3. Worksheet: Basic R

Kelsey Coates; Z620: Quantitative Biodiversity, Indiana University

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OVERVIEW

This worksheet introduces some of the basic features of the R computing environment (http://www.r-project.org). It is designed to be used along side the **3. RStudio** Handout in your binder. You will not be able to complete the exercises without the corresponding Handout. We recommend that you work through this Worksheet on the computer with your Handoutopen in your binder.

Directions:

1. In the Markdown version of this document in your cloned repo, change "Student Name" on line 3 (above) with your name.
2. Complete as much of the worksheet as possible during class.
3. Use the handout as a guide; it contains a more complete description of data sets along with examples of proper scripting needed to carry out the exercises.
4. Answer questions in the Worksheet. Space for your answers is provided in this document and is indicated by the ">" character. If you need a second paragraph be sure to start the first line with ">". You should notice that the answer is highlighted in green by RStudio (color may vary if you changed the editor theme).
5. Before you leave the classroom today, you may want to **push** this file to your GitHub repo, at whatever stage you are. This will enable you to pull your work onto your own computer.
6. When you have completed the worksheet, **Knit** the text and code into a single PDF file by pressing the `Knit` button in the RStudio scripting panel. This will save the PDF output in your Posit.cloud workspace: `/cloud/project/QB-2025/Week1-RStudio/`
7. After Knitting, please submit the worksheet by making a **push** to your GitHub repo and then create a **pull request** via GitHub. Your pull request should include this file (**3.RStudio_Worksheet.Rmd**) with all code blocks filled out and questions answered) and the PDF output of `Knitr` (**3.RStudio_Worksheet.pdf**).

The completed exercise is due on **Wednesday, January 22nd, 2025 before 12:00 PM (noon)**.

1) HOW WE WILL BE USING R AND OTHER TOOLS

You are working in an RMarkdown (.Rmd) file. This allows you to integrate text and R code into a single document. There are two major features to this document: 1) Markdown formatted text and 2) "chunks" of R code. Anything in an R code chunk will be interpreted by R when you *Knit* the document.

When you are done, you will *knit* your document together. However, if there are errors in the R code contained in your Markdown document, you will not be able to knit a PDF file. If this happens, you will need to review your code, locate the source of the error(s), and make the appropriate changes. Even if you are able to knit without issue, you should review the knitted document for correctness and completeness

before you submit the Worksheet. Next to the `Knit` button in the RStudio scripting panel there is a spell checker button (`ABC`) button.

2) SETTING YOUR WORKING DIRECTORY

In the R code chunk below, please provide the code to: 1) clear your R environment, 2) print your current working directory, and 3) set your working directory to your `Week1-RStudio/` folder.

```
rm(list=ls())
getwd()
```

```
## [1] "C:/Users/coatesk23/OneDrive - East Carolina University/Documents/QuantBio/QB2026_Coates/WeekOne
```

```
#setwd("C:\\Users\\coatesk23\\Documents\\QuantBio\\QB2026_Coates\\WeekOne")
```

3) USING R AS A CALCULATOR

To follow up on the pre-class exercises, please calculate the following in the R code chunk below. Feel free to reference the **1. Introduction to version control and computing tools** handout.

1) the volume of a cube with length, l, = 5 (volume = l^3)
2) the area of a circle with radius, r, = 2 (area = pi * r^2).
3) the length of the opposite side of a right-triangle given that the angle, theta, = pi/4. (radians, a.k.a. 45°) and with hypotenuse length sqrt(2) (remember: sin(theta) = opposite/hypotenuse).
4) the log (base e) of your favorite number.

```
# cube
5^3
```

```
## [1] 125
```

```
pi <- 3.14159
# circle
pi*(2^2)
```

```
## [1] 12.56636
```

```
# triangle
sin(45)
```

```
## [1] 0.8509035
```

```
#fav number
log(6)
```

```
## [1] 1.791759
```

4) WORKING WITH VECTORS

To follow up on the pre-class exercises, please perform the requested operations in the R-code chunks below.

Basic Features Of Vectors

In the R-code chunk below, do the following: 1) Create a vector `x` consisting of any five numbers. 2) Create a new vector `w` by multiplying `x` by 14 (i.e., "scalar"). 3) Add `x` and `w` and divide by 15.

```
x <- c(1, 2, 3, 4, 5)
w <- x * 14
(x + w)/15
```

```
## [1] 1 2 3 4 5
```

Now, do the following: 1) Create another vector (`k`) that is the same length as `w`. 2) Multiply `k` by `x`. 3) Use the combine function to create one more vector, `d` that consists of any three elements from `w` and any four elements of `k`.

```
k <- c(7, 6, 42, 10, 15)
k * x
```

```
## [1]   7  12 126  40  75
```

```
d <- c(sample(w, 3), sample(k, 4))
d
```

```
## [1] 70 14 28 15 10  6  7
```

Summary Statistics of Vectors

In the R-code chunk below, calculate the **summary statistics** (i.e., maximum, minimum, sum, mean, median, variance, standard deviation, and standard error of the mean) for the vector (`v`) provided.

```
v <- c(16.4, 16.0, 10.1, 16.8, 20.5, NA, 20.2, 13.1, 24.8, 20.2, 25.0, 20.5, 30.5, 31.4, 27.1)
max(v, na.rm = TRUE)
```

```
## [1] 31.4
```

```
min(v, na.rm = TRUE)
```

```
## [1] 10.1
```

```
sum(v,na.rm = TRUE)
```

```
## [1] 292.6
```

```r
mean(v,na.rm = TRUE)
```

```
## [1] 20.9
```

```r
median(v,na.rm = TRUE)
```

```
## [1] 20.35
```

```r
var(v,na.rm = TRUE)
```

```
## [1] 39.44
```

```r
sd(v,na.rm = TRUE)
```

```
## [1] 6.280127
```

```r
se <- sd(v, na.rm = TRUE) / sqrt(sum(!is.na(v)))
```

