

SUMMARY

The objective of this work is to **apply Bayesian inference on parameters** of a simple viral infection dynamics model, with **non-linear parameter dependence**, in order to determine if the parameter values used in generating the artificial experimental data are included in the bulk of posterior parameter distribution.

A model for viral infection dynamics [1] was developed in gPROMS, then artificial noisy-data points were created from the simulation results obtained with a set of known parameter values and initial conditions.

Parameter posterior distribution was estimated using an ensemble Markov chain Monte Carlo (MCMC) sampling technique [2] and a uniform parameter prior distribution.

MOTIVATION

In the Bayesian approach, the inference problem is formulated using Bayes theorem. On one hand, this presents the following advantages:

- * prior knowledge can be incorporated;
- * there is no limitation with respect to shape of inferred variable distribution.

... on the other hand, the methods required for the calculation of posterior distribution are very computationally expensive (many likelihood function evaluations).

$$p(\theta|Data, M) = \frac{p(Data|\theta, M) \cdot p(\theta|M)}{p(Data|M)}$$

Can capture a non-Gaussian distribution

Include prior knowledge

Denominator is not available in a closed-form

Figure 1: Bayes theorem applied to parameter estimation.

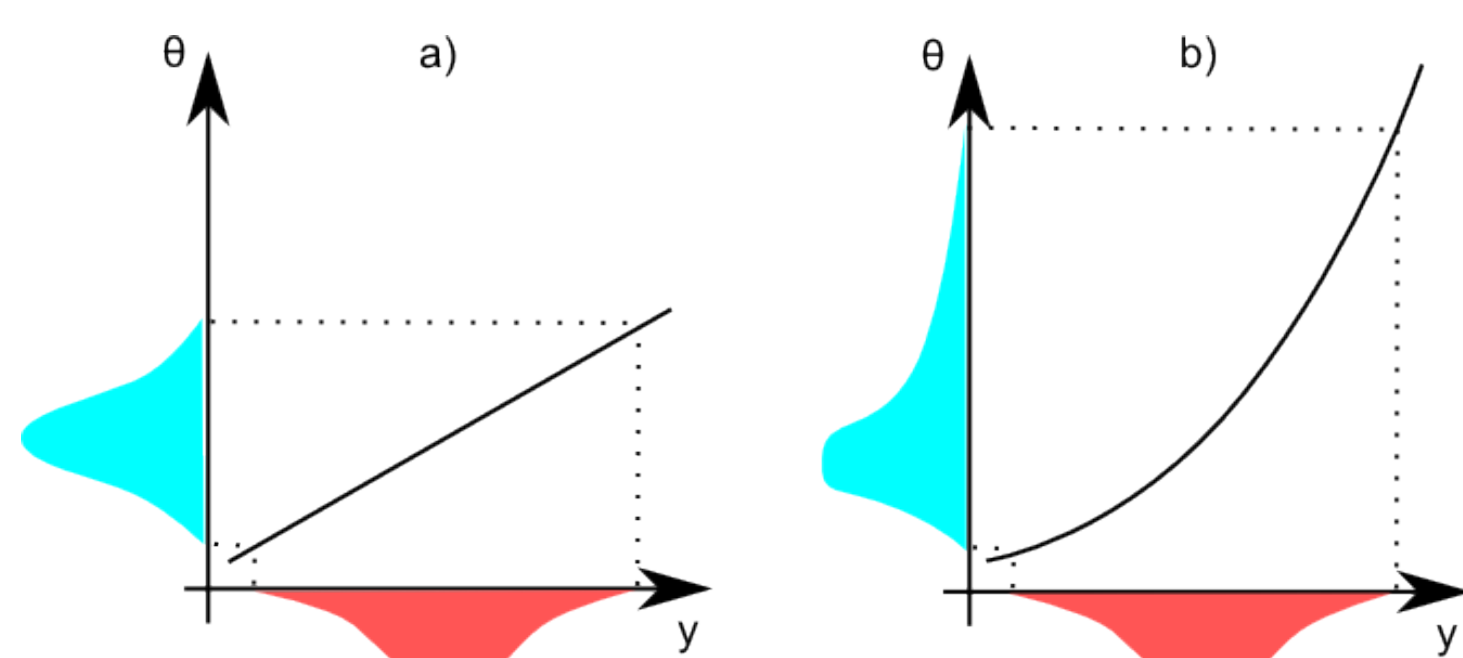


Figure 2: In case of models with non-linear parameter dependence (b), the degeneracy from normality manifests as skewness or/and multi-modality.

