

STOmics

**Stereo-seq
ANALYSIS WORKFLOW
FILE FORMAT
MANUAL**

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Manual Version: A3.1

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Note: Please download the latest version of the manual and use it with the software specific for this manual.

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CHAPTER 1

OVERVIEW

1.1. About Software

Stereo-seq Analysis Workflow¹ (SAW) software suite is a set of pipelines that are bundled to position sequenced reads to their spatial location on the tissue section, and quantify spatial gene expression.

SAW download (Docker Hub): <https://hub.docker.com/r/stomics/saw>

SAW Github: <https://github.com/STOmics/SAW>

1.2. About Manual

This manual includes descriptions of key files format generated from SAW, which help users better understand and make use of information from analysis results.

1.3. Terminologies and Concepts

Table 1-1 Terminologies and Concepts

Abbreviation	Full Name	Description
SN	Serial Number	Unique ID for Stereo-seq Chip T.
RIN	RNA Integrity Value	RNA integrity value measures the RNA degradation degree to indicate the integrity of RNA and evaluate the quality of the RNA sample. RIN values range from 1 (totally degraded) to 10 (intact). In Stereo-seq analysis, only tissue sample with a pre-measured RIN value greater than 7 should be used for further sequencing and bioinformatics analysis.
CID	Coordinate ID	Spatial position identifier, the artificially synthesized barcode sequence unique to each spot on the Stereo-seq Chip T.
MID	Molecular ID	Molecular identifier (same as UMI), the artificially synthesized sequence unique to each mRNA molecule captured from the sample which helps to differentiate the number of reads contributed by mRNA expression level due to amplification. Two copies of native transcripts from the same molecule captured on one DNB will result in two independent reads with the same CID but different MID. In contrast, two reads with identical CID and MID were originated from the same transcript but got amplified.
DNB	DNA Nanoball	DNA nanoball is the product of rolling-circle amplification (RCA) that is linearly amplified from the original circular single-stranded DNA template. DNB is the smallest capture unit on the Stereo-seq Chip T.
Bin	Bin	Bin (or Square Bin) is the analysis unit on the gene expression heat map. A bin is a fixed-sized square in which the expression value in this square is accumulated. Bins are not overlapped. The value followed by "Bin" represents the side length of the square. For bin 1, each DNB on the Stereo-seq Chip T is shown as a spot, which means one spot only contains the data from one DNB. Bin N means one spot on the heat map is an aggregation of data from N×N neighbor DNBs. For example, a spot of bin 100 covers data from 10,000 DNBs.
Cell Bin	Cell Bin	Similar to Square Bin, a cell bin stands for a region of cell on the expression map recognized by the algorithm (either from image or heat map). Expression within a cell bin region is accumulated, and neighbor cell bins are not overlapped.

CHAPTER 2

FILE FORMAT

2.1 FASTQ

FASTQ is a common format for storing sequencing reads and corresponding quality evaluation. The Stereo-seq method is PE (paired-end) sequencing. Read 1 contains CID and MID information while read 2 contains captured mRNA sequencing data. During multi-sample sequencing, an additional sequence (sample barcode) is added to identify samples. When sequencing data is off, low-quality MID (containing N bases in MID or having two or more bases with quality value lower than 10) filtration is performed on read 1 and paired read 2. CID and MID are added to readID of read 2 that has been filtered by MID quality, and single strands in read 2, containing mRNA information, are written into the file in FASTQ format as the original sequencing data, where sample barcode is removed.

Q40 FASTQ and Q4 FASTQ are two optional output formats for the original sequencing data. Q40 adopts an evaluation system that describes the quality of sequenced bases with 41 quality values. Q4 means a similar evaluation system but with 4 quality values. Q40 FASTQ, also called PE FASTQ, consists of a pair of read files, read 1 for CID, MID information and read 2 for captured mRNA sequencing data respectively.

Example of PE FASTQ:

```
# read 1
@E100026571L1C001R003000000000/1
TGTCCAACGGAGACGGCTCCGACAAGGCACTGGCA
+
>DG;<BGH=>*EFE8*G/3E@2:F0-GBGG188F<

# read 2
@E100026571L1C001R003000000000/2
GTCTCACCATACTTTTACAAAGTTATTTCACCCAAATCACAATTTAAGAATTATTTGTTCTACCTATGCCACACT
TTAAATAAATGTCTATTAAACCA
+
-GFEECG?ECBFF<=@A@<E@><;FGCF=>=E53FEF5>FGF@,0ADE9CEAG2GBE@
HF3EA<CE;G2F@=G8=?@G9FBGE.EG6G2;974E*D9DE9
```

Q4 FASTQ, also called SE FASTQ, is an output format with only one read file, which is split from a data set (containing 16 or 64 parts). ReadID in the file starts with "@" and contains read name and binary-coded CID and MID information. The sequence part contains captured mRNA sequencing data. Because of the combined output format and fewer kinds of quality values, the file storage space is greatly saved.

Example of SE FASTQ:

```
@FP3000000513L1C002R004000000218 CE242DF29A57 97D26
GTGTAGTGAACCCCATGGTAGTTTTCTGATTGTTGTTAAAAAAATGACTTAACATATTACATGGACACTCAAT
AAAAATGTTTTATTTCTGTTGAAAA
+
FFFFFFFFFFFFF8F8FFFFFFFFFFFFF8FFFFFFFFF8FF8FFF8FFFFFFFF,FFFFFFFFFFFF8FFFFFFFF8F
8F,F8FFFFFF,FFFFFFFF,FFF
```


2.2 BAM

The BAM² file format is a binary format for saving sequence alignment and gene annotation data. SAW **mapping** BAM adds custom tags in the BAM optional field to record reads coordinates, CID and MID information. **count** BAM adds annotation information in the tag field. Custom tags are described in Table 2-1.

