# Metagenome Analysis of Premature Birth

#### Jaewoong Lee Semin Lee

Department of Biomedical Engineering Ulsan National Institute of Science and Technology

jwlee230@unist.ac.kr

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#### Overview

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- Methods
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Introduction

#### Microbiome

- Microbiota: the microorganisms which live inside & on humans (Turnbaugh et al., 2007)
- Microbiome:  $10^{13}$  to  $10^{14}$  microorganisms whose which collective genome (Gill et al., 2006)



Figure: Concept of a core human microbiome (Turnbaugh et al., 2007)

#### rRNA

- Ribosomal RNA
- Well-known as a key to phylogeny (Olsen & Woese, 1993)

# Premature Birth (Preterm Birth)

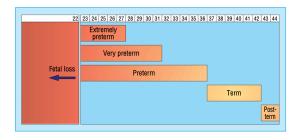


Figure: Definitions of Premature (J. Tucker & McGuire, 2004)

∴ Hence, in this study,

• Premature: < 37 weeks

• Normal:  $\geq$  37 weeks

### Materials

### 16S rRNA Sequencing

**16S rRNA sequencing** is the *reference method* for bacterial taxonomy & identification (Mignard & Flandrois, 2006)
Three main reasons (Janda & Abbott, 2007):

- 16S rRNA exists in almost all bacteria
- Functions of the 16S rRNA has not changed over time
- 16S rRNA is large enough for bioinformatics

# Train/Test Data vs. Validate Data

- JBNU/Helixco data
  - First data
  - Second data
  - Stool data

Table: Sample Information

Data	Participants	Samples	Remarks
First	24	107	-
Second	35	288	-
Stool	63	126	Stool

### Methods

#### Methods

Qiime 2 Workflow

### Qiime 2 Workflow



Figure: QIIME 2 workflow (Bolyen et al., 2019; Mandal, Van Treuren, White, Eggesbø, et al., 2015; McDonald et al., 2012)

# Filitering with Quality Score

#### Drawback between:

- Longer sequence read
- Higher quality value
- $\therefore$  Select the maximum length n where:

$$\forall n_i \in \{n_k | \text{MedianQualityScore} \ge 30\}$$
  
$$\exists! n \in \{n_i\} : n \ge n_i$$
 (1)

### **Denoising Techniques**

- DADA2: Amplicon Sequence Variants (ASVs) (Callahan et al., 2016)
- Deblur: Operational Taxonomic Units (OTUs) (Amir et al., 2017)



Figure: Denoising Algorithms

# Taxonomy Classification

- Greengenes (GG) (DeSantis et al., 2006)
- SILVA (Pruesse et al., 2007; Quast et al., 2012)



Figure: Taxonomy Classifications

"A **higher** performance at taxonomic levels above *genus level*; but performance appears to **drop** at *species level*" (Gihawi et al., 2019)

# Merging Denoising/Taxonomy

Merging multiple IDs (ASVs or OTUs) into one, which have

- Different IDs
- Identified as same taxonomy

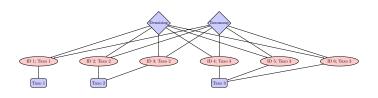


Figure: Example Diagram for Merging Denoising/Taxonomy

#### Methods

Abundance Test

#### **ANCOM**

- Analysis of composition of microbiome (Mandal, Van Treuren, White, Eggesbø, et al., 2015)
- ANCOM detects significantly abundant taxa, while maintain high statistical power
- Find taxa that can divide each classes

### Methods

**Diversity Indices** 



### **Diversity Indices**

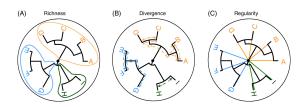


Figure: Three dimensions of phylogenic information (C. M. Tucker et al., 2017)

- A quantitative measure that shows richness, divergence, and regularity (C. M. Tucker et al., 2017)
- Alpha diversity indices: the richness of taxa at a single community
- Beta diversity indices: the taxonomic differentiation between communities

### Alpha Diversity Indices

- Evenness index
- Faith's Phylogenetic Diversity (Faith PD) index
- Oberseved Features index
- Shannon's Diversity index

### Beta Diversity Indices

- Bray-Curtis distance index
- Jaccard distnace index
- Unweighted UniFrac distance index
- Weighted UniFrac distance index

