



# **Advanced RNA-seq training**

## **Time course experiment analysis**

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# RNA-seq

whole transcriptome assessment

## novel and known transcripts

allele specific expression

gene fusion

lncRNA

eRNA

alternatively spliced

variants

## transcripts quantification

differential expression

# Time course experiments

development and growth

reaction to a treatment/condition over time

## single time series

one condition

all time points compared to  
the first one (control)

## multi time series

several conditions simultaneously

controls are sampled over time  
with the samples

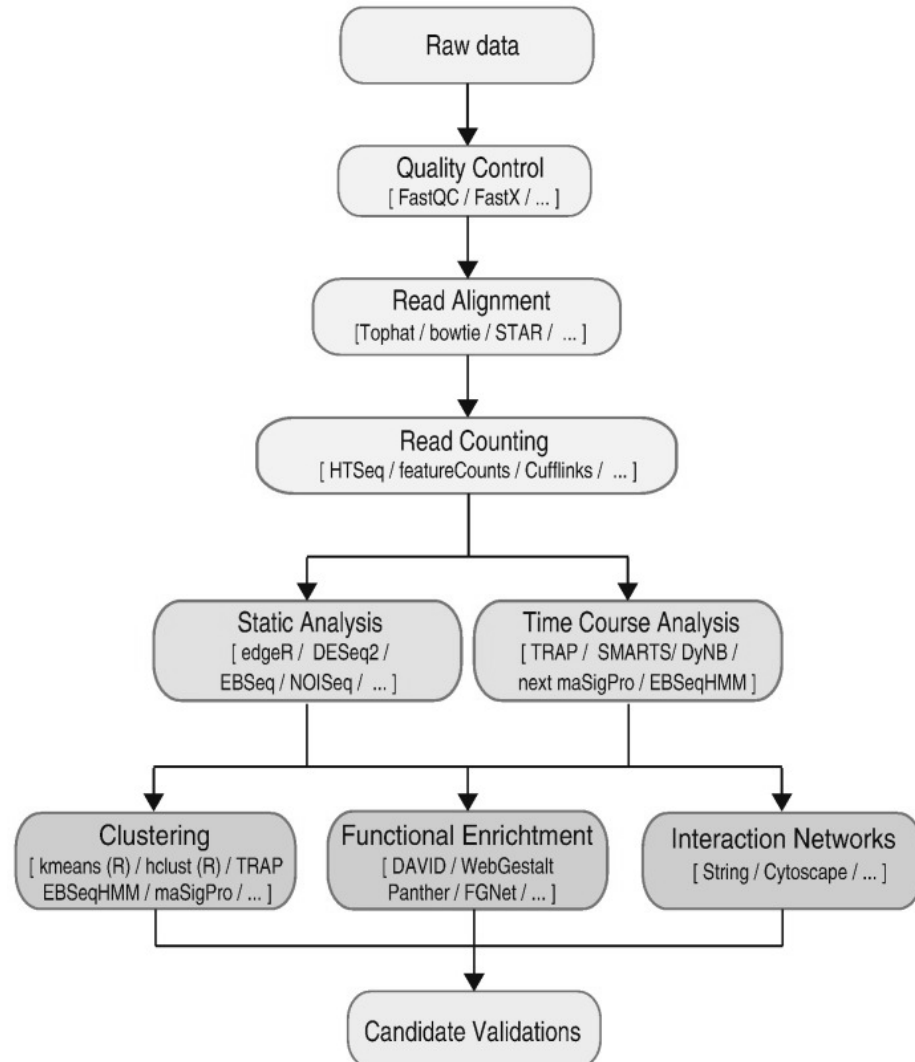
## periodic and cyclic time series

single or multiple conditions

reoccurring expression patterns and  
their difference between conditions

complex >> a lot of samples and synchronization needed

# Analysis workflow



similar to  
static RNA-seq experiments

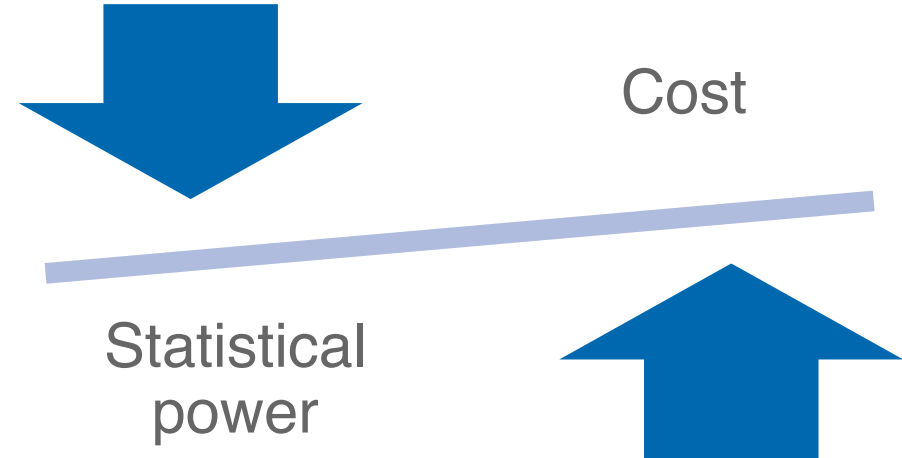
# Experimental design

## critical

number of time points

number of replicates

there are tools to estimate these parameters, but they don't consider multi-factor experiments



## when in doubt

more replicates better than greater sequencing depth

## bad design

👎 statistical power    👉 number of false positives

# Data analysis

## static tools

sequencing depth and  
library size  
batch effect >>  
protocol  
sequencing platform  
technical variability