5) WORKING WITH MATRICES

In the R-code chunk below, do the following: Using a mixture of Approach 1 and 2 from the **3. RStudio** handout, create a matrix with two columns and five rows. Both columns should consist of random numbers. Make the mean of the first column equal to 8 with a standard deviation of 2 and the mean of the second column equal to 25 with a standard deviation of 10.

```r
set.seed(123)   # for reproducibility

col1 <- rnorm(5, mean = 8, sd = 2)     # first column
col2 <- rnorm(5, mean = 25, sd = 10)   # second column

matrix <- cbind(col1, col2)
matrix
```

```
##           col1     col2
## [1,]  6.879049 42.15065
## [2,]  7.539645 29.60916
## [3,] 11.117417 12.34939
## [4,]  8.141017 18.13147
## [5,]  8.258575 20.54338
```

Question 1: What does the `rnorm` function do? What do the arguments in this function specify? Remember to use `help()` or type `?rnorm`.

> Answer 1: The rnorm function generates random numbers that are normally distributed and the arguments allow the mean and standard deviation to be specified.

In the R code chunk below, do the following: 1) Load `matrix.txt` from the **Week1-RStudio** data folder as matrix `m`. 2) Transpose this matrix. 3) Determine the dimensions of the transposed matrix.

```r
m <- as.matrix(read.table("matrix.txt"))
t(m)
```

```
##    [,1] [,2] [,3] [,4] [,5] [,6] [,7] [,8] [,9] [,10]
## V1    8    5    2    3    9   11    2    3    5     6
## V2    1    5    5    2    9    8    2    3    5     5
## V3    7    2    4    5    1    1    5    6    1     9
## V4    6    4    3    1    1    8    8    7    3     2
## V5    1    1    3    4    2    8    5    6    6     2
```

```r
dim(m)
```

```
## [1] 10  5
```

Question 2: What are the dimensions of the matrix you just transposed?

Answer 2: 10 x 5 meaning 10 columns and five rows

Indexing a Matrix

In the R code chunk below, do the following: 1) Index matrix `m` by selecting all but the third column. 2) Remove the last row of matrix `m`.

```r
# rows, columns
m_no_third <- m[, -3]
m_no_third
```

```
##       V1 V2 V4 V5
##  [1,]  8  1  6  1
##  [2,]  5  5  4  1
##  [3,]  2  5  3  3
##  [4,]  3  2  1  4
##  [5,]  9  9  1  2
##  [6,] 11  8  8  8
##  [7,]  2  2  8  5
##  [8,]  3  3  7  6
##  [9,]  5  5  3  6
## [10,]  6  5  2  2
```

```r
m_no_last_row <- m[-10,]
m_no_last_row
```

```
##       V1 V2 V3 V4 V5
##  [1,]  8  1  7  6  1
##  [2,]  5  5  2  4  1
##  [3,]  2  5  4  3  3
##  [4,]  3  2  5  1  4
##  [5,]  9  9  1  1  2
##  [6,] 11  8  1  8  8
##  [7,]  2  2  5  8  5
##  [8,]  3  3  6  7  6
##  [9,]  5  5  1  3  6
```

6) BASIC DATA VISUALIZATION AND STATISTICAL ANALYSIS

Load Zooplankton Data Set

In the R code chunk below, do the following: 1) Load the zooplankton data set from the **Week1-RStudio/** data folder. 2) Display the structure of this data set.

