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Accurate prediction of protein structures and interactions using a three-track neural network

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DeepMind presented remarkably accurate predictions at the recent CASP14 protein structure prediction assessment conference. We explored network architectures incorporating related ideas and obtained the best performance with a three-track network in which information at the 1D sequence level, the 2D distance map level, and the 3D coordinate level is successively transformed and integrated. The three-track network produces structure predictions with accuracies approaching those of DeepMind in CASP14, enables the rapid solution of challenging X-ray crystallography and cryo-EM structure modeling problems, and provides insights into the functions of proteins of currently unknown structure. The network also enables rapid generation of accurate protein-protein complex models from sequence information alone, short circuiting traditional approaches which require modeling of individual subunits followed by docking. We make the method available to the scientific community to speed biological research.

The prediction of protein structure from amino acid sequence information alone has been a longstanding challenge. The biannual Critical Assessment of Structure (CASP) meetings have demonstrated that deep learning methods such as AlphaFold (1, 2) and trRosetta (3), that extract information from the large database of known protein structures in the PDB, outperform more traditional approaches that explicitly model the folding process. The outstanding performance of DeepMind's AlphaFold2 in the recent CASP14 meeting (https://predictioncenter.org/casp14/zscores_final.cgi) left the scientific community eager to learn details beyond the overall framework presented and raised the question of whether such accuracy could be achieved outside of a world-leading deep learning company. As described at the CASP14 conference, the AlphaFold2 methodological advances

included 1) starting from multiple sequence alignments (MSAs) rather than from more processed features such as inverse covariance matrices derived from MSAs, 2) replacement of 2D convolution with an attention mechanism that better represents interactions between residues distant along the sequence, 3) use of a two-track network architecture in which information at the 1D sequence level and the 2D distance map level is iteratively transformed and passed back and forth, 4) use of an SE(3)-equivariant Transformer network to directly refine atomic coordinates (rather than 2D distance maps as in previous approaches) generated from the two-track network, and 5) end-to-end learning in which all network parameters are optimized by backpropagation from the final generated 3D coordinates through all network layers back to the input sequence.

Network architecture development

Intrigued by the DeepMind results, and with the goal of increasing protein structure prediction accuracy for structural biology research and advancing protein design (4), we explored network architectures incorporating different combinations of these five properties. In the absence of a published method, we experimented with a wide variety of approaches for passing information between different parts of the networks, as summarized in the methods and table S1. We succeeded in producing a “two-track” network with information flowing in parallel along a 1D sequence alignment track and a 2D distance matrix track with considerably better performance than trRosetta (BAKER-ROSETTASERVER and BAKER in Fig. 1B), the next best method after AlphaFold2 in CASP14 (https://predictioncenter.org/casp14/zscores_final.cgi).

We reasoned that better performance could be achieved by extending to a third track operating in 3D coordinate space to provide a tighter connection between sequence, residue-residue distances and orientations, and atomic coordinates. We constructed architectures with the two levels of the two-track model augmented with a third parallel structure track operating on 3D backbone coordinates as depicted in Fig. 1A (see methods and fig. S1 for details). In this architecture, information flows back and forth between the 1D amino acid sequence information, the 2D distance map, and the 3D coordinates, allowing the network to collectively reason about relationships within and between sequences, distances, and coordinates. In contrast, reasoning about 3D atomic coordinates in the two-track AlphaFold2 architecture happens after processing of the 1D and 2D information is complete (although end-to-end training does link parameters to some extent). Because of computer hardware memory limitations, we could not train models on large proteins directly as the three-track models have many millions of parameters; instead, we presented to the network many discontinuous crops of the input sequence consisting of two discontinuous sequence segments spanning a total of 260 residues. To generate final models, we combined and averaged the 1D features and 2D distance and orientation predictions produced for each of the crops and then used two approaches to generate final 3D structures. In the first, the predicted residue-residue distance and orientation distributions are fed into pyRosetta (5) to generate all-atom models. In the second, the averaged 1D and 2D features are fed into a final SE(3)-equivariant layer (6), and following end-to-end training from amino acid sequence to 3D coordinates, backbone coordinates are generated directly by the network (see methods). We refer to these networks, which also generate per residue accuracy predictions, as RoseTTAFold. The first has the advantage of requiring lower memory (for proteins over 400 residues, 8GB rather than 24GB) GPUs at inference time and producing full side chain models but requires CPU time for the pyRosetta

structure modeling step.

The three-track models with attention operating at the 1D, 2D, and 3D levels and information flowing between the three levels were the best models we tested (Fig. 1B), clearly outperforming the top 2 server groups (Zhang-server and BAKER-ROSETTASERVER), BAKER human group (ranked second among all groups), and our two-track attention models on CASP14 targets. As in the case of AlphaFold2, the correlation between multiple sequence alignment depth and model accuracy is lower for RoseTTAFold than for trRosetta and other methods tested at CASP14 (fig. S2). The performance of the three-track model on the CASP14 targets was still not as good as AlphaFold2 (Fig. 1B). This could reflect hardware limitations that limited the size of the models we could explore, alternative architectures or loss formulations, or more intensive use of the network for inference. DeepMind reported using several GPUs for days to make individual predictions, whereas our predictions are made in a single pass through the network in the same manner that would be used for a server; following sequence and template search (~1.5 hours), the end-to-end version of RoseTTAFold requires ~10 min on an RTX2080 GPU to generate backbone coordinates for proteins with less than 400 residues, and the pyRosetta version requires 5 min for network calculations on a single RTX2080 GPU and an hour for all-atom structure generation with 15 CPU cores. Incomplete optimization due to computer memory limitations and neglect of side chain information likely explain the poorer performance of the end-to-end version compared to the pyRosetta version (Fig. 1B; the latter incorporates side chain information at the all-atom relaxation stage); since SE(3)-equivariant layers are used in the main body of the three-track model, the added gain from the final SE(3) layer is likely less than in the AlphaFold2 case. We expect the end-to-end approach to ultimately be at least as accurate once the computer hardware limitations are overcome, and side chains are incorporated.

The improved performance of the three-track models over the two-track model with identical training sets, similar attention-based architectures for the 1D and 2D tracks, and similar operations in inference (prediction) mode suggests that simultaneously reasoning at the multiple sequence alignment, distance map, and three-dimensional coordinate representations can more effectively extract sequence-structure relationships than reasoning over only MSA and distance map information. The relatively low compute cost makes it straightforward to incorporate the methods in a public server and predict structures for large sets of proteins, for example, all human GPCRs, as described below.

Blind structure prediction tests are needed to assess any new protein structure prediction method, but CASP is held only once every two years. Fortunately, the Continuous Automated Model Evaluation (CAMEO) experiment (7) tests

structure prediction servers blindly on protein structures as they are submitted to the PDB. RoseTTAFold has been evaluated since May 15th, 2021 on CAMEO; over the 69 medium and hard targets released during this time (May 15th, 2021 ~ June 19th, 2021), it outperformed all other servers evaluated in the experiment including Robetta (3), IntFold6-TS (8), BestSingleTemplate (9), and SWISS-MODEL (10) (Fig. 1C).

We experimented with approaches for further improving accuracy by more intensive use of the network during sampling. Since the network can take as input templates of known structures, we experimented with a further coupling of 3D structural information and 1D sequence information by iteratively feeding the predicted structures back into the network as templates and random subsampling from the multiple sequence alignments to sample a broader range of models. These approaches generated ensembles containing higher accuracy models, but the accuracy predictor was not able to consistently identify models better than those generated by the rapid single pass method (fig. S3). Nevertheless, we suspect that these approaches can improve model performance and are carrying out further investigations along these lines.