Table 2-1 BAM custom tags

Tag	Description
Cx:i	x coordinate of CID.
Cy:i	y coordinate of CID.
UR:Z	The hexadecimal representation of uncorrected binary-encoded MID.
XF:Z	Mapping region on the reference genome. Valid value: 0=EXONIC, 1=INTRONIC, 2=INTERGENIC. 3=rRNA
GE:Z	Annotated gene name.
GS:Z	‘+’ or ‘-’, indicating forward/reverse strand respectively.
UB:Z	The hexadecimal representation of count corrected binary-encoded MID.

Example of mapping BAM:

```
E100026571L1C009R00301275185      16          1        3000095 255  
26M121066N74M       *           0            0     GGCTTTTTTTTTTTTTTTTTTTTCTAA  
ATATTGGGTTTTATTAGCACCATGATAACTGTATATTAATTTGCACTGACTGTCATAACAAAATAC         G+  
 :GFFFGGFGFFGFFGFGGF FFFFFFFCFGCFCGGGFGGFGFFFFGGFGGFGFFFGGFFGFFFGF GF FG FF GF  
GFFFFFFFFFFFFFFFFGGFFGGFFGEF    NH:i:1   HI:i:1    AS:i:88 nM:i:0   Cx:i:4826  
Cy:i:11598             UR:Z:6FA29
```

Example of **count** BAM:

```
E100026571L1C002R00703943265      1040      1      3082766 255
11M132671N89M      *      0      0      CTGCTGCAGCTTTTTTTTCTTTGAGATTTA
TTTTTATGCTATGTGTATGGGTATTTTGCCTGCATATATGTCTATGCACCATGTGTGTGCAGTGCTTGAG
FFFFFECGFDGDFGDFEE@EEGIBFGGCGFFGACGFCGFFDGDGFFFFFEGCDFCGFFGG@FFF=EFFDGGG
GGFDGFFFGGGGFFFGGGFFGGGDFG      NH:i:1 HI:i:1 AS:i:88 nM:i:0 Cx:i:7767
Cy:i:18052      UR:Z:7AE49      XF:i:0 GE:Z:Xkr4      GS:Z:- UB:Z:79E49
```

2.3 Mapped CID List with Reads Count File

mapping pipeline outputs mapped CID list file with reads counts for each CID. This file stores CID coordinates and reads count for each coordinate. The list does not have a header. The three columns are x coordinate, y coordinate and MID count.

Example of mapped CID list with reads count file:

```
14195      16619      1
19945      14450      2
14548      9438       1
```

2.4 Gene Expression File

Gene expression file (GEF) is a data management and storage format designed to support multi-dimensional data storage and high computation efficiency. Stereo-seq analysis workflow generates Square Bin GEF and Cell Bin GEF files. Square Bin file format is a hierarchically structured data model that stores one or bin combined gene expression matrices in different bin sizes. Cell Bin file format stores expression information within each cell.

Each GEF container organizes a collection of spatial gene expression matrices. It includes two primary data objects, Group and Dataset. A dataset is a multidimensional array of data elements. Group object is analogous to file system directory which organizes datasets and other groups in hierarchies.

2.4.1 Square Bin GEF

The first level of GEF includes four group objects: “geneExp” (required), “wholeExp” (optional), “wholeExpExon” (optional), and “stat” (optional). Group “geneExp” contains groups of gene spatial expression data in one or multiple bin size. Group “wholeExp” contains datasets that record expression level and gene type count of each coordinate in one or multiple bin sizes. Group “wholeExpExon” contains datasets that record exon level of each coordinate in one or multiple bin sizes. Group “stat” saves gene names, total MID count and spatial pattern enrichment score of each gene. “Attributes” of the file records the version of GEF format, software version, and omics information. “Attribute” in each dataset records the key metrics of that dataset. Check <https://github.com/STOmics/SAW/tree/main/Documents/FileFormat> to get the schematic diagram of square bin GEF. The field names and field data types of Square Bin GEF are described in Table 2-2. SAW outputs three GEF files in the whole process. Please check Table 2-3 to find the description.

Table 2-2 Square Bin Gene Expression File Text Fields Description

Attributes			
File Attributes	DataType	Example	Description
version	uint32	2	Gene expression file format version.
geftool_ver	uint32[3]	0,7,11	geftool version. It can be used as an individual tool to manipulate GEF files.
omics	S32	b'Transcriptomics'	Omics name.
gef_area	float32	4.4410855E10	Tissue or labeled tissue area in square nanometers.
/geneExp/binN/expression: Dataset “expression” is a 1D array which stores coordinates and MID counts of each gene in the bin size of N, aggregated by gene name.			
Dataset Attributes	DataType	Example (bin1)	Description
minX	int32	59820	Minimum x coordinate in bin N.
minY	int32	102086	Minimum y coordinate in bin N.
maxX	int32	73040	Maximum x coordinate in bin N.
maxY	int32	120539	Maximum y coordinate in bin N.
maxExp	uint32	28	Maximum MID count in a spot when the bin size is N. Data type for “maxExp” is dynamically changed for each sample.
resolution	uint32	500	Physical pitch (nm) between neighbor spots.
Dataset Data compound	DataType	Example (bin1)	Description
x	int32	71032	x coordinate in bin N.
y	int32	103180	y coordinate in bin N.
count	uint8/uint16/uint32	1	MID count at (x, y) when bin size is N. Data type for “count” is consistent with “maxExp” in the “Attributes.”

[optional] /geneExp/binN/exon:
Dataset “exon” is a 1D array which stores exon expression of each gene in the bin size of N, aggregated by gene name.

Dataset Attributes	DataType	Example (bin1)	Description
maxExon	int32	21	Max exon expression in binN.
Dataset DataType: 1D array	DataType	Example (bin1)	Description
count	uint8/uint16/uint32	0	Exon expression in binN at coordinate (x,y), the index is same to the index in the “expression” dataset. Data type for “count” is dynamically changed for each sample.

/geneExp/binN/gene:
Dataset “gene” is a 1D array which stores the gene names, the starting row indexes in dataset “expression”, and row counts.

