

Genome-Resolved Metagenomics of a Photosynthetic Bioreactor Performing Biological Nutrient Removal

 Elizabeth A. McDaniel,^a Renske Wever,^{c,d} Ben O. Oyserman,^{c,d} Daniel R. Noguera,^{b,e} Katherine D. McMahon^{a,b}

^aDepartment of Bacteriology, University of Wisconsin—Madison, Madison, Wisconsin, USA

^bDepartment of Civil and Environmental Engineering, University of Wisconsin—Madison, Madison, Wisconsin, USA

^cDepartment of Microbial Ecology, Netherlands Institute of Ecological Research, Wageningen, The Netherlands

^dBioinformatics Group, Wageningen University and Research, Wageningen, The Netherlands

^eDOE Great Lakes Bioenergy Research Center, Madison, Wisconsin, USA

ABSTRACT Enhanced biological phosphorus removal (EBPR) is an economically and environmentally significant wastewater treatment process for removing excess phosphorus by harnessing the metabolic physiologies of enriched microbial communities. We present a genome-resolved metagenomic data set consisting of 86 metagenome-assembled genome sequences from a photosynthetically operated lab-scale bioreactor simulating EBPR.

A photobioreactor performing light-supported enhanced biological phosphorus removal (EBPR) was seeded from two lab-scale wastewater treatment systems, a photosynthetic nitrifying enrichment culture and an activated sludge EBPR bioreactor designed to enrich for the polyphosphate-accumulating organism “*Candidatus Accumulibacter phosphatis*” (1, 2). The photobioreactor was designed to achieve phosphorus removal and denitrification without the addition of mechanical aeration by providing light (2). Three samples were collected during cycles 87, 103, and 129. DNA was extracted as described (2), and 100 ng of DNA was sheared to 300 bp using the Covaris LE220 ultrasonicator and size selected with solid-phase reversible immobilization (SPRI) beads (Beckman Coulter). The fragments were ligated with end repair, A-tailing, and Illumina-compatible adapters (IDT, Inc.) using the KAPA-Illumina library preparation kit (Kapa Biosystems). The libraries were quantified using the Kapa Biosystems next-generation sequencing library quantitative PCR (qPCR) kit and run on a Roche LightCycler 480 real-time PCR instrument. The quantified libraries were prepared using the v4 TruSeq paired-end cluster kit and the Illumina cBot instrument to create a clustered flow cell for sequencing. Shotgun metagenomic sequencing was performed on the Illumina HiSeq 2500 platform with the TruSeq sequencing-by-synthesis (SBS) sequencing kit, followed by 2 × 150-bp indexing. All metagenomic libraries consist of approximately 50 million 150-bp paired-end reads (table of metagenome information available at <https://doi.org/10.6084/m9.figshare.14164307>).

Default parameters were used for all software unless otherwise specified. The raw metagenomic reads were quality filtered and trimmed using bbdutk as part of the BBTools v38.07 suite (3). All three metagenomic samples were coassembled and individually assembled into contigs using SPAdes v3.9.0 with the metagenomic option (4). Metagenomic reads from each sample were mapped against all assemblies using BMap as part of the BBTools v38.07 suite with a 95% sequence identity cutoff (3). The assembled contigs from each assembly were binned into population genomes using MetaBAT2 v2.12.1 using only contigs larger than 1,000 bp (5). The bins were dereplicated using dRep v2.4.2 (6) to obtain the highest-quality representative set of genomes. This resulted in a total of 86 nonredundant species-resolved genome sequences, described in Table 1 and summarized in Fig. 1. All genome

Citation McDaniel EA, Wever R, Oyserman BO, Noguera DR, McMahon KD. 2021. Genome-resolved metagenomics of a photosynthetic bioreactor performing biological nutrient removal. Microbiol Resour Announc 10: e00244-21. <https://doi.org/10.1128/MRA.00244-21>.

Editor J. Cameron Thrash, University of Southern California

Copyright © 2021 McDaniel et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Elizabeth A. McDaniel, elizabethmcd93@gmail.com.