In developing RoseTTAFold, we found that combining predictions from multiple discontinuous crops generated more accurate structures than predicting the entire structure at once (fig. S4A). We hypothesized that this arises from selecting the most relevant sequences for each region from the very large number of aligned sequences often available (fig. S4B). To enable the network to focus on the most relevant sequence information for each region while keeping access to the full multiple sequence alignment in a more memory efficient way, we experimented with the Perceiver architecture (11), updating smaller seed MSAs (up to 100 sequences) with extra sequences (thousands of sequences) through cross-attention (fig. S4C). Current RoseTTAFold only uses the top 1000 sequences due to memory limitations; with this addition, all available sequence information can be used (often over 10,000 sequences). Initial results are promising (fig. S4D), but more training will be required for rigorous comparison.

Enabling experimental protein structure determination

With the recent considerable progress in protein structure prediction, a key question is what accurate protein structure models can be used for. We investigated the utility of the RoseTTAFold to facilitate experimental structure determination by X-ray crystallography and cryo-electron microscopy and to build models providing biological insights for key proteins of currently unknown structures.

Solution of X-ray structures by molecular replacement (MR) often requires quite accurate models. The much higher accuracy of the RoseTTAFold method than currently

available methods prompted us to test whether it could help solve previously unsolved challenging MR problems and improve the solution of borderline cases. Four recent crystallographic datasets (summarized, including resolution limits, in table S2), which had eluded solution by MR using models available in the PDB, were reanalyzed using RoseTTAFold models: glycine N-acetyltransferase (GLYAT) from *Bos taurus* (fig. S5A), a bacterial oxidoreductase (fig. S5B), a bacterial surface layer protein (SLP) (Fig. 2A) and the secreted protein Lrbp from the fungus *Phanerochaete chrysosporium* (Fig. 2B and fig. S5C). In all four cases, the predicted models had sufficient structural similarity to the true structures that led to successful MR solutions [see methods for details; the per-residue error estimates by DeepAccNet (12) allowed the more accurate parts to be weighted more heavily]. The increased prediction accuracy was critical for success in all cases, as models made with trRosetta did not yield MR solutions.

To determine why the RoseTTAFold models were successful, where PDB structures had previously failed, we compared the models to the crystal structures we obtained. The images in Fig. 2A and fig. S5 show that in each case, the closest homolog of the known structure was a much poorer model than the RoseTTAFold model; in the case of SLP, only a distant model covering part of the N-terminal domain (38% of the sequence) was available in the PDB, while no homologs of the C-terminal domain of SLP or any portion of Lrbp could be detected using HHsearch (13).

Building atomic models of protein assemblies from cryo-EM maps can be challenging in the absence of homologs with known structures. We used RoseTTAFold to predict the p101 G_{βγ} binding domain (GBD) structure in a heterodimeric PI3K_γ complex. The top HHsearch hit has a statistically insignificant E-value of 40 and only covers 14 residues out of 167 residues. The predicted structure could readily fit into the electron density map despite the low local resolution [Fig. 2C, top; trRosetta failed to predict the correct fold with the same MSA input (fig. S6)]. The C_α-RMSD between the predicted and the final refined structure is 3.0 Å over the beta-sheets (Fig. 2C, bottom).

Providing insights into biological function

Experimental structure determination can provide considerable insight into biological function and mechanism. We investigated whether structures generated by RoseTTAFold could similarly provide new insights into function. We focused on two sets of proteins: first, G protein-coupled receptors of currently unknown structure, and second, a set of human proteins implicated in disease. Benchmark tests on GPCR sequences with determined structures showed that RoseTTAFold models for both active and inactive states can be quite accurate even in the absence of close homologs with known structures [and better than those in current GPCR

model databases (14, 15); fig. S7] and that the DeepAccNet model quality predictor (12) provides a good measure of actual model accuracy (fig. S7D). We provide RoseTTAFold models and accompanying accuracy predictions for closed and open states of all human GPCRs of currently unknown structure.

Protein structures can provide insight into how mutations in key proteins lead to human disease. We identified human proteins without close homologs of known structure that contain multiple disease-causing mutations or have been the subject of intensive experimental investigation (see methods). We used RoseTTAFold to generate models for 693 domains from such proteins. Over one-third of these models have a predicted IDDT > 0.8, which corresponded to an average C_α-RMSD -RMSD of 2.6 Å on CASP14 targets (fig. S8). Here, we focus on three examples that illustrate the different ways in which structure models can provide insight into the function or mechanisms of diseases.

Deficiencies in TANGO2 (transport and Golgi organization protein 2) lead to metabolic disorders, and the protein plays an unknown role in Golgi membrane redistribution into the ER (16, 17). The RoseTTAFold model of TANGO2 adopts an N-terminal nucleophile aminohydrolase (Ntn) fold (Fig. 3A) with well-aligned active site residues that are conserved in TANGO2 orthologs (Fig. 3B). Ntn superfamily members with structures similar to the RoseTTAFold model suggest that TANGO2 functions as an enzyme that might hydrolyze a carbon-nitrogen bond in a membrane component (18). Based on the model, known mutations that cause disease (magenta spheres in Fig. 3A) could act by hindering catalysis (R26K, R32Q, and L50P, near active site) or produce steric clashes (G154R) (19) in the hydrophobic core. By comparison, a homology model based on very distant (<15% sequence identity) homologs had multiple alignment shifts that misplace key conserved residues (fig. S9 and table S3).

The ADAM (A Disintegrin And Metalloprotease) and ADAMTS families of metalloproteases are encoded by over 40 human genes, mediate cell-cell and cell-matrix interactions (20, 21) and are involved in a range of human diseases, including cancer metastasis, inflammatory disorders, neurological diseases and asthma (21, 22). The ADAMs contain prodomain and metalloprotease domains; the fold of the metalloprotease is known (23, 24), but not that of the prodomain, which has no homologs of known structure. The RoseTTAFold predicted structure of the ADAM33 prodomain has a lipocalin-like beta-barrel fold (Fig. 3C) belonging to an extended superfamily that includes metalloprotease inhibitors (MPIs) (25). There is a cysteine in an extension following the predicted prodomain barrel; taken together, these data are consistent with experimental data suggesting that the ADAM prodomain inhibits metalloprotease activity using a cysteine switch (26). Conserved residues within ADAM33

orthologs line one side of the barrel and likely interact with the metalloprotease (Fig. 3D).

Transmembrane spanning Ceramide synthase (CERS1) is a key enzyme in sphingolipid metabolism which uses acyl-CoA to generate ceramides with various acyl chain lengths that regulate differentiation, proliferation, and apoptosis (27). Structure information is not available for any of the CerS enzymes or their homologs, and the number and orientation of transmembrane helices (TMH) are not known (28). The RoseTTAFold CERS1 model for residues 98 to 304 (Pfam TLC domain) (29) includes six TMH that traverse the membrane in an up and down arrangement (Fig. 3E). A central crevice extends into the membrane and is lined with residues required for activity (His182 and Asp213) (30) or conserved (W298), as well as a pathogenic mutation (H183Q) found in progressive myoclonus epilepsy and dementia that decreases ceramide levels (31). This active site composition (His182, Asp 213, and potentially a neighboring Ser212) suggests testable reaction mechanisms for the enzyme (Fig. 3F).