Dataset DataType: compound	DataType	Example (bin1)	Description
gene	S32	b'Gm16045'	Gene name.
offset	uint32	21	The starting row index in dataset “expression” for the gene. In this example, the gene expression data for gene “Gm16045” starts from row 21 in the dataset “expression.”
count	uint32	2	Row count. In this example, expression data for gene “Gm16045” is recorded in row 21 and 22 (2 rows) in the dataset “expression.”

[optional] /wholeExp/binN:
Dataset “binN” is a 2D array (matrix) which stores the MID count and gene type count at each spot.

Dataset Attributes	DataType	Example (bin1)	Description
number	uint64	22879557	Number of non-zero spots in the dense matrix.
minX	int32	59820	Minimum x coordinate in bin N.
lenX	int32	13221	Length of x.
minY	int32	102086	Minimum y coordinate in bin N.
lenY	int32	18454	Length of y.
maxMID	uint32	2155	Maximum MID count in a spot.
maxGene	uint32	846	Maximum gene type count in a spot.
resolution	uint32	500	Pitch (nm) between neighbor spots.
Dataset DataType: 2D array (XxY), compound	DataType	Example (bin1)	Description
MIDcount	uint8/uint16/uint32	1	MID count in the spot. The spot coordinate can be identified from the row and column index of the 2D matrix plus the “minX” and “minY” specified in the attributes. Data type for “MIDcount” is dynamically changed for each sample.
genecount	uint16	1	Gene count in the spot. The spot coordinate can be identified from “Attributes” and the indexes of the 2D array.

[optional] /wholeExpExon/binN:

Dataset “binN” in “/wholeExpExon/” Group is a 2D array (matrix) which stores the exon expression count at each spot.

Dataset Attributes	DataType	Example (bin1)	Description
maxExon	uint32	21	Maximum exon expression count in a spot when the bin size is N.
Dataset Attributes	DataType	Example (bin1)	Description
MIDcount	uint8/uint16/uint32	0	MID count in the spot. The spot coordinate can be identified from the row and column index of the 2D matrix plus the “minX” and “minY” specified in the attributes. Data type for “MIDcount” is dynamically changed for each sample.

[optional] /stat/gene:

Dataset “gene” is a 1D array which stores the MID count and spatial pattern enrichment score (E10) of each gene. The array is order by the MID count in descending order.

Dataset Attributes	DataType	Example	Description
maxE10	float32	65.53	Maximum E10 score.
minE10	float32	0.	Minimum E10 score.
cutoff	float32	0.1	Threshold for filtering spots that will be used for computing E10. In this example, 0.1 means that the spots whose MID count is in the top 10% are used for calculating the spatial enrichment score.
Dataset Attributes	DataType	Example	Description
gene	S32	b'Ptgds'	Gene name.
MIDcount	uint32	229502	MID count for the gene.
E10	float32	65.53	The spatial pattern enrichment score (E10) for the gene.

The distinctions of each SAW output GEF files are explained in Table 2-3.

Table 2-3 SAW Output GEF Files Description

GEF Name	SAW Pipeline	Example	Description
SN.raw.gef	count	SS200000135TL_ D1.raw.gef	count output raw GEF, it only includes geneExp Group for the bin size of 1. The origin of expression matrix has been calibrated to (0,0).
SN.gef	tissueCut	SS200000135TL_ D1.gef	tissueCut output full GEF file. It contains geneExp Group and wholeExp Group for the bin size of 1, 10, 20, 50, 100, 200, and 500. SN.gef is also the only one that includes stat Group. The origin of expression matrix has been calibrated to (0,0), and its offsets are the same with SN.raw.gef. SN.gef is the input file for visualization.
SN.tissue.gef	tissueCut	SS200000135TL_ D1.tissue.gef	tissueCut output GEF file for the tissue-covered region. It only includes geneExp Group for the bin size of 1. The coordinates in the matrix and the offsets are all same with SN.raw.gef.

GEF Name	SAW Pipeline	Example	Description
SN.<label>.raw.label.gef	tissueCut	B01020C2.Label01.raw.label.gef	tissueCut output GEF file for the labeled ('aa' in the example) tissue-covered region. It only includes geneExp Group for the bin size of 1. The coordinates in the matrix and the offsets are all same with SN.raw.gef.
SN.<label>.label.gef	tissueCut	B01020C2.Label01.label.gef	tissueCut output full labeled tissue-covered GEF file.

2.4.2 Cell Bin GEF

The first layer of Cell Bin GEF contains one required group “cellBin” and multiple optional datasets. “Attributes” of the file records the version of GEF format, software version, and omics information. “Attribute” in each dataset records the key metrics of that dataset. Check <https://github.com/BGIResearch/SAW/tree/main/Documents/FileFormat> to get the schematic diagram of cell bin GEF. The field names and field data types of Cell Bin GEF are described in Table 2-4.

Table 2-4 Cell Bin Gene Expression File Text Fields Description

Attributes			
File Attributes	DataType	Example	Description
geftool_ver	uint32[3]	0,7,11	geftool version. It can be used as an individual tool to manipulate GEF files.
offsetX	int32	0	Minimum x coordinate in bin 1.
offsetY	int32	0	Minimum y coordinate in bin 1.
omics	S32	b‘Transcriptomics’	Omics name.
resolution	uint32	500	Pitch (nm) between neighbor spots.
version	uint32	2	Gene expression file format version.

/cellBin/cell: Dataset “cell” is a 1D array which stores basic information and indices information of cells and expression.			
Dataset Attributes	DataType	Example	Description
averageArea	float32	494.666	Average area for cells in pixel.
averageDnbCount	float32	194.299	Average number of mRNA-captured DNBs in a cell.
averageExpCount	float32	541.715	Average MID count in cell.
averageGeneCount	float32	310.157	Average gene count in cell.
maxArea	uint16	1925	Maximum area for cells in pixel.
maxDnbCount	uint16	883	Maximum number of mRNA-captured DNBs in a cell.
maxExpCount	uint16	3018	Maximum MID count in cell.
maxGeneCount	uint16	1415	Maximum gene count in cell.
maxX	int32	17658	Maximum x coordinate of the cell’s center of mass.
maxY	int32	19422	Maximum y coordinate of the cell’s center of mass.
medianArea	float32	474.	Median area for cells in pixel.
medianDnbCount	float32	183.	Median number of mRNA-captured DNBs in a cell.
medianExpCount	float32	491.	Median MID count in cell.