Received 12 March 2021

Accepted 31 March 2021

Published 6 May 2021

TABLE 1 Genome accession numbers and statistics^a

Isolate name	Assembly accession no.	Code	GTDB classification	Completeness (%)	Contamination (%)	Genome size (mbp)	No. of contigs	Avg. contig size (bp)	% GC	No. of rRNAs	No. of tRNAs	No. of 5S rRNAs	No. of 16S rRNAs	Relative abundance on date (mo/day/yr)			BioSample accession no.	GenBank accession no.	BioProject accession no.
														7/16/2015	8/6/2015	6/6/2015			
UWPOB_ACID01	GCA_017304135.1	ACID01	d_Bacterioplankton__Acidobacteriota; c_Acidobacteriota__Bryobacteriales; f_Bryobacteraceae_g__s	98.4	2.9	6.49	365	17,791	64.5	3	48	1	1	0.04386	0.06137	0.15309	SAMN18059832	JAFLEB0000000000	PRJNA704939
UWPOB_ACID02	GCA_017303955.1	ACID02	d_Bacterioplankton__Acidobacteriota; c_Acidobacteriota__UBA7656f_g__s	100	2.1	6.15	46	133,664	34.5	0	83	0	0	0.01576	0.0143	0.30104	SAMN18059833	JAFLEB0000000000	PRJNA704939
UWPOB_ARM01	GCA_017303935.1	ARMA1	d_Bacterioplankton__Armatimonadota; c_Armatimonadota; o_Fimbrimonadales; f_Fimbrimonadaceae_g__s	100	0	2.56	5	511,480	53.2	3	47	1	1	1.87686	0.06294	0.31483	SAMN18059834	JAFLEB0000000000	PRJNA704939
UWPOB_BACT1	GCA_017303995.1	BACT1	d_Bacterioplankton__Bacteroidetes; c_Bacteroidetes; o_Fimbrimonadales; f_Fimbrimonadaceae_g__s	100	0	4.31	137	31,460	41.5	2	40	2	0	0.12332	0.02724	0.08244	SAMN18059835	JAFLEB0000000000	PRJNA704939
UWPOB_BACT2	GCA_017303975.1	BACT2	d_Bacterioplankton__Bacteroidetes; c_Bacteroidetes; o_Fimbrimonadales; f_Fimbrimonadaceae_g__s	95.7	1	3.47	75	46,223	35.2	3	32	1	1	0.12227	0.02794	0.02315	SAMN18059837	JAFLEB0000000000	PRJNA704939
UWPOB_BACT3	GCA_017303895.1	BACT3	d_Bacterioplankton__Bacteroidetes; c_Bacteroidetes; o_Fimbrimonadales; f_Fimbrimonadaceae_g__s	99.1	0.7	3.61	35	103,001	37.3	3	33	1	1	0.10972	0.06722	0.02034	SAMN18059836	JAFLEB0000000000	PRJNA704939
UWPOB_CHIT1	GCA_017303905.1	CHIT1	d_Bacterioplankton__Chloroflexi; c_Chloroflexi; o_Fimbrimonadales; f_Fimbrimonadaceae_g__s	100	0	3.95	7	564,856	36.8	4	33	2	1	0.08039	0.12427	0.26855	SAMN18059838	JAFLEB0000000000	PRJNA704939
UWPOB_CHIT2	GCA_017303835.1	CHIT2	d_Bacterioplankton__Chloroflexi; c_Chloroflexi; o_Fimbrimonadales; f_Fimbrimonadaceae_g__s	99.5	0.3	3.26	2	1,631,117	42.5	3	36	1	1	2.73174	1.62266	3.05499	SAMN18059839	JAFLEB0000000000	PRJNA704939
UWPOB_CHIT3	GCA_017303415.1	CHIT3	d_Bacterioplankton__Chloroflexi; c_Chloroflexi; o_Fimbrimonadales; f_Fimbrimonadaceae_g__s	100	0	3.72	3	1,239,140	46	1	37	0	1	0.64668	0.61427	0.58909	SAMN18059840	JAFLEB0000000000	PRJNA704939
UWPOB_CHIT4	GCA_017303335.1	CHIT4	d_Bacterioplankton__Chloroflexi; c_Chloroflexi; o_Fimbrimonadales; f_Fimbrimonadaceae_g__s	99	0.3	4.01	7	572,270	49.5	3	42	1	1	0.64536	0.73815	1.22698	SAMN18059841	JAFLEB0000000000	PRJNA704939
UWPOB_CHIT5	GCA_017303455.1	CHIT5	d_Bacterioplankton__Chloroflexi; c_Chloroflexi; o_Fimbrimonadales; f_Fimbrimonadaceae_g__s	95.8	0	4.55	57	79,888	43	0	40	0	0	0.0768	0.06206	0.05275	SAMN18059843	JAFLEB0000000000	PRJNA704939
UWPOB_CHIT6	GCA_017303855.1	CHIT6	d_Bacterioplankton__Chloroflexi; c_Chloroflexi; o_Fimbrimonadales; f_Fimbrimonadaceae_g__s	99.5	0.7	4.62	31	149,027	46.1	1	37	1	0	0.23357	0.2022	0.15554	SAMN18059842	JAFLEB0000000000	PRJNA704939
UWPOB_CHIT7	GCA_017303395.1	CHIT7	d_Bacterioplankton__Chloroflexi; c_Chloroflexi; o_Fimbrimonadales; f_Fimbrimonadaceae_g__s	100	6.3	3.76	45	83,566	41.9	0	33	0	0	0.12824	0.08724	0.02842	SAMN18059844	JAFLEB0000000000	PRJNA704939
UWPOB_CHIT8	GCA_017303475.1	CHIT8	d_Bacterioplankton__Chloroflexi; c_Chloroflexi; o_Fimbrimonadales; f_Fimbrimonadaceae_g__s	98.5	0.5	6.04	53	114,008	47.9	3	40	1	1	0.83528	1.32521	1.47064	SAMN18059845	JAFLEB0000000000	PRJNA704939
UWPOB_CHIT9	GCA_017303875.1	CHIT9	d_Bacterioplankton__Chloroflexi; c_Chloroflexi; o_Fimbrimonadales; f_Fimbrimonadaceae_g__s	99	0.5	5.78	64	90,342	50.4	0	43	0	0	0.12036	0.15772	0.42621	SAMN18059846	JAFLEB0000000000	PRJNA704939
UWPOB_CYT01	GCA_017303805.1	CYT01	d_Bacterioplankton__Cyanobacteria; c_Cyanobacteria; o_Synechococcales; f_Synechococcaceae_g__s	98.5	0.3	4.29	13	329,632	45.1	1	35	1	0	0.03105	0.06252	0.27364	SAMN18059847	JAFLEB0000000000	PRJNA704939
UWPOB_CYT02	GCA_017303795.1	CYT02	d_Bacterioplankton__Cyanobacteria; c_Cyanobacteria; o_Synechococcales; f_Synechococcaceae_g__s	100	0	4.87	27	180,295	42.2	2	34	1	0	0.06209	0.0432	0.20485	SAMN18059848	JAFLEB0000000000	PRJNA704939
UWPOB_FLAV01	GCA_017302935.1	FLAV01	d_Bacterioplankton__Flavobacteriia; c_Flavobacteriia; o_Flavobacteriales; f_Flavobacteriaceae_g__s	93.2	0.8	3.09	264	11,718	34	2	40	2	0	0.04808	0.01753	0.05116	SAMN18059850	JAFLEB0000000000	PRJNA704939

