



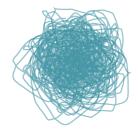
# BIOL 432

De novo assembly

# Whole Genome Shotgun (WGS) assembly







Extract



Fragment



Sequence

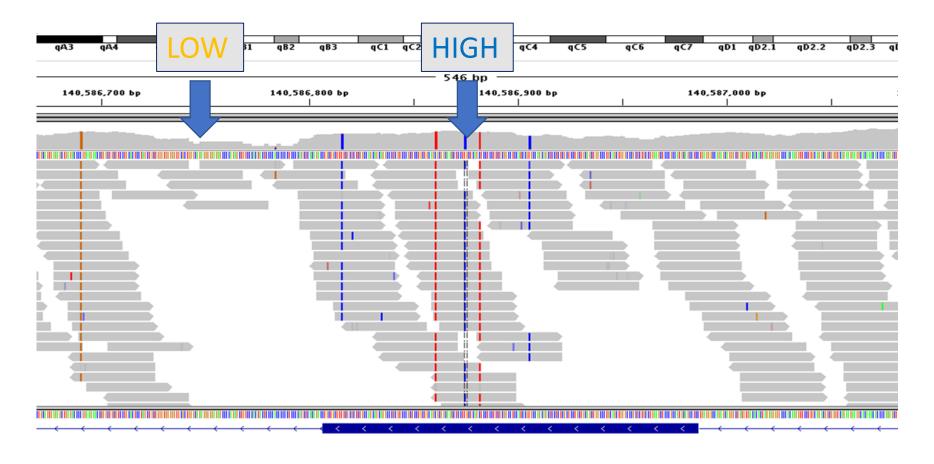


**Assemble** 

### Alignment coverage







- = average # reads at any given location (base pair)
- A 'random' sampling process
- (Higher is better)

### WGS without a reference genome





#### Alignment to reference

- sometimes called 'resequencing'
- common in 'model systems'
- e.g. human, mouse, *Drosophila*, *C. elegans*, *Arabidopsis*

#### de novo assembly

• 'non-model' organisms

Reference genome

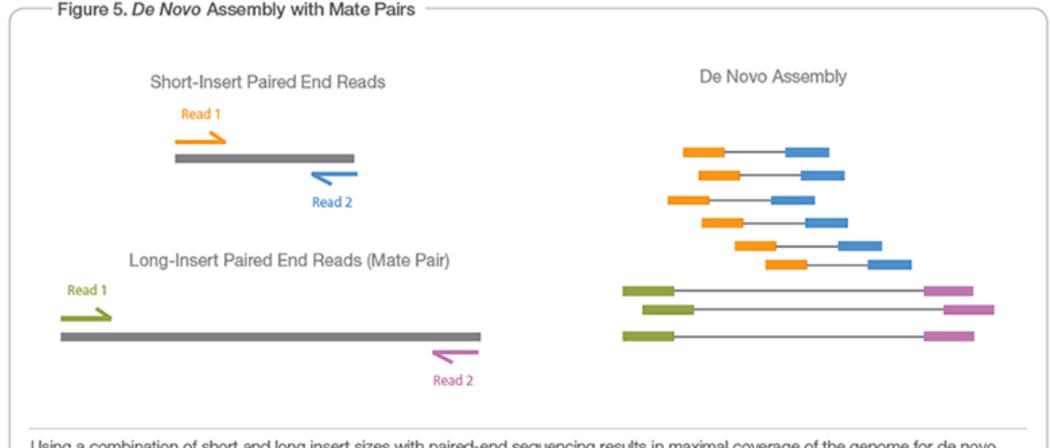
Consensus sequence

Contiguous consensus sequences (contigs)

### 'Paired End' vs 'Mate Pair'

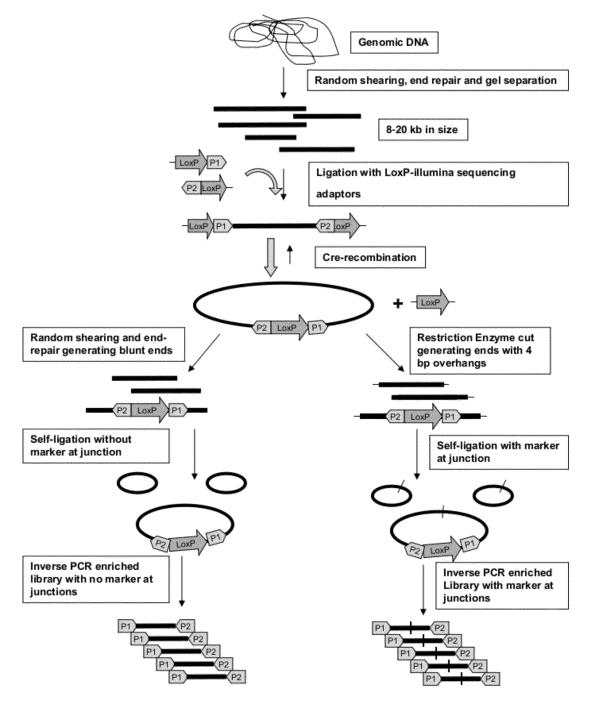






Using a combination of short and long insert sizes with paired-end sequencing results in maximal coverage of the genome for de novo assembly. Because larger inserts can pair reads across greater distances, they provide a better ability to read through highly repetitive sequences and regions where large structural rearrangements have occurred. Shorter inserts sequenced at higher depths can fill in gaps missed by larger inserts sequenced at lower depths. Thus a diverse library of short and long inserts results in better de novo assembly, leading to fewer gaps, larger contigs, and greater accuracy of the final consensus sequence.

