet*, the limma *MAList* does store gene annotation information, so we can use our newly created `gpl` of class *GPL* in a call to `GDS2MA` like so:

```
> MA <- GDS2MA(gds, GPL = gpl)
```

```
> MA
```

An object of class "MAList"

\$M

	GSM3	GSM4	GSM5	GSM6	GSM7	GSM8	GSM9	GSM10
[1,]	76820.87	71715.76	51430.49	139715.76	45027.85	69984.33	38569.01	55845.45
[2,]	39401.71	NA	37746.64	91150.39	29691.45	36329.52	30363.85	41058.05
[3,]	57422.25	18338.46	37134.59	75928.14	34181.67	42713.88	32090.47	45916.78
[4,]	46554.27	10928.63	34145.17	74550.27	28498.81	28617.40	33207.63	37744.81
[5,]	231790.33	341779.05	77703.83	99999.62	61151.98	65974.36	60665.36	69843.97

3451 more rows ...

\$A

NULL

\$targets

	sample	gender	tissue
1	GSM3	male	testis
2	GSM4	male	testis
3	GSM5	male	gonadectomized male
4	GSM6	male	gonadectomized male
5	GSM7	female	ovary
6	GSM8	female	ovary
7	GSM9	female	gonadectomized female
8	GSM10	female	gonadectomized female

	description
1	Value for GSM3: testis a; src: y w[67c1]/Y testis
2	Value for GSM4: testis b; src: y w[67c1]/Y testis
3	Value for GSM5: male a; src: y w[67c1]/Y male
4	Value for GSM6: male b; src: y w[67c1]/Y
5	Value for GSM7: ovary a; src: y w[67c1] ovary
6	Value for GSM8: ovary b; src: y w[67c1] ovary
7	Value for GSM9: female a; src: y w[67c1] female
8	Value for GSM10: female b; src: y w[67c1] female

\$genes

	ID	GB_ACC	BSCC_ID	CLONE_ID	SUB.ARRAY	DUPLICATE	ROW	COLUMN	PCR_QC	SPOT_ID
1	1	AI944549	bs03g07	FBgn0033989		1	a	1	passed	<NA>
2	2	AI944695	bs04c11	FBgn0032821		1	a	1	passed	<NA>
3	3	AI944741	bs04h01	FBgn0034374		1	a	1	passed	<NA>
4	4	AI944801	bs05f04	FBgn0039421		1	a	1	failed	<NA>
5	5	AI945043	bs08c11	FBgn0045370		1	a	1	passed	<NA>

1

2

3

4 gi|4505995|ref|NP_002697.1|PPPM1B| protein phosphatase 1B (formerly 2C), magnesium-dep

5

E_VAL SPOT_QC

1	2e-08	44364
2	<NA>	16957
3	<NA>	17896
4	1e-25	16363
5	<NA>	83502

3451 more rows ...

\$notes

[[1]]

```
[1] "able_begin" "able_end"
```

```
$channel_count
```

```
[1] "1"
```

```
$description
```

```
[1] "Adult testis gene expression profile and gene discovery. Examines testis, whole mal"
```

```
$feature_count
```

```
[1] "3456"
```

```
$order
```

```
[1] "none"
```

```
$platform
```

```
[1] "GPL5"
```

```
$platform_organism
```

```
[1] "Drosophila melanogaster"
```

```
$platform_technology_type
```

```
[1] "spotted DNA/cDNA"
```

```
$pubmed_id
```

```
[1] "11116097"
```

```
$reference_series
```

```
[1] "GSE462"
```

```
$sample_count
```

```
[1] "8"
```

```
$sample_organism
```

```
[1] "Drosophila melanogaster"
```

```
$sample_type
```

```
[1] "RNA"
```

```
$title
```

```
[1] "Testis gene expression profile"
```

```
$type
```



```
[1] "gene expression array-based"
```

```
$update_date
```

```
[1] "Aug 17 2006"
```

```
$value_type
```

```
[1] "count"
```

Now, `MA` is of class *MAList* and contains not only the data, but the sample information and gene information associated with GDS1.

4.4 Converting GSE to an `exprSet`

First, make sure that using the method described above in the section “Getting GSE Series Matrix files as an ExpressionSet” for using GSE Series Matrix files is not sufficient for the task, as it is much faster and simpler. If it is not (i.e., other columns from each GSM are needed), then this method will be needed.

Converting a *GSE* object to an *exprSet* object currently takes a bit of R data manipulation due to the varied data that can be stored in a *GSE* and the underlying *GSM* and *GPL* objects. However, using a simple example will hopefully be illustrative of the technique.

