# RWorksheet\_nandin#4b.Rmd

### Mary coolen nandin

### 2024-10-30

### **Using Loop Function**

1. Using the for loop, create an R script that will display a 5x5 matrix as shown in Figure 1. It must contain vector A = [1,2,3,4,5] and a  $5 \times 5$  zero matrix. Hint Use abs() function to get the absolute value

```
matrixx <- matrix(0, nrow= 5, ncol = 5)

for (i in 1:5) {
   for (j in 1:5) {
   matrixx[i, j] <- abs(i-j)
   }
}
print(matrixx)</pre>
```

```
[,1] [,2] [,3] [,4] [,5]
## [1,]
            0
                  1
                       2
                             3
                             2
## [2,]
                                   3
## [3,]
            2
                       0
                             1
                                   2
                  1
## [4,]
            3
                  2
                       1
                             0
                                   1
## [5,]
                  3
                       2
                             1
                                   0
```

2. Print the string "\*" using for() function. The output should be the same as shown in Figure

```
rows <- 5

for (i in 1:rows) {
   cat(rep("*", i), "\n")
}</pre>
## *
```

3. Get an input from the user to print the Fibonacci sequence starting from the 1st input up to 500. Use repeat and break statements. Write the R Scripts and its output.

```
start <- 10
a <- 0
b <- 1
repeat {
newfibonacci <- a + b
if (newfibonacci > 500) {
   break
}
```

```
if (newfibonacci >= start) {
cat(newfibonacci, "\n")
}
a <- b
b <- newfibonacci
}

## 13
## 21
## 34
## 55
## 89
## 144
## 233
## 377</pre>
```

### Using Basic Graphics (plot(),barplot(),pie(),hist())

- 4. Import the dataset as shown in Figure 1 you have created previously.
- a. What is the R script for importing an excel or a csv file? Display the first 6 rows of the dataset? Show your codes and its result

```
library(readr)
data <- read_csv("/cloud/project/shoesize.csv")</pre>
## Rows: 13 Columns: 6
## -- Column specification -------
## Delimiter: ","
## chr (2): F, M
## dbl (4): 6.5, 66, 13, 77
##
## i Use `spec()` to retrieve the full column specification for this data.
## i Specify the column types or set `show_col_types = FALSE` to quiet this message.
data
## # A tibble: 13 x 6
                       `13`
     `6.5`
           `66` F
                             `77` M
     <dbl> <dbl> <chr> <dbl> <dbl> <chr>
```

```
##
##
##
    1
        9
              68
                    F
                            11.5
                                     72 M
##
    2
        8.5
              64.5 F
                             8.5
                                     59 F
    3
        8.5
              65
                    F
                             5
                                     62 F
##
                                     72 M
##
    4
       10.5
              70
                    М
                            10
        7
                    F
##
    5
              64
                             6.5
                                     66 F
##
    6
        9.5
              70
                    F
                             7.5
                                     64 F
##
    7
        9
              71
                    F
                             8.5
                                     67 M
##
    8
       13
              72
                    М
                            10.5
                                     73 M
##
    9
        7.5
              64
                    F
                             8.5
                                     69 F
       10.5
                                     72 M
## 10
              74
                    М
                            10.5
## 11
        8.5
              67
                    F
                            11
                                     70 M
## 12
       12
              71
                    М
                             9
                                     69 M
## 13
                            13
                                     70 M
       10.5
                    М
```

b. Create a subset for gender(female and male). How many observations are there in Male? How about in Female? Write the R scripts and its output.

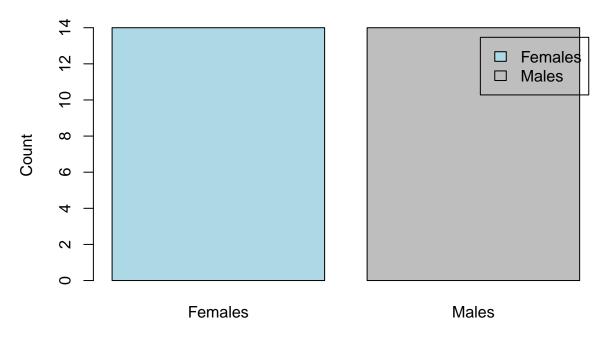
#### ## [1] 14

c. Create a graph for the number of males and females for Household Data. Use plot(), chart type = barplot. Make sure to place title, legends, and colors. Write the R scripts and its result.

```
gender_counts <- c(Females = 14, Males = 14)

barplot(
   gender_counts,
   main = "Number of Males and Females in Household Data",
   xlab = "Gender",
   ylab = "Count",
   col = c("lightblue", "grey"),
   legend.text = c("Females", "Males"),
   beside = TRUE
)</pre>
```

# **Number of Males and Females in Household Data**



## Gender

- 5. The monthly income of Dela Cruz family was spent on the following: Food Electricity Savings Miscellaneous  $60\ 10\ 5\ 25$
- a. Create a piechart that will include labels in percentage. Add some colors and title of the chart. Write the R scripts and show its output.