CALIBRATION DATA

In order to test if the “true” parameter values are included in the bulk of parameter posterior distribution, artificial calibration data was generated through simulation of the model with parameter values from table 1 and addition of Gaussian noise.

The number of uninfected cells, infected cells, antibodies and virus particles were recorded every 6 hours for 30 days.

The errors were applied to measured variables and considered normally distributed around recorded value with a spread given by a standard deviation equal to $0.3 \cdot \bar{Y}$, where \bar{Y} is any of the measured variables.

Parameter	Value	Units
a	0.4	day^{-1}
b	0.5	day^{-1}
β	2×10^{-7}	day^{-1}
c	10	day^{-1}
d	0.102	day^{-1}
k	100	day^{-1}
λ	1×10^5	day^{-1}
p	5×10^{-6}	—
s	0.3	—
u	5	day^{-1}

Table 1: Parameters used to generate the calibration data. Initial conditions were: $x(0) = 10^6$, $y(0) = 0$, $z(0) = 100$, and $v(0) = 10$.

ACKNOWLEDGEMENTS

This research received financial support from the European Union’s Horizon 2020 research and innovation programme under grant agreement No 675585 (Marie-Curie ITN “SyMBioSys”). Author is a Marie-Curie Early Stage Researcher at Process Systems Enterprise Ltd and a PhD student at Imperial College London, being involved within SyMBioSys project.

REFERENCES

- [1] M. A. Nowak and R. M. May. *Virus dynamics: Mathematical principles of immunology and virology*. Oxford University Press, 1st edition, 2000.
- [2] Daniel Foreman-MacKey. emcee: The MCMC Hammer. *Chicago Journals*, 125(925):306–312, March 2013.
- [3] John Skilling. Nested sampling for general bayesian computation. *Bayesian Analysis*, 1(4):833–860, 2006.

MODEL

Model description:

The uninfected cells within an analysed tissue are replenished from external sources with the constant rate, λ , follow a logistic growth together with infected cells, and die at a rate $d \cdot x$.

Free viruses infect cells at a rate $\beta \cdot x \cdot v$ and decay with rate $u \cdot v$ in absence of any immune system reaction.

Infected cells release new free viruses at a rate $k \cdot y$ and die with a rate $(d + a)y$.

As a response to viral infection, the immune system produces antibodies, which remove free viruses with a rate $p \cdot v \cdot z$ and their number reaches a saturation level. In absence of free virus particles, the antibodies would decay at a rate $b \cdot z$.

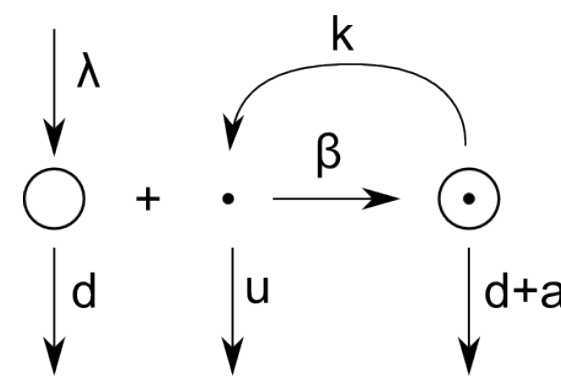


Figure 3: Basic viral infection mechanism [1].

Mathematical model:

$$\dot{x} = \lambda + r \cdot x \left(1 - \frac{x+y}{K}\right) - d \cdot x - \beta \cdot x \cdot v \quad (1)$$

$$\dot{y} = r \cdot y \left(1 - \frac{x+y}{K}\right) + \beta \cdot x \cdot v - (d + a)y \quad (2)$$

$$\dot{z} = \frac{c \cdot v \cdot z}{1 + s \cdot z} - b \cdot z \quad (3)$$

$$\dot{v} = k \cdot y - u \cdot v - p \cdot v \cdot z \quad (4)$$

Notations:

x , y , z , v - number of uninfected cells, infected cells, antibodies, respectively free viruses within analysed tissue;
 a - additional death rate constant of infected cells;
 b - antibodies decay rate in absence of stimulation;
 β - infection effectiveness;
 c - antibodies responsiveness;
 d - uninfected cells death rate constant;
 k - free viruses generation rate constant;
 λ - replenishment rate of uninfected cells from external sources;
 K - maximum proliferation capacity;
 p - free viruses removal rate constant;
 r - proliferation rate constant;
 s - antibodies saturation level;
 u - virus death rate constant.

