Tangram Examples

Shawn Garbett 2018-08-07

A Grammar of Tables

This package is meant to implement the concept of a grammar of tables. It allows for a simple formula expression and a data frame to create a rich summary table in a variety of formats. It is designed for extensibility at each step of the process, so that one is not limited by the authors choice of table statistics, output format. The grammar however is an integral part of the package, and as such is not modifiable.

Here's an example similar to summary M from Hmisc to get us started:

Context Aware Compilation

	N	D-penicillamine (N=154)	placebo (N=158)	not randomized (N=106)	Test Statistic
Serum Bilirubin	418	0.70 *1.30* 3.60	0.80 *1.40* 3.22	0.70 *1.40* 3.12	F_{2,415}=0.03, P=0.972^1
Albumin	418	3.34 *3.54* 3.78	3.21 *3.56* 3.83	3.12 *3.47* 3.73	F_{2,415}=2.13, P=0.120^1
Histologic Stage, Ludwig Criteria	412				X^2_6=5.33, P=0.502^2
1		0.026 4/154	0.076 12/158	0.050 5/100	
2		0.208 32/154	0.222 35/158	0.250 25/100	
3		0.416 64/154	0.354 56/158	0.350 35/100	
4		0.351 54/154	0.348 55/158	0.350 35/100	
Prothrombin Time	416	10.0 *10.6* 11.4	10.0 *10.6* 11.0	10.1 *10.6* 11.0	F_{2,413}=0.23, P=0.795 ¹
sex : female	418	0.903 139/154	0.867 137/158	0.925 98/106	X^2_2=2.38, P=0.304^2
Age	418	41.4 *48.1* 55.8	42.9 *51.9* 59.0	46.0 *53.0* 61.1	F_{2,415}=6.10, P=0.002^1
spiders : present	312	0.292 45/154	0.285 45/158		X^2_1=0.02, P=0.885^2

N is the number of non-missing value. ^1 Kruskal-Wallis. ^2 Pearson. ^3 Wilcoxon.

Notice that stage in the formula wasn't stored as a factor, i.e. Categorical variable, so by adding a type specifier in the formula given, it is treated as a Categorical. There is no preconversion applied to the data frame, nor is there a guess based on the number of unique values. Full direct control of typing is provided in the formula specification.

It also supports HTML5, with styling fragments.

Hmisc Style Example

	N	D-penicillamine	placebo	not randomized	Test Statistic
		(N=154)	(N=158)	(N=106)	
Serum Bilirubin mg/dl	418	0.30 0.70 **1.30** 3.60 28.00 3.65±5.28	0.30 0.80 **1.40** 3.22 20.00 2.87±3.63	0.40 0.70 **1.40** 3.12 18.00 3.12±4.04	F _{2,415} =0.03, P=0.972 ¹
Albumin gm/dl	418	$1.96\ 3.34\ **3.54**\ 3.78\ 4.38\ 3.52{\pm}0.40$	$2.10\ 3.21\ ^{**}3.56^{**}\ 3.83\ 4.64\ 3.52{\pm}0.44$	$2.31\ 3.12\ **3.47**\ 3.73\ 4.52\ 3.43{\pm}0.43$	$F_{2,415}=2.13, P=0.120^1$
Histologic Stage, Ludwig Criteria	412				χ^2_6 =5.33, P=0.502 ²
1		$0.026 \frac{4}{154}$	$0.076 \frac{12}{158}$	$0.050 \frac{5}{100}$	
2		$0.208 \frac{32}{154}$	$0.222 \frac{35}{158}$	$0.250 \frac{25}{100}$	
3		$0.416 \frac{64}{154}$	$0.354 \frac{56}{158}$	$0.350 \frac{35}{100}$	
4		$0.351 \frac{54}{154}$	$0.348 \frac{55}{158}$	$0.350 \frac{35}{100}$	
Prothrombin Time sec.	416	9.2 10.0 **10.6** 11.4 17.1 10.8±1.1	9.0 10.0 **10.6** 11.0 14.1 10.7±0.9	9.0 10.1 **10.6** 11.0 18.0 10.8±1.1	$F_{2,413}=0.23, P=0.795^1$
sex : female	418	$0.903 \frac{139}{154}$	$0.867 \frac{137}{158}$	$0.925 \frac{98}{106}$	χ^2_2 =2.38, P=0.304 ²
Age	418	$30.6\ 41.4\ **48.1**\ 55.8\ 74.5\ 48.6{\pm}10.0$	26.3 42.9 **51.9** 59.0 78.4 51.4±11.0	$33.0\ 46.0\ ^{**}53.0^{**}\ 61.1\ 75.0\ 52.9\pm9.8$	$F_{2,415}$ =6.10, P =0.002 ¹
spiders : present	312	$0.292 \frac{45}{154}$	$0.285 \frac{45}{158}$		χ^2_1 =0.02, P=0.885 ²

