

Contents

Diário de bordo da geração dos shinyCircos	1
Configuração inicial	1
Pre processamento	2
Obter os cromossomos	2
Obtendo a densidade dos cromossomos	2
Exemplos de uso	2
Problemas encontrados até o momento	2
Corrigindo os arquivos para o uso do DensityMap	2
Obter os CODING (Camada 1)	3
Obter os CODING com densidade (Camada 1)	4
Obter os NON CODING (Camada 2)	4
Obter os NON CODING com densidade (Camada 2)	4
Obter os TEs (Camada 3)	4
Obter os TEs com densidade (Camada 3)	5
Anotações	5

Diário de bordo da geração dos shinyCircos

Considerar apenas os C147-Phased

Camadas:

1. Cromossomos
2. Coding
3. Non Coding
4. TE

Configuração inicial

```
1 mkdir -pv {tmp,output}
```

Pre processamento

O arquivo `ResultadoFinal_C174-PHASED.gff` possui strand +, -, plus e minus.
O correto é apenas + e -.

```
1 awk 'BEGIN {FS="\t";OFS="\t"} {if ($7=="plus"){ $7="+"}else  
    if($7=="minus"){ $7="-"} {print}}'  
    ResultadoFinal_C174-PHASED.gff >  
    tmp/ResultadoFinal_C174-PHASED-corrigido.gff
```

Obter os cromossomos

```
1 # Print sequence length and names (no sequences)  
2 mkdir -pv output  
3 echo "chr;start;end" > output/chr.csv  
4 seqkit fx2tab C174-PHASED.fa -l -n | awk 'BEGIN {OFS = ","}  
    {print $1,0,$2}' >> output/chr.csv
```

Obtendo a densidade dos cromossomos

Exemplos de uso

```
1 DensityMap.pl -i ./annot-genes/Tgrand_C174P-Annotation.gff3 -ty  
    "ncRNA=fused=10" -o 2R.svg -for -v  
2  
3 DensityMap.pl -i ../DensityMap/dmel/dmel.gff3 -o egn.svg -ty  
    'gene=fused;exon=fused;ncRNA=fused=10' -gc 12 -sc 40000 -ba  
    white -str_s 15 -str_w 25 -sp 35 -sh 50 -title "Density Map  
    of Gene, Exon, ncRNA and GC%" -la -15 -ro ceil -v
```

Problemas encontrados até o momento

1. A marcação `##sequence-region` do GFF precisa obrigatoriamente iniciar em 1. Caso contrário o DensityMap não identifica a linha. Foi apresentado uma solução no Pull Request (<https://github.com/sguizard/DensityMap/pull/5>). Aguardando o autor aceitar.
2. Se o GFF possuir mais de uma região (`##sequence-region`), a marcação da região deve vir imediatamente antes dos seus registros.

Corrigindo os arquivos para o uso do DensityMap

```
1 # ./annot-genes/Tgrand_C174P-Annotation.gff3  
2  
3 mkdir -pv tmpx
```

```

4 awk '/##sequence-region/ {print "##gff-version 3" >
    "tmpx/"$2".gff3"}' ./annot-genes/Tgrand_C174P-Annotation.gff3
5 awk '/##sequence-region/ {print $1,$2,1,$4 >> "tmpx/"$2".gff3"}'
    ./annot-genes/Tgrand_C174P-Annotation.gff3
6 awk '/^chr[0-9]{1,2}[ab]/ {print $0 >> "tmpx/"$1".gff3"}'
    ./annot-genes/Tgrand_C174P-Annotation.gff3
7 cat tmpx/*.gff3 > tmp/Tgrand_C174P-Annotation-for-DensityMap.gff3
8 rm -rv tmpx

```

```

1 # ResultadoFinal_C174-PHASED.gff
2 mkdir -pv tmpx
3 awk 'BEGIN {FS=";";OFS=" "} /^chr[0-9]+[ab]/ {print
    "##gff-version 3" > "tmpx/"$1".gff3"}' output/chr.csv
4 awk 'BEGIN {FS=";";OFS=" "} /^chr[0-9]+[ab]/ {print
    "##sequence-region", $1, "1", $3 >> "tmpx/"$1".gff3"}'
    output/chr.csv
5 awk 'BEGIN {FS="\t";OFS="\t"} /^chr[0-9]{1,2}[ab]/ {if ($3 ~
    /RNA$/) {$3="ncRNA"; print $0 >> "tmpx/"$1".gff3"}}'
    tmp/ResultadoFinal_C174-PHASED-corrigido.gff
6 cat tmpx/*.gff3 >
    tmp/ResultadoFinal_C174-PHASED-for-DensityMap.gff3
7 rm -rv tmpx