RESULTS

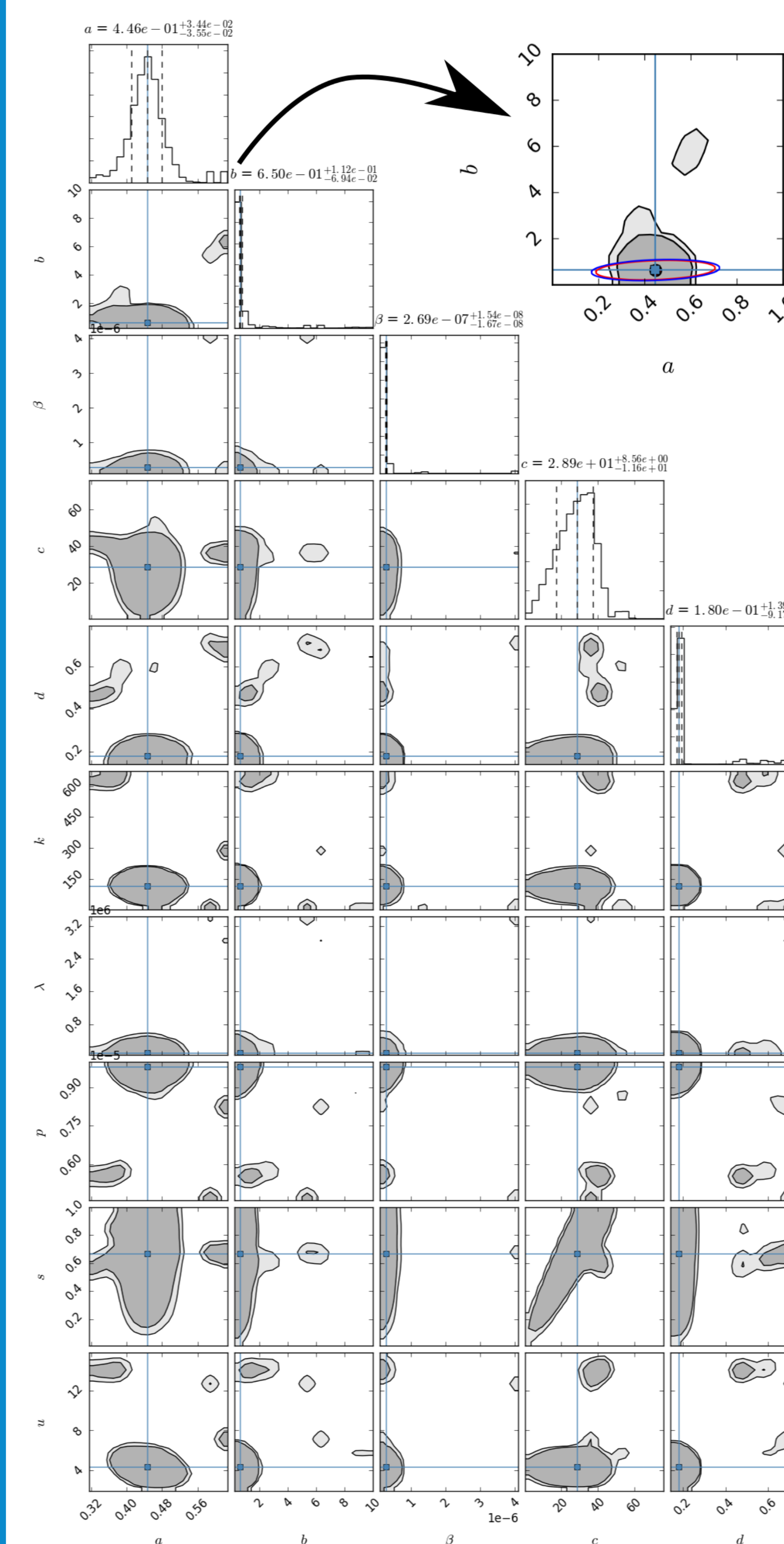


Figure 4: Corner plot and comparison with linearization around MLE.