First, we need to make sure that all of the *GSMs* are from the same platform:

```
> gsmplatforms <- lapply(GSMList(gse), function(x) {  
+   Meta(x)$platform  
+ })  
> gsmplatforms
```

```
$GSM10
```

```
[1] "GPL5"
```

```
$GSM3
```

```
[1] "GPL5"
```

```
$GSM4
```

```
[1] "GPL5"
```

```
$GSM5
```

```
[1] "GPL5"
```

```
$GSM6
```

```
[1] "GPL5"
```

```
$GSM7
```

```
[1] "GPL5"
```

```
$GSM8
```

```
[1] "GPL5"
```

```
$GSM9
```

```
[1] "GPL5"
```

Indeed, they all used GPL5 as their platform (which we could have determined by looking at the GPLList for `gse`, which shows only one GPL for this particular GSE.). So, now we would like to know what column represents the data that we would like to extract. Looking at the first few rows of the Table of a single GSM will likely give us an idea (and by the way, GEO uses a convention that the column that contains the single “measurement” for each array is called the “VALUE” column, which we could use if we don’t know what other column is most relevant).

```
> Table(GSMList(gse)[[1]])[1:5, ]
```

	ID_REF	SIGNAL_RAW	BKD_FORM	NORM_FORM	BKD_RAW	NORM_VALUE	CONST	VALUE
1	1	4486.49	0	0	3379.579	23337.54	39542	55845.45
2	2	3482.51	0	0	3379.579	23337.54	39542	41058.05
3	3	3812.39	0	0	3379.579	23337.54	39542	45916.78
4	4	3257.56	1	0	3379.579	23337.54	39542	37744.81
5	5	5436.91	0	0	3379.579	23337.54	39542	69843.97

```
> Columns(GSMList(gse)[[1]])[1:5, ]
```

	Column	Description
1	ID_REF	
2	SIGNAL_RAW	raw signal
3	BKD_FORM	
4	NORM_FORM	
5	BKD_RAW	raw background

We will indeed use the “VALUE” column. We then want to make a matrix of these values like so:

```
> probesets <- Table(GPLList(gse)[[1]])$ID
> data.matrix <- log2(do.call("cbind", lapply(GSMList(gse), function(x) {
+   tab <- Table(x)
+   mymatch <- match(probesets, tab$ID_REF)
+   return(tab$VALUE[mymatch])
+ })))
> data.matrix[1:5, ]
```

	GSM10	GSM3	GSM4	GSM5	GSM6	GSM7	GSM8	GSM9
[1,]	15.76915	16.22921	16.13000	15.65034	17.09214	15.45853	16.09474	15.23515
[2,]	15.32538	15.26597	NaN	15.20406	16.47596	14.85776	15.14885	14.89007
[3,]	15.48673	15.80932	14.16259	15.18048	16.21235	15.06094	15.38242	14.96986
[4,]	15.20399	15.50663	13.41582	15.05939	16.18593	14.79861	14.80460	15.01923
[5,]	16.09185	17.82246	18.38270	16.24570	16.60964	15.90011	16.00962	15.88859

Note that we do a “match” to make sure that the values and the platform information are in the same order. Finally, to make the *ExpressionSet* object:

```
> require(Biobase)
> rownames(data.matrix) <- probesets
> colnames(data.matrix) <- names(GSMList(gse))
> pdata <- data.frame(samples = names(GSMList(gse)))
> rownames(pdata) <- names(GSMList(gse))
> pheno <- as(pdata, "AnnotatedDataFrame")
> eset2 <- new("ExpressionSet", exprs = data.matrix, phenoData = pheno)
> eset2
```

```
ExpressionSet (storageMode: lockedEnvironment)
assayData: 3456 features, 8 samples
  element names: exprs
phenoData
  sampleNames: GSM10, GSM3, ..., GSM9 (8 total)
  varLabels and varMetadata description:
    samples: NA
featureData
  featureNames: 1, 2, ..., 3456 (3456 total)
  fvarLabels and fvarMetadata description: none
experimentData: use 'experimentData(object)'
Annotation:
```

So, using a combination of `lapply` on the `GSMList`, one can extract as many columns of interest as necessary to build the data structure of choice. Because the GSM data from the GEO website are fully downloaded and included in the *GSE* object, one can extract foreground and background as well as quality for two-channel arrays, for example. Getting array annotation is also a bit more complicated, but by replacing “platform” in the `lapply` call to get platform information for each array, one can get other information associated with each array. Future work with this package will likely focus on better tools for manipulating *GSE* data.

5 Conclusion

The `GEOquery` package provides a bridge to the vast array resources contained in the NCBI GEO repositories. By maintaining the full richness of the GEO data rather than focusing on

getting only the “numbers”, it is possible to integrate GEO data into current Bioconductor data structures and to perform analyses on that data quite quickly and easily. These tools will hopefully open GEO data more fully to the array community at large.

6 sessionInfo

- R version 2.7.0 Under development (unstable) (2007-11-28 r43553), x86_64-unknown-linux-gnu
- Locale: LC_CTYPE=en_US.UTF-8;LC_NUMERIC=C;LC_TIME=en_US.UTF-8;LC_COLLATE=en_US.UTF-8;LC_MONETARY=en_US.UTF-8;LC_MESSAGES=en_US.UTF-8;LC_PAPER=en_US.UTF-8;LC_NAME=C;LC_ADDRESS=C;LC_IDENTIFICATION=C
- Base packages: base, datasets, graphics, grDevices, methods, stats, tools, utils
- Other packages: Biobase 1.17.6, GEOquery 2.3.0, limma 2.13.1, RCurl 0.8-3