(Continued on next page)

Downloaded from <http://mra.asm.org/> on May 6, 2021 at Univ of Wisconsin - Madison

(Continued on next page)

Volume 10 Issue 18 e00244-21

Isolate name	Assembly accession no.	Code	GTDB classification	Completeness (%)	Contamination (%)	Genome size (mbp)	No. of contigs	Avg. contig size (bp)	% GC content	No. of rRNAs	No. of 5S rRNAs	No. of 16S rRNAs	Relative abundance on date (mo/day/yr)				Avg abundance	BioSample accession no.	GenBank accession no.	BioProject accession no.
													7/16/2015	7/24/2015	8/6/2015	11/05/21				
JWPOB_OB51	GCA_017303615.1	OB51	d__Bacteri__Cyanobacteria; c__Vampirovibrionia; o__Obscuribacteriales; f__Obscuribacteraceae; g__Obscuribacter; d__Bacteri__Mycrococota; c__Polyangia__Polyangiales; f__Sindaracinaeae; s__	96.6	0.9	5.06	70	72,324	49	3	42	1	1	0.22519	1.10521	0.64416	SAMN18059867	JAFLC0000000000	PRJNA704939	
JWPOB_DELTAS	GCA_017303575.1	DELTA5	d__Bacteri__Mycrococota; c__Polyangia__Polyangiales; f__Sindaracinaeae; s__	95.8	0	9.39	540	17,383	70.1	3	71	1	1	0.04541	0.10081	0.34331	SAMN18059868	JAFLC0000000000	PRJNA704939	
JWPOB_PLANC1	GCA_017303565.1	PLANC1	d__Bacteri__Planctomycetota; c__Phycisphaerae__Phycisphaerales; f__SM1A02g__s__	88.3	6.3	2.93	530	5,521	68.6	1	37	0	1	0.02712	0.0326	0.0993	SAMN18059869	JAFLC0000000000	PRJNA704939	
JWPOB_PLANC2	GCA_017303555.1	PLANC2	d__Bacteri__Planctomycetota; c__Phycisphaerae__Phycisphaerales; f__SM1A02g__UBA2402s__	95.8	0	4.06	42	96,645	63.5	3	48	1	1	0.35589	0.44712	0.11479	SAMN18059870	JAFLCN0000000000	PRJNA704939	
JWPOB_PLAC3	GCA_017303535.1	PLANC3	d__Bacteri__Planctomycetota; c__Planctomycetes__Pirellulales; f__s__	93.5	1.3	6.61	641	10,313	61.8	3	71	1	1	0.02337	0.03888	0.08353	SAMN18059871	JAFLC0000000000	PRJNA704939	
JWPOB_PLANC4	GCA_017303505.1	PLANC4	d__Bacteri__Planctomycetota; c__Planctomycetes__Pirellulales; f__Pirellulaceae__Pirellula_Bs__	96.4	0.2	7.83	308	25,424	50.3	0	98	0	0	0.02843	0.10226	0.04436	SAMN18059872	JAFLC0000000000	PRJNA704939	
JWPOB_PLANC5	GCA_017302815.1	PLANC5	d__Bacteri__Planctomycetota; c__UBA11346o__UBA11346; f__UBA11346g__UBA11346s__	78	2	4.28	568	7,540	69.7	0	74	0	0	0.14524	0.27428	0.04468	SAMN18059873	JAFLC0000000000	PRJNA704939	
JWPOB_ALPHA1	GCA_017302755.1	ALPHA1	d__Bacteri__Proteobacteria; c__Alphaproteobacteria__f__g__s__	94.8	0	2.12	275	7,722	59.4	2	35	1	0	1	0.00365	0.0307	0.09852	SAMN18059874	JAFLC0000000000	PRJNA704939
JWPOB_CAULO1	GCA_017302335.1	CAULO1	d__Bacteri__Proteobacteria; c__Alphaproteobacteria; o__Caulobacteriales; f__Caulobacteraceae; g__Brevundimonas; s__	96.6	0.7	2.6	168	15,478	67.9	1	44	0	1	0	0.29751	0.15141	0.02773	SAMN18059876	JAFLC0000000000	PRJNA704939
JWPOB_CAULO2	GCA_017302105.1	CAULO2	d__Bacteri__Proteobacteria; c__Alphaproteobacteria; o__Caulobacteriales; f__Caulobacteraceae; g__Brevundimonas; s__	100	0	2.57	12	214,253	65.7	3	38	1	1	1.58091	0.90968	0.11758	SAMN18059875	JAFLC0000000000	PRJNA704939	
JWPOB_CAULO3	GCA_017303355.1	CAULO3	d__Bacteri__Proteobacteria; c__Alphaproteobacteria; o__Caulobacteriales; f__Caulobacteraceae; g__Brevundimonas; s__	98.4	0.7	3.67	34	107,997	63.9	3	47	1	1	0.9416	0.71847	0.41979	SAMN18059877	JAFLC0000000000	PRJNA704939	
JWPOB_CAULO4	GCA_017303435.1	CAULO4	d__Bacteri__Proteobacteria; c__Alphaproteobacteria; o__Caulobacteriales; f__Hyphomonadaceae__UBA1942; s__	75	6.3	3.49	362	9,650	39.2	0	35	0	0	0.09806	0.03402	0.00279	SAMN18059878	JAFLCV0000000000	PRJNA704939	
JWPOB_MICAV1	GCA_017302165.1	MICAV1	d__Bacteri__Proteobacteria; c__Alphaproteobacteria; o__Micavibrionales; f__Micavibrionaceae; s__	96	0.2	2	6	333,677	49.3	2	40	0	1	1	0.02039	0.01146	0.35063	SAMN18059879	JAFLCV0000000000	PRJNA704939
JWPOB_RHZO1	GCA_017302635.1	RHZO1	d__Bacteri__Proteobacteria; c__Alphaproteobacteria; o__Rhizobiales__Beijerinckaceae_A; g__PAR1s__	97.3	0.6	3.1	6	516,587	62.1	3	47	1	1	0.21701	0.24399	0.35215	SAMN18059880	JAFLC0000000000	PRJNA704939	
JWPOB_RHODOB1	GCA_017303485.1	RHODOB1	d__Bacteri__Proteobacteria; c__Alphaproteobacteria; o__Rhodobacteriales; f__Rhodobacteraceae__Tabrizicola; s__	96.6	8.6	4.39	272	16,121	65.3	0	53	0	0	0.06267	0.10359	0.03424	SAMN18059881	JAFLCV0000000000	PRJNA704939	
JWPOB_RICK1	GCA_017302665.1	RICK1	d__Bacteri__Proteobacteria; c__Alphaproteobacteria; o__Rickettsiales__Rickettsiaceae; s__	98.6	7.1	1.65	29	56,773	33.3	4	34	1	1	2	0.00855	0.16939	0.03581	SAMN18059882	JAFLC2000000000	PRJNA704939