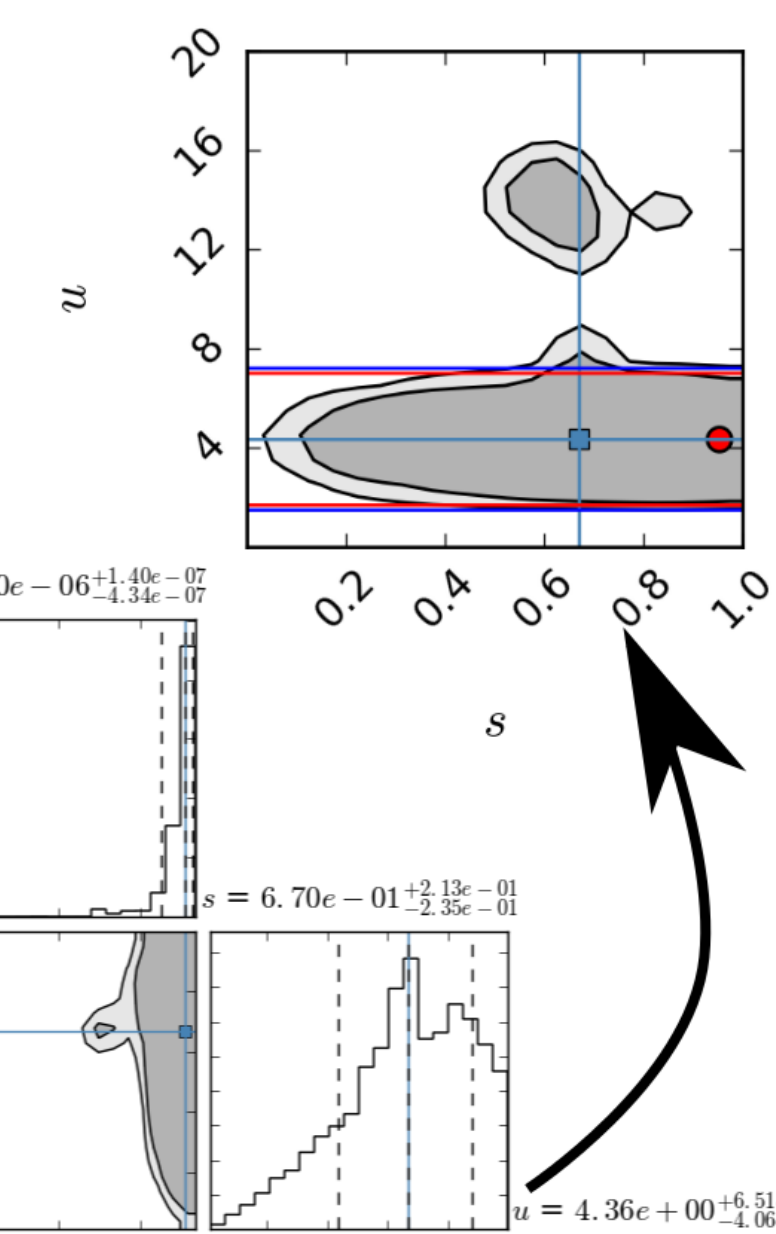
The 90% and 95% credibility-level regions are presented as 2D-projections (dark, respectively light grey surfaces), while marginal distribution of each parameter is shown as a 20 bins histogram together with its median (blue line) ± 0.16 quantile, respectively 0.84 quantile (dashed lines).

For comparison, two projections of the 90% (red contour), respectively 95% (blue contour) confidence-level ellipsoids are shown for parameter-pairs $a - b$ and $s - u$. The maximum likelihood estimate is marked with a red dot.

The MCMC sampling used an ensemble of 60 walkers with a stretching parameter of 2 such that the acceptance rate was on average equal to 0.31 and the total number of steps was 3,500.

The **total number of likelihood function evaluations was equal to 210,000** (wall clock time was around 2.0 hours on one core of a i7-5600U CPU).

The number of burnt points per each walker (starting positions were spread within the prior uniform box) was equal to 1,000 and decided based on trace-plots.



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