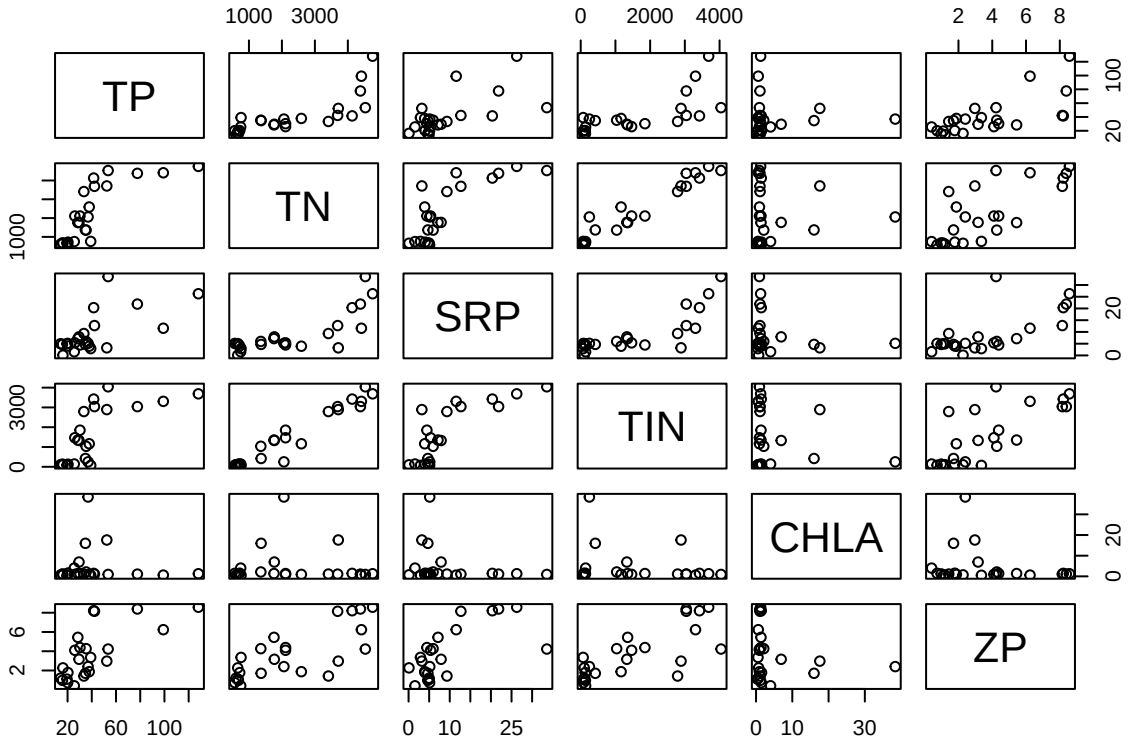
```
zoop <- read.table("zoops.txt", header = TRUE)
zoop_nuts <- read.table("zoop_nuts.txt", header = TRUE)

str(zoop)
```

```
## 'data.frame':    24 obs. of  11 variables:
##  $ TANK: int  5 14 16 21 23 25 27 34 12 15 ...
##  $ NUTS: chr  "L" "L" "L" "L" ...
##  $ CAL : num  70.5 27.1 5.3 79.2 31.4 22.7 0 35.7 74.8 5.3 ...
##  $ DIAP: num  0 19.2 8.8 17.9 0 ...
##  $ CYCL: num  66.1 129.6 12.7 141.3 11 ...
##  $ BOSM: num  2.2 0 0 3.4 0 0 0 0 0 0 ...
##  $ SIMO: num  417.8 0 73.1 0 482 ...
##  $ CERI: num  159.8 79.4 107.5 199 101.9 ...
##  $ NAUP: num  0 0 1.2 0 0 1.2 1.6 3.1 0 1.4 ...
##  $ DLUM: num  0 0 0 0 0 6.6 0 0 0 0 ...
##  $ CHYD: num  267 159 3158 298 580 ...
```

Correlation

In the R-code chunk below, do the following: 1) Create a matrix with the numerical data in the `meso` dataframe. 2) Visualize the pairwise **bi-plots** of the six numerical variables. 3) Conduct a simple **Pearson's correlation** analysis.

```
meso.num <- zoop_nuts[,3:8]
pairs(meso.num)
```




```
cor1 <- cor(meso.num)
cor1
```

```
##               TP           TN         SRP         TIN         CHLA         ZP
## TP    1.00000000  0.786510407  0.6540957  0.7171143 -0.016659593  0.6974765
## TN    0.78651041  1.000000000  0.7841904  0.9689999 -0.004470263  0.7562474
## SRP   0.65409569  0.784190400  1.0000000  0.8009033 -0.189148017  0.6762947
## TIN   0.71711434  0.968999866  0.8009033  1.0000000 -0.156881463  0.7605629
## CHLA -0.01665959 -0.004470263 -0.1891480 -0.1568815  1.000000000 -0.1825999
## ZP    0.69747649  0.756247384  0.6762947  0.7605629 -0.182599904  1.0000000
```

Question 3: Describe some of the general features based on the visualization and correlation analysis above.

> Answer 3: Zooplankton communities were positively correlated with TN, SRP, TIN, and ZIP. Zooplankton communities were negatively correlated with chlorophyll A.

In the R code chunk below, do the following: 1) Redo the correlation analysis using the `corr.test()` function in the `psych` package with the following options: method = "pearson", adjust = "BH". 2) Now, redo this correlation analysis using a non-parametric method. 3) Use the print command from the handout to see the results of each correlation analysis.

```
install.packages("psych", repos="http://cran.rstudio.com/")
```

```
## Installing package into 'C:/Users/coatesk23/AppData/Local/R/win-library/4.5'
## (as 'lib' is unspecified)
```

```
## package 'psych' successfully unpacked and MD5 sums checked
##
## The downloaded binary packages are in
##  C:\Users\coatesk23\AppData\Local\Temp\Rtmpsh6ghJ\downloaded_packages
```

```
require("psych")
```

```
## Loading required package: psych
```

```
## Warning: package 'psych' was built under R version 4.5.2
```

```
cor2 <- corr.test(meso.num, method = "pearson", adjust = "BH")
print(cor2, digits = 3)
```

```
## Call:corr.test(x = meso.num, method = "pearson", adjust = "BH")
## Correlation matrix
##          TP     TN    SRP    TIN   CHLA     ZP
## TP    1.000  0.787  0.654  0.717 -0.017  0.697
## TN    0.787  1.000  0.784  0.969 -0.004  0.756
## SRP   0.654  0.784  1.000  0.801 -0.189  0.676
## TIN   0.717  0.969  0.801  1.000 -0.157  0.761
## CHLA -0.017 -0.004 -0.189 -0.157  1.000 -0.183
## ZP    0.697  0.756  0.676  0.761 -0.183  1.000
## Sample Size
## [1] 24
## Probability values (Entries above the diagonal are adjusted for multiple tests.)
##         TP    TN   SRP   TIN  CHLA    ZP
## TP   0.000 0.000 0.001 0.000 0.983 0.000
## TN   0.000 0.000 0.000 0.000 0.983 0.000
## SRP  0.001 0.000 0.000 0.000 0.491 0.000
## TIN  0.000 0.000 0.000 0.000 0.536 0.000
## CHLA 0.938 0.983 0.376 0.464 0.000 0.491
## ZP   0.000 0.000 0.000 0.000 0.393 0.000
##
##  To see confidence intervals of the correlations, print with the short=FALSE option
```

```
install.packages("corrplot", repos="http://cran.rstudio.com/")
```

```
## Installing package into 'C:/Users/coatesk23/AppData/Local/R/win-library/4.5'
## (as 'lib' is unspecified)
```

```
## package 'corrplot' successfully unpacked and MD5 sums checked
##
## The downloaded binary packages are in
##  C:\Users\coatesk23\AppData\Local\Temp\Rtmpsh6ghJ\downloaded_packages
```

