

Introduction to scRNA-seq integration

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scRNA-seq Integration with GABA1 and GABA2 data sets

```
library(Seurat)
library(patchwork)
```

Create Seurat objects from the filtered cell ranger data

```
experiment_name <- "C. elegans scRNA-seq"
dataset_loc <- "../ackley_data"
ids <- c("GABA1", "GABA2")

d10x.data <- lapply(ids, function(i) {
  data <- Read10X(data.dir = file.path(dataset_loc, i, "outs", "filtered_feature_bc_matrix"))
})
names(d10x.data) <- ids

subject.list <- lapply(ids, function(i) {
  subject <- CreateSeuratObject(counts = d10x.data[[i]])
  # remember which id the data belongs to (useful after integration)
  subject[["group"]] <- rep(i, times = length(subject@meta.data$nCount_RNA))
  # Does the 'MT' prefix signify mitochondrial DNA in C. elegans and does it need to be
  # filtered out?
  subject[["percent.mt"]] <- PercentageFeatureSet(subject, pattern = "^MT")
  subject
})
names(subject.list) <- ids
```

Normalization

```
# normalize and identify variable features for each dataset independently
subject.list <- lapply(X = subject.list, FUN = function(x) {
  x <- NormalizeData(x)
  x <- FindVariableFeatures(x, selection.method = "vst", nfeatures = 2000)
})

# select features that are repeatedly variable across datasets for integration
features <- SelectIntegrationFeatures(object.list = subject.list)
```

Perform integration

```
grouped.anchors <- FindIntegrationAnchors(object.list = subject.list, anchor.features = features)
```

```
# this command creates an 'integrated' data assay  
grouped.combined <- IntegrateData(anchorset = grouped.anchors)
```

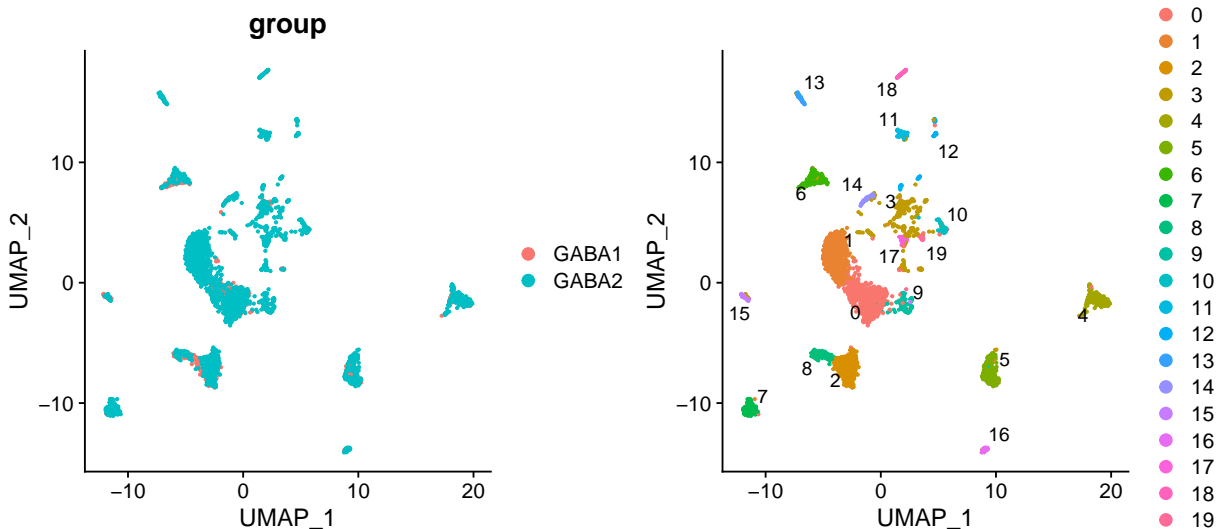
Perform an integrated analysis

Now we can run a single integrated analysis on all cells!

```
# specify that we will perform downstream analysis on the corrected data note that the  
# original unmodified data still resides in the 'RNA' assay  
DefaultAssay(grouped.combined) <- "integrated"
```