Downloaded from <http://mra.asm.org/> on May 6, 2021 at Univ of Wisconsin - Madison

Volume 10 Issue 18 e00244-21

Isolate name	Assembly accession no.	Code	GTDB classification	Completeness (%)	Contamination (%)	Genome size (mbp)	No. of contigs	Avg contig size (bp)	% GC content	No. of rRNAs	No. of tRNAs	Relative abundance on date (mo/day/yr)									
												2/25	7/16/2015	7/24/2015	8/6/2015	Avg abundance	BioSample accession no.	GenBank accession no.	BioProject accession no.		
JWPOB_RICK2	GCA_017302595.1	RICK2	d__Bacteriap__Proteobacteria; c__Alphaproteobacteria; o__Rickettsialesf__Rickettsiaceae; g__GCA_2402195f__d_Bacteriap__Proteobacteria;	99.5	1	1.33	5	264,993	34.7	0	34	0	0	0	0.03107	0.20695	0.08658	0.1082	SAMN18059883	JAFLEX000000000	PRINA704939
JWPOB_SPHING1	GCA_017302775.1	SPHING1	d__Bacteriap__Proteobacteria; c__Alphaproteobacteria; o__Sphingomonadales; f__Sphingomonadaceae; g__Novosphingobiumsf__d_Bacteriap__Proteobacteria;	78.5	5.1	3.16	556	5,684	61.6	0	48	0	0	0	0.06166	0.0559	0.01391	0.04516	SAMN18059885	JAFLEX000000000	PRINA704939
JWPOB_SPHING2	GCA_017302615.1	SPHING2	c__Alphaproteobacteria; o__Sphingomonadales; f__Sphingomonadaceae; g__Novosphingobiumsf__d_Bacteriap__Proteobacteria;	99.2	0.1	3.02	33	91,486	65.7	3	46	1	1	1	0.6247	0.62665	0.23261	0.49465	SAMN18059884	JAFLEX000000000	PRINA704939
JWPOB_ALPHA2	GCA_017302573.1	ALPHA2	f__Burkholderiales; o__Burkholderiales; f__Burkholderiaceae; g__Ottowiaf__Burkholderiaceae; s__Ottowia	98.9	0	1.86	1	1,861,815	50.8	3	39	1	1	1	0.47594	0.19689	0.04249	0.23844	SAMN18059886	JAFLEX000000000	PRINA704939
JWPOB_BURK1	GCA_017302655.1	BURK1	c__Alphaproteobacteria; o__UBA998; f__UBA3002;q__S	100	0	4.44	203	21,857	67.3	2	48	1	1	0	0.37852	0.42244	0.55223	0.45106	SAMN18059887	JAFLEX000000000	PRINA704939
JWPOB_OTTO1	GCA_017302725.1	OTTO1	c__Gammaproteobacteria; o__Burkholderiales; f__Burkholderiaceae; g__Ottowiaf__Burkholderiaceae; s__Ottowia	95.8	0	2.87	325	8,835	68.6	3	52	1	1	1	0.76562	0.83948	0.30961	0.63823	SAMN18059888	JAFLEX000000000	PRINA704939
JWPOB_RUBR1	GCA_017302495.1	RUBR1	d__Bacteriap__Proteobacteria; c__Gammaproteobacteria; o__Burkholderiales; f__Burkholderiaceae; g__Ottowiaf__Burkholderiaceae; s__Ottowia	81	0.8	4.51	553	8,152	71.1	3	56	1	1	1	1.62184	1.49067	0.45613	1.18955	SAMN18059889	JAFLEX000000000	PRINA704939
JWPOB_RUBR2	GCA_017302505.1	RUBR2	f__Burkholderiales; o__Burkholderiales; f__Burkholderiaceae; g__Rubrivivaxsf__Burkholderiaceae; s__Rubrivivax	100	0	5.5	555	9,913	68.4	4	52	2	1	1	0.21114	0.24105	0.10683	0.18634	SAMN18059890	JAFLEX000000000	PRINA704939
JWPOB_VITRI1	GCA_017302485.1	VITRI1	d__Bacteriap__Proteobacteria; c__Gammaproteobacteria; o__Burkholderiales; f__Burkholderiaceae; g__Vitreoscilla_A; s__Vitreoscilla_A	95.8	0	3.68	513	7,177	68.4	1	42	1	0	0	0.07383	0.05699	0.17722	0.10268	SAMN18059891	JAFLEX000000000	PRINA704939
JWPOB_BURK2	GCA_017302475.1	BURK2	d__Bacteriap__Proteobacteria; c__Gammaproteobacteria; o__Burkholderialesf__Palaeo-1005;g__S	79.2	0	2.71	498	5,448	69.6	0	34	0	0	0	0.10158	0.12158	0.11145	0.11154	SAMN18059892	JAFLEX000000000	PRINA704939
JWI10-POB	GCA_017302555.1	CAPIB	d__Bacteriap__Proteobacteria; c__Gammaproteobacteria; o__Burkholderiales; f__Rhodocyclaceae; g__Accumulibacterf__d_Bacteriap__Proteobacteria;	100	0	4.4	88	49,949	62.5	0	41	0	0	0	0.52449	0.60697	0.65232	0.59459	SAMN18059893	JAFLEX000000000	PRINA704939
JWI12-POB	GCA_017302435.1	CAPID	c__Gammaproteobacteria; o__Burkholderiales; f__Rhodocyclaceae; g__Accumulibacterf__d_Bacteriap__Proteobacteria;	98.1	0.7	4.56	91	50,155	62.7	0	46	0	0	0	0.35414	0.32156	0.05167	0.24246	SAMN18059894	JAFLEX000000000	PRINA704939
JWI13-POB	GCA_017302415.1	CAPIF	c__Gammaproteobacteria; o__Burkholderiales; f__Rhodocyclaceae; g__Accumulibacterf__d_Bacteriap__Proteobacteria;	97.1	4.4	5.45	245	22,232	65.9	1	48	0	1	0	1.65004	0.70231	0.09466	0.81567	SAMN18059895	JAFLEX000000000	PRINA704939