/cellBin/cell:
Dataset “cell” is a 1D array which stores basic information and indices information of cells and expression.

medianGeneCount	float32	289.	Median gene count in cell.
minArea	uint16	2	Minimum area for cells in pixel.
minDnbCount	uint16	0	Minimum number of mRNA-captured DNBs in a cell.
minExpCount	uint16	0	Minimum MID count in cell.
minGeneCount	uint16	0	Minimum gene count in cell.
minX	int32	2933	Minimum x coordinate of the cell’s center of mass.
minY	int32	5568	Minimum y coordinate of the cell’s center of mass.
Dataset DataType: compound			
	DataType	Example	Description
id	uint32	10	Cell ID index, the start ID is 0. In the Example, 10 represent the 10th cell in the data-set.
x	int32	541	The x coordinate of the cell’s center of mass. In the Example, the x coordinate of the 10th cell’s center of mass is 541.
y	int32	190	The y coordinate of the cell’s center of mass. In the Example, the x coordinate of the 10th cell’s center of mass is 190.
offset	uint32	494	The start row index of the cell in the “/cellBin/cellExp” dataset. The example represents that the gene ID index and total MID count information of the 10th cell in the “/cellBin/cellExp” dataset start from the 494th row.
geneCount	uint16	100	Gene count in the cell. In the example, 100 represents that the 100 rows in the “/cellBin/cellExp”, start from the 494th to the 593th row, contains the gene ID indices and total MID count of the gene for the 10th cell in “/cellBin/cell” dataset.
expCount	uint16	500	Cell MID count.
dnbCount	uint16	200	mRNA-captured DNBs of the cell.
area	uint16	474	Cell area in pixel.
cellTypeID	uint32	0	Cell type ID.
clusterID	uint32	20	Cell cluster ID.

/cellBin/cellBorder:

Dataset “cellBorder” is a 3D array which stores the lists of points for the bounding polygons of the cell.

Dataset Attributes	DataType	Example	Description
maxX	int32	16127	Maximum x coordinate of the bounding box of the cell.
maxY	int32	16663	Maximum y coordinate of the bounding box of the cell.
minX	int32	11129	Minimum x coordinate of the bounding box of the cell.
minY	int32	12784	Minimum y coordinate of the bounding box of the cell.
Dataset DataType: 3D array	DataType	Example	Description
-	32*(int16,int16)	[[-17,-11],[-15,-5]... [32767,32767]]	A list of 32 coordinates recording the differences between cell bounding points and the cell’s center of mass (0,0). The real coordinate of cell’s center of mass (x, y) can be obtained from “cell” dataset using cellID.

/cellBin/cellExp:

Dataset “cellExp” is a 1D array which stores the expression information of each cell.

Dataset Attributes	DataType	Example	Description
maxCount	uint16	336	Maximum MID count of a gene in a cell.
Dataset DataType: compound	DataType	Example	Description
geneID	uint32	1610	Gene IDs of the genes detected in the cell. ID is the index of “gene” dataset. In the example, 1610 represents the 1610th item in the “gene” dataset, and the name of the gene can be acquired in “gene” dataset.
count	uint16	3	MID count for the gene. In the example, (assume this is the 0th item in the “cellExp” dataset, from the “offset” and “geneCount” record in the “cell” dataset we can know that the 0th item in the “cellExp” belongs to the cell whose cellID=0) the MID count for the gene (geneID=1610) in the cell (cellID=0) is 3.

[optional] /cellBin/cellExon:

Dataset “cellExon” is a 1D array which stores the exon information for each cell.

Dataset Attributes	DataType	Example	Description
maxExon	uint16	5793	Maximum exon count of a gene in all cells.
minExon	uint16	0	Minimum exon count of a gene in all cells.
Dataset DataType: 1D array	DataType	Example	Description
-	uint16	16	Exon count in a cell, the index of the array is same to the cellID in the “cell” dataset.

[optional] /cellBin/cellExpExon:

Dataset “cellExpExon” is a 1D array which stores exon expression information for each cell.

Dataset Attributes	DataType	Example	Description
maxExon	uint16	336	Maximum exon count of a gene in a cell.
Dataset DataType: 1D array	DataType	Example	Description
-	uint16	3	Exon count (MID) for the gene. The index is same to the “cellExp” dataset. In the example, (assume this is the 0th item in the “cellExpExon” dataset, since the index is same to “cellExp” dataset, from the “offset” and “geneCount” record in the “cell” dataset we can know that the 0th item in the “cellExpExon” belongs to the cell whose cellID=0) the exon count (MID) for the gene (geneID=1610) in the cell (cellID=0) is 3.

/cellBin/cellTypeList:

Dataset “cellTypeList” is a 1D array which stores cell types of each cell.

Dataset DataType: 1D array	DataType	Example	Description
-	S32	b'default'	Cell type, “default” stands for undefined cell type.

/cellBin/gene:

Dataset “gene” is a 1D array which stores the indices of cell and expression information of each gene.