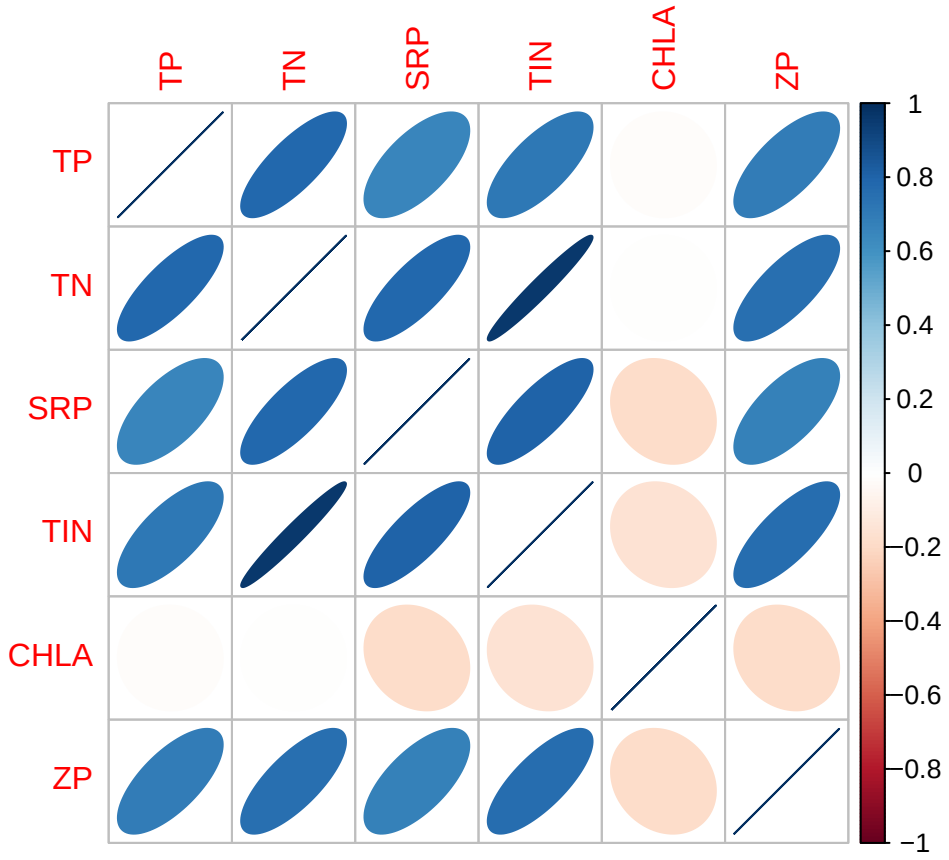
```
require("corrplot")
```

```
## Loading required package: corrplot
```

```
## Warning: package 'corrplot' was built under R version 4.5.2
```

```
## corrplot 0.95 loaded
```

```
corrplot(cor1, method = "ellipse")
```




```
dev.off()
```

```
## null device
##           1
```

```
cor2
```

```
## Call:corr.test(x = meso.num, method = "pearson", adjust = "BH")
## Correlation matrix
##          TP   TN   SRP   TIN  CHLA    ZP
## TP    1.00 0.79  0.65  0.72 -0.02  0.70
## TN    0.79 1.00  0.78  0.97  0.00  0.76
## SRP   0.65 0.78  1.00  0.80 -0.19  0.68
## TIN   0.72 0.97  0.80  1.00 -0.16  0.76
## CHLA -0.02 0.00 -0.19 -0.16  1.00 -0.18
## ZP    0.70 0.76  0.68  0.76 -0.18  1.00
## Sample Size
## [1] 24
## Probability values (Entries above the diagonal are adjusted for multiple tests.)
##        TP   TN  SRP  TIN CHLA   ZP
```

```
## TP   0.00 0.00 0.00 0.00 0.98 0.00
## TN   0.00 0.00 0.00 0.00 0.98 0.00
## SRP  0.00 0.00 0.00 0.00 0.49 0.00
## TIN  0.00 0.00 0.00 0.00 0.54 0.00
## CHLA 0.94 0.98 0.38 0.46 0.00 0.49
## ZP   0.00 0.00 0.00 0.00 0.39 0.00
##
##  To see confidence intervals of the correlations, print with the short=FALSE option
```

Question 4: Describe what you learned from `corr.test`. Specifically, are the results sensitive to whether you use parametric (i.e., Pearson's) or non-parametric methods? When should one use non-parametric methods instead of parametric methods? With the Pearson's method, is there evidence for false discovery rate due to multiple comparisons? Why is false discovery rate important?

> Answer 4: This correlation test also shows that zooplankton communities were positively correlated with TN, SRP, TIN, and ZIP and negatively correlated with chlorophyll A with about the same values as the first test considering rounding to the three digits. These results were not sensitive to parametric vs non-parametric methods. Parametric methods should be used over non-parametric methods when the data is normally distributed and measured on an interval/ratio scale. Non-parametric tests should be used when the data is not normally distributed and/or ordinal/rank data. Yes, Pearson's method is likely to have false positives if a correction for multiple comparisons is not completed. FDR is important because large correlation matrices that are common in ecological data are likely to produce false positives due to the t-test's way of testing the null hypothesis of each variable independently.