Downloaded from <http://mra.asm.org/> on May 6, 2021 at Univ of Wisconsin - Madison

Volume 10 Issue 18 e00244-21

Isolate name	Assembly accession no.	Code	GTDB classification	Completeness (%)	Contamination (%)	Genome size (mbp)	No. of contigs	Avg. contig size (bp)	% GC content	No. of rRNAs	No. of tRNAs	No. of 5S rRNAs	No. of 16S rRNAs	Relative abundance on date (mo/day/yr)					Avg abundance	BioSample accession no.	GenBank accession no.	BioProject accession no.				
														7/16/2015	7/24/2015	8/6/2015	10/23/2015	11/4/2015								
JW11-POB	GCA_017302385.1	CAPIC	d__Bacteriopl__Proteobacteria; o__Burkholderiales; f__Rhodocyclaceae; g__Accumulibacter__Accumulibacter	100	0	4.63	144	32,127	61.2	0	47	0	0	12.2117	11.8669	10.2307	11.4364		SAMN18059898	JAFLDP0000000000	PRJNA704939					
JW8-POB	GCA_017302345.1	CAPIA	d__Bacteriopl__Proteobacteria; c__Gammaproteobacteria; o__Burkholderiales; f__Rhodocyclaceae; g__Accumulibacter__Accumulibacter	100	0	4.26	49	86,993	64.2	0	51	0	0	6.34328	6.19256	0.4336	4.32315		SAMN18059896	JAFLDN0000000000	PRJNA704939					
JW9-POB	GCA_017302455.1	CAPIA	d__Bacteriopl__Proteobacteria; c__Gammaproteobacteria; o__Burkholderiales; f__Rhodocyclaceae; g__Accumulibacter__Accumulibacter	100	0	4.93	126	39,138	64.1	1	46	1	0	0.55782	0.54654	0.15933	0.42123		SAMN18059897	JAFLDQ0000000000	PRJNA704939					
JWPOB_DECH1	GCA_017302355.1	DECH1	d__Bacteriopl__Proteobacteria; c__Gammaproteobacteria; o__Burkholderiales; f__Rhodocyclaceae; g__Accumulibacter__Accumulibacter	91.5	8.1	3.69	279	13,228	65	2	40	0	1	2.09698	3.33708	3.17482	2.86963		SAMN18059899	JAFLDQ0000000000	PRJNA704939					
JWPOB_SULF1	GCA_017302275.1	SULF1	d__Bacteriopl__Proteobacteria; c__Gammaproteobacteria; o__Burkholderiales; f__Rhodocyclaceae; g__Dechloromonas	98.3	1.1	4.41	88	50,106	64	3	47	2	1	0	0.76164	0.92706	2.2433	1.31067		SAMN18059900	JAFLDR0000000000	PRJNA704939				
JWPOB_THAU1	GCA_017302295.1	THAU1	d__Bacteriopl__Proteobacteria; c__Gammaproteobacteria; o__Burkholderiales; f__Rhodocyclaceae; g__Sulfuritalea	98.7	1.4	4.11	56	73,417	66.1	3	52	3	0	0	0.5635	0.57917	0.40828	0.51699		SAMN18059901	JAFLD5000000000	PRJNA704939				
JWPOB_ZOOG1	GCA_017309145.1	ZOOG1	d__Bacteriopl__Proteobacteria; c__Gammaproteobacteria; o__Burkholderiales; f__Rhodocyclaceae; g__Thauera	83.3	4.2	4.91	428	11,461	65.7	0	58	0	0	0	0.06958	0.06569	0.07688	0.07071		SAMN18059903	JAFLDU0000000000	PRJNA704939				
JWPOB_ZOOG2	GCA_017302315.1	ZOOG2	d__Bacteriopl__Proteobacteria; c__Gammaproteobacteria; o__Burkholderiales; f__Rhodocyclaceae; g__Zoogloea	95.2	1.8	4.52	165	27,407	64.5	0	55	0	0	0	0.12938	0.14704	0.21305	0.16316		SAMN18059902	JAFLDT0000000000	PRJNA704939				
JWPOB_PARA1	GCA_017302255.1	PARA1	d__Bacteriopl__Proteobacteria; c__Gammaproteobacteria; o__Enterobacterales; f__Alteromonadaceae; g__Parahelminthina	100	4.2	4.03	77	52,358	46.2	0	50	0	0	0	0.20334	0.02657	0.00492	0.07828		SAMN18059904	JAFLDV0000000000	PRJNA704939				
JWPOB_GAMMA1	GCA_017302205.1	GAMMA1	d__Bacteriopl__Proteobacteria; c__Gammaproteobacteria; o__Xanthomonadales; f__Rhodanobacteraceae; g__Dokdonella	79.2	1	2.51	456	5,508	69.2	1	34	1	0	0	0.05103	0.07089	0.10094	0.07429		SAMN18059905	JAFLDW0000000000	PRJNA704939				
JWPOB_RHODAN1	GCA_017302215.1	RHODAN1	d__Bacteriopl__Proteobacteria; c__Gammaproteobacteria; o__Xanthomonadales; f__Rhodanobacteraceae; g__Dokdonella	91.7	0	3.5	323	10,843	68.8	0	49	0	0	0	0.144	0.20595	0.16775	0.17257		SAMN18059906	JAFLDX0000000000	PRJNA704939				
JWPOB_AQU1	GCA_017302155.1	AQU1	d__Bacteriopl__Proteobacteria; c__Gammaproteobacteria; o__Xanthomonadales; f__Xanthomonadaceae; g__Aquimona	98.3	1.6	3.98	113	35,212	61.5	0	46	0	0	0	0.28393	0.24393	0.06593	0.19793		SAMN18059907	JAFLDY0000000000	PRJNA704939				
JWPOB_LYSO1	GCA_017302135.1	LYSO1	d__Bacteriopl__Proteobacteria; c__Gammaproteobacteria; o__Xanthomonadales; f__Xanthomonadaceae; g__Lysobacter	99.6	1	4.34	35	123,959	64.1	2	48	1	1	0	1.21091	1.20058	0.96205	1.12451		SAMN18059908	JAFLDZ0000000000	PRJNA704939				