Table 1: Table Hmisc Style

N is the number of non-missing value. $^1\mathrm{Kruskal\text{-}Wallis.}\ ^2\mathrm{Pearson.}\ ^3\mathrm{Wilcoxon.}$

NEJM Style Example

Fragments can have localized style sheets specified by given id. Notice the specification of 2 digits for bilirubin in the formula.

Table NEJM Style					
	N	D-penicillamine	placebo	not randomized	Test Statistic
		(N=154)	(N=158)	(N=106)	
Serum Bilirubin mg/dl	418				F _{2,415} =0.03, P=0.972
Median (interquartile range)		1.30 (0.70—3.60)	1.40 (0.80—3.22)	1.40 (0.70—3.12)	
Range		0.30—28.00	0.30—20.00	0.40—18.00	
Albumin gm/dl	418				F _{2,415} =2.13, P=0.120
Median (interquartile range)		3.54 (3.34—3.78)	3.56 (3.21—3.83)	3.47 (3.12—3.73)	
Range		1.96—4.38	2.10-4.64	2.31—4.52	
Histologic Stage, Ludwig Criteria	412				$\chi^2_6 = 5.33$, P=0.502 ²
1		4/154 (2.6)	12/158 (7.6)	5/100 (5.0)	
2		32/154 (20.8)	35/158 (22.2)	25/100 (25.0)	
3		64/154 (41.6)	56/158 (35.4)	35/100 (35.0)	
4		54/154 (35.1)	55/158 (34.8)	35/100 (35.0)	
Prothrombin Time sec.	416				F _{2,413} =0.23, P=0.795
Median (interquartile range)		10.6 (10.0—11.4)	10.6 (10.0—11.0)	10.6 (10.1—11.0)	
Range		9.2—17.1	9.0—14.1	9.0—18.0	
sex : female	418	139/154 (90.3)	137/158 (86.7)	98/106 (92.5)	$\chi^2_2 = 2.38$, P=0.304 ²
Age	418				F _{2,415} =6.10, P=0.002
Median (interquartile range)		48.1 (41.4—55.8)	51.9 (42.9—59.0)	53.0 (46.0—61.1)	
Range		30.6—74.5	26.3—78.4	33—75	
spiders : present	312	45/154 (29.2)	45/158 (28.5)		$\chi^2_1 = 0.02$, P=0.885 ²

Table 2: Table NEJM Style

N is the number of non-missing value. $^1{
m Kruskal-Wallis}$. $^2{
m Pearson}$. $^3{
m Wilcoxon}$.

Lancet Style Example

Fragments can have localized style sheets specified by given id. Specifications of digit output can also be c specifiers if enclosed in quotes.

	N	D-penicillamine	placebo	not randomized	Test Statistic
		(N=154)	(N=158)	(N=106)	
Serum Bilirubin mg/dl	418	3.649 (5.282)	2.873 (3.629)	3.117 (4.043)	F _{2,415} =0.03, P=0.97248 ¹
Albumin gm/dl	418	3.52 (0.40)	3.52 (0.44)	3.43 (0.43)	F _{2,415} =2·13, P=0·11996 ¹
Histologic Stage, Ludwig Criteria	412				$\chi^2_6 = 5.33$, P=0.50235 ²
1		4 (2.6%)	12 (7.6%)	5 (5.0%)	
2		32 (20.8%)	35 (22.2%)	25 (25.0%)	
3		64 (41.6%)	56 (35.4%)	35 (35.0%)	
4		54 (35·1%)	55 (34.8%)	35 (35.0%)	
Prothrombin Time sec.	416	10.8 (1.1)	10.7 (0.9)	10.8 (1.1)	F _{2,413} =0.23, P=0.79472 ¹
sex : female	418	139 (90.3%)	137 (86.7%)	98 (92.5%)	$\chi^2_2 = 2.38$, P=0.30387 ²
Age	418	48.6 (10.0)	51.4 (11.0)	52.9 (9.8)	$F_{2,415} = 6.10, P = 0.00245^1$
spiders : present	312	45 (29.2%)	45 (28.5%)		$\chi^2_1 = 0.02$, P=0.88534 ²