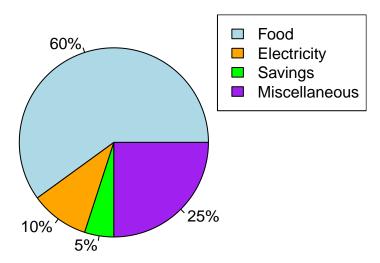
```
categories <- c("Food", "Electricity", "Savings", "Miscellaneous")
expenses <- c(60, 10, 5, 25)

percent_labels <- pasteO(round((expenses / sum(expenses)) * 100), "%")

colors <- c("lightblue", "orange", "green", "purple")

pie(expenses, labels = percent_labels, col = colors, main = "Dela Cruz Family Monthly Expenses")
legend("topright", legend = categories, fill = colors)</pre>
```

# **Dela Cruz Family Monthly Expenses**



- 6. Use the iris dataset. data(iris)
- a. Check for the structure of the dataset using the str() function. Describe what you have seen in the output.

```
data(iris)
str(iris)
```

```
## 'data.frame': 150 obs. of 5 variables:
## $ Sepal.Length: num 5.1 4.9 4.7 4.6 5 5.4 4.6 5 4.4 4.9 ...
## $ Sepal.Width : num 3.5 3 3.2 3.1 3.6 3.9 3.4 3.4 2.9 3.1 ...
## $ Petal.Length: num 1.4 1.4 1.3 1.5 1.4 1.7 1.4 1.5 1.4 1.5 ...
## $ Petal.Width : num 0.2 0.2 0.2 0.2 0.2 0.4 0.3 0.2 0.2 0.1 ...
## $ Species : Factor w/ 3 levels "setosa", "versicolor", ..: 1 1 1 1 1 1 1 1 1 1 ...
```

b. Create an R object that will contain the mean of the sepal.length, sepal.width, petal.length, and petal.width. What is the R script and its result?

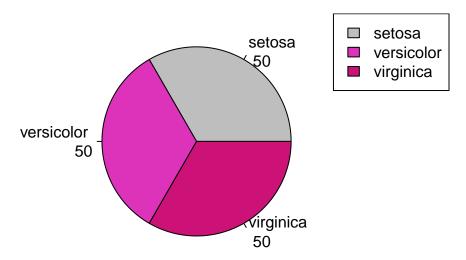
```
mean_values <- colMeans(iris[, 1:4])
mean_values</pre>
```

```
## Sepal.Length Sepal.Width Petal.Length Petal.Width ## 5.843333 3.057333 3.758000 1.199333
```

c. Create a pie chart for the Species distribution. Add title, legends, and colors. Write the R script and its result.

```
data(iris)
species_counts <- table(iris$Species)
pie(species_counts,
main = "Species Distribution in Iris Dataset",
col = c("grey", "#D3B", "#C17"),
labels = paste(names(species_counts), "\n", species_counts))
legend("topright", legend = names(species_counts), fill = c("grey", "#D3B", "#C17"))</pre>
```

# **Species Distribution in Iris Dataset**



d. Subset the species into setosa, versicolor, and virginica. Write the R scripts and show the last six (6) rows of each species.

```
setosa_subset <- iris[iris$Species == "setosa", ]
versicolor_subset <- iris[iris$Species == "versicolor", ]
virginica_subset <- iris[iris$Species == "virginica", ]
last_six_setosa <- tail(setosa_subset, 6)
last_six_versicolor <- tail(versicolor_subset, 6)
last_six_virginica <- tail(virginica_subset, 6)
last_six_setosa</pre>
```