Dataset Attributes	DataType	Example	Description
maxCellCount	uint32	5718	Maximum number of cells a gene can be detected.
maxExpCount	uint32	55361	Maximum MID count of a gene.
minCellCount	uint32	1	Minimum number of cells a gene can be detected.
minExpCount	uint32	1	Minimum MID count of a gene.
Dataset DataType: compound	DataType	Example	Description
geneName	S32	b'AC149090.1'	Gene name.
offset	uint32	0	The start row index of the gene in “/cellBin/geneExp” dataset. In the example, 0 means that start from the 0th item in “/cellBin/geneExp” dataset records the cellIDs and total MID count information of “AC149090.1”.
cellcount	uint32	60	Number of cells a gene can be detected. In the example, 60 represents that start from the 0th item to the 59th item records the information of gene “AC149090.1”.
expCount	uint32	100	Sum of MID count for the gene. In the example, the total MID count of “AC149090.1” is 100.
maxMIDcount	uint16	4	Maximum MID count of a gene in a cell. In this case, the maximum MID count of gene “AC149090.1” in a cell is 4.

/cellBin/geneExp:

Dataset “geneExp” is a 1D array which stores cell and expression information of each gene.

Dataset Attributes	DataType	Example	Description
maxCount	uint16	10	Maximum MID count of a gene.
Dataset DataType: compound	DataType	Example	Description
cellID	uint32	1247	cellID that contains the gene whose index is same to the index in “gene” dataset. In the example, (assume we use the 0th item in “geneExp” dataset) 1247 shows that the gene “AC149090.1” appears in the cell whose cellID is 1247.
count	uint16	3	The MID count of the gene, whose index is same to the index in “gene” dataset, in the cellID. In the example, the MID count of gene “AC149090.1” in the cell (cellID=1247) is 3.

[optional] /cellBin/geneExon:

Dataset “geneExon” is a 1D array which stores the exon expression information of each gene.

Dataset Attributes	DataType	Example	Description
maxExon	uint32	55361	Maximum exon count of a gene.
minExon	uint32	0	Minimum exon count of a gene.
Dataset DataType: 1D array	DataType	Example	Description
-	uint32	97	Total exon count of a gene, the index of “geneExon” dataset is same to the “gene” dataset. In the example, (assume this is the 0th item in the “geneExon” dataset, and gene “AC149090.1” is the 0th item in the “gene” dataset) the exon count of gene “AC149090.1” is 97.

[optional] /cellBin/geneExpExon:

Dataset “geneExpExon” is a 1D array which stores the exon expression information in cells of each gene.

Dataset Attributes	DataType	Example	Description
maxExon	uint16	336	Maximum exon expression of a gene in a cell.
Dataset DataType: 1D array	DataType	Example	Description
-	uint16	3	Exon count of a gene in a cell. The index of “geneExpExon” dataset is same to the “geneExp” dataset. In the example, (assume this is the 0th item in the “geneExpExon” dataset, since the index is same to “geneExp” dataset, from the “offset” and “cellCount” record in the “gene” dataset we can know that the 0th item in the “geneExpExon” dataset belongs to the gene “AC149090.1”) 3 stands for the exon count of gene “AC149090.1” in cell 1247 is 3.

/cellBin/bockIndex:

Dataset “bockIndex” is a 1D array which stores the matrix block partition information.

Dataset	DataType	Example	Description
1D array			
-	uint32	0	Cell count in each partition block. cnt=blockIndex[i+1]-blockIndex[i]

/cellBin/bockSize:

Dataset “bockSize” is a 1D array which stores the block size of partition.

Dataset	DataType	Example	Description
1D array			
-	uint32	256, 256, 104, 104	4-element array. The 4 items represent the block length in x-axis, block length in y-axis, block count in x-axis, and block count in y-axis, respectively.

2.5 Gene Expression Matrix

Gene expression matrix stores genes spatial expression data. SAW generates multiple gene expression matrix files in the workflow, the basic format requires four columns with a header row that show the column names. The four columns are gene name, x coordinate, y coordinate, and MID count. The origin of **tissueCut** generated gene expression matrices have been calibrated to (0, 0). The header of expression matrix for maximum area enclosing rectangle region has six annotation rows start with “#” before the column rows. The header field names and field types are described in Table 2-5.

Table 2-5 Gene Expression Matrix Header Fields Description

Fields	DataType	Example	Description
#FileFormat	string	GEMv0.1	Gene expression matrix file format version.
#SortedBy	string	None	Gene expression matrix sorting strategy. Valid values: “geneID”, “x”, “y”, “MIDCount”, “None”.
#BinSize	uint16	1	(Please check 1.3 Terminologies and Concepts Bin)
#STOmicsChip	string	SS200000135TL_D1	Stereo-seq Chip T serial number.
#OffsetX	uint32	1	X coordinate of the origin before calibration.
#OffsetY	uint32	1	Y coordinate of the origin before calibration.
geneID	string	Cr2	Gene name.
x	uint32	16809	X coordinate of the spot.
y	uint32	8546	Y coordinate of the spot.
MIDCount	uint32	1	Number of MIDs at (x, y) for the gene in the corresponding row.
ExonCount	uint32	0	(Optional) Number of exon count at (x, y) for the gene in the corresponding row.
CellID	uint32	55892	(Optional) CellID for (x, y).

Example of GEM:

```
#FileFormat=GEMv0.1
#SortedBy=None
#BinSize=1
#STOmicsChip=SS200000135TL_D1
#OffsetX=0
#OffsetY=0
geneID      x      y      MIDCount      ExonCount      CellID
Ptgds      7585      19729      1      1      55892
Cdk8      7582      19730      2      0      55892
1500011K16Rik      7585      19730      2      2      55892
```

2.6 Image Process Record File

Image process record (IPR) file is designed to record the whole-life information of a microscopic staining image from photo-taking to processing. Each staining image (ssDNA/DAPI, IF, HE) includes six basic groups, “ImageInfo”, “QCInfo”, “Stitch”, “TissueSeg”, “CellSeg”, and “Register”, which are used to store microscopy photo-taking information, image quality control information, image stitching records, tissue segmentation records, cell segmentation records (optional), and registration records. Check <https://github.com/STOmics/SAW/tree/main/Documents/FileFormat> to get the schematic diagram of IPR.

