

Using reticulate to access CPTAC data in R

`reticulate` is an R package that allows you to create Python objects and convert them into R objects for use in your R environment.

Thus, even though `cptac` is a Python package, with the help of `reticulate` you can load and work with the datasets in R.

Step 1: Environment setup

First, you need to load the `reticulate` package, and then tell it which Python environment you want to use to access the `cptac` package.

```
# Install reticulate if necessary
if (!require(reticulate)) install.packages("reticulate")

# Load the package
library(reticulate)

# Specify to use the environment where the cptac package is installed. Replace
# "dev" with the name of your environment. If you use an environment manager
# besides conda, you'll want this command: use_virtualenv("myenv")
use_condaenv("dev", required = TRUE)
```

Step 2: Load cptac and access data

Now we will load `cptac` and get the dataset we want.

```
# Import the package. We pass convert = FALSE so that objects won't be
# converted from Python to R until we explicitly ask for it. This is necessary
# to properly prepare a multiindex dataframe for conversion to R (see below).
cptac <- import("cptac", convert = FALSE)

# Load the dataset
en <- cptac$Endometrial()
```

Load a dataframe and convert it to an R object

This dataframe just has a regular column index (not a multiindex), so we can directly convert it into an R object after loading it.

When `reticulate` converts a `pandas.DataFrame` into R, it converts it to the R `data.frame` type.

```
# Load the table
prot_py <- en$get_proteomics()

# Convert into R
prot <- py_to_r(prot_py)

print(prot[1:10, 1:8])
```

	A1BG	A2M	A2ML1	A4GALT	AAAS	AACS	AADAT	AAED1
## C3L-00006	-1.180	-0.8630	-0.802	0.2220	0.2560	0.6650	1.2800	-0.3390
## C3L-00008	-0.685	-1.0700	-0.684	0.9840	0.1350	0.3340	1.3000	0.1390
## C3L-00032	-0.528	-1.3200	0.435	NaN	-0.2400	1.0400	-0.0213	-0.0479
## C3L-00090	-1.670	-1.1900	-0.443	0.2430	-0.0993	0.7570	0.7400	-0.9290
## C3L-00098	-0.374	-0.0206	-0.537	0.3110	0.3750	0.0131	-1.1000	NaN
## C3L-00136	-1.080	-0.7080	-0.126	-0.4260	-0.1140	-0.1110	0.8950	1.2600
## C3L-00137	-1.320	-0.7080	-0.808	-0.0709	0.1380	0.6560	-0.2800	-0.1280
## C3L-00139	-0.467	0.3700	-0.339	NaN	0.4340	0.0358	-0.1750	0.1810
## C3L-00143	-1.120	-1.3100	0.912	0.4180	-0.0768	0.8460	-0.1210	NaN
## C3L-00145	-0.716	-0.8850	2.820	-0.3430	0.1470	0.4450	-0.0565	-0.8380

Load a multiindex dataframe

If it's a dataframe with a column multiindex, you need to first flatten the index before converting it to an R object.

See [tutorial 4](#) for more information on column multiindexes.

```
# Load the table
phos_py_multiindex <- en$get_phosphoproteomics()

# Load cptac.utils so we can access a helper function to convert the multiindex
# to a single level index.
utils <- import("cptac.utils", convert = FALSE)

# Convert the multiindex to a single level index. This works because we passed
# convert = FALSE when we imported the cptac package, so the dataframe is still
# a Python object at this point (it wasn't automatically converted into an R
# object on loading) and can be passed to this Python function.
phos_py_single_index <- utils$reduce_multiindex(phos_py_multiindex, flatten = TRUE)

# Convert to R
phos <- py_to_r(phos_py_single_index)

print(phos[1:10, 1:6])
```

	AAAS_S495	AAAS_S541	AAAS_Y485	AACS_S618	AAED1_S12	AAGAB_S310
## C3L-00006	NaN	NaN	NaN	-0.881	-1.8100	NaN
## C3L-00008	NaN	NaN	NaN	NaN	0.0840	NaN
## C3L-00032	-0.202	NaN	NaN	NaN	-1.8800	NaN
## C3L-00090	-0.002	NaN	-0.407	NaN	NaN	NaN
## C3L-00098	0.556	-0.0461	NaN	NaN	0.9410	NaN
## C3L-00136	NaN	NaN	NaN	NaN	0.0796	NaN
## C3L-00137	0.300	NaN	NaN	-0.371	-1.1400	NaN
## C3L-00139	0.490	NaN	NaN	NaN	NaN	NaN
## C3L-00143	-0.231	0.0108	NaN	NaN	0.2420	NaN
## C3L-00145	0.268	NaN	NaN	NaN	-0.1120	NaN