## time course

do not consider correlation  
between neighbouring time  
points

# Choosing the right method

questions to address

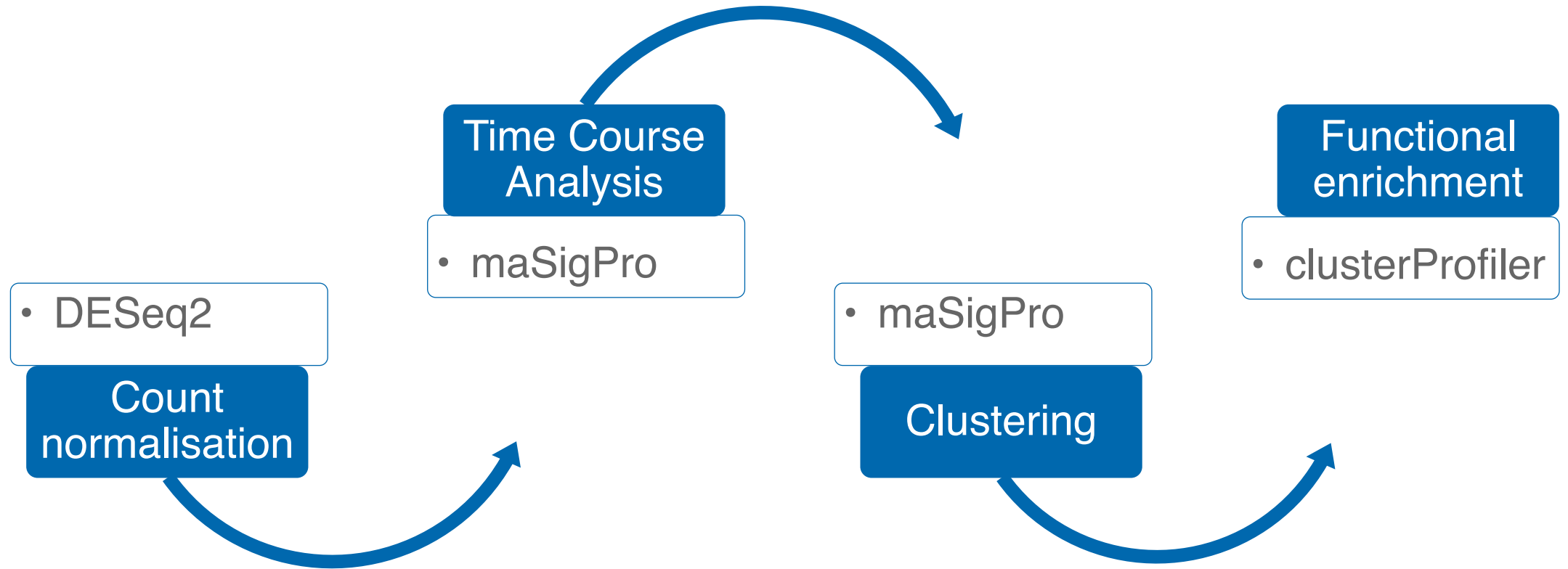
**number of  
replicates**

**experimental  
design**

two-way or multi factor

**differential  
expression of  
RNA isoforms**

# Training steps





# Workflow

## data normalisation with DESeq2

size factor correction drawn  
from the negative binomial  
distribution

## maSigPro

GLM with negative binomial distribution as a linking  
function

$$y_i = \beta_0 + \beta_1 t_i + \beta_2 t_i^2 + \beta_3 z_{1i} + \beta_4 t_i z_{1i} + \beta_5 t_i^2 z_{1i} + \varepsilon_i$$

## functional enrichment with clusterProfiler

hypergeometric distribution testing  
finds functional terms occurring significantly more than  
expected

# Data used in this training

Christelle Etard, Olivier Armant, Urmas Roostalu, Victor Gourain, Marco Ferg and Uwe Strähle

**Loss of function of myosin chaperones triggers Hsf1-mediated transcriptional response in skeletal muscle cells**

*Genome Biology* 2015 **16**:267

<https://doi.org/10.1186/s13059-015-0825-8>

RNA-seq\_Strahle\_Lab\_0005AS.<SequencingID>.USERvgourain.R.ReadsPerGene.out.tab



**Hpf**

**24**

**48**

**72**

**wt**

DCD001548SQ

DCD001546SQ

DCD001547SQ

DCD001559SQ

DCD001558SQ

DCD001545SQ

**unc45b**

DCD001560SQ

DCD001564SQ

DCD001565SQ

DCD001554SQ

DCD001555SQ

DCD001551SQ

All the libraries were:

unstranded

paired-ended

sequenced on Illumina HiSeq 2000  
producing 50 bp long reads.

# References

- [1] Daniel Spies and Constance Ciaudo.  
**Dynamics in Transcriptomics: Advancements in RNA-seq Time Course and Downstream Analysis.** *Comput Struct Biotechnol J.* 2015; 13: 469–477.
- [2] Michael I Love, Wolfgang Huber and Simon Anders.  
**Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2.** *Genome Biology.* 2014; 15: 550.
- [3] María José Nueda, Sonia Tarazona and Ana Conesa.  
**Next maSigPro: updating maSigPro bioconductor package for RNA-seq time series.** *Bioinformatics.* 2014; 30(18): 2598–2602.
- [4] Guangchuang Yu, Li-Gen Wang, Yanyan Han and Qing-Yu He.  
**clusterProfiler: an R Package for Comparing Biological Themes Among Gene Clusters.** *OMICS: A Journal of Integrative Biology.* 2012; 16(5): 284-287.