Table 2-6 IPR File Format and Text Fields Description

File Attribute & Dataset	
Attributes	Description
IPRVersion	IPR file format version.
Dateset	Description
Preview	A 2D matrix merges stitched image, tissue segmentation boundary and cell segmentation boundary.
/<stainType>/ImageInfo: Group records basic image information.	
Attributes	Description
AppFileVer	Microscope software version.
BackgroundBalance	Background balance.
BitDepth	The bit-depth of a camera sensor describes its ability to transform the analog signal coming from the pixel array into a digital signal.
Brightness	Relative intensity affecting a person or sensor.
ChannelCount	Number of RGB channels.
ColorEnhancement	Whether enhanced image color display or not.

/<stainType>/ImageInfo: Group records basic image information.

Contrast	The difference in color and intensity of the depicted object from its background.
DeviceSN	Microscope device serial number.
DistortionCorrection	Whether fixed distortion or not.
ExposureTime	Exposure time in ms.
FOVHeight	Height of an individual FOV in pixel.
FOVWidth	Width of an individual FOV in pixel.
Gain	Amplification applied to the signal by the image sensor.
Gamma	The coefficient links between the human eye and the digital camera.
GammaShift	Whether adapt the digital image taken with the help of a linearly re-cording camera to the nonlinear perception of the human eye or not.
Illuminance	Intensity of light.
Manufacture	Microscope manufacture.
Model	Microscope model.
Overlap	Overlapping pixels between single tiles.
Pitch	Physical pitch (nm) between neighbor spots.
PixelSizeX	Size of pixel in x direction.
PixelSizeY	Size of pixel in y direction.
QCResultFile	Prefix of ImageQC/ImageStudio result file, the unique identifier of the image.
ScanChannel	Fluorescence channel.
ScanCols	Number of columns scanned.
ScanObjective	Magnification power of the scan objective lens.
ScanRows	Number of rows scanned.
ScanTime	Scan date and time.
Sharpness	Degree of clarity of the edge(s) of the image.
StereoResepVersion	Stereo-resep version.
StitchedImage	Whether the corresponding image is a panorama image (true) or a set of tiled images (false).
STOmicsChipSN	Stereo-seq Chip T serial number.
WhiteBalance	An adjustment in electronic and film imaging that corrects the color balance of the lighting.

Dataset DataType: 1D array

Description

RGBScale RGB color.

/<stainType>/QCInfo:**Group records the QC information of the image.**

Attributes	Description
ClarityScore	Reference score for evaluating the clearness of cell boundaries.
Experimenter	Email of the experimenter who did QC for the image.
GoodFOVCount	Number of FOVs that have identified more than 3 track cross points.
ImageQCVersion	ImageQC/imageStudio version.
QCPassFlag	Whether the corresponding image passed QC.
RemarkInfo	Any remarks, notes, comments on the image.
StainType	Stain type of microscopy image.
TotalFOVCount	Total number of FOVs.
TrackLineScore	Reference score for evaluating whether the detected track lines can be used as references for image stitching and registering with gene expression matrix. (This score only evaluate whether the program detected track lines on the image, it does not infer the clarity of the lines or the images).
TrackLineChannel	Shooting channel of track lines.
TrackCrossQCPassFlag	Whether the QC of the track-cross point is passed.
ScopeStitchQCScore	Score to assess the stability of microscope overlap.
ScopeStitchQCMatrix	Matrix of overlap deviation of each microscope FOV.
ScopeStitchQCPassFlag	Whether the QC of microscope stitch is passed.
Dataset DataType: 2D array	Description
ScopeStitchQCMatrix	Matrix of overlap deviation of each microscope FOV.
CrossPoints/row_col*n	Group of datasets for each FOV that records the track cross point coordinates. (Row and col stand for the FOV row and column index number, and n stands for number of FOVs). Each dataset is a 2D array records, (x, y) coordinates of track cross points in each FOV.

/<stainType>/Calibration**Group records the calibration information (only <protein>_IF).**

Attributes	Description
CalibrationQCPassFlag	Whether the calibration QC of IF image is passed.

/<stainType>/Calibration/Scope

Attributes	Description
Confidence	Calibration confidence of microscope-stitched tiled image vs. IF.
OffsetX	Horizontal offset.
OffsetY	Vertical offset.

[optional]<stainType>/Calibration/BGI

Attributes	Description
Confidence	Calibration confidence of BGI-stitched image vs. IF.
OffsetX	Horizontal offset.
OffsetY	Vertical offset.

**/<stainType>/Stitch:
Group records the stitching information.**

Attributes	Description
StitchingScore	Reference score for stitching.
TemplateSource	The reference FOV for deriving the template used for rotating and scaling the microscopic images.
Dataset DataType: 2D array	Description
TemplatePoint	Center coordinates for deriving template lines.
TransformTemplate	Coordinates of template points registered with expression matrix.

**/<stainType>/Stitch/BGIStitch:
Group records the image stitching information processed by BGI program.**

Attributes	Description
StitchedGlobalHeight	Height of stitched tiled images using BGI stitching algorithm. Tiled image only.
StitchedGlobalWidth	Width of stitched tiled images using BGI stitching algorithm. Tiled image only.
Dataset DataType: 2D array	Description
StitchedGlobalLoc	Coordinates for the BGI stitched tiled image.

**/<stainType>/Stitch/ScopeStitch:
Group records the image stitching information processed by microscope imaging software.**

Attributes	Description
GlobalHeight	Height of panorama image.
GlobalWidth	Width of panorama image.
Dataset DataType: 2D array	Description
GlobalLoc	Coordinates for the stitched tiled image (either program stitched or microscope stitched).

/<stainType>/Stitch/StitchEval: Group records the evaluation result of stitching.

Attributes	Description
MaxDeviation	Maximum stitching deviation.
Dataset DataType: 2D array	Description
GlobalDeviation	Global stitching deviation matrix.
StitchEvalH	Stitching deviation matrix for the horizontal axes.
StitchEvalV	Stitching deviation matrix for the vertical axes.

/<stainType>/TissueSeg: Group records the tissue segmentation information.