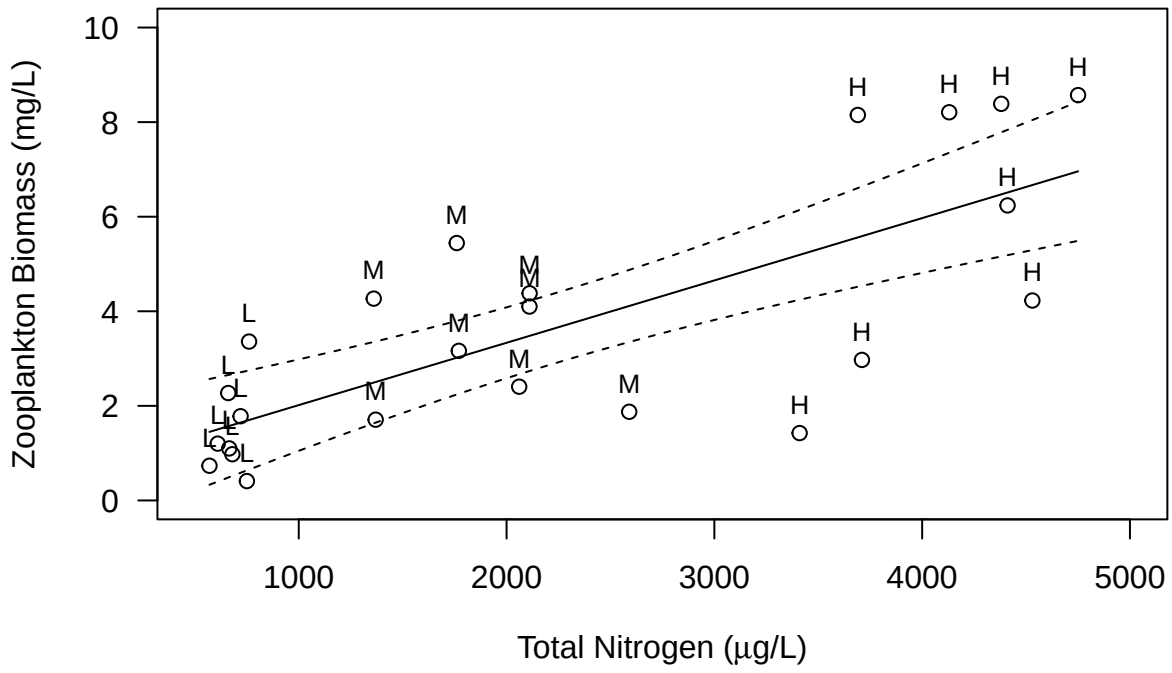
Linear Regression

In the R code chunk below, do the following: 1) Conduct a linear regression analysis to test the relationship between total nitrogen (TN) and zooplankton biomass (ZP). 2) Examine the output of the regression analysis. 3) Produce a plot of this regression analysis including the following: categorically labeled points, the predicted regression line with 95% confidence intervals, and the appropriate axis labels.

```
fitreg <- lm(ZP ~ TN, data = zoop_nuts)
summary(fitreg)
```

```
##
## Call:
## lm(formula = ZP ~ TN, data = zoop_nuts)
##
## Residuals:
##     Min      1Q  Median      3Q     Max
## -3.7690 -0.8491 -0.0709  1.6238  2.5888
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept) 0.6977712  0.6496312   1.074    0.294
## TN          0.0013181  0.0002431   5.421 1.91e-05 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 1.75 on 22 degrees of freedom
## Multiple R-squared:  0.5719, Adjusted R-squared:  0.5525
## F-statistic: 29.39 on 1 and 22 DF,  p-value: 1.911e-05
```

```
newTN <- seq(min(zoop_nuts$TN), max(zoop_nuts$TN), 10)
regline <- predict(fitreg, newdata = data.frame(TN = newTN))

conf95 <- predict(fitreg, newdata = data.frame(TN = newTN),
                  interval = c("confidence"), level = 0.95, type = "response")


p <- plot(zoop_nuts$TN, zoop_nuts$ZP, ylim = c(0, 10), xlim = c(500, 5000),
     xlab = expression(paste("Total Nitrogen (", mu,"g/L)")),
     ylab = "Zooplankton Biomass (mg/L)", las = 1) + text(zoop_nuts$TN, zoop_nuts$ZP, zoop_nuts$NUTS, p
```




```
p
```

```
## integer(0)
```