# PE protocol



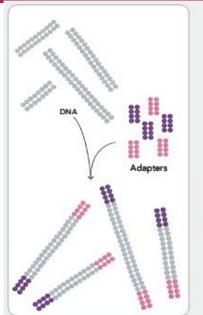


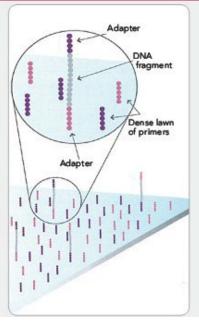


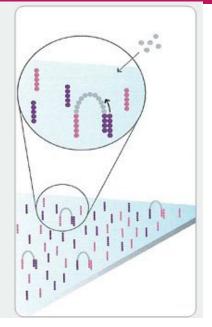
# Illumina sequencing (formerly Solexa)

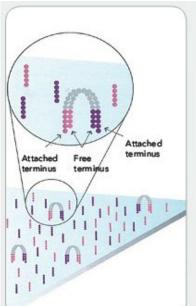


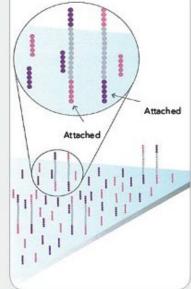


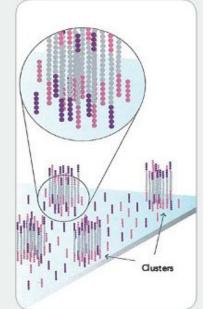










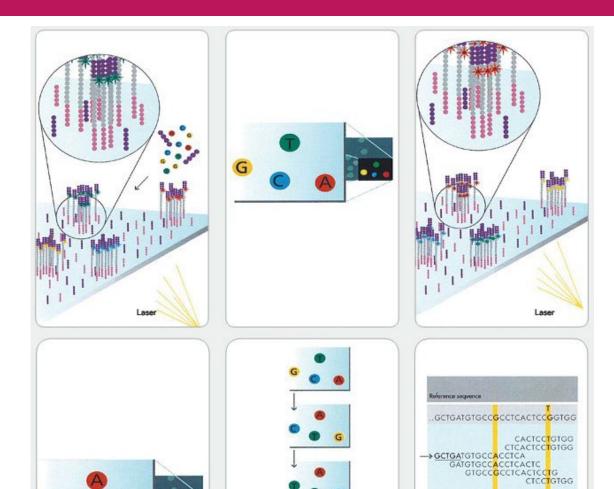


- 1. Prepare genomic DNA
- 2. Attach DNA to surface
- 3. Bridge amplification
- 4. Fragment become double stranded
- 5. Denature the double stranded molecules
- 6. Complete amplification

# Illumina sequencing







- 7. Determine first base
- 8. Image first base
- 9. Determine second base
- 10. Image second base
- 11. Sequence reads over multiple cycles
- 12. Align data

### A few de novo assemblers





- ABySS
- ALLPATHS-LG
- CORTEX
- CLC Genomics Workbench
- DISCOVAR de novo
- Geneious
- IDBA
- MaSuRCA
- MIRA
- PLATANUS
- RAY
- SOAP de novo

### Group Activity:





Assemble a genome by hand

What are the 'read lengths', 'coverage' and 'genome size' (aka assembly size) of your genome assembly?

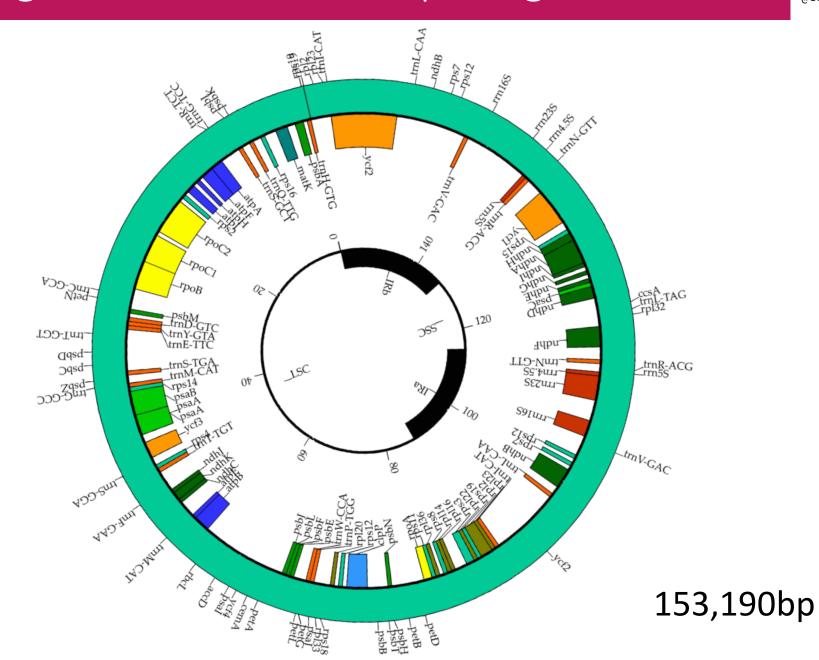
Why/how would these three factors affect the quality of the genome assembly?

What else might affect assembly quality?

## Assemble the garlic mustard chloroplast genome







### Post-its





Think about the previous tutorial from Hartmut from CAC, compared to Colautti tutorials

Pink (improve)

Thing(s) about **CAC tutorial** that **Colautti** tutorial should consider.

Thing(s) **shared** by CAC & Colautti tutorials that should be **changed** 

Green (keep)

Things(s) about **Colautti** tutorial does that **CAC** should consider.

Thing(s) **shared** by CAC & Colautti tutorials that should **not** be changed