Table 3: Table Lancet Style

N is the number of non-missing value. $^1{\rm Kruskal\text{-}Wallis.}$ $^2{\rm Pearson.}$ $^3{\rm Wilcoxon.}$

Indexing

It is also capable of producing an index of contents inside a table for traceability.

```
key
 [1,] "MGNk" "tangram:bili:drug:cell_n1"
 [2,] "MDQ4" "tangram:bili:drug[D-penicillamine]:cell1"
 [3,] "ZDMz" "tangram:bili:drug[placebo]:cell1"
 [4,] "NjRm" "tangram:bili:drug[not randomized]:cell1"
 [5,] "MGVi" "tangram:bili:drug:statistics1"
 [6,] "MTY2" "tangram:albumin:drug:cell_n1"
 [7,] "Y2E4" "tangram:albumin:drug[D-penicillamine]:cell1"
 [8,] "Mjc3" "tangram:albumin:drug[placebo]:cell1"
[9,] "ZTc3" "tangram:albumin:drug[not randomized]:cell1"
[10,] "YmM3" "tangram:albumin:drug:statistics1"
[11,] "ZjIx" "tangram:stage:drug:cell1"
[12,] "ZjIx" "tangram:stage:drug:cell1"
[13,] "ZjIx" "tangram:stage:drug:cell1"
[14,] "ZjIx" "tangram:stage:drug:cell1"
[15,] "MWIx" "tangram:stage:drug:statistics1"
[16,] "ZGIz" "tangram:stage[ 1]:drug[D-penicillamine]:cell1"
[17,] "YTU5" "tangram:stage[ 2]:drug[D-penicillamine]:cell1"
[18,] "ZDRk" "tangram:stage[
                              3]:drug[D-penicillamine]:cell1"
[19,] "ZjIx" "tangram:stage:drug:cell1"
[20,] "NTBh" "tangram:stage[ 1]:drug[placebo]:cell1"
```

```
value
 [1,] "418"
 [2,] "0.70 **1.30** 3.60"
 [3,] "0.80 **1.40** 3.22"
 [4,] "0.70 **1.40** 3.12"
 [5,] "F~2,415~=0.03, P=0.972^1^"
 [6.] "418"
 [7,] "3.34 **3.54** 3.78"
 [8,] "3.21 **3.56** 3.83"
 [9,] "3.12 **3.47** 3.73"
[10,] "F~2,415~=2.13, P=0.120^1^"
[11,] "412"
[12,] ""
[13,] ""
[14,] ""
[15,] "X^2^~6~=5.33, P=0.502^2^"
[16,] "0.026 \\frac{ 4}{154}"
[17,] "0.076 \\frac{ 12}{158}"
[18,] "0.050 \\frac{ 5}{100}"
[19,] ""
[20,] "0.208 \\frac{ 32}{154}"
```

Intercept Model Example

	N	All
Global score, 3m	375	73 **78** 85
Global score, 12m	374	74 **80** 86

Table 4: Intercept Example

N is the number of non-missing value. ¹Kruskal-Wallis. ²Pearson. ³Wilcoxon.

Types

The Hmisc default style recognizes 3 types: Categorical, Bionimial, and Numerical. Then for each product of these two, a function is provided to generate the corresponding rows and columns. As mentioned before, the user can declare any type in a formula, and one is not limited to the Hmisc defaults. This is completely customizable, which will be covered later.

Let's cover the phases of table generations.

1. Syntax. The formula is parsed into an abstract syntax tree (AST), and factors are right distributed, and the data frame is split into appropriate pieces attached to each node in the AST. The syntax and

- parser are the only portions of this library that are fixed, and not customizable. The grammar may expand with time, but cautiously as to not create an overly verbose set of possibilities to interpret. The goal is to create a clean grammar that describes the bold areas of a table to fill in.
- 2. Semantics. The elements of the AST are examined, and passed to compilation functions. The compilation function function is chosen by determining the type of the row variable, and the type of column variable. For example, drug ~ stage::Categorical, is a Categorical×Categorical which references the summarize_chisq for compiling. One can easily specify different compilers for a formula and get very different results inside a formula. Note: the application of multiplication * cannot be done in the previous phase, because this involves semantic meaning of what multiplication means. In one context it might be an interaction, in another simple multiplication. Handling multiplicative terms can be tricky. Once compiling is finished a table object composed of cells (list of lists) which are one of a variety of S3 types is the result.
- 3. Rendering. With a compiled table object in memory, the final stage is conversion to an output format which could be plain text, HTML5, LaTeX or anything. These are overrideable via S3 classes representing the different possible types of cells that are present inside a table. User specified rendering is possible as well.