Question 5: Interpret the results from the regression model

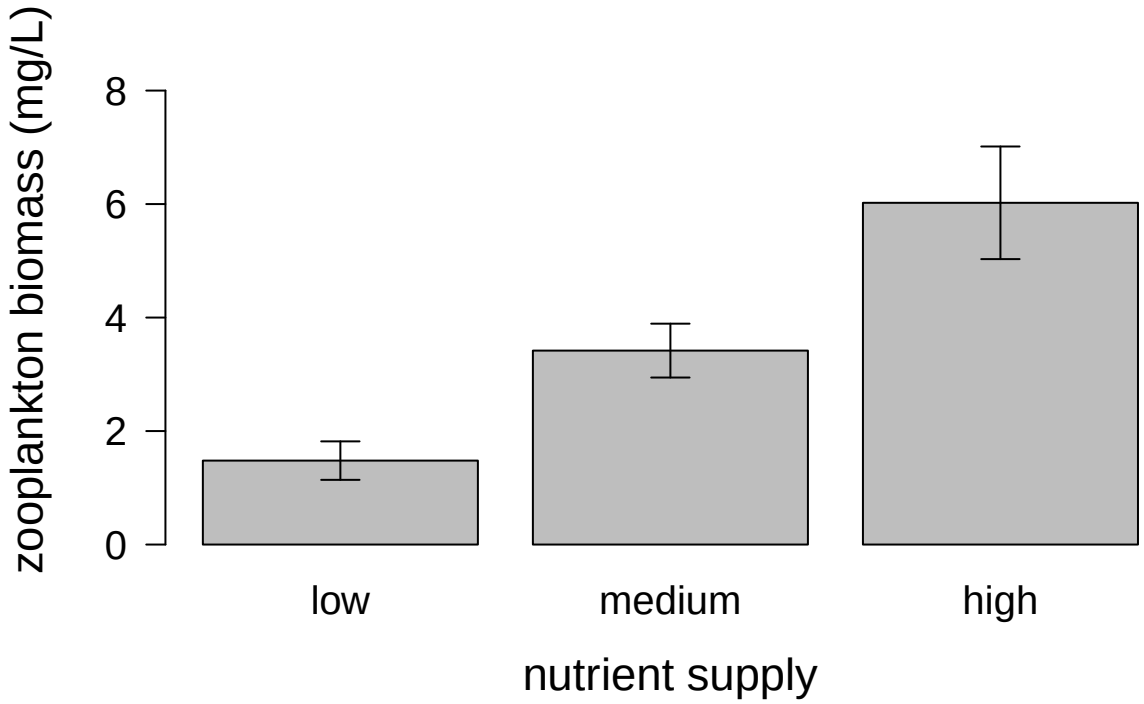
> Answer 5: The regression model shows that total nitrogen is a significant predictor of zooplankton communities with a pvalue of 1.91e-05.

Analysis of Variance (ANOVA)

Using the R code chunk below, do the following: 1) Order the nutrient treatments from low to high (see handout). 2) Produce a barplot to visualize zooplankton biomass in each nutrient treatment. 3) Include

error bars (+/- 1 sem) on your plot and label the axes appropriately. 4) Use a one-way analysis of variance (ANOVA) to test the null hypothesis that zooplankton biomass is affected by the nutrient treatment.

```
NUTS <- factor(zoop_nuts$NUTS, levels = c('L', 'M', 'H'))

zp.means <- tapply(zoop_nuts$ZP, NUTS, mean)

sem <- function(x){
  sd(na.omit(x))/sqrt(length(na.omit(x)))
}



zp.sem <- tapply(zoop_nuts$ZP, NUTS, sem)

bp <- barplot(zp.means, ylim =c(0, round(max(zoop_nuts$ZP), digits = 0)),
              pch = 15, cex = 1.25, las = 1, cex.lab = 1.4, cex.axis = 1.25,
              xlab = "nutrient supply",
              ylab = "zooplankton biomass (mg/L)",
              names.arg = c("low", "medium", "high"))
arrows(x0 = bp, y0 = zp.means, y1 = zp.means - zp.sem, angle = 90,
       length = 0.1, lwd = 1)
arrows(x0 = bp, y0 = zp.means, y1 = zp.means + zp.sem, angle = 90,
       length = 0.1, lwd = 1)
```



```r
bp
```

```
##      [,1]
## [1,]  0.7
## [2,]  1.9
## [3,]  3.1
```

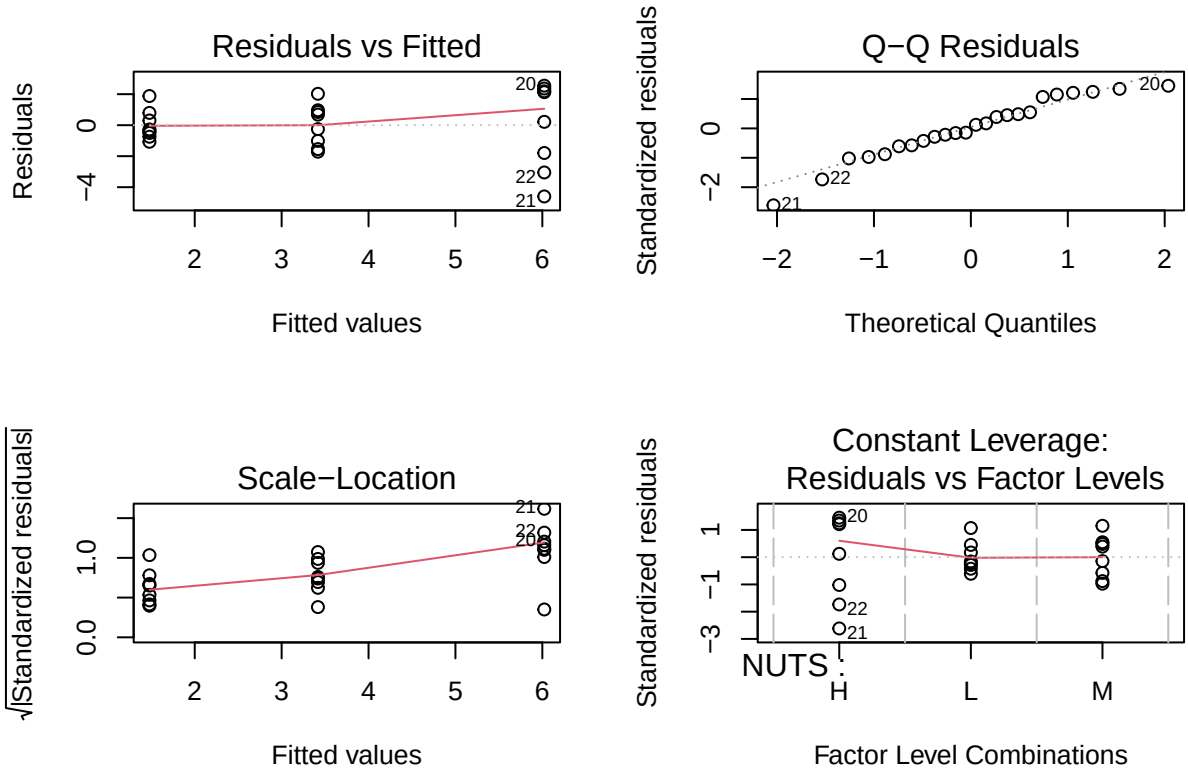
```r
fitanova <- aov(ZP ~ NUTS, data = zoop_nuts)

summary(fitanova)
```

```
##             Df Sum Sq Mean Sq F value   Pr(>F)
## NUTS         2  83.15   41.58   11.77 0.000372 ***
## Residuals   21  74.16    3.53
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```r
TukeyHSD(fitanova)
```

```
##   Tukey multiple comparisons of means
##     95% family-wise confidence level
##
## Fit: aov(formula = ZP ~ NUTS, data = zoop_nuts)
##
## $NUTS
##          diff        lwr        upr     p adj
## L-H -4.543175 -6.9115094 -2.1748406 0.0002512
## M-H -2.604550 -4.9728844 -0.2362156 0.0294932
## M-L  1.938625 -0.4297094  4.3069594 0.1220246
```

```r
par(mfrow = c(2, 2), mar = c(5.1, 4.1, 4.1, 2.1))
plot(fitanova)
```