Attributes	Description
TissueSegScore	Reference score for tissue segmentation.
TissueSegShape	Image shape for tissue segmentation mask image.
Dataset DataType: 2D array	Description
TissueMask	Encoded tissue segmentation mask file (before registration with gene expression matrix).

/<stainType>/TissueSeg/Labeling: Group records the labeling tissue segmentation information, specifically labeled areas.

Dataset DataType: 2D array	Description
TissueMask_<label>	Encoded labeling tissue segmentation mask file.
Attributes (within Dataset)	Description
Shape	Reference score for tissue segmentation.
Offset	Image shape for tissue segmentation mask image.
Color	Color of the labeling area.
Description	Description for the labeling area.

/<stainType>/CellSeg: Group records the cell segmentation information.

Attributes	Description
CellSegShape	Reference score for cell segmentation.
Dataset DataType: 2D array	Description
<stainType>/CellSeg/CellMask	Encoded cell segmentation mask file (before register with gene expression matrix).

/<stainType>/Register: Group records the information that align images with gene expression matrix.

Attributes	Description
CounterRot90	Count of counter-clockwise rotation of 90 degrees.
Flip	Whether horizontally flipped or not.
MatrixShape	Height and width of the gene expression matrix.
OffsetX	Offset between microscope image and gene expression matrix in x-axis.
OffsetY	Offset between microscope image and gene expression matrix in y-axis.
RegistrationScore	Reference score for registration.
Rotation	Rotation degree between raw image and deviation template.
ScaleX	Scale between raw image and deviation template in horizontal direction.
ScaleY	Scale between raw image and deviation template in vertical direction.
XStart	Gene expression matrix offset x (GEF geneExp/binN/expression attribute minX).
YStart	Gene expression matrix offset y (GEF geneExp/binN/expression attribute minY).
ManualRotation	Manual rotation degree of the raw image around the center point.
ManualScaleX	Manual scale of the raw image in horizontal direction based on center point.
ManualScaleY	Manual scale of the raw image in vertical direction based on center point.
Dataset DataType: 2D array	Description
MatrixTemplate	List of track cross points derived from gene expression matrix.

/StereoResepSwitch: Group stores the state of each module that whether the module need to be performed.

Attributes	Description
stitch	Switch for performing stitching.
tissueseg	Switch for performing tissue segmentation.
cellseg	Switch for performing cell segmentation.
register	Switch for performing registration.

/ManualState: Group stores the state of each module that whether the module has been manually processed.

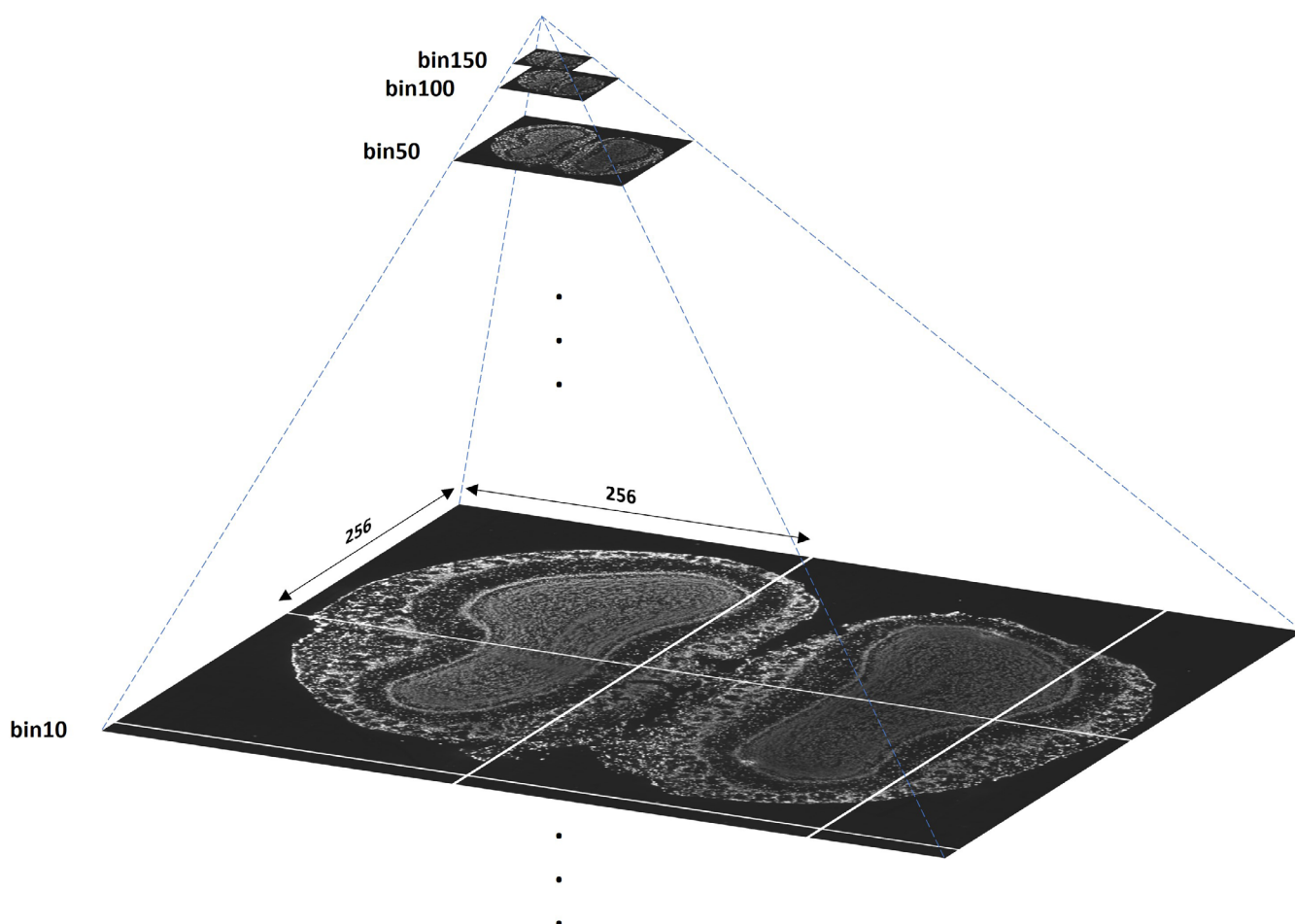
Attributes	Description
stitch	Whether manually stitched the tiled images.
tissueseg	Whether manually delineated the tissue coverage region.
cellseg	Whether manually delineated the cell coverage regions.
register	Whether manually aligned microscope image and gene expression matrix.
calibration	Whether manually calibrated two images. For example, manually match IF image with DAPI image.