Direct generation of protein-protein complex models

The final layer of the end-to-end version of our three-track network generates 3D structure models by combining features from discontinuous crops of the protein sequence (two segments of the protein with a chain break between them). We reasoned that because the network can seamlessly handle chain breaks, it might be able to predict the structure of protein-protein complexes directly from sequence information. Rather than providing the network the sequence of a single protein, with or without possible template structures, two or more sequences (and possible templates for these) can be input, with the output the backbone coordinates of two or more protein chains. Thus, the network enables the direct building of structure models for protein-protein complexes from sequence information, short circuiting the standard procedure of building models for individual subunits and then carrying out rigid-body docking. In addition to the great reduction in compute time required (complex models are generated from sequence information in ~30 min on a 24G TITAN RTX GPU), this approach implements “flexible backbone” docking almost by construction as the structures of the chains are predicted in the context of each other. We tested the end-to-end three-track network on paired sequence alignments for complexes of known structures (32) (see methods and table S4 for details) containing two (Fig. 4A) or three (Fig. 4B) chains, and in many cases, the resulting models were very close to the actual structures [TM-score (33) > 0.8]. Information on residue-residue co-evolution between the paired sequences likely contributes to the accuracy of the rigid body placement as more accurate complex structures were generated when more sequences were available (fig. S10). The network was trained on monomeric proteins, not complexes, so there may be some

training set bias in the monomer structures, but there is none for the complexes.

To illustrate the application of RoseTTAFold to complexes of unknown structure with more than three chains, we used it to generate models of the complete four-chain human IL-12R/IL-12 complex (Fig. 4C and fig. S11). A previously published cryo-EM map of the IL-12 receptor complex indicated a similar topology to that of the IL-23 receptor; however, the resolution was not sufficient to observe the detailed interaction between IL-12R β 2 and IL-12p35 (34). Such an understanding is important for dissecting the specific actions of IL-12 and IL-23 and generating inhibitors that block IL-12 without impacting IL-23 signaling. The RoseTTAFold model fits the experimental cryo-EM density well and identified a shared interaction between Y189 in IL-12p35 and G115 in IL-12R β 2 analogous to the packing between W156 in IL-23p19 with G116 in IL-23R. In addition, the model suggests a role for the IL-12R β 2 N-terminal peptide (residue 24–31) in IL-12 binding not observed in the IL-12 cryo-electron microscopy (IL-12R β 2 D26 may interact with nearby K190 and K194 in IL-12p35), which may provide an avenue to target the interaction between IL-12 and IL-12R β 2 specifically.

Conclusions

RoseTTAFold enables solutions of challenging X-ray crystallography and cryo-EM modeling problems, provides insight into protein function in the absence of experimentally determined structures, and rapidly generates accurate models of protein-protein complexes. Further training on protein-protein complex datasets will likely further improve the modeling of the structures of multiprotein assemblies. The approach can be readily coupled with existing small molecule and protein binder design methodology to improve computational discovery of new protein and small molecule ligands for targets of interest. The simultaneous processing of sequence, distance, and coordinate information by the three-track architecture opens the door to new approaches incorporating constraints and experimental information at all three levels for problems ranging from cryo-EM structure determination to protein design.

REFERENCES AND NOTES

- A. W. Senior, R. Evans, J. Jumper, J. Kirkpatrick, L. Sifre, T. Green, C. Qin, A. Žídek, A. W. R. Nelson, A. Bridgland, H. Penedones, S. Petersen, K. Simonyan, S. Crossan, P. Kohli, D. T. Jones, D. Silver, K. Kavukcuoglu, D. Hassabis, Improved protein structure prediction using potentials from deep learning. *Nature* **577**, 706–710 (2020). doi:10.1038/s41586-019-1923-7 Medline
- J. Jumper, R. Evans, A. Pritzel, T. Green, M. Figurnov, K. Tunyasuvunakool, O. Ronneberger, R. Bates, A. Žídek, A. Bridgland, C. Meyer, S. A. A. Kohl, A. Potapenko, A. J. Ballard, A. Cowie, B. Romera-Paredes, S. Nikolov, R. Jain, J. Adler, T. Back, S. Petersen, D. Reiman, M. Steinegger, M. Pacholska, D. Silver, O. Vinyals, A. W. Senior, K. Kavukcuoglu, P. Kohli, D. Hassabis, "High accuracy protein structure prediction using deep learning" in *Fourteenth Critical Assessment of Techniques for Protein Structure Prediction: CASP14 Abstract Book* (Protein Structure Prediction Center, 2020), pp.22–24.
- J. Yang, I. Anishchenko, H. Park, Z. Peng, S. Ovchinnikov, D. Baker, Improved protein structure prediction using predicted interresidue orientations. *Proc. Natl. Acad. Sci. U.S.A.* **117**, 1496–1503 (2020). doi:10.1073/pnas.1914677117 Medline
- I. Anishchenko, T. M. Chidyausiku, S. Ovchinnikov, S. J. Pellock, D. Baker, De novo protein design by deep network hallucination. *bioRxiv* 2020.07.22.211482 [Preprint] (2020); <https://doi.org/10.1101/2020.07.22.211482>.
- S. Chaudhury, S. Lyskov, J. J. Gray, PyRosetta: A script-based interface for implementing molecular modeling algorithms using Rosetta. *Bioinformatics* **26**, 689–691 (2010). doi:10.1093/bioinformatics/btq007 Medline
- F. B. Fuchs, D. E. Worrall, V. Fischer, M. Welling, SE(3)-Transformers: 3D roto-translation equivariant attention networks. arXiv:2006.10503 [cs.LG] (2020).
- J. Haas, A. Barbato, D. Behringer, G. Studer, S. Roth, M. Bertoni, K. Mostaguir, R. Gumieny, T. Schwede, Continuous Automated Model EvaluatiOn (CAMEO) complementing the critical assessment of structure prediction in CASP12. *Proteins* **86**, 387–398 (2018). doi:10.1002/prot.25431 Medline
- L. J. McGuffin, R. Adiyaman, A. H. A. Maghrabi, A. N. Shuid, D. A. Brackenridge, J. O. Nealon, L. S. Philomina, IntFOLD: An integrated web resource for high performance protein structure and function prediction. *Nucleic Acids Res.* **47**, W408–W413 (2019). doi:10.1093/nar/gkz322 Medline
- J. Haas, R. Gumieny, A. Barbato, F. Ackermann, G. Tauriello, M. Bertoni, G. Studer, A. Smolinski, T. Schwede, Introducing "best single template" models as reference baseline for the Continuous Automated Model Evaluation (CAMEO). *Proteins* **87**, 1378–1387 (2019). doi:10.1002/prot.25815 Medline
- A. Waterhouse, M. Bertoni, S. Bienert, G. Studer, G. Tauriello, R. Gumieny, F. T. Heer, T. A. P. de Beer, C. Remppfer, L. Bordoli, R. Lepore, T. Schwede, SWISS-MODEL: Homology modelling of protein structures and complexes. *Nucleic Acids Res.* **46**, W296–W303 (2018). doi:10.1093/nar/gky427 Medline
- A. Jaegle, F. Gimeno, A. Brock, A. Zisserman, O. Vinyals, J. Carreira, Perceiver: General perception with iterative attention. arXiv:2103.03206 [cs.CV] (2021).
- N. Hiranuma, H. Park, M. Baek, I. Anishchenko, J. Dauparas, D. Baker, Improved protein structure refinement guided by deep learning based accuracy estimation. *Nat. Commun.* **12**, 1340 (2021). doi:10.1038/s41467-021-21511-x Medline
- M. Steinegger, M. Meier, M. Mirdita, H. Vöhringer, S. J. Haunsberger, J. Söding, HH-suited3 for fast remote homology detection and deep protein annotation. *BMC Bioinformatics* **20**, 473 (2019). doi:10.1186/s12859-019-3019-7 Medline
- A. J. Kooistra, S. Mordalski, G. Pády-Szekeres, M. Esguerra, A. Mamyrbekov, C. Munk, G. M. Keserű, D. E. Gloriam, GPCRdb in 2021: Integrating GPCR sequence, structure and function. *Nucleic Acids Res.* **49**, D335–D343 (2021). doi:10.1093/nar/gkaa1080 Medline
- B. J. Bender, B. Marlow, J. Meiler, Improving homology modeling from low-sequence identity templates in Rosetta: A case study in GPCRs. *PLOS Comput. Biol.* **16**, e1007597 (2020). doi:10.1371/journal.pcbi.1007597 Medline
- L. S. Kremer, F. Distelmaier, B. Alhaddad, M. Hempel, A. Iuso, C. Küpper, C. Mühlhausen, R. Kovacs-Nagy, R. Satanovskij, E. Graf, R. Berutti, G. Eckstein, T. Durbin, S. Sauer, G. F. Hoffmann, T. M. Strom, R. Santer, T. Meitinger, T. Klopstock, H. Prokisch, T. B. Haack, Bi-allelic truncating mutations in *TANGO2* cause infancy-onset recurrent metabolic crises with encephalocardiomyopathy. *Am. J. Hum. Genet.* **98**, 358–362 (2016). doi:10.1016/j.ajhg.2015.12.009 Medline
- C. Rabouille, V. Kondylis, TANGOing along the protein secretion pathway. *Genome Biol.* **7**, 213 (2006). doi:10.1186/gb-2006-7-4-213 Medline
- M. P. Milev, D. Saint-Dic, K. Zardoui, T. Klopstock, C. Law, F. Distelmaier, M. Sacher, The phenotype associated with variants in *TANGO2* may be explained by a dual role of the protein in ER-to-Golgi transport and at the mitochondria. *J. Inherit. Metab. Dis.* **44**, 426–437 (2021). doi:10.1002/jimd.12312 Medline
- S. R. Lalani, P. Liu, J. A. Rosenfeld, L. B. Watkin, T. Chiang, M. S. Leduc, W. Zhu, Y. Ding, S. Pan, F. Vetrini, C. Y. Miyake, M. Shinawi, T. Gambin, M. K. Eldomery, Z. H. C. Akdemir, L. Emrick, Y. Wilnai, S. Schelley, M. K. Koenig, N. Memori, L. S. Farach, B. P. Coe, M. Azamian, P. Hernandez, G. Zapata, S. N. Jhangiani, D. M. Muzny, T. Lotze, G. Clark, A. Wilfong, H. Northrup, A. Adesina, C. A. Bacino, F. Scaglia, P. E. Bonnen, J. Crosson, J. Duis, G. H. B. Maegawa, D. Coman, A. Inwood, J. McGill, E. Boerwinkle, B. Graham, A. Beaudet, C. M. Eng, N. A. Hanchard, F. Xia, J. S. Orange, R. A. Gibbs, J. R. Lupski, Y. Yang, Recurrent muscle weakness with rhabdomyolysis, metabolic crises, and cardiac arrhythmia due to bi-allelic *TANGO2* mutations. *Am. J. Hum. Genet.* **98**, 347–357 (2016). doi:10.1016/j.ajhg.2015.12.008 Medline