SYNTHESIS: SITE-BY-SPECIES MATRIX

In the R code chunk below, load the zoops.txt data set in your **3.RStudio** data folder. Create a site-by-species matrix (or dataframe) that does *not* include TANK or NUTS. The remaining columns of data refer to the biomass (µg/L) of different zooplankton taxa:

- CAL = calanoid copepods
- DIAP = *Diaphanasoma* sp.
- CYL = cyclopoid copepods
- BOSM = *Bosmina* sp.
- SIMO = *Simocephallus* sp.
- CERI = *Ceriodaphnia* sp.
- NAUP = naupuli (immature copepod)
- DLUM = *Daphnia lumholtzi*
- CHYD = *Chydorus* sp.

Question 6: With the visualization and statistical tools that we learned about in the **3. RStudio** handout, use the site-by-species matrix to assess whether and how different zooplankton taxa were responsible for the total biomass (ZP) response to nutrient enrichment. Describe what you learned below in the "Answer" section and include appropriate code in the R chunk.

Answer 6: Zooplankton taxa varied in abundance by site. SIMO shown in pink was present in most sites with a relatively high abundance. BOSM shown in coral was a rare species not present in many sites and had low abundance when present.

```
zoops <- read.table("zoops.txt")
zoops_trim <- zoops[,-c(1,2)]
zoops_trim_df <- as.data.frame(zoops_trim)
rownames(zoops_trim[1,]) <- rownames(zoops_trim_df[1,])
colnames(zoops_trim_df) <- zoops_trim_df[1,]
zoops_trim_df
```

```
##      CAL  DIAP  CYCL BOSM   SIMO  CERI NAUP DLUM   CHYD
## 1    CAL  DIAP  CYCL BOSM   SIMO  CERI NAUP DLUM   CHYD
## 2   70.5   0.0  66.1  2.2  417.8 159.8  0.0  0.0  266.9
## 3   27.1  19.2 129.6  0.0    0.0  79.4  0.0  0.0  158.7
## 4    5.3   8.8  12.7  0.0   73.1 107.5  1.2  0.0 3158.2
## 5   79.2  17.9 141.3  3.4    0.0 199.0  0.0  0.0  298.5
## 6   31.4   0.0  11.0  0.0  482.0 101.9  0.0  0.0  580.2
## 7   22.7 285.1 153.0  0.0  241.5 135.5  1.2  6.6  262.4
## 8    0.0   2.3  11.0  0.0   73.1 185.0  1.6  0.0 2004.4
## 9   35.7  65.9 102.9  0.0    0.0 318.5  3.1  0.0 1260.7
## 10  74.8 178.7 266.5  0.0    0.0   1.9  0.0  0.0 1190.9
## 11   5.3   4.9  87.8  0.0 1099.2 136.4  1.4  0.0 2939.6
## 12  18.4   2.3  29.4  0.0  393.8 147.6  1.2  0.0 4857.3
## 13  14.0   2.3  37.7  0.0 1251.5  74.8  0.0  0.0 2725.5
## 14  14.0   2.3 132.9  0.0  818.6  98.1  1.2  0.0  814.5
## 15  48.8   2.3 107.9  2.2    9.0 132.7  0.0  0.0 2867.5
## 16   0.0   0.0  17.7  0.0  145.3  19.7  0.0  0.0 4201.6
## 17 292.0 269.5 373.4 10.7    0.0   8.5  1.2  0.0 1456.8
## 18   9.7   0.0  41.1  0.0 2397.8   9.4  0.0  0.0 5697.9
## 19   0.0   2.3   0.0  0.0  225.5  24.3  0.0  0.0 8323.2
## 20   5.3   0.0  86.2  0.0  465.9 527.7  1.2  0.0 3146.9
## 21  14.0   7.5  69.5  0.0  594.2  78.5  0.0  0.0 7629.2
## 22   0.0  24.4 101.2  0.0  313.6 176.6  0.0  0.0 7597.6
## 23   0.0   7.5 253.2  8.3    0.0 112.1  1.6  0.0 2594.8
## 24   5.3   2.3  96.2  0.0  786.6  76.6  0.0  0.0  463.0
## 25   0.0   2.3  66.1  0.0  826.7  85.1  0.0  0.0 5263.0
```

```
zoops_trim_df <- zoops_trim_df[-1,]
zoops_matrix <- as.matrix(zoops_trim_df)

library(dplyr)
```

```
##
## Attaching package: 'dplyr'

## The following objects are masked from 'package:stats':
##
##     filter, lag

## The following objects are masked from 'package:base':
##
##     intersect, setdiff, setequal, union
```