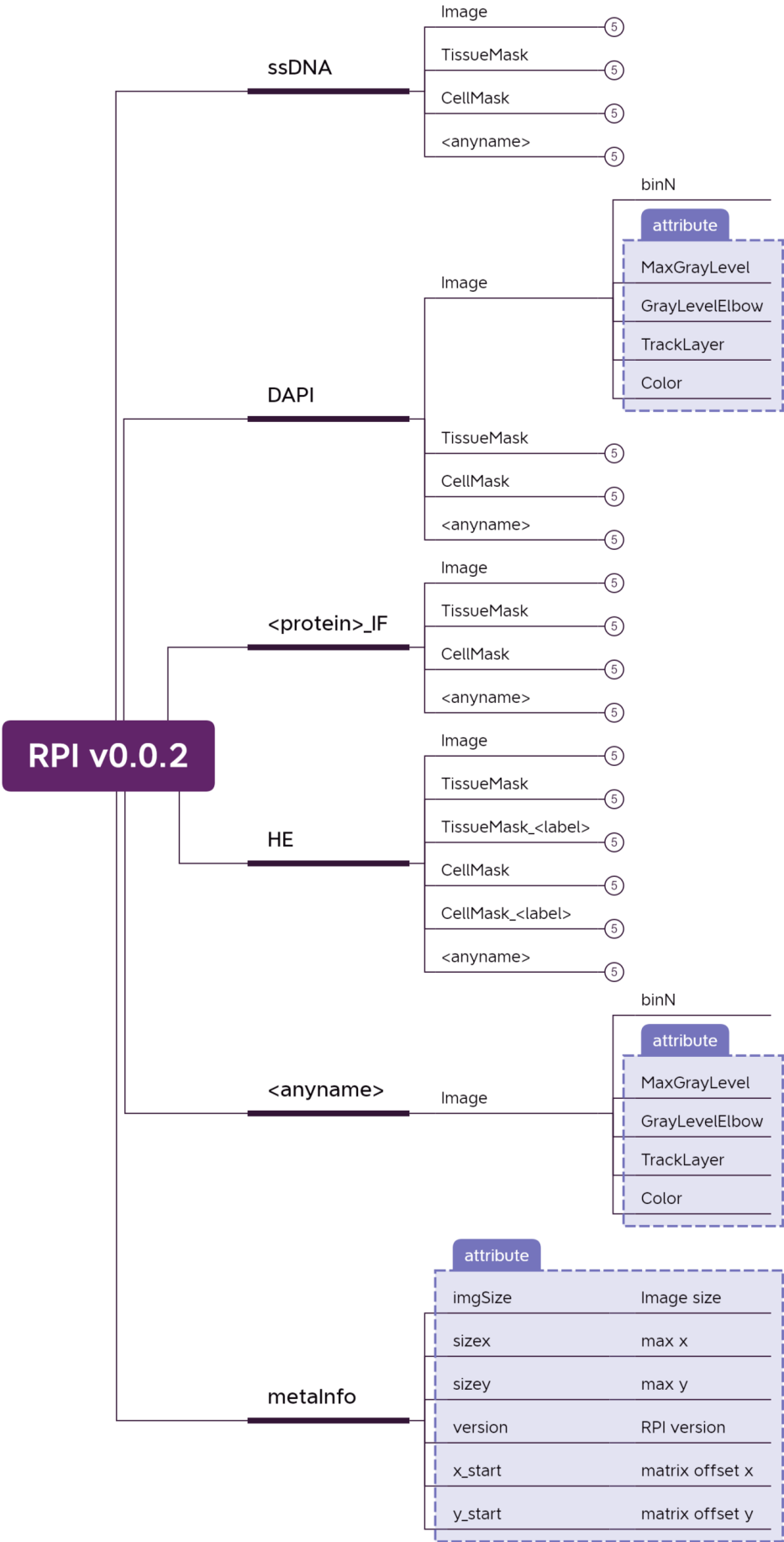
2.7 Image Pyramid

The image pyramid model is a multi-resolution hierarchical model that is used to store and display images in different resolutions. For the same field of view, the layer of the image pyramid that is closest to the bottom includes the most detailed information and has the largest scale. **register** pipeline performs the down-sampling step on the registered image, and the resulted images are layered to construct a pyramid with the suffix “.rpi”. For each resolution layer, the intact registered image is split into 256 pixels x 256 pixels tiles. If the size of a layer is smaller than 256 x 256, the image will then remain intact.

The outer layer group of RPI file is defined according to its stain type, generally including ssDNA, DAPI, IF (immunofluorescence image especially for protein, group name as <protein_IF>), and HE. In each group of stain types, multiple image results, including an image subgroup (registered microscopy image), a TissueMask subgroup (registered mask boundary for the tissue coverage area), and a CellMask subgroup (registered mask boundaries for the cell coverage area, optional), could be saved respectively.

Schematic diagram of image pyramid:





References

1. STOmics/SAW. Accessed April 17, 2023. <https://github.com/STOmics/SAW>
2. Sequence Alignment/Map Format Specification. Accessed May 21, 2021. <https://github.com/samtools/hts-specs>.

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Please raise GitHub issues for reporting bugs and requesting features:

SAW GitHub Issue Page: <https://github.com/STOmics/SAW/issues>