20. T. G. Wolfsberg, P. Primakoff, D. G. Myles, J. M. White, ADAM, a novel family of membrane proteins containing A Disintegrin And Metalloprotease domain: Multipotential functions in cell-cell and cell-matrix interactions. *J. Cell Biol.* **131**, 275–278 (1995). doi:10.1083/jcb.131.2.275 Medline
21. T. Klein, R. Bischoff, Active metalloproteases of the A Disintegrin and Metalloprotease (ADAM) family: Biological function and structure. *J. Proteome Res.* **10**, 17–33 (2011). doi:10.1021/pr05056z Medline
22. S. Zhong, R. A. Khalil, A Disintegrin and Metalloproteinase (ADAM) and ADAM with thrombospondin motifs (ADAMTS) family in vascular biology and disease. *Biochem. Pharmacol.* **164**, 188–204 (2019). doi:10.1016/j.bcp.2019.03.033 Medline
23. P. Orth, P. Reichert, W. Wang, W. W. Prosite, T. Yarosh-Tomaine, G. Hammond, R. N. Ingram, L. Xiao, U. A. Mirza, J. Zou, C. Strickland, S. S. Taremi, H. V. Le, V. Madison, Crystal structure of the catalytic domain of human ADAM33. *J. Mol. Biol.* **335**, 129–137 (2004). doi:10.1016/j.jmb.2003.10.037 Medline
24. S. Takeda, T. Igarashi, H. Mori, S. Araki, Crystal structures of VAP1 reveal ADAMs' MDC domain architecture and its unique C-shaped scaffold. *EMBO J.* **25**, 2388–2396 (2006). doi:10.1038/sj.emboj.7601131 Medline
25. D. R. Flower, A. C. North, C. E. Sansom, The lipocalin protein family: Structural and sequence overview. *Biochim. Biophys. Acta* **1482**, 9–24 (2000). doi:10.1016/S0167-4838(00)00148-5 Medline
26. H. E. Van Wart, H. Birkedal-Hansen, The cysteine switch: A principle of regulation of metalloproteinase activity with potential applicability to the entire matrix metalloproteinase gene family. *Proc. Natl. Acad. Sci. U.S.A.* **87**, 5578–5582 (1990). doi:10.1073/pnas.87.14.5578 Medline
27. M. Levy, A. H. Futerman, Mammalian ceramide synthases. *IUBMB Life* **62**, 347–356 (2010). Medline
28. J. L. Kim, B. Mestre, S.-H. Shin, A. H. Futerman, Ceramide synthases: Reflections on the impact of Dr. Lina M. Obeid. *Cell. Signal.* **82**, 109958 (2021). doi:10.1016/j.cellsig.2021.109958 Medline
29. E. Winter, C. P. Ponting, TRAM, LAG1 and CLN8: Members of a novel family of lipid-sensing domains? *Trends Biochem. Sci.* **27**, 381–383 (2002). doi:10.1016/S0968-0004(02)02154-0 Medline
30. S. Spassieva, J.-G. Seo, J. C. Jiang, J. Bielawski, F. Alvarez-Vasquez, S. M. Jazwinski, Y. A. Hannun, L. M. Obeid, Necessary role for the Lag1p motif in (dihydro)ceramide synthase activity. *J. Biol. Chem.* **281**, 33931–33938 (2006). doi:10.1074/jbc.M608092200 Medline
31. N. Vanni, F. Fruscione, E. Ferlazzo, P. Striano, A. Robbiani, M. Traverso, T. Sander, A. Falace, E. Gazzero, P. Bramanti, J. Bielawski, A. Fassio, C. Minetti, P. Genton, F. Zara, Impairment of ceramide synthesis causes a novel progressive myoclonus epilepsy. *Ann. Neurol.* **76**, 206–212 (2014). doi:10.1002/ana.24170 Medline
32. Q. Cong, I. Anishchenko, S. Ovchinnikov, D. Baker, Protein interaction networks revealed by proteome coevolution. *Science* **365**, 185–189 (2019). Medline
33. Y. Zhang, J. Skolnick, Scoring function for automated assessment of protein structure template quality. *Proteins* **57**, 702–710 (2004). doi:10.1002/prot.20264 Medline
34. C. R. Glassman, Y. K. Mathiharan, K. M. Jude, L. Su, O. Panova, P. J. Lupardus, J. B. Spangler, L. K. Ely, C. Thomas, G. Skiniotis, K. C. Garcia, Structural basis for IL-12 and IL-23 receptor sharing reveals a gateway for shaping actions on T versus NK cells. *Cell* **184**, 983–999.e24 (2021). doi:10.1016/j.cell.2021.01.018 Medline
35. E. F. Pettersen, T. D. Goddard, C. C. Huang, E. C. Meng, G. S. Couch, T. I. Croll, J. H. Morris, T. E. Ferrin, UCSF ChimeraX: Structure visualization for researchers, educators, and developers. *Protein Sci.* **30**, 70–82 (2021). doi:10.1002/pro.3943 Medline
36. M. Baek, F. DiMaio, I. Anishchenko, J. Dauparas, S. Ovchinnikov, J. Wang, D. Baker, RoseTTAFold: The first release of RoseTTAFold. Zenodo (2021); <https://zenodo.org/record/5068265>.
37. A. Vaswani, N. Shazeer, N. Parmar, J. Uszkoreit, L. Jones, A. N. Gomez, L. Kaiser, I. Polosukhin, Attention is all you need. arXiv:1706.03762 [cs.CL] (2017).
38. J. Ho, N. Kalchbrenner, D. Weissenborn, T. Salimans, Axial attention in multidimensional transformers. arXiv:1912.12180 [cs.CV] (2019).
39. K. Choromanski, V. Likhoshesterov, D. Dohan, X. Song, A. Gane, T. Sarlos, P. Hawkins, J. Davis, A. Mohiuddin, L. Kaiser, D. Belanger, L. Colwell, A. Weller, Rethinking attention with Performers. arXiv:2009.14794 [cs.LG] (2020).