##		Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
##	45	5.1	3.8	1.9	0.4	setosa
##	46	4.8	3.0	1.4	0.3	setosa
##	47	5.1	3.8	1.6	0.2	setosa
##	48	4.6	3.2	1.4	0.2	setosa
##	49	5.3	3.7	1.5	0.2	setosa
##	50	5.0	3.3	1.4	0.2	setosa

last\_six\_versicolor

##		Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
##	95	5.6	2.7	4.2	1.3	versicolor
##	96	5.7	3.0	4.2	1.2	versicolor
##	97	5.7	2.9	4.2	1.3	versicolor
##	98	6.2	2.9	4.3	1.3	versicolor
##	99	5.1	2.5	3.0	1.1	versicolor
##	100	5.7	2.8	4.1	1.3	versicolor

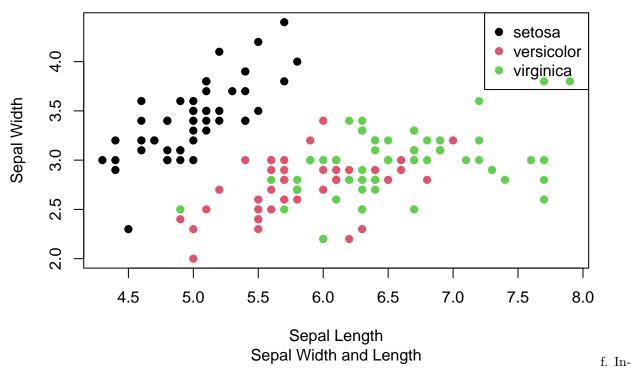
last\_six\_virginica

##		Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
##	145	6.7	3.3	5.7	2.5	virginica
##	146	6.7	3.0	5.2	2.3	virginica
##	147	6.3	2.5	5.0	1.9	virginica
##	148	6.5	3.0	5.2	2.0	virginica
##	149	6.2	3.4	5.4	2.3	virginica
##	150	5.9	3.0	5.1	1.8	virginica

e. Create a scatterplot of the sepal.length and sepal.width using the different species(setosa, versicolor, virginica). Add a title = "Iris Dataset", subtitle = "Sepal width and length, labels for the x and y axis, the pch symbol and colors should be based on the species. Male? How about in Female? Write the R scripts and its output.

```
plot(iris$Sepal.Length, iris$Sepal.Width,
main = "Iris Dataset",
sub = "Sepal Width and Length",
xlab = "Sepal Length",
ylab = "Sepal Width",
pch = 19,
col = iris$Species)
legend("topright", legend = levels(iris$Species),
col = 1:3, pch = 19)
```

### **Iris Dataset**



terpret the result. The number of each iris species in the dataset is displayed in the pie chart. Every slice shows the size of each group that determines the most prevalent species. The subsets show the final six records for every species, enabling us to examine their particular measurements, such as sepal and petal sizes, and recognize their differences. The scatterplot displays the connection between sepal and width and length. In the event that setosa clusters at lower measures, this species often has smaller blooms, but if virginica and versicolor overlap, it indicates that their sizes are comparable, which makes it more difficult to distinguish between them.

### BASIC CLEANING AND TRANSFORMATION OF OBJECTS

- 7. Import the alexa-file.xlsx. Check on the variations. Notice that there are ex- tra whitespaces among black variants (Black Dot, Black Plus, Black Show, Black Spot). Also on the white variants (White Dot, White Plus, White Show, White Spot).
- a. Rename the white and black variants by using gsub() function.

```
library(readxl)
alexa_data <- read_excel("/cloud/project/alexa_file.xlsx")</pre>
unique(alexa_data$variation)
    [1] "Charcoal Fabric"
                                         "Walnut Finish"
                                         "Sandstone Fabric"
##
    [3] "Heather Gray Fabric"
##
    [5] "Oak Finish"
                                         "Black"
   [7] "White"
##
                                         "Black Spot"
## [9] "White Spot"
                                         "Black Show"
                                         "Black Plus"
## [11] "White Show"
## [13] "White Plus"
                                         "Configuration: Fire TV Stick"
## [15] "Black Dot"
                                         "White Dot"
alexa data$variation <- gsub("Black Dot", "BlackDot", alexa data$variation)
alexa_data$variation <- gsub("Black Plus", "BlackPlus", alexa_data$variation)</pre>
alexa_data$variation <- gsub("Black Show", "BlackShow", alexa_data$variation)</pre>
alexa_data$variation <- gsub("Black Spot", "BlackSpot", alexa_data$variation)</pre>
alexa_data$variation <- gsub("White Dot", "WhiteDot", alexa_data$variation)</pre>
alexa_data$variation <- gsub("White Plus", "WhitePlus", alexa_data$variation)</pre>
alexa_data$variation <- gsub("White Show", "WhiteShow", alexa_data$variation)</pre>
alexa_data$variation <- gsub("White Spot", "WhiteSpot", alexa_data$variation)
unique(alexa_data$variation)
   [1] "Charcoal Fabric"
                                         "Walnut Finish"
   [3] "Heather Gray Fabric"
                                         "Sandstone Fabric"
##
##
  [5] "Oak Finish"
                                         "Black"
## [7] "White"
                                         "Black Spot"
   [9] "White Spot"
                                         "Black Show"
                                         "Black Plus"
## [11] "White Show"
## [13] "White Plus"
                                         "Configuration: Fire TV Stick"
## [15] "Black Dot"
                                         "White Dot"
  b. Get the total number of each variations and save it into another object. Save the object as varia-
     tions.RData. Write the R scripts. What is its result? Hint: Use the dplyr package. Make sure to install
     it before loading the package.
library(dplyr)
##
## Attaching package: 'dplyr'
## The following objects are masked from 'package:stats':
##
##
       filter, lag
## The following objects are masked from 'package:base':
##
       intersect, setdiff, setequal, union
variations <- alexa data %>%
count(variation)
print(variations)
## # A tibble: 16 x 2
##
      variation
##
      <chr>
                                     <int>
```

