# wind: wORKFLOW FOR PiRNAs AnD BEYONd

Computational workflow for the preprocessing of selected samples from of TCGA regarding Breast Cancer

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## The Data set

We will work with a dataset from TCGA which is a subset of 18 samples from TCGA-BRCA. In detail, we used 9 Primary Solid Tumors versus 9 Solid Tissue Normal matched samples. ## Data acquisition and preprocessing

#### i. Downloading the samples

To obtain the data from TCGA we used a manifest with the selected samples and followed the instructions on GDC website

## ii. BAM to FASTQ

The acquired files were in BAM format but in order to do the whole workflow we transformed them to fastq format.

```
docker run --rm -ti -v $(pwd):/home/my_data congelos/sncrna_workflow

mkdir my_data/fastq_files
for file in my_data/bam_files/*.bam;
do regex="${file%.bam}"; samp=`basename ${regex}`;
echo "Processing sample ${samp}";
samtools bam2fq -0 6 $file > my_data/fastq_files/${samp}.fastq
echo "pigz sample ${samp}";
pigz --best "my_data/fastq_files/${samp}.fastq"
done
```

### iii. Preprocessing of the samples

We perform quality control(QC) on the fastq files to get basic information about the samples. We work with the **Fastqc** tool to perform QC.

```
mkdir -p my_data/Breast_Cancer_TCGA/qc_first

'fastqc' --threads 6 --outdir=my_data/Breast_Cancer_TCGA/qc_first/ my_data/myscratch/fastq_files/*.fastq.gz;

for file in my_data/myscratch/fastq_files/*.fastq.gz;

do ./spar_prepare/smrna_adapter_cut.sh $file 6;

done

mkdir my_data/Breast_Cancer_TCGA/qc_after

'fastqc' --threads 6 --outdir=my_data/Breast_Cancer_TCGA/qc_after/ my_data/myscratch/fastq_files/*.trimeexit
```

## Alignment and Quantification

#### i. Transcript abundances with Salmon

We will use a public docker image to run salmon

```
# run the docker
docker run --rm -it -v $(pwd):/home/my_data combinelab/salmon

# create the index
salmon index -t ncRNA_transcripts_100bp_RNA_Central_piRNAbank_hg38.fa -i genome_transc_human/ncRNA_Cent
mkdir my_data/smallRNA-breast-cancer/GSE129076/quants/

# run the samples

#!/bin/bash

for fn in my_data/smallRNA-breast-cancer/Breast_Cancer_TCGA/fastq_files/*trimmed.fastq.gz;
do samp=`basename ${fn}`;
echo "Processing sample ${samp}";
salmon quant -i my_data/genome_
```

```
transc_human/ncRNA_Central_piRNAB_hg38_index -1 A -r ${fn} --seqBias --gcBias --numBootstraps 100 -p 6
done

#save as bam files
for file in my_data/smallRNA-breast-cancer/Breast_Cancer_TCGA/quants/*.sam;
do
regex="${file%%.sam}";
echo "Processing sample ${regex}";
echo samtools view -0 bam -o ${regex}.bam -@ 6 ${file};
done
exit
```

#### ii. Alignment and quantification of sequenced reads with STAR and Featurecounts

We use the **STAR** aligner and then perform quantification with featureCounts from **Rsubread** package. With the docker image that contains STAR and **Samtools** we get sorted BAM files and use them for quantification / annotation for smallRNAs.

## iii. Alignment with STAR

```
# run docker
docker run --rm -ti -v "$PWD":/home/my_data congelos/sncrna_workflow

# index generation
STAR --runMode genomeGenerate --genomeDir my_data/human_data/GRCh38 --genomeFastaFiles my_data/human_da
mkdir my_data/smallRNA-breast-cancer/Breast_Cancer_TCGA/star_results

# alignment
for file in my_data/smallRNA-breast-cancer/Breast_Cancer_TCGA/fastq_files/*.trimmed.fastq.gz;
do samp=`basename ${file}`;
regex="${samp\%.trimmed.fastq.gz}";
echo "Processing sample ${samp} start: $(date)";
STAR --genomeDir my_data/genome_transc_human/human_data/GRCh38_2_7_4a --genomeLoad LoadAndKeep --readFidone
exit
```

Next, we run a docker image which includes varius R packages that will be used futhermore in the downstream analysis following featurecounts for the exploratory data analysis of piRNA data

#### iv. R docker

```
docker run --rm -v $(pwd):/home/0 -p 8787:8787 -e PASSWORD=12345 -e USER=$UID congelos/rocker_tidyverse
```

From here on we work in R using a browser. we input http://localhost:8787/ on browser, 0 for username and 12345 for password.