Downloaded from <http://mra.asm.org/> on May 6, 2021 at Univ of Wisconsin - Madison

TABLE 1 (Continued)

Isolate name	Assembly accession no.	Code	GTDB classification	Completeness (%)	Contamination (%)	Genome size (mbp)	No. of contigs	Avg contig size (bp)	% GC content	No. of rRNAs	No. of tRNAs	Relative abundance on date (mo/day/yr)					Avg abundance	BioSample accession no.	GenBank accession no.	BioProject accession no.	
												7/16/2015	7/24/2015	8/6/2015	235						
UWPOB_PSEUDO1	GCA_017302035.1	PSEUDO1	d__Bacteriaph__Proteobacteria; c__Gammaproteobacteria; o__Xanthomonadales; f__Xanthomonadaceae; g__Pseudoxanthomonas;	95.8	0	2.91	125	23,300	69.3	1	48	1	0	0	1.50336	2.17593	0.47299	1.38409	SAMN18059909	JAFLEA0000000000	PRJNA704939
UWPOB_THERMO1	GCA_017302095.1	THERMO1	d__Bacteriaph__Proteobacteria; c__Gammaproteobacteria; o__Xanthomonadales; f__Xanthomonadaceae; g__Thermomonas;	76.4	2.1	2.31	299	7,734	69.7	0	39	0	0	0	0.21965	0.25626	0.05906	0.17832	SAMN18059911	JAFLEC0000000000	PRJNA704939
UWPOB_THERMO2	GCA_017302075.1	THERMO2	d__Bacteriaph__Proteobacteria; c__Gammaproteobacteria; o__Xanthomonadales; f__Xanthomonadaceae; g__Thermomonas;	87.5	0.5	2.77	245	11,303	69.9	0	41	0	0	0	0.79368	1.14049	0.4586	0.79759	SAMN18059910	JAFLEB0000000000	PRJNA704939
UWPOB_SPIRO1	GCA_017302045.1	SPIRO1	d__Bacteriaph__Spirochaetota; c__Leptospiraeo__Turnerellales; f__Turneriellaceae; g__Turneriella;	86.5	0	3.78	412	9,184	53.8	2	31	2	0	0	0.03758	0.03209	0.0621	0.04392	SAMN18059912	JAFLED0000000000	PRJNA704939
UWPOB_SPIRO2	GCA_017302015.1	SPIRO2	d__Bacteriaph__Spirochaetota; c__UBA12135to__f__g__s__	87.5	0	5.43	173	31,368	65.2	2	41	0	1	1	0.09083	0.09013	0.07872	0.08656	SAMN18059913	JAFLEE0000000000	PRJNA704939
UWPOB_VERRUCO1	GCA_017302195.1	VERRUCO1	d__Bacteriaph__Verrucomicrobiota; c__Verrucomicrobiae; o__Pedosphaerales; f__UBA9464; g__UBA9464s;	81	8.3	4.86	848	5,726	67.1	0	35	0	0	0	0.04712	0.07195	0.01614	0.04507	SAMN18059915	JAFLEG0000000000	PRJNA704939
UWPOB_VERRUCO2	GCA_017309115.1	VERRUCO2	d__Bacteriaph__Verrucomicrobiota; c__Verrucomicrobiae; o__Pedosphaerales; f__UBA9464; g__UBA9464s;	98.6	3.5	7.53	112	67,228	61.5	3	50	1	1	1	0.14886	0.18964	0.1474	0.16196	SAMN18059914	JAFLEF0000000000	PRJNA704939
UWPOB_VERRUCO3	GCA_017301995.1	VERRUCO3	d__Bacteriaph__Verrucomicrobiota; c__Verrucomicrobiae; o__Verrucomicrobiales; f__Alkermansiaaceae; g__UBA1315s;	100	0	7.04	68	103,585	62.3	2	60	2	0	0	0.30515	0.26629	0.07792	0.21646	SAMN18059916	JAFLEH0000000000	PRJNA704939
UWPOB_VERRUCO4	GCA_017301975.1	VERRUCO4	d__Bacteriaph__Verrucomicrobiota; c__Verrucomicrobiae; o__Verrucomicrobiales; f__Verrucomicrobiaceae; g__Prostheobacteria;	97	0	6.9	97	71,172	60.3	0	54	0	0	0	0.17845	0.15859	0.07204	0.13636	SAMN18059917	JAFLEI0000000000	PRJNA704939