40. R. Rao, J. Liu, R. Verkuil, J. Meier, J. F. Canny, P. Abbeel, T. Sercu, A. Rives, MSA Transformer, bioRxiv 2021.02.12.430858 [Preprint] (2021); <https://doi.org/10.1101/2021.02.12.430858>.
41. F. Ju, J. Zhu, B. Shao, L. Kong, T.-Y. Liu, W.-M. Zheng, D. Bu, CopulaNet: Learning residue co-evolution directly from multiple sequence alignment for protein structure prediction. *Nat. Commun.* **12**, 2535 (2021). doi:10.1038/s41467-021-22869-8 Medline
42. Y. Shi, Z. Huang, S. Feng, H. Zhong, W. Wang, Y. Sun, Masked label prediction: Unified message passing model for semi-supervised classification. arXiv:2009.03509 [cs.LG] (2020).
43. V. Mariani, M. Biasini, A. Barbato, T. Schwede, IDDT: A local superposition-free score for comparing protein structures and models using distance difference tests. *Bioinformatics* **29**, 2722–2728 (2013). doi:10.1093/bioinformatics/btt473 Medline
44. M. Mirdita, L. von den Driesch, C. Galiez, M. J. Martin, J. Söding, M. Steinegger, Uniclust databases of clustered and deeply annotated protein sequences and alignments. *Nucleic Acids Res.* **45**, D170–D176 (2017). doi:10.1093/nar/gkw1081 Medline
45. M. Steinegger, M. Mirdita, J. Söding, Protein-level assembly increases protein sequence recovery from metagenomic samples manyfold. *Nat. Methods* **16**, 603–606 (2019). doi:10.1038/s41592-019-0437-4 Medline
46. L. Zimmermann, A. Stephens, S.-Z. Nam, D. Rau, J. Kübler, M. Lozajic, F. Gabler, J. Söding, A. N. Lupas, V. Alva, A completely reimplemented MPI bioinformatics toolkit with a new HHpred server at its core. *J. Mol. Biol.* **430**, 2237–2243 (2018). doi:10.1016/j.jmb.2017.12.007 Medline
47. G. Bunkóczki, R. J. Read, Improvement of molecular-replacement models with Sculptor. *Acta Crystallogr. D Biol. Crystallogr.* **67**, 303–312 (2011). doi:10.1107/S090744910051218 Medline
48. G. Bunkóczki, R. J. Read, phenix.ensemble: A tool for multiple superposition. *Comput. Crystallogr. Newslett.* **2**, 8–9 (2011).
49. A. J. McCoy, R. W. Grosse-Kunstleve, P. D. Adams, M. D. Winn, L. C. Storoni, R. J. Read, Phaser crystallographic software. *J. Appl. Crystallogr.* **40**, 658–674 (2007). doi:10.1107/S0021889807021206 Medline
50. A. Vagin, A. Lebedev, MoRDa, an automatic molecular replacement pipeline. *Acta Crystallogr. A Found. Adv.* **A71**, S19 (2015). doi:10.1107/S2053273315099672.
51. Y. Wang, J. Virtanen, Z. Xue, Y. Zhang, I-TASSER-MR: Automated molecular replacement for distant-homology proteins using iterative fragment assembly and progressive sequence truncation. *Nucleic Acids Res.* **45**, W429–W434 (2017). doi:10.1093/nar/gkx349 Medline
52. A. J. McCoy, R. D. Oeffner, A. G. Wrobel, J. R. M. Ojala, K. Tryggvason, B. Lohkamp, R. J. Read, Ab initio solution of macromolecular crystal structures without direct methods. *Proc. Natl. Acad. Sci. U.S.A.* **114**, 3637–3641 (2017). doi:10.1073/pnas.1701640114 Medline
53. G. Bunkóczki, B. Wallner, R. J. Read, Local error estimates dramatically improve the utility of homology models for solving crystal structures by molecular replacement. *Structure* **23**, 397–406 (2015). doi:10.1016/j.str.2014.11.020 Medline
54. T. C. Terwilliger, Maximum-likelihood density modification. *Acta Crystallogr. D Biol. Crystallogr.* **56**, 965–972 (2000). doi:10.1107/S0907444900005072 Medline
55. D. Liebschner, P. V. Afonine, M. L. Baker, G. Bunkóczki, V. B. Chen, T. I. Croll, B. Hintze, L. W. Hung, S. Jain, A. J. McCoy, N. W. Moriarty, R. D. Oeffner, B. K. Poon, M. G. Prisant, R. J. Read, J. S. Richardson, D. C. Richardson, M. D. Sammito, O. V. Sobolev, D. H. Stockwell, T. C. Terwilliger, A. G. Urzhumtsev, L. L. Videau, C. J. Williams, P. D. Adams, Macromolecular structure determination using x-rays, neutrons and electrons: Recent developments in Phenix. *Acta Crystallogr. D Struct. Biol.* **75**, 861–877 (2019). doi:10.1107/S2059798319011471 Medline
56. T. C. Terwilliger, R. W. Grosse-Kunstleve, P. V. Afonine, N. W. Moriarty, P. H. Zwart, L. W. Hung, R. J. Read, P. D. Adams, Iterative model building, structure refinement and density modification with the PHENIX AutoBuild wizard. *Acta Crystallogr. D Biol. Crystallogr.* **64**, 61–69 (2008). doi:10.1107/S090744490705024X Medline
57. P. Emsley, B. Lohkamp, W. G. Scott, K. Cowtan, Features and development of Coot. *Acta Crystallogr. D Biol. Crystallogr.* **66**, 486–501 (2010). doi:10.1107/S0907444910007493 Medline
58. P. V. Afonine, R. W. Grosse-Kunstleve, N. Echols, J. J. Headd, N. W. Moriarty, M. Mustyakimov, T. C. Terwilliger, A. Urzhumtsev, P. H. Zwart, P. D. Adams, Towards