```

```

1 # TEs/Tgrand_C174P-EDTA.TEanno.gff3
2 mkdir -pv tmpx
3 awk 'BEGIN {FS=";";OFS=" "} /^chr[0-9]+[ab]/ {print
    "##gff-version 3" > "tmpx/"$1".gff3"}' output/chr.csv
4 awk 'BEGIN {FS=";";OFS=" "} /^chr[0-9]+[ab]/ {print
    "##sequence-region", $1, "1", $3 >> "tmpx/"$1".gff3"}'
    output/chr.csv
5 awk 'BEGIN {FS="\t";OFS="\t"} /^chr[0-9]{1,2}[ab]/ {$3="TE";
    print $0 >> "tmpx/"$1".gff3"}'
    TEs/Tgrand_C174P-EDTA.TEanno.gff3
6 cat tmpx/*.gff3 >
    tmp/Tgrand_C174P-EDTA.TEanno-for-DensityMap.gff3
7 rm -rv tmpx

```

Obter os CODING (Camada 1)

```

1 echo "chr;start;end;stack" > output/chart1_mRNA.csv
2 awk 'BEGIN {FS = "\t";OFS = ";"} /^chr[0-9]{1,2}[ab]/ {if
    ($3=="mRNA") {print $1,$4,$5,"mRNA"}}'
    ./annot-genes/Tgrand_C174P-Annotation.gff3 >>
    output/chart1_mRNA.csv
3
4 wc -l output/*_mRNA.csv

```

Obter os CODING com densidade (Camada 1)

```
1 DensityMap.pl -i tmp/Tgrand_C174P-Annotation-for-DensityMap.gff3
  -ty "mRNA=fused" -o tmp/chart1_mRNA.svg -sc 500000 -ro ceil
  -for -v
2
3 echo "chr;start;end;value" > output/chart1_mRNA-DensityMap.csv
4 awk 'BEGIN {FS = "\t";OFS = ";"} /^chr[0-9]{1,2}[ab]/ {print $1,
  $3, $4-1, $5}' tmp/chart1_mRNA.csv >>
  output/chart1_mRNA-DensityMap.csv
```

Obter os NON CODING (Camada 2)

```
1 echo "chr;start;end;stack" > output/chart1_ncRNA.csv
2 awk 'BEGIN {FS = "\t";OFS = ";"} /^chr[0-9]{1,2}[ab]/ {if ($3 ~
  /RNA$/) {print $1,$4,$5,"ncRNA"}}}'
  tmp/ResultadoFinal_C174-PHASED-corrigido.gff >>
  output/chart1_ncRNA.csv
3
4 grep -P "RNA\t" tmp/ResultadoFinal_C174-PHASED-corrigido.gff |
  wc -l
5 wc -l output/*_ncRNA.csv
```

Obter os NON CODING com densidade (Camada 2)

```
1 DensityMap.pl -i
  tmp/ResultadoFinal_C174-PHASED-for-DensityMap.gff3 -ty
  "ncRNA=fused" -o tmp/chart1_ncRNA.svg -sc 1000000 -ro ceil
  -for -v
2
3 echo "chr;start;end;value" > output/chart1_ncRNA-DensityMap.csv
4 awk 'BEGIN {FS = "\t";OFS = ";"} /^chr[0-9]{1,2}[ab]/ {print $1,
  $3, $4-1, $5}' tmp/chart1_ncRNA.csv >>
  output/chart1_ncRNA-DensityMap.csv
```

Obter os TEs (Camada 3)

```
1 echo "chr;start;end;stack" > output/chart1_TE.csv
2 awk 'BEGIN {FS = "\t";OFS = ";"} /^chr[0-9]{1,2}[ab]/ {print
  $1,$4,$5,"TE"}}' ./TEs/Tgrand_C174P-EDTA.TEanno.gff3 >>
  output/chart1_TE.csv
3
4 wc -l ./TEs/Tgrand_C174P-EDTA.TEanno.gff3
5 wc -l output/*_TE.csv
```

Obter os TEs com densidade (Camada 3)

```
1 DensityMap.pl -i
    tmp/Tgrand_C174P-EDTA.TEanno-for-DensityMap.gff3 -ty
    "TE=fused" -o tmp/chart1_TE.svg -sc 500000 -ro ceil -for -v
2
3 echo "chr;start;end;value" > output/chart1_TE-DensityMap.csv
4 awk 'BEGIN {FS = "\t";OFS = ";"} /^chr[0-9]{1,2}[ab]/ {print $1,
    $3, $4-1, $5}' tmp/chart1_TE.csv >>
    output/chart1_TE-DensityMap.csv
```

Anotações

```
1 awk '/^chr[0-9]{1,2}[ab]/ {print $3}' <
    ResultadoFinal_C174-PHASED.gff | sort | head
2 awk '/^chr[0-9]{1,2}[ab]/ {if ($3 ~ /RNA$/) {print $3}}'
    ResultadoFinal_C174-PHASED.gff | sort | uniq
3
4 # Obter tipos de TEs
5 awk '/^chr[0-9]{1,2}[ab]/ {print $3}'
    ./TEs/Tgrand_C174P-EDTA.TEanno.gff3 | sort | uniq
```