Set 17 - Multiple comparisons

STAT 401 (Engineering) - Iowa State University

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Mice diet effect on lifetimes

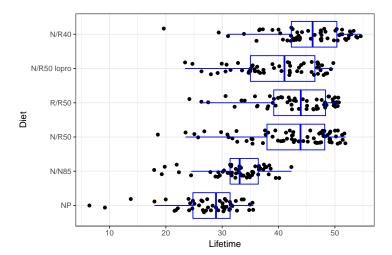
Female mice were randomly assigned to six treatment groups to investigate whether restricting dietary intake increases life expectancy. Diet treatments were:

- NP mice ate unlimited amount of nonpurified, standard diet
- N/N85 mice fed normally before and after weaning. After weaning, ration was controlled at 85 kcal/wk
- N/R50 normal diet before weaning and reduced calorie diet (50 kcal/wk) after weaning
- R/R50 reduced calorie diet of 50 kcal/wk both before and after weaning
- N/R50 lopro normal diet before weaning, restricted diet (50 kcal/wk) after weaning and dietary protein content decreased with advancing age
- N/R40 normal diet before weaning and reduced diet (40 Kcal/wk) after weaning.

Exploratory analysis

```
library("Sleuth3")
# head(case0501)
summary(case0501)
   Lifetime
                  Diet
Min. : 6.4 N/N85:57
1st Qu.:31.8 N/R40:60
Median:39.5
              N/R50:71
     :38.8
Mean
              NP :49
3rd Qu.:46.9
              R/R50:56
Max. :54.6
              lopro:56
case0501 <- case0501 %>%
 mutate(Diet = factor(Diet, c("NP", "N/N85", "N/R50", "R/R50", "lopro", "N/R40")),
        Diet = recode(Diet, lopro = "N/R50 lopro"))
case0501 %% group_by(Diet) %>% summarize(n=n(), mean = mean(Lifetime), sd = sd(Lifetime))
# A tibble: 6 4
        Diet
                       mean
                                  sd
      <fctr> <int>
                      <dbl>
                              <dbl>
              49 27.40204 6.133701
      N/N85
             57 32.69123 5.125297
3
      N/R50
             71 42.29718 7.768195
       R/R50 56 42.88571 6.683152
5 N/R50 lopro 56 39.68571 6.991695
6
       N/R40
                60 45.11667 6.703406
```

```
ggplot(case0501, aes(x=Diet, y=Lifetime)) +
geom_jitter(width=0.2) +
geom_boxplot(fill=NA, color='blue', outlier.color = NA) +
coord_flip() +
theme_bw()
```



Are the data compatible with a common mean?

Let Y_{ij} represent the lifetime of mouse j in diet i for $i=1,\ldots,I$ and $j=1,\ldots,n_i$. Assume $Y_{ij} \overset{ind}{\sim} N(\mu_i,\sigma^2)$ and calculate a pvalue for $H_0: \mu_i = \mu$ for all i.

```
bartlett.test(Lifetime ~ Diet, data = case0501)
Bartlett test of homogeneity of variances
data: Lifetime by Diet
Bartlett's K-squared = 10.996, df = 5, p-value = 0.05146
oneway.test(Lifetime ~ Diet, data = case0501, var.equal = TRUE)
One-way analysis of means
data: Lifetime and Diet
F = 57.104, num df = 5, denom df = 343, p-value < 2.2e-16
oneway.test(Lifetime ~ Diet, data = case0501, var.equal = FALSE)
One-way analysis of means (not assuming equal variances)
data: Lifetime and Diet
F = 64.726, num df = 5.00, denom df = 157.84, p-value < 2.2e-16
```

Statistical testing errors

Definition

A type I error occurs when a true null hypothesis is rejected.

Definition

A type II error occurs when a false null hypothesis is not rejected. Power is one minus the type II error probability.

We set our significance level a to control the type I error probability. If we set a=0.05, then we will incorrectly reject a true null hypothesis 5% of the time.

Statistical testing errors

	Truth	
Decision	H_0 true	H_0 false
	Type I error	Correct (power)
H_0 true	Correct	Type II error

Definition

The familywise error rate is the probability of rejecting at least one true null hypothesis.

Type I error for all pairwise comparisons of J groups

How many combinations when choosing 2 items out of J?

$$\binom{J}{2} = \frac{J!}{2!(J-2)!}.$$

If J=6, then there are 15 different comparison of means. If we set a=0.05 as our significance level, then individually each test will only incorrectly reject 5% of the time.