- automated crystallographic structure refinement with phenix.refine. *Acta Crystallogr. D Biol. Crystallogr.* **68**, 352–367 (2012). doi:10.1107/S0907444912001308 Medline
59. C. J. Williams, J. J. Headd, N. W. Moriarty, M. G. Prisant, L. L. Viedau, L. N. Deis, V. Verma, D. A. Keedy, B. J. Hintze, V. B. Chen, S. Jain, S. M. Lewis, W. B. Arendall III, J. Snoeyink, P. D. Adams, S. C. Lovell, J. S. Richardson, D. C. Richardson, MolProbity: More and better reference data for improved all-atom structure validation. *Protein Sci.* **27**, 293–315 (2018). doi:10.1002/pro.3330 Medline
60. R. J. Read, A. J. McCoy, Using SAD data in Phaser. *Acta Crystallogr. D Biol. Crystallogr.* **67**, 338–344 (2011). doi:10.1107/S0907444910051371 Medline
61. J. Xu, M. McPartlon, J. Li, Improved protein structure prediction by deep learning irrespective of co-evolution information. *Nat. Mach. Intell.* (2021). doi:10.1038/s42256-021-00348-5
62. J. Yang, Y. Zhang, I-TASSER server: New development for protein structure and function predictions. *Nucleic Acids Res.* **43**, W174–W181 (2015). doi:10.1093/nar/gkv342 Medline
63. D. Xu, Y. Zhang, Toward optimal fragment generations for ab initio protein structure assembly. *Proteins* **81**, 229–239 (2013). doi:10.1002/prot.24179 Medline
64. The UniProt Consortium, UniProt: The universal protein knowledgebase. *Nucleic Acids Res.* **45**, D158–D169 (2017). doi:10.1093/nar/gkw1099 Medline
65. J. Pei, N. V. Grishin, The DBSAV database: Predicting deleteriousness of single amino acid variations in the human proteome. *J. Mol. Biol.* **433**, 166915 (2021). doi:10.1016/j.jmb.2021.166915 Medline
66. L. S. Johnson, S. R. Eddy, E. Portugaly, Hidden Markov model speed heuristic and iterative HMM search procedure. *BMC Bioinformatics* **11**, 431 (2010). doi:10.1186/1471-2105-11-431 Medline
67. S. El-Gebali, J. Mistry, A. Bateman, S. R. Eddy, A. Luciani, S. C. Potter, M. Qureshi, L. J. Richardson, G. A. Salazar, A. Smart, E. L. L. Sonnhammer, L. Hirsh, L. Paladin, D. Piovesan, S. C. E. Tosatto, R. D. Finn, The Pfam protein families database in 2019. *Nucleic Acids Res.* **47**, D427–D432 (2019). doi:10.1093/nar/gky995 Medline
68. S. Bienert, A. Waterhouse, T. A. P. de Beer, G. Tauriello, G. Studer, L. Bordoli, T. Schwede, The SWISS-MODEL Repository-new features and functionality. *Nucleic Acids Res.* **45**, D313–D319 (2017). doi:10.1093/nar/gkw1132 Medline
69. B. Mészáros, G. Erdos, Z. Dosztányi, IUPred2A: Context-dependent prediction of protein disorder as a function of redox state and protein binding. *Nucleic Acids Res.* **46**, W329–W337 (2018). doi:10.1093/nar/gky384 Medline
70. J. Hanson, K. K. Paliwal, T. Litfin, Y. Zhou, SPOT-Disorder2: Improved protein intrinsic disorder prediction by ensembled deep learning. *Genomics Proteomics Bioinformatics* **17**, 645–656 (2019). doi:10.1016/j.gpb.2019.01.004 Medline
71. F. Gabler, S.-Z. Nam, S. Till, M. Mirdita, M. Steinegger, J. Söding, A. N. Lupas, V. Alva, Protein sequence analysis using the MPI Bioinformatics Toolkit. *Curr. Protoc. Bioinformatics* **72**, e108 (2020). doi:10.1002/cpb.108 Medline
72. H. Cheng, R. D. Schaeffer, Y. Liao, L. N. Kinch, J. Pei, S. Shi, B.-H. Kim, N. V. Grishin, ECOD: An evolutionary classification of protein domains. *PLOS Comput. Biol.* **10**, e1003926 (2014). doi:10.1371/journal.pcbi.1003926 Medline
73. R. Ayoub, Y. Lee, RUPEE: A fast and accurate purely geometric protein structure search. *PLOS ONE* **14**, e0213712 (2019). doi:10.1371/journal.pone.0213712 Medline
74. J. Pei, N. V. Grishin, AL2CO: Calculation of positional conservation in a protein sequence alignment. *Bioinformatics* **17**, 700–712 (2001). doi:10.1093/bioinformatics/17.8.700 Medline
75. K. Katoh, D. M. Standley, MAFFT multiple sequence alignment software version 7: Improvements in performance and usability. *Mol. Biol. Evol.* **30**, 772–780 (2013). doi:10.1093/molbev/mst010 Medline
76. A. M. Altenhoff, C.-M. Train, K. J. Gilbert, I. Mediratta, T. Mendes de Farias, D. Moi, Y. Nevers, H.-S. Radoykova, V. Rossier, A. Warwick Vesztrocy, N. M. Glover, C. Dessimoz, OMA orthology in 2021: Website overhaul, conserved isoforms, ancestral gene order and more. *Nucleic Acids Res.* **49**, D373–D379 (2021). doi:10.1093/nar/gkaa1007 Medline
77. P. Benkert, M. Biasini, T. Schwede, Toward the estimation of the absolute quality of individual protein structure models. *Bioinformatics* **27**, 343–350 (2011). doi:10.1093/bioinformatics/btq662 Medline
78. L. Holm, Using Dali for protein structure comparison. *Methods Mol. Biol.* **2112**, 29–42 (2020). doi:10.1007/978-1-0716-0270-6_3 Medline
79. S. J. Hubbard, J. M. Thornton, “naccess,” computer program (Department of Biochemistry and Molecular Biology, University College London, 1993).
80. A. Lafita, S. Bliven, A. Kryshtafovych, M. Bertoni, B. Monastyrskyy, J. M. Duarte, T. Schwede, G. Capitani, Assessment of protein assembly prediction in CASP12. *Proteins* **86** (Suppl 1), 247–256 (2018). doi:10.1002/prot.25408 Medline
81. P. Conway, M. D. Tyka, F. DiMaio, D. E. Konnerding, D. Baker, Relaxation of backbone bond geometry improves protein energy landscape modeling. *Protein Sci.* **23**, 47–55 (2014). doi:10.1002/pro.2389 Medline
82. M. A. Larkin, G. Blackshields, N. P. Brown, R. Chenna, P. A. McGgettigan, H. McWilliam, F. Valentini, I. M. Wallace, A. Wilim, R. Lopez, J. D. Thompson, T. J. Gibson, D. G. Higgins, Clustal W and Clustal X version 2.0. *Bioinformatics* **23**, 2947–2948 (2007). doi:10.1093/bioinformatics/btm404 Medline