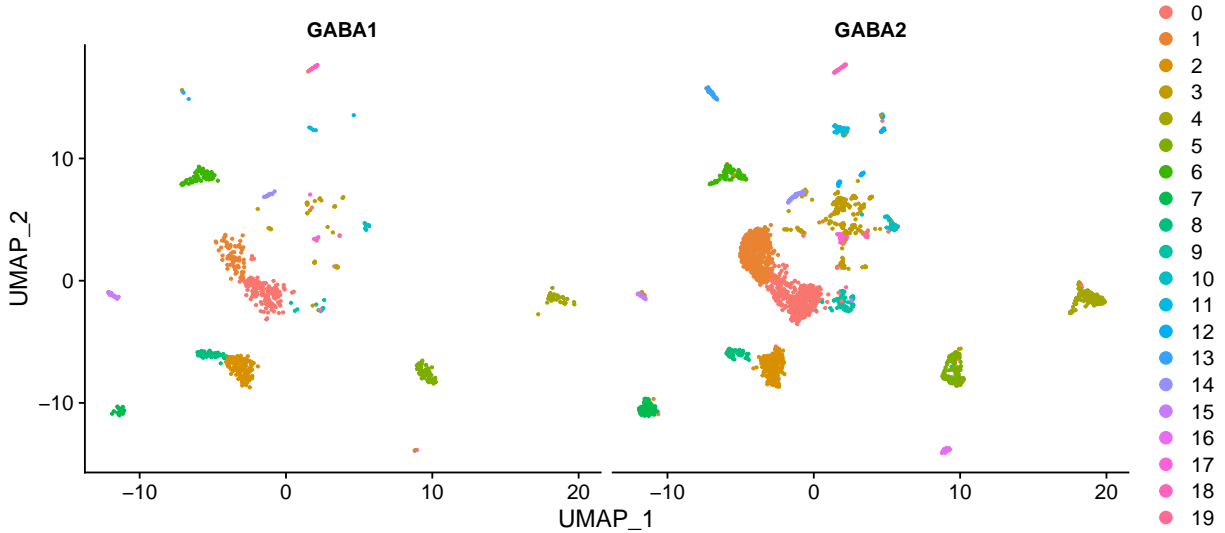
```
# Run the standard workflow for visualization and clustering  
grouped.combined <- ScaleData(grouped.combined, verbose = FALSE)  
grouped.combined <- RunPCA(grouped.combined, npcs = 30, verbose = FALSE)  
grouped.combined <- RunUMAP(grouped.combined, reduction = "pca", dims = 1:30)  
grouped.combined <- FindNeighbors(grouped.combined, reduction = "pca", dims = 1:30)  
grouped.combined <- FindClusters(grouped.combined, resolution = 0.5)
```

```
# Visualization  
p1 <- DimPlot(grouped.combined, reduction = "umap", group.by = "group")  
p2 <- DimPlot(grouped.combined, reduction = "umap", label = TRUE, repel = TRUE)  
p1 + p2
```



To visualize the two conditions side-by-side, we can use the `split.by` argument to show each condition colored by cluster.

```
DimPlot(grouped.combined, reduction = "umap", split.by = "group")
```



Identify conserved cell type markers

For performing differential expression after integration, we switch back to the original data

```
DefaultAssay(grouped.combined) <- "RNA"
nk.markers <- FindConservedMarkers(grouped.combined, ident.1 = 6, grouping.var = "group", verbose = FALSE)
head(nk.markers)
```

```
##           GABA1_p_val GABA1_avg_log2FC GABA1_pct.1 GABA1_pct.2 GABA1_p_val_adj
## flp-10  8.658042e-151      5.921891      1.000      0.015      4.062700e-146
## vab-23   1.775101e-86      2.742712      0.580      0.004      8.329485e-82
## nlp-40   4.223174e-101      5.859277      1.000      0.107      1.981682e-96
## D1007.19 5.318342e-137      4.355848      0.943      0.018      2.495579e-132
## T05A8.3  1.289278e-76      6.213582      1.000      0.223      6.049808e-72
## srv-7    4.384392e-66      2.211071      0.432      0.001      2.057332e-61
##           GABA2_p_val GABA2_avg_log2FC GABA2_pct.1 GABA2_pct.2 GABA2_p_val_adj
## flp-10  0.000000e+00      4.651373      0.877      0.008      0.000000e+00
## vab-23   2.797923e-198      2.533769      0.438      0.003      1.312897e-193
## nlp-40   6.358671e-198      4.830521      1.000      0.057      2.983743e-193
## D1007.19 1.812119e-150      3.346140      0.699      0.032      8.503187e-146
## T05A8.3  5.047622e-148      6.947285      1.000      0.093      2.368546e-143
## srv-7    1.806037e-118      1.888279      0.260      0.002      8.474649e-114
##           max_pval minimum_p_val
## flp-10  8.658042e-151  0.000000e+00
## vab-23   1.775101e-86  5.595845e-198
## nlp-40   4.223174e-101  1.271734e-197
## D1007.19 5.318342e-137  3.624238e-150
## T05A8.3  1.289278e-76  1.009524e-147
## srv-7    4.384392e-66  3.612074e-118
```