If we have 15 tests and use a=0.05, what is the familywise error rate?

$$1 - (1 - 0.05)^{15} = 1 - (0.95)^{15} = 1 - 0.46 = 0.54$$

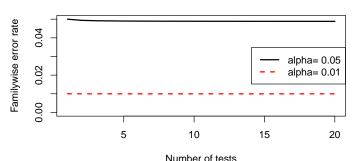
So there is a greater than 50% probability of falsely rejecting at least one true null hypothesis!

Bonferroni correction

Definition

If we do m tests and want the familywise error rate to be a, the Bonferroni correction uses a/m for each individual test. The familywise error rate, for independent tests, is $1-(1-a/m)^m$.

Bonferroni familywise error rate



Pairwise comparisons

If we want to consider all pairwise comparisons of the average lifetimes on the 6 diets, we have 15 tests. In order to maintain a familywise error rate of 0.05, we need a significance level of 0.05/15 = 0.0033333.

Pairwise comparisons

If we want to consider all pairwise comparisons of the average lifetimes on the 6 diets, we have 15 tests. Alternatively, you can let R do the adjusting for you, but now you need to compare with the original significance level a.

Comments on the Bonferroni correction

The Bonferroni correction can be used in any situation. In particular, it can be used on unadjusted pvalues reported in an article that has many tests by comparing their pvalues to a/m where m is the number of tests they perform.

The Bonferroni correction is (in general) the **most** conservative multiple comparison adjustment, i.e. it will lead to the least null hypothesis rejections.

Constructing multiple confidence intervals

A 100(1-a)% confidence interval should contain the true value 100(1-a)% of the time when used with different data sets.

An error occurs if the confidence interval does not contain the true value.

Just like the Type I error and familywise error rate, we can ask what is the probability at least one confidence interval does not cover the true value.

The procedures we will talk about for confidence intervals have equivalent approaches for hypothesis testing (pvalues). Within these procedures we still have the equivalence between pvalues and Cls.

Constructing multiple confidence intervals

Confidence interval for the difference between group j and group j':

$$\overline{Y}_j - \overline{Y}_{j'} \pm M \, s_p \sqrt{\frac{1}{n_j} + \frac{1}{n_{j'}}}$$

where M is a multiplier that depends on the adjustment procedure:

Procedure	M	Use
LSD	$t_{n-J}(1-a/2)$	After significant F -test
		(no adjustment)
Dunnett	multivariate $\it t$	Compare all groups to control
Tukey-Kramer	$q_{J,n-J}(1-a)/\sqrt{2}$	All pairwise comparisons
Scheffé	$\sqrt{(J-1)F_{(J-1,n-J)}(1-a)}$	All contrasts
Bonferroni	$t_{n-J}(1-(a/m)/2)$	m tests
		(most generic)

Tukey for all pairwise comparisons

```
TukevHSD(aov(Lifetime " Diet, data = case0501))
  Tukey multiple comparisons of means
    95% family-wise confidence level
Fit: aov(formula = Lifetime ~ Diet, data = case0501)
$Diet
                       diff
                                   lwr
                                             upr
                                                     p adj
N/N85-NP
                5.2891873 1.5606269 9.0177476 0.0008380
N/R50-NP 14.8951423 11.3405719 18.4497127 0.0000000
R/R50-NP
            15.4836735 11.7397556 19.2275913 0.0000000
N/R50 lopro-NP 12.2836735 8.5397556 16.0275913 0.0000000
N/R40-NP
                 17.7146259 14.0294069 21.3998448 0.0000000
N/R50-N/N85 9.6059550 6.2021702 13.0097399 0.0000000
R/R50-N/N85
            10.1944862 6.5934168 13.7955556 0.0000000
N/R50 lopro-N/N85 6.9944862 3.3934168 10.5955556 0.0000008
N/R40-N/N85
                 12.4254386 8.8854359 15.9654413 0.0000000
R/R50-N/R50
                0.5885312 -2.8320696 4.0091319 0.9963976
N/R50 lopro-N/R50 -2.6114688 -6.0320696 0.8091319 0.2460200
N/R40-N/R50
                  2.8194836 -0.5367684 6.1757356 0.1564608
N/R50 lopro-R/R50 -3.2000000 -6.8169683 0.4169683 0.1167873
N/R40-R/R50
                 2.2309524 -1.3252222 5.7871269 0.4684413
N/R40-N/R50 lopro 5.4309524 1.8747778 8.9871269 0.0002306
```

False Discovery Rate

Not wanting to make a single mistake is pretty conservative. In high-throughput fields a more common multiple comparison adjustment is false discovery rate.

Definition

False discovery rate procedures try to control the expected proportion of incorrectly rejected null hypotheses.

How to incorporate multiple comparison adjustments

- 1. Determine what tests are going to be run (before looking at the data) or what confidence intervals are going to be constructed.
- 2. Determine which multiple comparison adjustment is the most relevant
- 3. Use/state that adjustment and interpret your results