

A Bioinformatics Pipeline Reveals a Shared $\text{I}\kappa\text{B}\alpha$ Interface for NF- κB and Histone Binding



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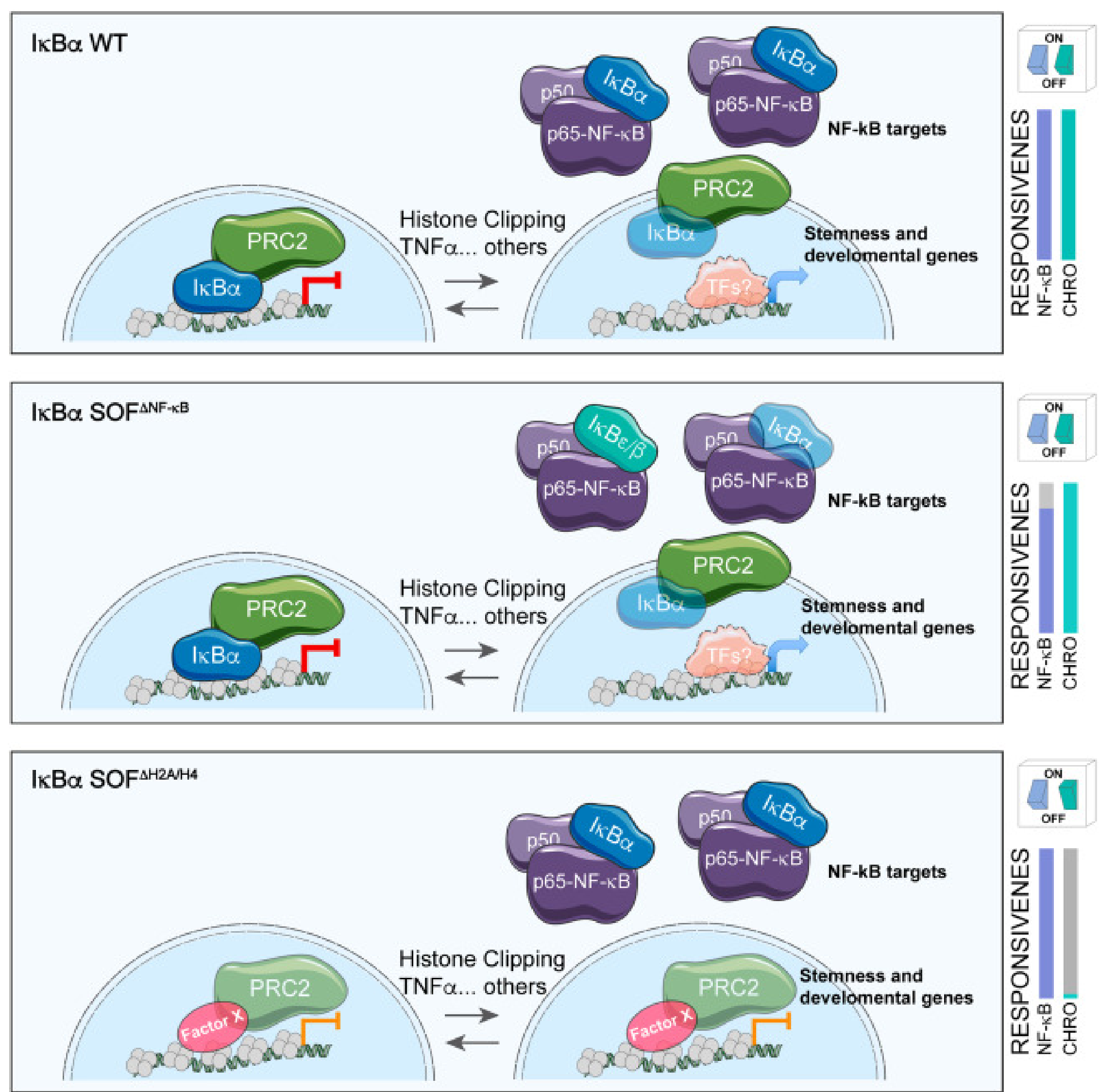
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SUMMARY

