Kasdin et al. (2025) show that dopamine in the brains of young zebra finches acts as a learning signal, increasing when they sing closer to their adult song and decreasing when they sing further away, effectively guiding their vocal development through trial-and-error. This suggests that complex natural behaviors, like learning to sing, are shaped by dopamine-driven reinforcement learning, similar to how artificial intelligence learns. You can find the paper at this link: https://www.nature.com/articles/s41586-025-08729-1..

Note they measure dopamine using fibre photometry, changes in the fluorescence indicate dopamine changes in realtime. Their specific measurement considers changes in flourescence in 100-ms windows between 200 and 300 ms from the start of singing, averaged across development.

1. Using the pwr package for R (Champely, 2020), conduct a power analysis. How many observations would the researchers need to detect a moderate-to-large effect (d=0.65) when using  $\alpha=0.05$  and default power (0.80) for a two-sided one sample t test.

```
pwr.t.test(d=0.65,
           sig.level=0.05,
           type="one.sample",
           alternative = "two.sided",
           power=0.8)
##
##
        One-sample t test power calculation
##
                 n = 20.58039
##
                 d = 0.65
##
        sig.level = 0.05
##
            power = 0.8
##
       alternative = two.sided
##
#n=20.58 - need at least 21 observations
```

They would need to conduct at least 21 observations.

2. Click the link to go to the paper. Find the source data for Figure 2. Download the Excel file. Describe what you needed to do to collect the data for Figure 2(g). Note that you only need the closer\_vals and further\_vals. Ensure to mutate() the data to get a difference (e.g., closer\_vals - further\_vals).

```
#Pulling data from figure 2
fig2_further = read_csv("further_data.csv", col_names = "Further")
## Rows: 25 Columns: 1
## -- Column specification
## Delimiter: "."
## dbl (1): Further
\#\# i Use 'spec()' to retrieve the full column specification for this data.
## i Specify the column types or set 'show_col_types = FALSE' to quiet this message.
fig2_closer = read_csv("closer_data.csv", col_names = "Closer")
## Rous: 25 Columns: 1
## -- Column specification
## Delimiter:
## dbl (1): Closer
\textit{## i Use `spec()' to retrieve the full column specification for this data}.
\#\# i Specify the column types or set 'show_col_types = FALSE' to quiet this message.
#Combining 2 values into a single tibble
fig2_tibble = bind_cols(fig2_further, fig2_closer)
#Mutating a new column to show the difference between columns
fig2 tibble = fig2 tibble |>
 mutate(Difference = Closer-Further)
```

To collect the data, I had to first download the source data from the research paper. I could then save the desired variables as .csv files (further and closer), before combining them all into a single

tibble. Generating the difference between further and closer just required using mutate(). With these variables, it would be possible to generate Figure 2(g) - the closer values would be on the left, further values on the right, and the difference would represent the lines between them.

- 3. Summarize the data.
  - (a) Summarize the further data. Do the data suggest that dopamine in the brains of young zebra finches decreases when they sing further away?

```
PART 3
dopamine_summ = fig2_tibble |>
  summarize(
   further = mean(Further),
closer = mean(Closer),
   difference = mean(Difference)
#view(dopamine_summ)
#Summarizing further
further_summary = ggplot() +
  geom_histogram(aes(fig2_tibble$Further, y=after_stat(density))) +
  theme_bw() +
  geom_hline(yintercept=0) +
  xlab("Dopamine Effect") +
 ylab("Density")
#Summarizina closer
closer_summary = ggplot() +
  geom_histogram(aes(fig2_tibble$Closer, y=after_stat(density))) +
  theme bw() +
  geom_hline(yintercept=0) +
  xlab("Dopamine Effect") +
  ylab("Density")
#Summarizing difference
difference_summary = ggplot() +
  geom_histogram(aes(fig2_tibble$Difference, y=after_stat(density))) +
  theme_bw() +
  geom_hline(yintercept=0) +
  xlab("Dopamine Effect") +
  ylab("Density")
summaries_plot = (further_summary | closer_summary | difference_summary)
#Saving the plot to be used in Sweave
ggsave("summaries_plot.pdf", plot = summaries_plot, width = 6, height = 4)
## 'stat_bin()' using 'bins = 30'. Pick better value with 'binwidth'.
## 'stat_bin()' using 'bins = 30'. Pick better value with 'binwidth'.
## 'stat_bin()' using 'bins = 30'. Pick better value with 'binwidth'.
```

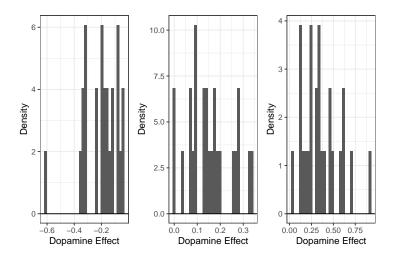


Figure 1: Histogram of Recorded Dopamine

Yes, the data suggests dopamine decreases when birds sing further away. Figure 1 (Pedersen, 2024; Wickham et al., 2019) demonstrates that data for birds that sing further is below the baseline, with a recorded mean of -0.203.