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SUPPLEMENTARY MATERIALS

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Materials and Methods

Figs. S1 to S17

Tables S1 to S4

References (37–82)

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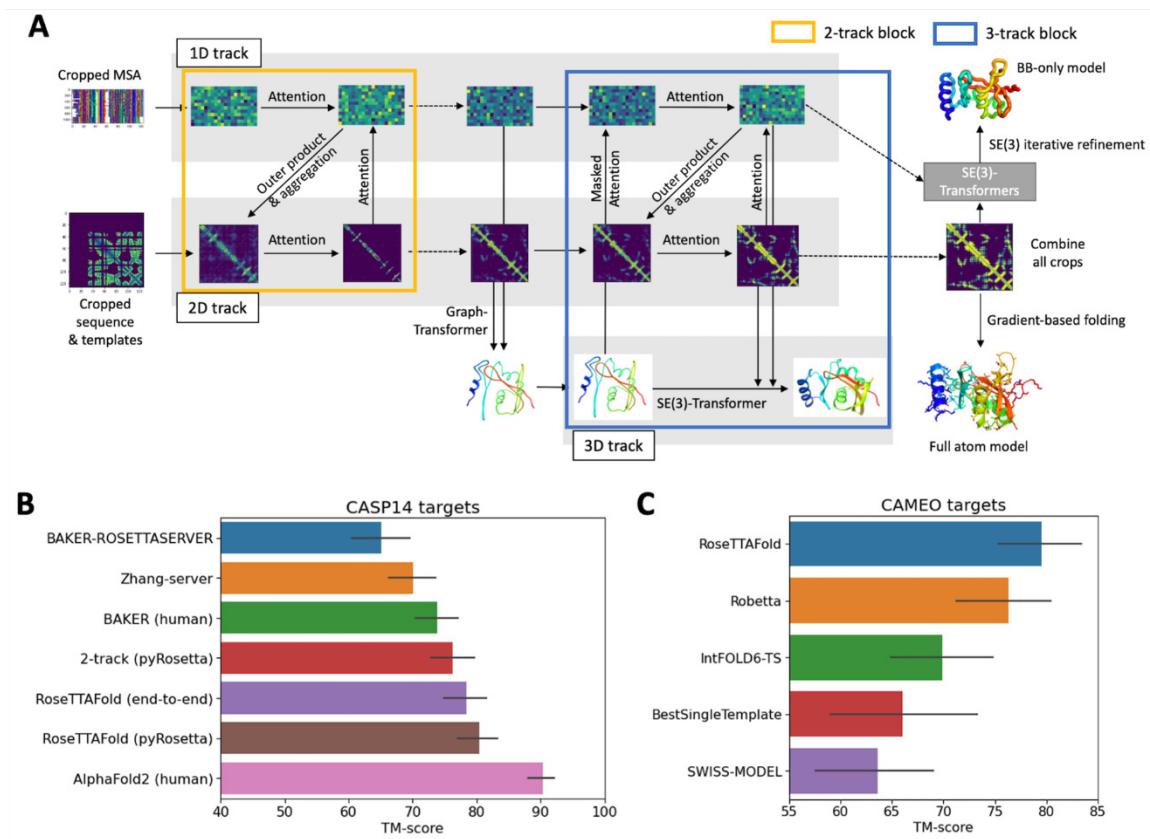


Fig. 1. Network architecture and performance. (A) RoseTTAFold architecture with 1D, 2D, and 3D attention tracks. Multiple connections between tracks allow the network to simultaneously learn relationships within and between sequences, distances, and coordinates (see methods and fig. S1 for details). (B) Average TM-score of prediction methods on the CASP14 targets. Zhang-server and BAKER-ROSETTASERVER were the top 2 server groups while AlphaFold2 and BAKER were the top 2 human groups in CASP14; BAKER-ROSETTASERVER and BAKER predictions were based on trRosetta. Predictions with the two-track model and RoseTTAFold (both end-to-end and pyRosetta version) were completely automated. (C) Blind benchmark results on CAMEO medium and hard targets; model accuracies are TM-score values from the CAMEO website (<https://cameo3d.org/>).

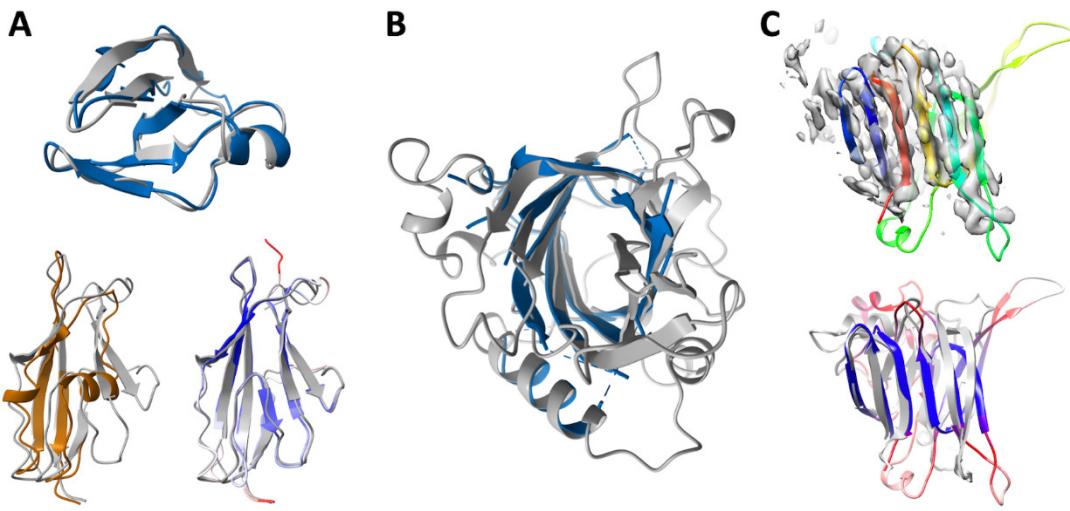


Fig. 2. Enabling experimental structure determination with RoseTTAFold. (A and B) Successful molecular replacement with RoseTTAFold models. (A) SLP. (top) C-terminal domain: comparison of final refined structure (gray) to RoseTTAFold model (blue); there are no homologs with known structure. (bottom) N-terminal domain: refined structure is in gray, and RoseTTAFold model is colored by the estimated RMS error (ranging from blue for 0.67 Å to red for 2 Å or greater). 95 C_α atoms of the RoseTTAFold model can be superimposed within 3 Å of C_α atoms in the final structure, yielding a C_α-RMSD of 0.98 Å. In contrast, only 54 C_α atoms of the closest template (4I3a, brown) can be superimposed (with a C_α-RMSD of 1.69 Å). (B) Refined structure of Lrbp (gray) with the closest RoseTTAFold model (blue) superimposed; residues having estimated RMS error greater than 1.3 Å are omitted (full model is in fig. S5C). (C) Cryo-EM structure determination of p101 G_β_γ binding domain (GBD) in a heterodimeric PI3K_γ complex using RoseTTAFold. (top) RoseTTAFold models colored in a rainbow from the N terminus (blue) to the C terminus (red) have a consistent all-beta topology with a clear correspondence to the density map. (bottom) Comparison of the final refined structure to the RoseTTAFold model colored by predicted RMS error ranging from blue for 1.5 Å or less to red 3 Å or greater. The actual C_α-RMSD between the predicted structure and final refined structure is 3.0 Å over the beta-sheets. Figure prepared with ChimeraX (35).