#### Methods

Miscellaneous

# t-distributed Stochastic Neighbor Embedding (t-SNE)



Figure: t-SNE with handwritten data (Maaten & Hinton, 2008)

# Python Packages

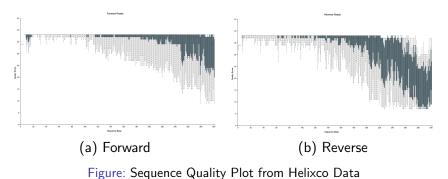
- Pandas (McKinney et al., 2011)
- Scikit-Learn (Pedregosa et al., 2011)
- SciPy (Virtanen et al., 2020)
- Matplotlib (Hunter, 2007)
- Seaborn (Waskom et al., 2020)
- Statannot

### Results

### Results

Filtering Results

# Quality Score from First Data



Tigure: Sequence Quanty Flot from Fleixes Bata

Maximum Length:  $n_{Forward} = 300$ ,  $n_{Reverse} = 265$ 

# Quality Score from Second Data

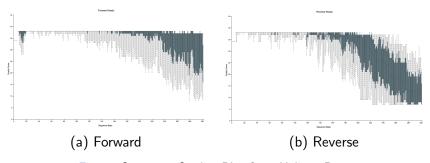


Figure: Sequence Quality Plot from Helixco Data

Maximum Length:  $n_{Forward} = 300$ ,  $n_{Reverse} = 222$ 

# Quality Score from Stool Data

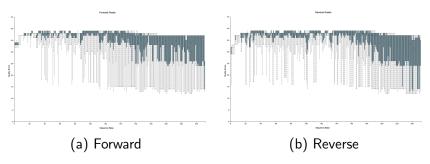


Figure: Sequence Quality Plot from Stool Data

Maximum Length:  $n_{Forward} = 250$ ,  $n_{Reverse} = 251$ 

#### Results

t-SNE with Site/Premature Information

# Workflow for t-SNE with Site/Premature Information

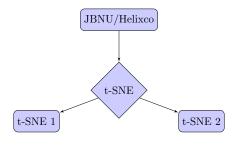


Figure: Workflow of t-SNE for Site/Premature Information

#### t-SNE with Site Information I

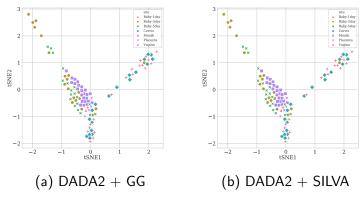


Figure: t-SNE with Site by DADA2

#### t-SNE with Site Information II

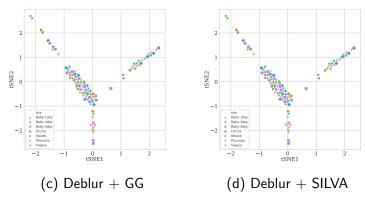


Figure: t-SNE with Site by Deblur

#### t-SNE with Premature Information I

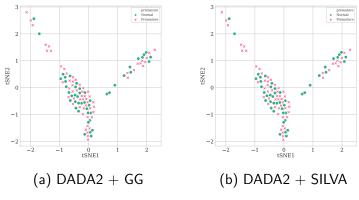


Figure: t-SNE with Premature by DADA2

#### t-SNE with Premature Information II

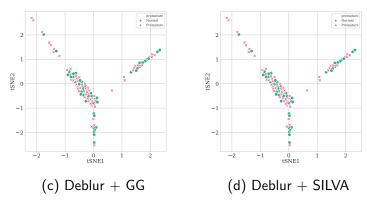


Figure: t-SNE with Premature by Deblur

# Results

Bacterial Abundance Test with ANCOM

## ANCOM?

- Analysis of composition of microbiomes
- ANCOM can be used for analyzing the composition of microbiomes in multiple populations (Mandal, Van Treuren, White, Eggesbø, et al., 2015)
- Differential abundance testing
- clr: Centered log(Ratio)
- W: a count of the number of sub-hypothesis which have passed for given species

#### ANCOM with ...

- Site where get the microbiome
- Premature Before 37 weeks and After 37 weeks
- Detailed Premature Before 34 weeks, After 37 weeks, and the other
- C-section
- PROM Premature rupture of membranes
- Using Steroid?
- Using anti-biotic?