- (b) Summarize the closer data. Do the data suggest that dopamine in the brains of young zebra finches increases when they sing closer to their adult song?
  - Yes, the data suggests dopamine increases when birds sing further away. Figure 1 demonstrates that data for birds that sing further is above the baseline, with a recorded mean of 0.156.
- (c) Summarize the paired differences. Do the data suggest that there is a difference between dopamine in the brains of young zebra finches when they sing further away compared to closer to their adult song?
  - Yes, the data suggests there is a difference between dopamine when the young zebra finches sing further away compared to closer. Fiugre 1 highlights that the birds tends to have a higher recorded dopamine when singing closer to the song, with a recorded mean of a 0.359 difference in dopamine.
- (d) **Optional Challenge:** Can you reproduce Figure 2(g)? Note that the you can use geom\_errorbar() to plot the range created by adding the mean  $\pm$  one standard deviation.
- 4. Conduct the inferences they do in the paper. Make sure to report the results a little more comprehensively that is your parenthetical should look something like: (t = 23.99, p < 0.0001; g = 1.34; 95% CI: 4.43, 4.60).

**Note:** Your numbers may vary slightly as they performed some unclear correction of their *p*-values. I'm waiting to hear back from them via email!

(a) "The close responses differed significantly from 0 ( $p=1.63\times10^{-8}$ )." I calculated similar values as the researchers reported:  $t=8.3024; df=24; p=8.132\times10^{-9}; 95\%CI:0.1173875, 0.1950586$ ).

```
mu0 <- 0
#Calculating for closer (right-sided)
#Manually calculating statistics
x <- fig2_tibble$Closer
xbar <- mean(x)
s <- sd(x)
n <- length(x)
t.stat <- (xbar - mu0)/(s/sqrt(n))
p.val <- pt(q=-abs(t.stat), df = n-1)
#Calculating hedges value
closer_hedges_vals = hedges_g(x = x, mu = mu0, alternative = "greater")</pre>
```

```
#having t.test calculate values automatically and ensuring they match
(closer_t_test = t.test(x=x, mu = mu0, alternative = "greater"))
## One Sample t-test
##
## data: x
## t = 8.3024, df = 24, p-value = 8.132e-09
## alternative hypothesis: true mean is greater than 0
## 95 percent confidence interval:
## 0.1240301
                   Inf
## sample estimates:
## mean of x
## 0.1562231
({\tt closer\_CI = t.test(x=x)\$conf.int}) \ \textit{\#Caluclating the CI using a two sided test}
## [1] 0.1173875 0.1950586
## attr(,"conf.level")
## [1] 0.95
```

(b) "The far responses differed significantly from 0  $(p = 5.17 \times 10^{-8})$ ." I calculated similar values as the researchers reported: t = -7.778; df = 24;  $p = 2.587 \times 10^{-8}$ ; 95%CI : -0.2565176, -0.1489313

```
#Calculating for further (left sided)
#Manually calculating statistics
x <- fig2_tibble$Further
xbar <- mean(x)
s \leftarrow sd(x)
n <- length(x)
t.stat <- (xbar - mu0)/(s/sqrt(n))
p.val \leftarrow pt(q=-abs(t.stat), df = n-1)
#Calculating hedges value
further_hedges_vals = hedges_g(x = x, mu = mu0, alternative = "less")
#having t.test calculate values automatically and ensuring they match
(further_t_test = t.test(x=x, mu = mu0, alternative = "less"))
##
## One Sample t-test
##
## data: x
## t = -7.778, df = 24, p-value = 2.587e-08
\mbox{\tt\#\#} alternative hypothesis: true mean is less than 0
## 95 percent confidence interval:
##
          -Inf -0.1581322
## sample estimates:
## mean of x
## -0.2027244
(further\_CI = t.test(x=x)\$conf.int) \ \#Caluclating \ the \ CI \ using \ a \ two \ sided \ test
## [1] -0.2565176 -0.1489313
## attr(,"conf.level")
## [1] 0.95
```

