

Canadian Bioinformatics Workshops

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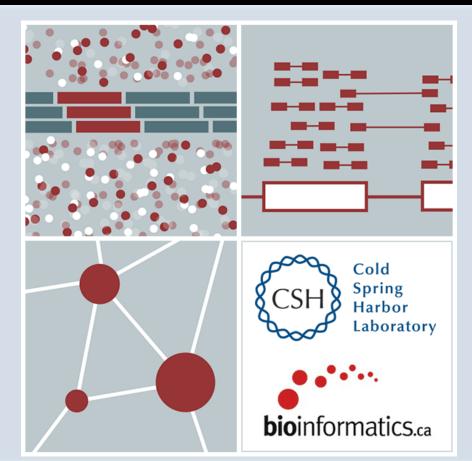
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RNA-Seq Module 4 Isoform Discovery and Alternative Expression (tutorial)

Malachi Griffith, Obi Griffith, Fouad Yousif High-Throughput Biology: From Sequence to Networks March 20-26, 2017







Learning Objectives of Tutorial

- Learn how to run StringTie in 'reference only', 'reference guided', and 'de novo' modes
- Learn how to use Cuffmerge to combine transcriptomes from multiple Cufflinks runs and compare assembled transcripts to known transcripts
- Learn how to perform differential splicing analysis with Cuffdiff
- Examine junctions counts and Cufflinks differential splicing files at the command line
- Visualize TopHat junction counts and Cufflinks assembled transcripts in IGV

5-i,ii. Running cuffinks in 'ref-guided' and 'de-novo' mode

- In Module 3 we ran cufflinks in 'ref-only' mode. This mode gives us an expression estimate for each known gene/transcript
- Now we want to be able to potentially identify novel genes, and novel isoforms of known genes
- To accomplish this we will re-run cufflinks in 'ref-guided' and 'de-novo' modes
 - In 'ref-guided' mode a known transcriptome will be used as a guide
 - In 'de-novo' mode no knowledge of the transcriptome will be used at all

'-g', '-G' woe is me...

- tophat has a '-G' option
 - Used to supply a transcriptome GTF file
 - This will be used to assist the alignment step by allowing alignment to both transcriptome and genome sequences
 - Coordinates from alignments to transcriptomes will be converted back to genome coordinates
 - Even though we supply a transcriptome, tophat will not be limited in anyway to known transcripts
- tophat also has a '-g' option
 - Used to specify the maximum number of multiple mappings for a single read
- cufflinks has a '-G' option
 - Used to supply a transcriptome GTF file
 - If specified, cufflinks will quantitate against reference transcript annotations
 - We call this the 'ref-only' analysis mode
- cufflinks also has a '-g' option
 - Use to supply a transcriptome GTF file
 - Use reference transcript annotations to guide assembly
 - We call this 'reference-guided' analysis mode
- Running cufflinks with neither '-G' or '-g'
 - We call this 'de-novo' analysis mode
- cuffdiff requires a GTF file but it is not specified with a '-G' or '-g' option, but rather is simply supplied as a file path when you run cuffdiff