#### v. FeatureCounts

```
library(Rsubread)
library(tidyverse)
list.BAM <- list.files(path = "Breast_Cancer_TCGA/star_results",</pre>
                       pattern = ".bam$",
                       recursive = TRUE,
                        full.names = T)
path_gtf <- "../genome_transc_human/ncRNA_transcripts_100bp_RNA_Central_piRNAbank_hg38.gtf"
todate <- format(Sys.time(), "%d_%b_%Y")</pre>
fc <- featureCounts(files = list.BAM,</pre>
                    annot.ext = path_gtf,
                    isGTFAnnotationFile = TRUE,
                    GTF.featureType = "exon",
                    GTF.attrType.extra = c("gene_type", "sRNA_id", "seq_RNA"),
                    nthreads = 8,
                    useMetaFeatures = TRUE,
                    allowMultiOverlap = TRUE,
                    minOverlap = 10,
                    largestOverlap = TRUE,
                    fraction = TRUE,
                    strandSpecific = 0,
                    verbose = TRUE,
                    reportReads = "BAM",
                    reportReadsPath = "Breast_Cancer_TCGA/star_results/")
fc %>% write_rds(str_glue("Breast_Cancer_TCGA/feature_counts_BRCA_TCGA_{todate}.rds"))
```

Next we will follow the workflow of data exploration salmon fc.RMD

## R Session Info

```
sessionInfo()
R Under development (unstable) (2019-12-06 r77536)
Platform: x86_64-pc-linux-gnu (64-bit)
Running under: Debian GNU/Linux 10 (buster)
Matrix products: default
BLAS/LAPACK: /usr/lib/x86_64-linux-gnu/libopenblasp-r0.3.5.so
locale:
 [1] LC_CTYPE=en_US.UTF-8
                               LC_NUMERIC=C
 [3] LC_TIME=en_US.UTF-8
                               LC_COLLATE=en_US.UTF-8
 [5] LC_MONETARY=en_US.UTF-8
                               LC_MESSAGES=C
 [7] LC_PAPER=en_US.UTF-8
                               LC NAME=C
 [9] LC_ADDRESS=C
                               LC_TELEPHONE=C
[11] LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C
attached base packages:
[1] stats
          graphics grDevices utils
                                         datasets methods
                                                               base
```

```
other attached packages:
 [1] forcats_0.5.0
                                             dplyr_0.8.4
                                                                 purrr_0.3.3
                         stringr_1.4.0
 [5] readr 1.3.1
                        tidyr_1.0.2
                                             tibble_2.1.3
                                                                 ggplot2_3.2.1
 [9] tidyverse 1.3.0
                        Rsubread 2.1.2
                                             BiocManager 1.30.10
loaded via a namespace (and not attached):
 [1] Rcpp_1.0.3
                     cellranger_1.1.0 pillar_1.4.3
                                                        compiler_4.0.0
 [5] dbplyr 1.4.2
                     tools 4.0.0
                                       lubridate 1.7.4 jsonlite 1.6.1
[9] lifecycle_0.1.0 nlme_3.1-144
                                       gtable_0.3.0
                                                        lattice_0.20-40
[13] pkgconfig_2.0.3 rlang_0.4.5
                                       reprex_0.3.0
                                                        Matrix_1.2-18
[17] cli_2.0.2
                     DBI_1.1.0
                                       rstudioapi_0.11
                                                       xfun_0.12
[21] haven_2.2.0
                     knitr_1.28
                                       withr_2.1.2
                                                        xml2_1.2.2
[25] httr_1.4.1
                                       generics_0.0.2
                                                        vctrs_0.2.3
                     fs_1.3.1
[29] hms_0.5.3
                     grid_4.0.0
                                       tidyselect_1.0.0 glue_1.3.1
[33] R6_2.4.1
                     fansi_0.4.1
                                       readxl_1.3.1
                                                        modelr_0.1.6
[37] magrittr_1.5
                     backports_1.1.5 scales_1.1.0
                                                        rvest_0.3.5
[41] assertthat_0.2.1 colorspace_1.4-1 stringi_1.4.6
                                                        lazyeval_0.2.2
[45] munsell_0.5.0
                     broom_0.5.5
                                       crayon_1.3.4
```

## We work on:

```
[root@localhost GSE124507_brain_project]# cat /etc/*-release
CentOS Linux release 7.8.2003 (Core)
NAME="CentOS Linux"
VERSION="7 (Core)"
ID="centos"
ID_LIKE="rhel fedora"
VERSION_ID="7"
PRETTY_NAME="CentOS Linux 7 (Core)"
ANSI_COLOR="0;31"
CPE NAME="cpe:/o:centos:centos:7"
[root@localhost GSE124507_brain_project]# docker version
Client: Docker Engine - Community
Version:
                    19.03.8
 API version:
                    1.40
Go version:
                    go1.12.17
 Git commit:
                    afacb8b
                    Wed Mar 11 01:27:04 2020
 Built:
 OS/Arch:
                    linux/amd64
 Experimental:
                    false
Server: Docker Engine - Community
 Engine:
  Version:
                    19.03.8
  API version:
                    1.40 (minimum version 1.12)
                    go1.12.17
  Go version:
  Git commit:
                    afacb8b
                    Wed Mar 11 01:25:42 2020
  Built:
```

OS/Arch: linux/amd64

Experimental: false

containerd:

Version: 1.2.13
GitCommit: 7ad184331fa3e55e52b890ea95e65ba581ae3429

runc:

Version:

1.0.0-rc10 dc9208a3303feef5b3839f4323d9beb36df0a9dd GitCommit:

docker-init:

0.18.0 Version: GitCommit: fec3683

[root@localhost GSE124507\_brain\_project]# git version git version 1.8.3.1

[root@localhost GSE124507\_brain\_project]# pigz --version pigz 2.3.4