^a Genome information for 86 metagenome-assembled genomes is shown. The column labeled "Isolate name" lists the name submitted to NCBI, and that labeled "Code" lists the codes used in Fig. 1. Classifications were assigned with GTDB-tk and "Candidatus Accumulibacter phosphatis" clade designations based on comparisons to *ppk1* clones and publicly available reference genome sequences. "Candidatus Accumulibacter phosphatis" genomes are named alphanumerically according to previous nomenclature; all other genomes are named UWPOB_CODE. The genome statistics for completeness, contamination, GC content, genome size, and number of contigs were performed with CheckM v1.1.2. The rRNA and tRNA genes were predicted with Barrnap v0.9, as part of the Prokka v1.13.7 package. The relative abundance was calculated using coverM v0.4.0 using the relative abundance method for the three metagenomic samples, taken on 16 July 2015, 24 July 2015, and 6 August 2015 toward the end of the enrichment period, and averaged together across all three samples for an average relative abundance calculation.

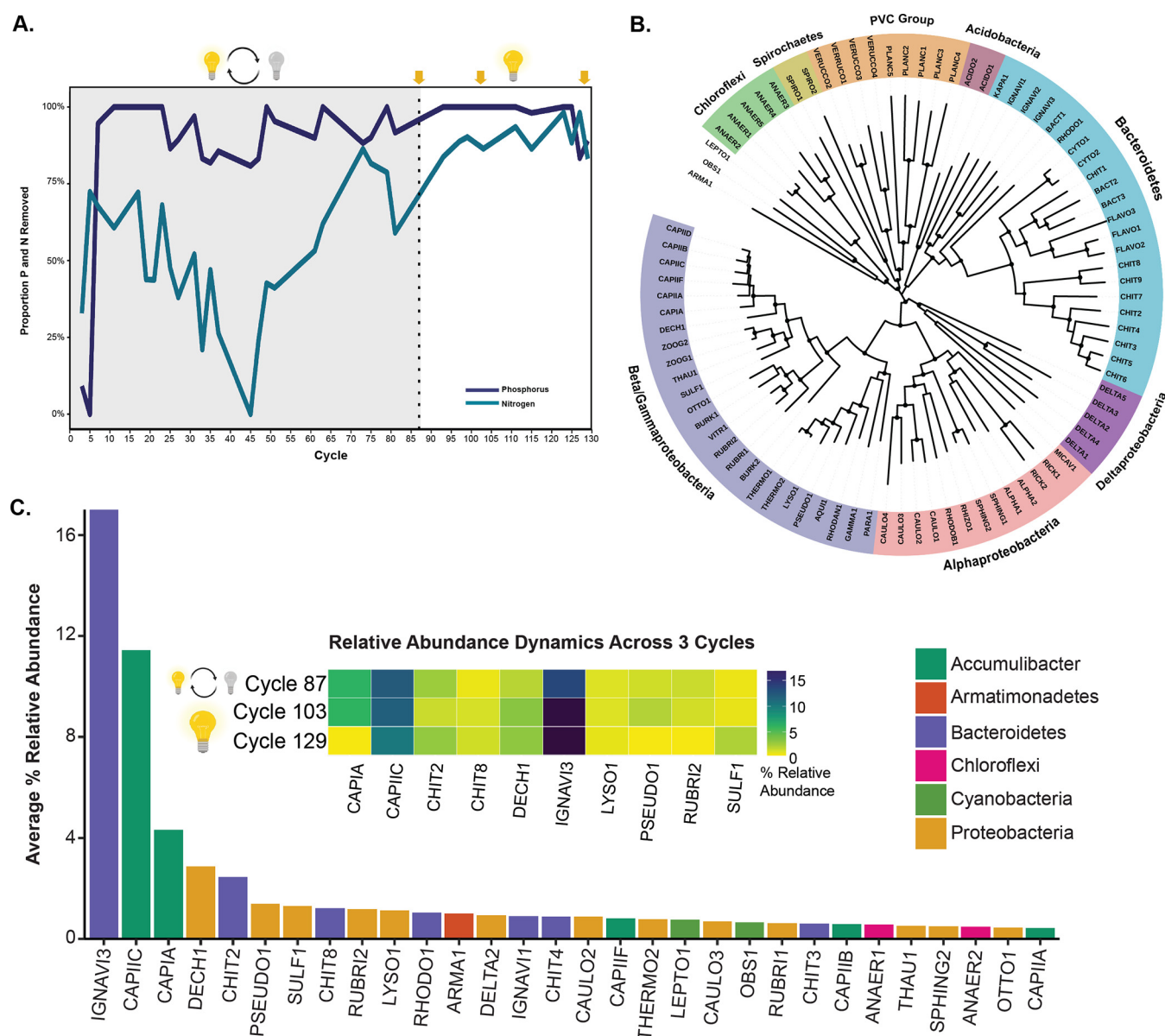


FIG 1 Overview of reactor performance and assembled MAGs. (A) Proportions of nitrogen (blue) and phosphorus (purple) removed during reactor operation. Cycle length refers to the hydraulic residence time of 0.625 days (2). The reactor was operated in two phases—in the first phase (gray), light was cycled on and off, and in the second phase (white), light was provided continuously. The arrows denote the time points at which metagenomic sequencing was performed. (B) Phylogenetic tree of all 86 assembled MAGs constructed from a concatenated alignment of 120 single-copy marker genes from GTDB-tk v0.3.2 (9) that were identified and aligned using HMMER v3.2.1 (11). The tree was constructed using RAxML v8.2.12 with 100 rapid bootstraps (12) and viewed using iTOL v5.7 (13). (C) Relative abundance and population dynamics of abundant species represented by assembled MAGs. The bar graph represents the average relative abundance of the top 30 most abundant species across all three samples. Relative abundance calculations were made using coverM v0.4.0 with the relative_abundance method and averaged across all three samples. The inset heat map shows the population dynamics of the top 10 most abundant species, representing the relative abundance of that MAG in that sample.

sequences were checked for uniform differential coverage using uBin v0.9.14 (7). All genome statistics were calculated with CheckM v1.1.2 (8). Taxonomical classifications were made based on the Genome Taxonomy Database (GTDB) using GTDB-Tk v0.3.2 (9, 10). Relative abundance calculations were performed with coverM v0.4.0 using the relative_abundance calculation method. The methods and phylogenetic tree for assigning clades of six “*Ca. Accumulibacter phosphatis*” draft genome sequences based on the *ppk1* locus are available at <https://doi.org/10.6084/m9.figshare.14164478>.

Data availability. The raw metagenomes for the 3 samples and genome assemblies for all 86 metagenome-assembled genomes (MAGs) are available at NCBI GenBank