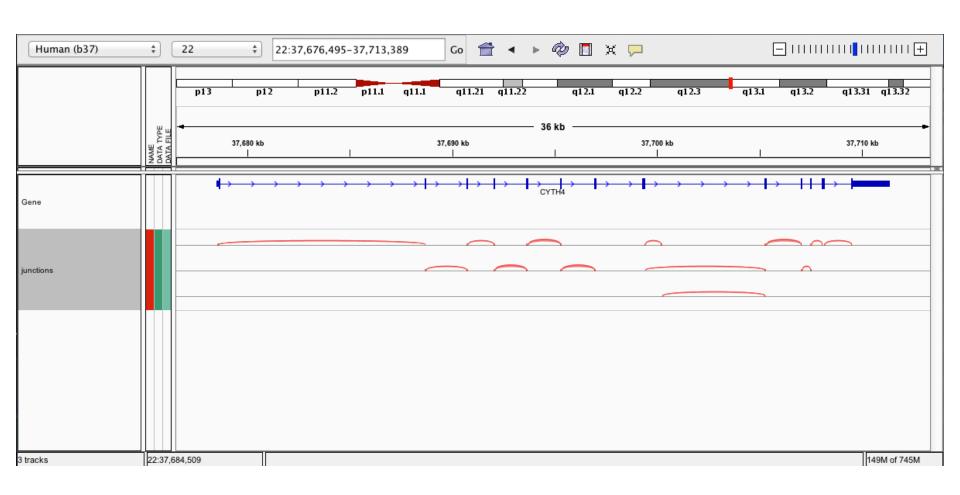
The tophat 'junctions.bed' file

- After alignment, tophat creates a summary of all reads that support exon-exon junctions
 - e.g. exon1-exon2 has 5 reads
 - e.g. exon1-exon3 has 9 reads
- This file reports all of the unique exon-exon junctions observed and the read counts for each
 - In BED format

track	name=junctions	description="To	pHat junctions"	-			-			
22	17062079	17063415	JUNC00000001	3	-	17062079	17063415	255,0,0 2	98,19	0,1317
22	17092740	17095057	JUNC00000002	5	+	17092740	17095057	255,0,0 2	43,91	0,2226
22	17117940	17119543	JUNC00000003	6	+	17117940	17119543	255,0,0 2	40,75	0,1528
22	17152466	17156100	JUNC00000004	3	-	17152466	17156100	255,0,0 2	12,88	0,3546
22	17525819	17528242	JUNC00000005	1	+	17525819	17528242	255,0,0 2	71,29	0,2394
22	17528261	17538007	JUNC00000006	1	+	17528261	17538007	255,0,0 2	55,45	0,9701
22	17566071	17577976	JUNC00000007	10	+	17566071	17577976	255,0,0 2	48,25	0,11880
22	17577951	17578785	JUNC00000008	24	+	17577951	17578785	255,0,0 2	25,99	0,735
22	17578093	17578710	JUNC00000009	1	+	17578093	17578710	255,0,0 2	76,24	0,593