Summary columns

A simple example of using an intercept in a formula, with some post processing to remove undesired columns.

```
d1 <- iris
d1$A <- d1$Sepal.Length > 5.1
attr(d1$A,"label") <- "Sepal Length > 5.1"
tangram(
    Species + 1 ~ A + Sepal.Width,
    data = d1,
    id="tbl6",
    style="nejm",
    caption="Example All Summary"
) %>%
drop_statistics() %>%
del_col(6)
```

Example All Summary							
	N	setosa	versicolor	virginica	All		
		(N=50)	(N=50)	(N=50)	(N=150)		
Sepal Length > 5.1 : TRUE	150	14/50 (28.0)	46/50 (92.0)	49/50 (98.0)	109/150 (72.7)		
Sepal.Width	150						
Median (interquartile range)		3.40 (3.19—3.70)	2.80 (2.50—3.00)	3.00 (2.80—3.20)	3.00 (2.80—3.31)		
Range		2.30—4.40	2.00—3.40	2.20—3.80	2.00—4.40		

Table 5: Example All Summary

N is the number of non-missing value. $^1{
m Kruskal-Wallis}$. $^2{
m Pearson}$. $^3{
m Wilcoxon}$.

Extensibility

The library is designed to be extensible, in the hopes that more useful summary functions can generate results into a wide variety of formats. This is done by the translator functions, which given a row and column from a formula will process the data into a table.

This example shows how to create a function that given a row and column, to construct summary entries for a table.

```
### Make up some data, which has events nested within an id
n <- 1000
df <- data.frame(id = sample(1:250, n*3, replace=TRUE), event = as.factor(rep(c("A", "B", "C"), n)))</pre>
attr(df$id, "label") <- "ID"</pre>
### Now create custom function for counting events with a category
summarize count <- function(table, row, column, ...)</pre>
  ### Getting Data for row column ast nodes, assuming no factors
  datar <- row$data
  datac <- column$data
  ### Grabbing categories
  col_categories <- levels(datac)</pre>
 n_labels <- lapply(col_categories, FUN=function(cat_name){</pre>
   x <- datar[datac == cat_name]</pre>
    cell_n(length(unique(x)), subcol=cat_name)
  })
  # Test a poisson model
  test <- summary(aov(glm(x ~ treatment,
                      aggregate(datar, by=list(id=datar, treatment=datac), FUN=length),
                      family=poisson)))[[1]]
  test <- hmisc_fstat(f = render_f(test$'F value'[1], "%.2f"),</pre>
                      df1 = test$Df[1], df2 = test$Df[2],
                      p = hmisc_p(test$'Pr(>F)'[1]))
  # Build the table
  table
                                                       %>%
  # Create Headers
  row_header(derive_label(row))
                                                       %>%
  col_header("N", col_categories, "Test Statistic")
                                                      %>%
                                   11.11
  col_header("", n_labels,
  # Add the First column of summary data as an N value
  add_col(cell_n(length(unique(datar))))
  # Now add quantiles for the counts
  table_builder_apply(col_categories, FUN=
    function(tbl, cat_name) {
      # Compute each data set
      x <- datar[datac == cat_name]</pre>
      xx <- aggregate(x, by=list(x), FUN=length)$x
      # Add a column that is a quantile
      add_col(tbl, hmisc_iqr(xx, row$format, na.rm=TRUE))
  })
                                                       %>%
  # Now add a statistical test for the final column
  add_col(test)
tangram(event ~ id["%1.0f"], df, id="tbl7", transforms=summarize_count)
```

	N	A	В	\mathbf{C}	Test Statistic
		245	247	249	
ID	250	2 **4** 5	3 **4** 5	3 **4** 5	$F_{2,738}=0.07, P=0.935^1$

Table 6:

And that concludes the basic demonstration of formula capabilities and writing one's own simple transform bundle.