(c) "The difference between populations was significant  $(p=1.04\times 10^{-8})$ ." I calculated similar values as the researchers reported:  $t=8.5109; df=24; p=1.037\times 10^{-8}; 95\% CI: 0.2719028, 0.4459921$ 

```
#Manually calculating statistics
x <- fig2_tibble$Difference
xbar <- mean(x)
s <- sd(x)
n <- length(x)
t.stat <- (xbar - mu0)/(s/sqrt(n))
p.val <- 2*pt(q=-abs(t.stat), df = n-1)

#Calculating hedges value
difference_hedges_vals = hedges_g(x = x, mu = mu0, alternative = "two.sided")

#having t.test calculate values automatically and ensuring they match
(difference_t_test = t.test(x=x, mu = mu0, alternative = "two.sided"))</pre>
```

```
##
## One Sample t-test
##
## data: x
## t = 8.5109, df = 24, p-value = 1.037e-08
## alternative hypothesis: true mean is not equal to 0
## 95 percent confidence interval:
## 0.2719028 0.4459921
## sample estimates:
## mean of x
## 0.3589475
```

- 5. Reverse engineer the hypothesis test plot from Lecture 20 to create accurate hypothesis testing plots for each part of the previous question.
  - (a) Question 4, part(a).

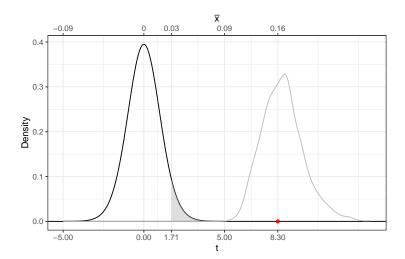


Figure 2: Hypothesis Plot for Closer

```
ggdat.t <- tibble(t=seq(-5,5,length.out=1000))|>
mutate(pdf.null = dt(t, df=n-1))
#Making plot for closer
#Pulling t value
obs_t=closer_t_test$statistic[[1]]
{\it \# Resampling \ to \ approximate \ the \ sampling \ distribution}
# on the data
R <- 1000
resamples <- tibble(t=numeric(R))</pre>
for(i in 1:R){
curr.sample <- sample(x=fig2_tibble$Closer,</pre>
                       size=n,
                       replace=T)
resamples\$t[i] = (mean(curr.sample) - mu0) / (sd(curr.sample) / sqrt(n))
s <- sd(fig2_tibble$Closer)
t.breaks \leftarrow c(-5, 0,
            qt(0.95, df = n-1), 5, # rejection region (right)
obs_t) # t-statistic observed
xbar.breaks <- t.breaks * s/(sqrt(n)) + mu0
```

```
# Create Plot
closer_plot = ggplot() +
# null distribution
geom_line(data=ggdat.t,
         aes(x=t, y=pdf.null))+
geom_hline(yintercept=0)+
# rejection regions
geom_ribbon(data=subset(ggdat.t, t>=qt(0.95, df=n-1)),
            aes(x=t, ymin=0, ymax=pdf.null),
fill="grey", alpha=0.5)+
# plot p-value (not visible)
geom_ribbon(data=subset(ggdat.t, t>=t.stat),
            aes(x=t, ymin=0, ymax=pdf.null),
            fill="reg", alpha=0.25)+
# plot observation point
geom_point(data=ggdat.obs, aes(x=t, y=y), color="red")+
# Resampling Distribution
stat_density(data=resamples,
             aes(x=t),
             geom="line", color="grey")+
# clean up aesthetics
theme_bw()+
ylab("Density")+
scale_x_continuous("t",
                   breaks = round(t.breaks,2),
                    sec.axis = sec_axis(~.,
                                         name = bquote(bar(x)),
                                        breaks = t.breaks,
labels = round(xbar.breaks,2)))
```