Junction read count

Viewing the junctions.bed in IGV

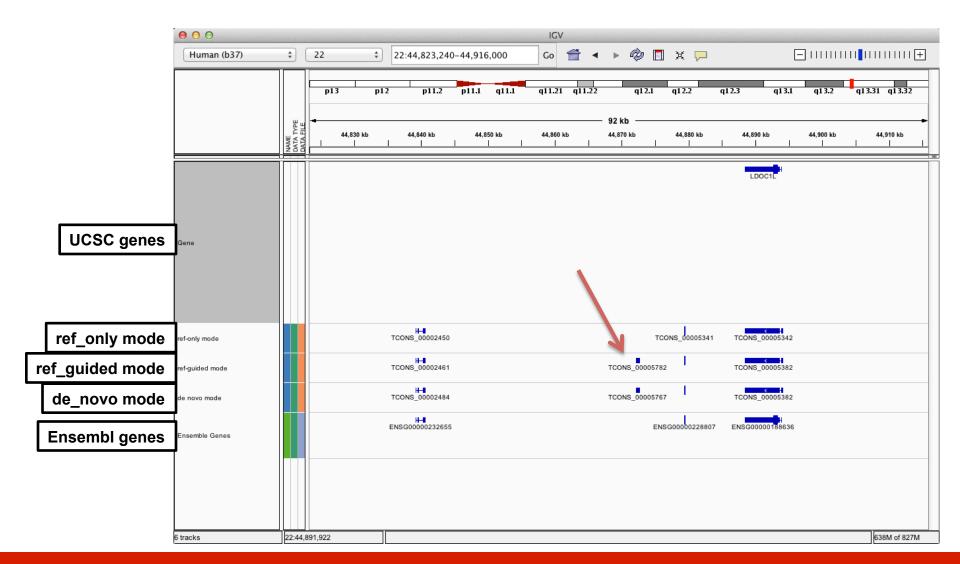


5-iii,iv. Cuffmerge

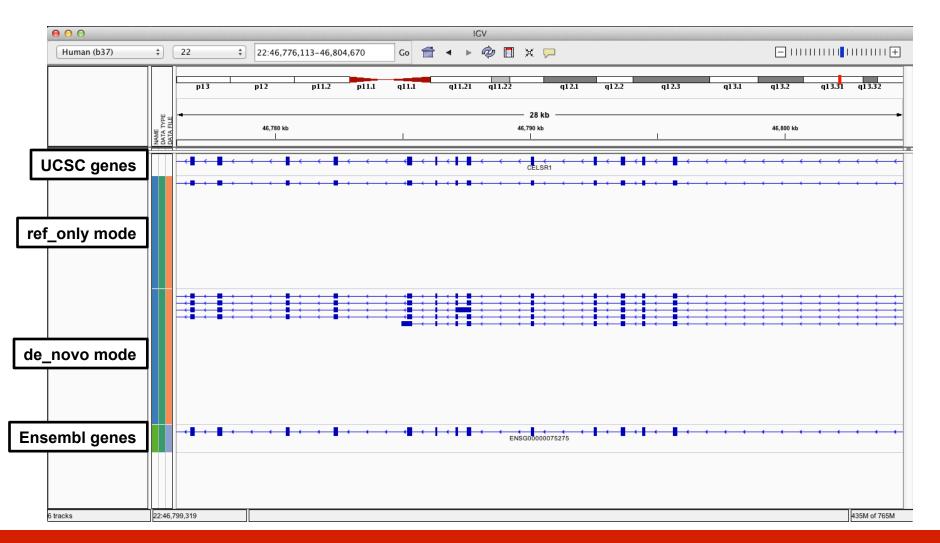
http://cufflinks.cbcb.umd.edu/manual.html#cuffmerge

- Cuffmerge combines transcripts predicted from multiple RNA-seq data sets into one view of the transcriptome
 - Do this before running cuffdiff to compare between multiple conditions
- Cuffmerge can also simultaneously compare transcripts to the known transcripts GTF file from Ensembl, etc.
 - http://cufflinks.cbcb.umd.edu/manual.html#class_codes

5-v. Comparison of merged GTFs from each cufflinks mode



Comparison of merged GTFs from each cufflinks mode



What if I return to my lab and can not get this to work on my own data?

- Refer to the materials provided with this course for clues
- Refer to the Nature Protocols tutorial (Trapnell et al. 2012)
 - In particular refer to the troubleshooting table (next slide)
- Search BioStars, SeqAnswers, and Google
 - http://www.biostars.org/
 - http://www.seqanswers.com
- If your question is not already answered on BioStars...
 - Ask it! Then follow up so that others that have the same problem in the future know whether this solution worked

TopHat/Cufflinks/Cuffdiff troubleshooting table

TABLE 2 | Troubleshooting table.

Step	Problem	Possible reason	Solution
1	TopHat cannot find Bowtie or the SAM tools	Bowtie and/or SAM tools binary executables are not in a directory listed in the PATH shell environment variable	Add the directories containing these executables to the PATH environment variable. See the man page of your UNIX shell for more details
2	Cufflinks crashes with a 'bad_alloc' error Cufflinks takes excessively long to finish	Machine is running out of memory trying to assemble highly expressed genes	Pass the -max-bundle-frags option to Cufflinks with a value of <1,000,000 (the default). Try 500,000 at first, and lower values if the error is still thrown
5	Cuffdiff crashes with a 'bad_ alloc' error Cuffdiff takes excessively long to finish	Machine is running out of memory trying to quantify highly expressed genes	Pass the -max-bundle-frags option to Cuffdiff with a value of <1,000,000 (the default). Try 500,000 at first, and lower values if the error is still thrown
	Cuffdiff reports FPKM = 0 for all genes and transcripts	Chromosome names in GTF file do not match the names in the BAM alignment files	Use a GTF file and alignments that has matching chromosome names (e.g., the GTF included with an iGenome index)

We are on a Coffee Break & Networking Session