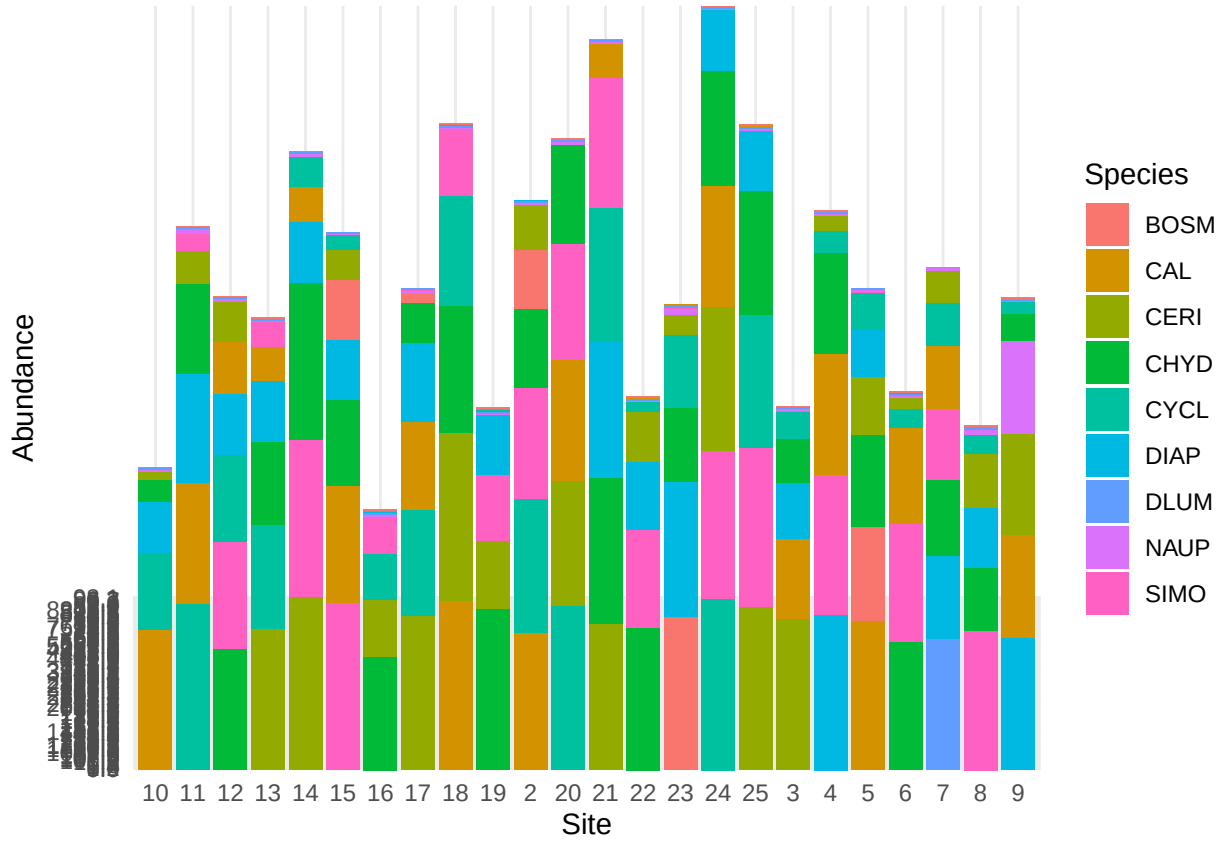
```
library(ggplot2)
```

```
## Warning: package 'ggplot2' was built under R version 4.5.2
```

```
##
## Attaching package: 'ggplot2'
```

```
## The following objects are masked from 'package:psych':
##
##     %+%, alpha
```

```
library(tidyr)

zoops_trim_df <- zoops_trim_df %>% mutate(Site = c(1:24))
df_long <- as.data.frame(zoops_trim_df) %>% mutate(Site = rownames(.)) %>%
  pivot_longer(-Site, names_to = "Species", values_to = "Abundance")

ggplot(df_long, aes(x= Site, y = Abundance, fill = Species)) +
  geom_bar(stat = "identity") +
  theme_minimal()
```



SUBMITTING YOUR WORKSHEET

Use Knitr to create a PDF of your completed **3.RStudio_Worksheet.Rmd** document, push the repo to GitHub, and create a pull request, but *do not* self-merge it. In a collaboration when using github, you will

often assign a pull request to *another* person. This alerts your collaborator to the proposed changes and allows them to look them over before accepting them. To formally **assign** a pull request to someone, you will see an option on the right hand side to choose a reviewer or a assignee. If you click on the gear icon, a list of collaborators will come up. Your primary instructor (`jaytlennon`) should be one of those on the QBstudent organization. Click on his username to assign before you finalize the pull request.

Please make sure your updated repo include both the PDF and RMarkdown files.

This assignment is due on **Wednesday, January 22nd, 2025 at 12:00 PM (noon)**.