$\text{I}\kappa\text{B}\alpha$, traditionally a negative regulator of NF- κB , has recently been linked to chromatin-dependent transcriptional control (Marruecos et al., 2021). We developed a bioinformatics pipeline to identify conserved, surface-exposed residues mediating protein interactions. Our Fold-Excluded Evolutionary Conservation (FEEC) metric integrates structural packing (WCN) and evolutionary conservation (ConSurf) to prioritize $\text{I}\kappa\text{B}\alpha$ binding residues. FEEC highlighted an ANK-repeat surface engaging both the p65 NLS and histone H4 tail. Guided by FEEC, we engineered separation-of-function (SOF) mutants disrupting either NF- κB binding ($\text{SOF}^{\Delta\text{NF-}\kappa\text{B}}$) or histone association ($\text{SOF}^{\Delta\text{H2A/H4}}$). Structural modeling, mutagenesis, and functional assays confirmed their specificity. Transcriptomic analysis of $\text{SOF}^{\Delta\text{NF-}\kappa\text{B}}$ cells revealed repression of intestinal stem-cell genes independent of NF- κB , underscoring a chromatin-related role for $\text{I}\kappa\text{B}\alpha$. FEEC thus provides a generalizable framework to resolve multifunctional protein interfaces. (Álvarez-Villanueva et al., 2025)



Regulation of NF- κB and PRC2 target genes by $\text{I}\kappa\text{B}\alpha$ WT and SOF mutants

METHODOLOGY

The **Weighted Contact Number (WCN)** quantifies atomic contact density from interatomic distances as

$$\text{WCN}_i = \sum_{j \neq i} \frac{1}{r_{ij}^2}, \quad \text{iWCN}_i = \frac{1}{\text{WCN}_i}.$$

Sequence conservation scores ($S_{C,i}$) were obtained from the CONSURF server and expressed as Z-scores. The **Fold-Excluded Evolutionary Conservation (FEEC)** score identifies residues with conservation beyond structural constraints:

$$\text{FEEC}_i = \text{iWCN}_i - S_{C,i}.$$

Interface energies were computed from Rosetta-generated conformations weighted by Boltzmann probabilities and expected binding energies:

$$P_i = \frac{e^{-E_i/kT}}{\sum_{i=1}^N e^{-E_i/kT}},$$
$$\langle E_{\text{binding}} \rangle = \sum_i P_i E_{b,i}, \quad E_{b,i} = E_{\text{complex},i} - (E_{A,i} + E_{B,i}).$$