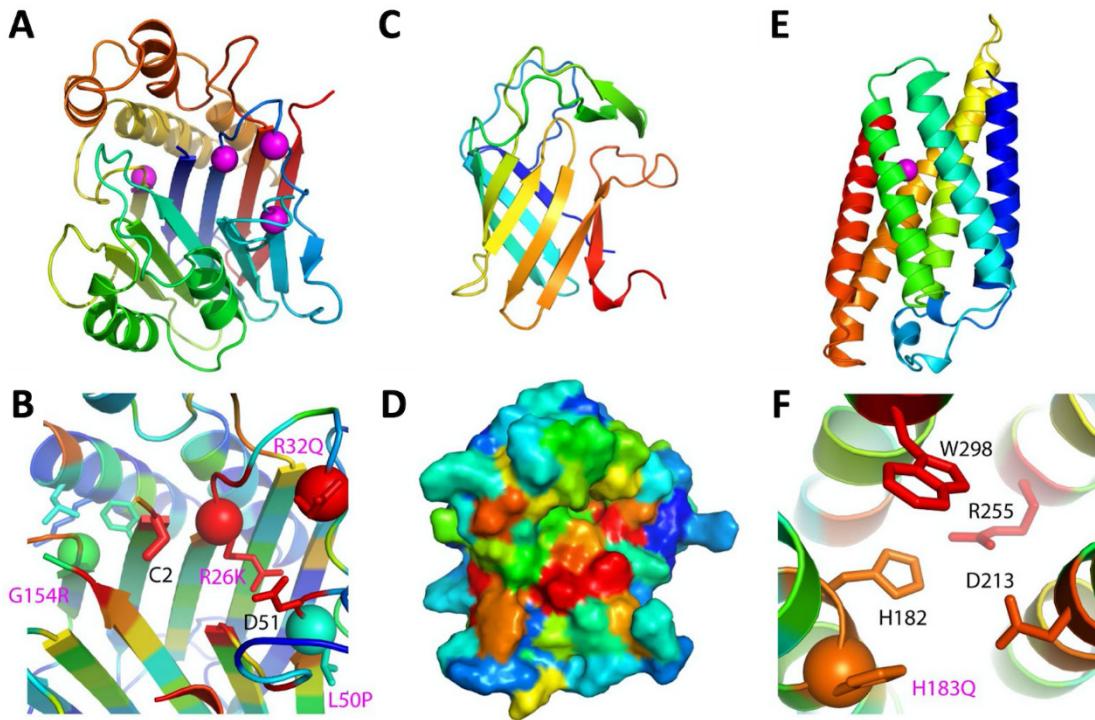


Fig. 3. RoseTTAFold models provide insights into function. (A) TANGO2 model, colored in a rainbow from the N terminus (blue) to the C terminus (red), adopts an Ntn hydrolase fold. Pathogenic mutation sites are in magenta spheres. (B) Predicted TANGO2 active site colored by ortholog conservation in rainbow scale from variable (blue) to conserved (red) with conserved residues in stick and labeled. Pathogenic mutations (spheres with wild-type side chains in the sticks) are labeled in magenta; select neighboring residues are depicted in the sticks. (C) ADAM33 prodomain adopts a lipocalin-like barrel shown in a rainbow from N terminus (blue) to C terminus (red). (D) ADAM33 model surface rendering colored by ortholog conservation from blue (variable) to red (conserved), highlighting a conserved surface patch. (E) CERS1 transmembrane structure prediction is colored from N terminus (blue) to C terminus (red), with a pathogenic mutation in TMH2 near a central cavity in magenta. (F) Zoom of CERS1 active site with residues colored by ortholog conservation from variable (blue) to conserved (red). Residues that contribute to catalysis (H182 and D213) or are conserved (W298 and D213) line the cavity. The conserved pathogenic mutation is adjacent to the active site.

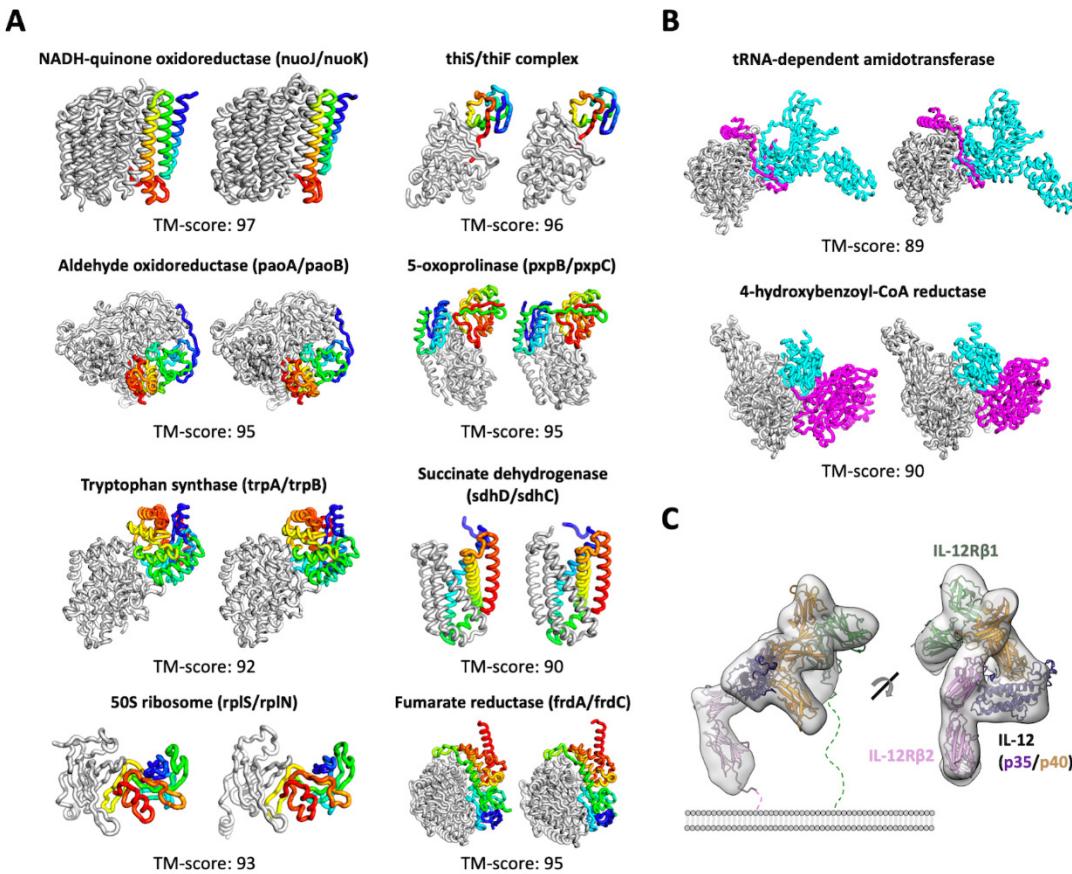


Fig. 4. Complex structure prediction using RoseTTAFold. (A and B) Prediction of structures of *E.coli* protein complexes from sequence information. Experimentally determined structures are on the left, RoseTTAFold models, on the right; the TM-scores below indicate the extent of structural similarity. (A) Two chain complexes. The first subunit is colored in gray, and the second subunit is colored in a rainbow from blue (N-terminal) to red (C-terminal). (B) Three chain complexes. Subunits are colored in gray, cyan, and magenta. (C) IL-12R/IL-12 complex structure generated by RoseTTAFold fits the previously published cryo-EM density (EMD-21645).

Accurate prediction of protein structures and interactions using a three-track neural network

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