## (b) Question 4, part(b).

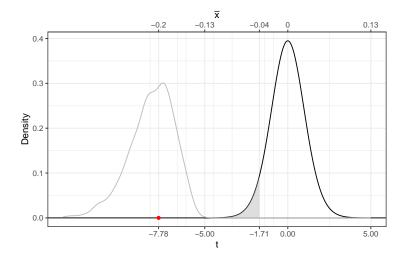


Figure 3: Hypothesis Plot for Further

```
s <- sd(fig2_tibble$Further)
t.breaks \leftarrow c(-5, qt(0.05, df = n-1), # rejection region (left)
            0, 5,
            obs_t)
                                      # t-statistic observed
xbar.breaks <- t.breaks * s/(sqrt(n)) + mu0
# Create Plot
further_plot = ggplot() +
# null distribution
geom_line(data=ggdat.t,
         aes(x=t, y=pdf.null))+
geom_hline(yintercept=0)+
# rejection regions
geom_ribbon(data=subset(ggdat.t, t<=qt(0.05, df=n-1)),</pre>
aes(x=t, ymin=0, ymax=pdf.null),
    fill="grey", alpha=0.5)+
# plot p-value (not visible)
geom_ribbon(data=subset(ggdat.t, t>=t.stat),
            aes(x=t, ymin=0, ymax=pdf.null),
            fill="reg", alpha=0.25)+
# plot observation point
geom_point(data=ggdat.obs, aes(x=t, y=y), color="red")+
# Resampling Distribution
stat_density(data=resamples,
             aes(x=t),
             geom="line", color="grey")+
{\it \# clean up aesthetics}
theme_bw()+
ylab("Density")+
scale_x_continuous("t",
                    breaks = round(t.breaks,2),
                    sec.axis = sec_axis(~.,
                                         name = bquote(bar(x)),
                                         breaks = t.breaks,
                                         labels = round(xbar.breaks,2)))
```

## (c) Question 4, part(c).

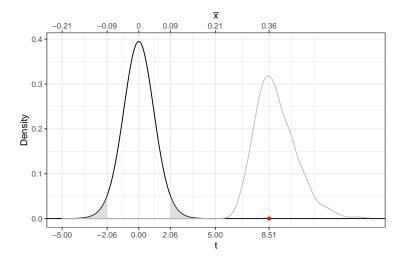


Figure 4: Hypothesis Plot for Difference

```
R <- 1000
resamples <- tibble(t=numeric(R))</pre>
for(i in 1:R){
curr.sample <- sample(x=fig2_tibble$Difference;</pre>
                      size=n,
                       replace=T)
resamples$t[i] = (mean(curr.sample)-mu0)/(sd(curr.sample)/sqrt(n))
s <- sd(fig2_tibble$Difference)
t.breaks \leftarrow c(-5, qt(0.025, df = n-1), # rejection region (left)
            qt(0.975, df = n-1), 5,  # rejection region (right)
                                      # t-statistic observed
xbar.breaks <- t.breaks * s/(sqrt(n)) + mu0
# Create Plot
difference_plot = ggplot() +
# null distribution
geom_line(data=ggdat.t,
         aes(x=t, y=pdf.null))+
geom_hline(yintercept=0)+
# rejection regions
geom_ribbon(data=subset(ggdat.t, t>=qt(0.975, df=n-1)),
            aes(x=t, ymin=0, ymax=pdf.null),
            fill="grey", alpha=0.5)+
geom_ribbon(data=subset(ggdat.t, t<=qt(0.025, df=n-1)),</pre>
            aes(x=t, ymin=0, ymax=pdf.null),
fill="grey", alpha=0.5)+
# plot p-value (not visible)
geom_ribbon(data=subset(ggdat.t, t>=t.stat),
            aes(x=t, ymin=0, ymax=pdf.null),
            fill="reg", alpha=0.25)+
# plot observation point
geom_point(data=ggdat.obs, aes(x=t, y=y), color="red")+
# Resampling Distribution
stat_density(data=resamples,
             aes(x=t).
             geom="line", color="grey")+
# clean up aesthetics
theme_bw()+
vlab("Densitv")+
scale_x_continuous("t",
                   breaks = round(t.breaks,2),
                    sec.axis = sec_axis(~.,
                                         name = bquote(bar(x)),
                                         breaks = t.breaks,
labels = round(xbar.breaks,2)))
```

## References

Champely, S. (2020). pwr: Basic Functions for Power Analysis. R package version 1.3-0.

Kasdin, J., Duffy, A., Nadler, N., Raha, A., Fairhall, A. L., Stachenfeld, K. L., and Gadagkar, V. (2025). Natural behaviour is learned through dopamine-mediated reinforcement. *Nature*, pages 1–8.

Pedersen, T. L. (2024). patchwork: The Composer of Plots. R package version 1.3.0.

Wickham, H., Averick, M., Bryan, J., Chang, W., McGowan, L. D., François, R., Grolemund, G., Hayes, A., Henry, L., Hester, J., Kuhn, M., Pedersen, T. L., Miller, E., Bache, S. M., Müller, K., Ooms, J., Robinson, D., Seidel, D. P., Spinu, V., Takahashi, K., Vaughan, D., Wilke, C., Woo, K., and Yutani, H. (2019). Welcome to the tidyverse. *Journal of Open Source Software*, 4(43):1686.