261

## 1 Black

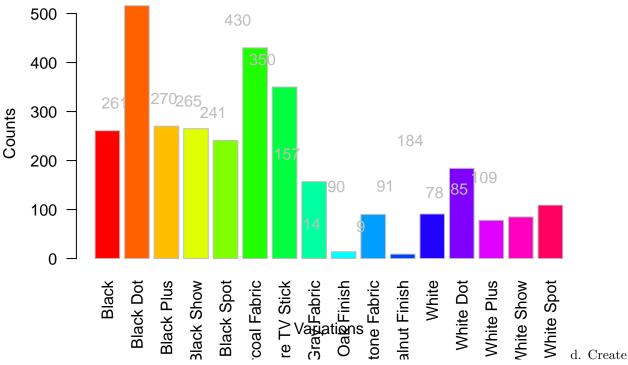
```
## 2 Black Dot
                                    516
## 3 Black Plus
                                    270
## 4 Black Show
                                    265
## 5 Black Spot
                                    241
## 6 Charcoal Fabric
                                    430
## 7 Configuration: Fire TV Stick
                                    350
## 8 Heather Gray Fabric
                                    157
## 9 Oak Finish
                                     14
## 10 Sandstone Fabric
                                     90
## 11 Walnut Finish
                                      9
## 12 White
                                     91
## 13 White Dot
                                    184
## 14 White Plus
                                     78
## 15 White Show
                                     85
## 16 White Spot
                                    109
save(variations, file = "variations.RData")
```

### SAMPLE OUTPUT

c. From the variations.RData, create a barplot(). Complete the details of the chart which include the title, color, labels of each bar.

```
library(dplyr)
load("variations.RData")
variations$variation <- gsub(" +", " ", variations$variation)</pre>
variations$variation <- trimws(variations$variation)</pre>
bardata <- variations$n</pre>
barnames <- variations$variation</pre>
barplot(
bardata,
main = "Counts of Variations",
col = rainbow(length(bardata)),
names.arg = barnames,
xlab = "Variations",
ylab = "Counts",
las = 2,
border = "grey"
)
text(
x = seq_along(bardata),
y = bardata + max(bardata) * 0.05,
labels = bardata,
pos = 3,
cex = 1,
col = "grey"
```

## **Counts of Variations**



a barplot() for the black and white variations. Plot it in 1 frame, side by side. Complete the details of the chart.

```
library(ggplot2)
library(dplyr)
load("variations.RData")
variations$variation <- gsub(" +", " ", variations$variation)</pre>
variations$variation <- trimws(variations$variation)</pre>
bwvariations <- variations %>%
filter(grepl("Black|White", variation))
bardata <- as.matrix(bwvariations$n)</pre>
barnames <- bwvariations$variation</pre>
barplot(
bardata,
beside = TRUE,
main = "Counts of Black and White Variations",
col = c("lightblue", "pink", "yellow", "grey"),
names.arg = barnames,
xlab = "Variations",
ylab = "Counts",
las = 2,
border = "black"
)
text(x = seq_along(bardata), y = bardata, labels = bardata, pos = 3, cex = 0.8, col = "black")
```

# **Counts of Black and White Variations**

