Modelling Competition for Nutrients between Microbial Cultures Growing on Solid Agar Surfaces

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ABSTRACT

Motivation: The fitness of microbial strains can be estimated from the growth of cultures inoculated onto solid agar. In high-throughput procedures, an array of cultures is grown on the same agar plate and competition for nutrients between cultures is likely to affect growth. However, analysis assumes that cultures grow independently. We test a model of nutrient dependent growth and diffusion and try to correct for competition to provide more accurate and reliable fitness estimates.

Results: What should we say?

Availability and Implementation: CANS, a Python2 package developed for the analysis in this paper, is freely available from github pypi.

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2 METHODS

2.1 Subsection

1 INTRODUCTION

Dummy Lawless *et al.* (2010) citations (Heydari *et al.*, 2016) (Addinall *et al.*, 2008).

1.1 Subsection

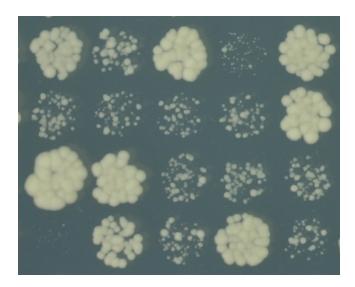


Figure 1: A section of a plate from a QFA experiment (Addinall et al., 2011).

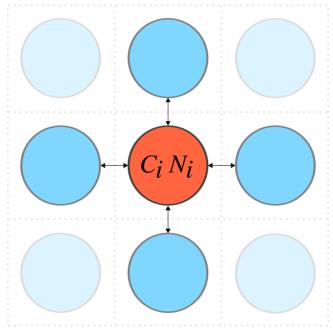


Figure 2: Schematic of the modelling approach. Each circle represents a culture, indexed i, on solid agar and arrows represent diffusion of nutrients. C_i - amount of cells; N_i - amount of nutrients; darker blue circles- neighbourhood δ_i .

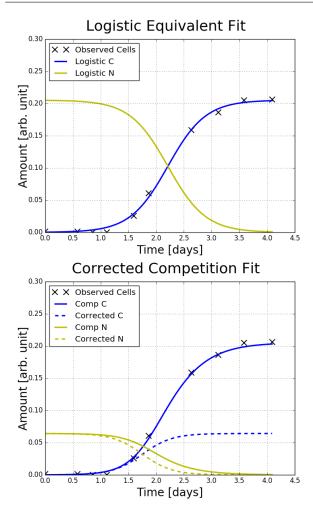


Figure 3: PUT KN VALUES (and r and K) ON THE PLOT. (Could even put obj fun values). Fits to culture (R10, C3) of P15 (Addinall et al., 2011) illustrating how the competition model can be seen as a correction to the logistic model. C-Cells; N - Nutrients. Top - Logistic Equivalent Fit; Bottom - Competition Fit (solid) and Corrected Logistic Equivalent Fit (dashed).

3 RESULTS

(Palková et al., 1997)

3.1 Subsection

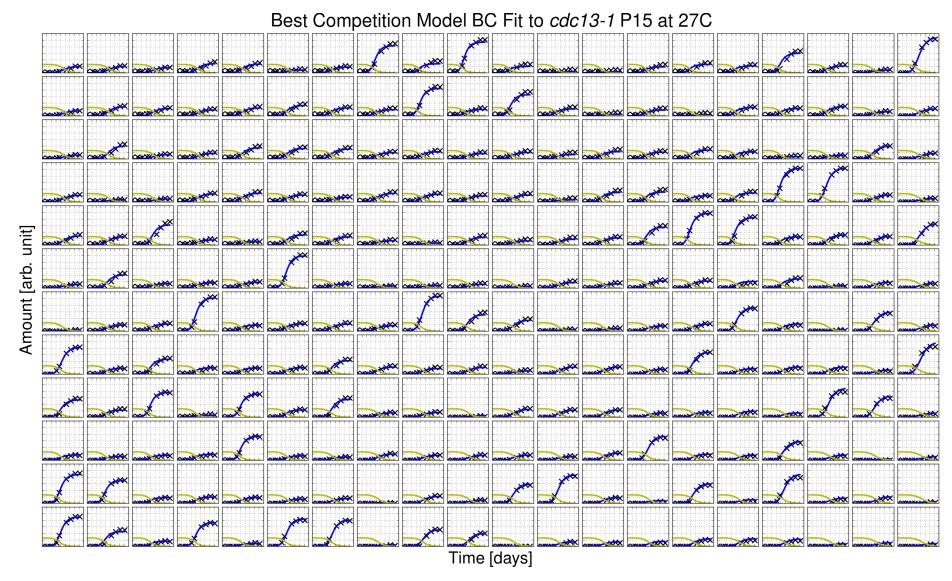


Figure 4: (R5, C18) P15

Best Competition Model BC Fit to cdc13-1 P15 at 27C (R5, C18) 0.25 0.15 0.10 0.05 0.00 Tiun 0.25 Amount [arb. content of the content 0.20 Obs. Cells 0.25 Est. Cells 0.20 Est. Nutrients 0.15 0.05 3.0 2.0 2.5 Time [days]

Figure 5: (R5, C18) P15

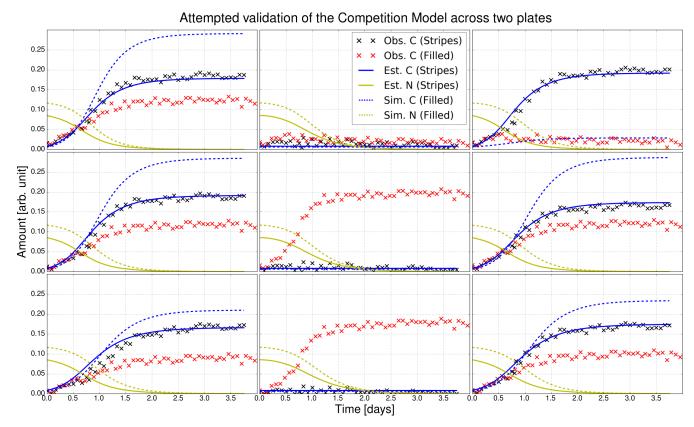


Figure 6: (R9, C10) top-right possible pinning mistake. Bottom-left not close to any such mistake.

4 DISCUSSION

4.1 Subsection REFERENCES

Addinall, S.G. *et al.* (2008) A genomewide suppressor and enhancer analysis of cdc13-1 reveals varied cellular processes influencing telomere capping in saccharomyces cerevisiae. *Genetics*, **180**, 4, 2251–2266.

Addinall, S.G. *et al.* (2011) Quantitative fitness analysis shows that nmd proteins and many other protein complexes suppress or enhance distinct telomere cap defects. *PLoS Genet*, **7**, 4, 1–16.

Heydari, J. et al. (2016) Bayesian hierarchical modelling for inferring genetic interactions in yeast. *Journal of the Royal Statistical Society: Series C (Applied Statistics)*, **65**, 3, 367–393.

Lawless, C. *et al.* (2010) Colonyzer: automated quantification of micro-organism growth characteristics on solid agar. *BMC Bioinformatics*, **11**, 1, 1–12.

Palková, Z. *et al.* (1997) Ammonia mediates communication between yeast colonies. *Nature*, **390**, 6659, 532–536.