#### ANCOM with Detailed Premature

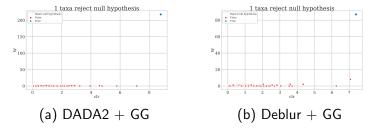


Figure: ANCOM results with Detailed Premature

- Ureaplasma genus
- Aerococcus genus

#### ANCOM with Site

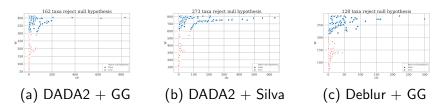


Figure: ANCOM results with Site

#### ANCOM with PROM

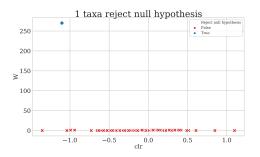
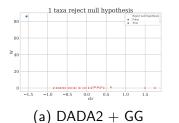
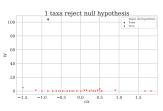


Figure: ANCOM result with PROM (DADA2 + GG)

• Campylobacteraceae genus Campylobacter species

# ANCOM with Using Steroid





(b) Deblur + GG

Figure: ANCOM results with Using Steroid

- Ureaplasma genus
- Aerococcus genus

# Results

Alpha-Diversity

# Alpha-Diversity

# Results

Beta-Diversity

# Beta-Diversity

# Results

#### Classification

#### Workflow for Classification

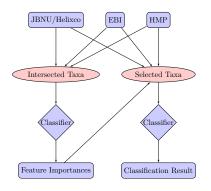


Figure: Workflow with Classification

#### Random Forest Classifier I

Input Data was treated with **Deblur** and **SILVA**.

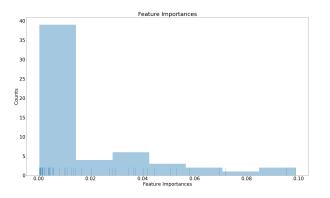


Figure: Feature Importance derived by Random Forest Classifier

#### Random Forest Classifier II



Figure: Number of Features vs. Accuracy

#### Random Forest Classifier III

- Bacteria Firmicutes Bacilli Lactobacillales Lactobacillaceae Lactobacillus Lactobacillus iners
- Bacteria Fusobacteriota Fusobacteria Fusobacteriales Leptotrichiaceae Leptotrichia
- Bacteria Actinobacteriota Actinobacteria
- Bacteria Firmicutes Bacilli Lactobacillales Lactobacillaceae Lactobacillus
- Bacteria Firmicutes Clostridia Peptostreptococcales-Tissierellales Peptostreptococcaceae Romboutsia
- Bacteria Firmicutes Bacilli Mycoplasmatales Mycoplasmataceae Ureaplasma
- Bacteria Actinobacteriota Actinobacteria Corynebacteriales
   Corynebacteriaceae Corynebacterium Corynebacterium matruchotii

# Random Forest Classifier IV

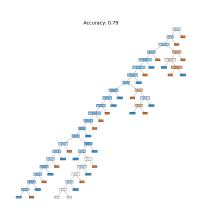


Figure: Random Forest Classifier

# Random Forest Classifier V

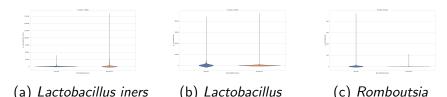


Figure: Violin Plot of Taxonomy

- Bacteria Firmicutes Bacilli Lactobacillales Lactobacillaceae Lactobacillus Lactobacillus iners
- Bacteria Firmicutes Bacilli Lactobacillales Lactobacillaceae Lactobacillus
- Bacteria Firmicutes Clostridia Peptostreptococcales-Tissierellales Peptostreptococcaceae Romboutsia

# Lactobacillus (Lb.)

- Vaginal Lb. may be clinically useful tools at PTB under 33 weeks.
   (Usui et al., 2002)
- Presence of Lb. sp (odds ratio 0.2) was negatively associated. (MARTIUS et al., 1988)
- Lb. crispatus/gasseri could decrease the risk of PTB. (Stafford et al., 2017)
- Lb. were associated with decreased risk of PTB. (Tabatabaei et al., 2019)

# Romboutsia

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