under BioProject accession number [PRJNA704939](https://ncbi.nlm.nih.gov/bioproject/PRJNA704939). The metagenomes are available under the SRA accession numbers [SRR13786854](https://ncbi.nlm.nih.gov/sra/SRR13786854), [SRR13786855](https://ncbi.nlm.nih.gov/sra/SRR13786855), and [SRR13786856](https://ncbi.nlm.nih.gov/sra/SRR13786856).

ACKNOWLEDGMENTS

This work was supported by funding from the National Science Foundation (MCB-1518130) to K.D.M. and D.R.N.

We thank the University of Wisconsin—Madison Biotechnology Center DNA Sequencing Facility for Illumina sequencing services. This research was performed in part using the Wisconsin Energy Institute computing cluster, which is supported by the Great Lakes Bioenergy Research Center as part of the U.S. Department of Energy Office of Science (DE-SC0018409).

REFERENCES

- Martin HG, Ivanova N, Kunin V, Warnecke F, Barry KW, McHardy AC, Yeates C, He S, Salamov AA, Szeto E, Dalin E, Putnam NH, Shapiro HJ, Pangilinan JL, Rigoutsos I, Kyrpides NC, Blackall LL, McMahon KD, Hugenholtz P. 2006. Metagenomic analysis of two enhanced biological phosphorus removal (EBPR) sludge communities. *Nat Biotechnol* 24:1263–1269. <https://doi.org/10.1038/nbt1247>.
- Oyserman BO, Martirano JM, Wipperfurth S, Owen BR, Noguera DR, McMahon KD. 2017. Community assembly and ecology of activated sludge under photo-synthetic feast-famine conditions. *Environ Sci Technol* 51:3165–3175. <https://doi.org/10.1021/acs.est.6b03976>.
- Bushnell B, Rood J, Singer E. 2017. BBMerge—accurate paired shotgun read merging via overlap. *PLoS One* 12:e0185056. <https://doi.org/10.1371/journal.pone.0185056>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Kang DD, Froula J, Egan R, Wang Z. 2015. MetaBAT, an efficient tool for accurately reconstructing single genomes from complex microbial communities. *PeerJ* 3:e1165. <https://doi.org/10.7717/peerj.1165>.
- Olm MR, Brown CT, Brooks B, Banfield JF. 2017. dRep: a tool for fast and accurate genomic comparisons that enables improved genome recovery from metagenomes through de-replication. *ISME J* 11:2864–2868. <https://doi.org/10.1038/ismej.2017.126>.
- Bornemann TLV, Esser SP, Stach TL, Burg T, Probst AJ. 2020. uBin—a manual refining tool for metagenomic bins designed for educational purposes. *bioRxiv*. <https://doi.org/10.1101/2020.07.15.204776>.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
- Chaumeil P-A, Mussig AJ, Hugenholtz P, Parks DH. 2019. GTDB-Tk: a tool-kit to classify genomes with the Genome Taxonomy Database. *Bioinformatics* 36:1925–1927. <https://doi.org/10.1093/bioinformatics/btz848>.
- Parks DH, Chuvochina M, Waite DW, Rinke C, Skarshewski A, Chaumeil P-A, Hugenholtz P. 2018. A standardized bacterial taxonomy based on genome phylogeny substantially revises the tree of life. *Nat Biotechnol* 36:996–1004. <https://doi.org/10.1038/nbt.4229>.
- Eddy SR. 2011. Accelerated profile HMM searches. *PLoS Comput Biol* 7:e1002195. <https://doi.org/10.1371/journal.pcbi.1002195>.
- Stamatakis A. 2014. The RAxML v8.0.X manual. <https://cme.h-its.org/exelixis/resource/download/NewManual.pdf>.
- Letunic I, Bork P. 2016. Interactive tree of life (iTOL) v3: an online tool for the display and annotation of phylogenetic and other trees. *Nucleic Acids Res* 44:W242–W245. <https://doi.org/10.1093/nar/gkw290>.