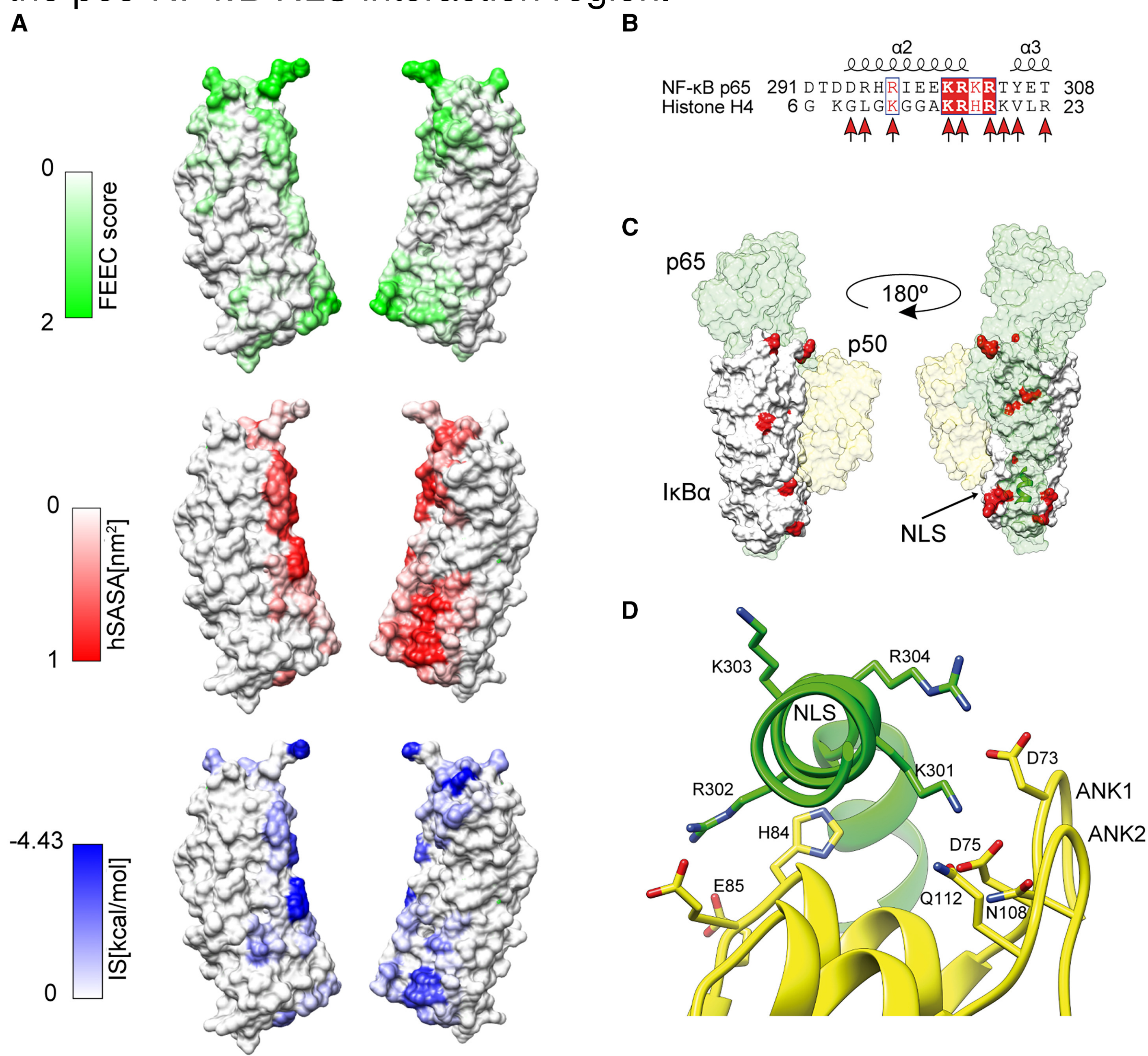
Mutational effects were estimated after repacking and minimizing residues within 8 Å of each mutation site.

REFERENCES

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- Marruecos, L. et al. (2021). "Dynamic chromatin association of $\text{I}\kappa\text{B}\alpha$ is regulated by acetylation and cleavage of histone H4". In: *EMBO Reports* 22.8, e52649. doi: 10.15252/embr.202152649.

COMMON DOMAIN OF $\text{I}\kappa\text{B}\alpha$ BINDING TO P65-NF- κB AND H2A/H4

FEEC scores accurately predicted most $\text{I}\kappa\text{B}\alpha$ /NF- κB interface residues (83% for p65, 73% for p50), validating FEEC as a reliable predictor of binding sites. Mapping positive FEEC residues revealed strong overlap with known interface regions from structural data. Negatively charged FEEC-positive residues in the ANK1-ANK2 domains (e.g., D73, D75, E85, E86) likely form the binding site for the histone H4 N-terminal tail, overlapping with the p65-NF- κB NLS interaction region.



(A) Surface mapping of $\text{I}\kappa\text{B}\alpha$ residues with positive FEEC scores (capped at 2), solvent-accessible area buried upon NF- κB binding, and per-residue interface scores. (B) Alignment of the p65-NF- κB NLS region with the H4 N-terminal tail; conserved positions (blue), similar/identical residues (red/white), and $\text{I}\kappa\text{B}\alpha$ -contacting residues (red arrows) are indicated. (C) $\text{I}\kappa\text{B}\alpha$ (white)-NF- κB (p65, green; p50, yellow) complex showing negatively charged $\text{I}\kappa\text{B}\alpha$ residues with FEEC>0 (red). (D) Interaction of the p65 NLS motif (KRKR, green) with $\text{I}\kappa\text{B}\alpha$ ANK1-2 repeats; polar interacting residues in yellow. (PDB: 1NFI, 3UW9; UniProt Q04206/p65 and P25963/ $\text{I}\kappa\text{B}\alpha$).

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