Seed-bank contribution in maintaining phytoplanktonic biodiversity

April 30, 2020

Introduction

How the high biodiversity of plant communities maintains is still an unresolved question for both experimental and theoretical ecologists. Terrestrial plants and phytoplanktonic communities can present hundreds of species relying on similar resources. Early theoreticians have proposed that environmental fluctuations only [ref] could sustain coexistence but further research showed that this could not explain the order of magnitude of species richness [ref]. Other mechanisms such as niche differentiation[ref], demography [ref] and life history traits [ref] have completed explanation by the stochastic environmental variations and demographic processes.

Analyses of coexistence in terrestrial plant communities often take into account several life stages [refs]. Considering at least two stages, seeds/juveniles and adults, different models have uncovered mechanisms that might explain long-term coexistence. Examples of such mechanisms are the bet-hedging strategy, the storage effect and the Janzen-Connell effect. Bet-hedging is a long-term strategy relying on the creation of seeds which can remain dormant for a long period of time (over a year, often much longer). Dormant seeds can tolerate harsher years during which adults cannot maintain, but they also reduce part of the population that could germinate from one year to another (in case of an annual plant). The storage effect has first been defined by the presence of a long-lived life stage and temporal variation in recruitment from this long-lived life stage that helps escape interspecific competition (Chesson, 1986; Cáceres, 1997). This has been later generalized as a negative correlation between the effect of the environment and the effect of competition (Ellner et al., 2016). In good environmental conditions, competition between individuals is stronger as seeds might germinate and therefore use the same resources at the same time. Finally, models and experiments suggest that adults can have a negative effect on seed survival, through the Janzen-Connell effect (Comita et al., 2014). Therefore, neglecting explicit modeling of this life stage can modify the understanding we have of the dynamics of the populations (Nguyen et al., 2019).

Even though different coexistence mechanisms have been unveiled through the use of several life stages and a focus on the youngest stage (seeds) for terrestrial plants, aquatic plants, and more specifically phytoplanktonic algae, have not been modeled with the same precision. Although ecologists have proposed for a long time that the blooms (peaks in abundances that can cover several orders of magnitude) may initiate after the resuspension and germination of seeds (Patrick, 1948; Marcus & Boero, 1998), it is unusal to see an explicit model of such process (but see Hinners $et\ al.$, 2019). The classical view behind phytoplankton dynamics is that bloom formation is mostly seasonal, due to the variation in light and temperature, assuming vegetative cells remaining in the environment can duplicate enough to attain bloom amplitude. However, a recent review (Ellegaard & Ribeiro, 2018) suggests that seeds/cysts might another player .

Phytoplankton communities in coastal environments may benefit from seed banks even more than the oceanic communities [REF-find back], as the distance to the sea bottom is smaller allowing frequently recolonization of the pelagic environment from the shallow sea bottom. Also, and similarly to the seed bank approach in the terrestrial

plant literature, Smayda (2002) has proposed the term "pelagic seed bank" to characterize the contribution of the ocean to coastal communities. This has been noticed for dinoflagellate especially [[ref Dinophysis, check what we have on diatoms]]. Conversely, we can wonder to which extent the seed banks can contribute to the biodiversity in the ocean, especially in the long term. Indeed, spores are able to germinate again after tens of years (McQuoid et al., 2002; Ellegaard & Ribeiro, 2018) or even thousands of years (Sanyal et al., 2018) of dormancy, which can have a long-term effect on biodiversity in both oceanic and coastal environments.

Here we build on the discrete metacommunity model by Shoemaker & Melbourne (2016), completed by Wisnoski et al. (2019) who included exchanges with a seed bank. The growth rate in this model is temperature-dependent and inspired by a metabolism-based model (Scranton & Vasseur, 2016), which we adapted to the specific phenology of modeled phytoplankton groups (Picoche & Barraquand, 2020). Empirical interactions between organisms are informed by previous analyses of field data (Picoche & Barraquand, 2019, 2020). In a first step, we use linear interaction effects, then tweak the Beverton-Holt model to introduce saturating interactions. With these different elements, we aim to examine the effect of exchanges between different compartments (ocean and coastal water column and the seed bank at the bottom of the water column) on coexistence.

Methods

Models

Our models builds atop those developed by Shoemaker & Melbourne (2016) and Wisnoski *et al.* (2019). These discrete-time models are designed for metacommunities with multiple competing populations and unfold as follow: first, individuals increase with a Beverton-Holt (BH) density-dependence (eqs. 1 and 3), then exchanges occur between the different compartments or patches constituting the metacommunity (eq. 4).

In this paper, individuals are phytoplanktonic cells that move between the upper layer of coastal water, its bottom layer where a seed bank accumulates (hereafter called a cyst bank to remain consistent with phytoplankton terminology) and the ocean. Only oceanic and coastal cells are subject to BH-density dependence. Cysts are only affected by mortality m and burial due to sedimentation ζ . The different populations are field-inspired morphotypes accounting for the most frequent genera observed along the French coast (Picoche & Barraquand, 2020) and will hereafter be called taxa. Parameters and state variables are defined in Table 1.

The BH formulation of growth is the Lotka-Volterra equivalent for discrete-time models and is often used to represent terrestrial plant growths. In this model, the maximum achievable growth rate is modified by positive (competitive) and negative (facilitative) interactions α_{ij} . In the most classical formulation of the BH model, which is also our first model, interaction effects increase linearly with the abundances (model I, eq. 1). We subsequently defined saturating interactions (model II, eq. 3). More specifically, in our case, the first step of the first model is written as

$$\begin{cases}
N_{t',i,c} = \frac{\exp(r_i(T))N_{t,i,c}}{1+\sum_j \alpha_{ij}N_{t,j,c}} - lN_{t,i,c} \\
N_{t',i,o} = \frac{\exp(r_i(T))N_{t,i,o}}{1+k_{c2o}\sum_j \alpha_{ij}N_{t,j,o}} - lN_{t,i,o} \\
N_{t',i,b} = N_{t,i,b}(1-m-\zeta)
\end{cases}$$
(1)

where the growth rate $r_i(T)$ is a taxon-specific function of the temperature (see eq. 2), the interaction coefficients α_{ij} are the strength of the effect of species j on species i, and the loss term l accounts for lethal processes such a natural mortality, predation or parasitism. Interactions are inferred from a previous work on coastal data with a Multivariate AutoRegressive (MAR) model (Picoche & Barraquand, 2020). The shift from MAR to BH- interaction matrices is described in the SI. In this model, as we assume that competition for nutrients is stronger in the ocean than along the coast [ref], a coefficient $k_{c2o} > 1$ is applied to competitive interactions.

The growth rate $r_i(T)$ is a modified version of the formula by Scranton & Vasseur (2016) (eq. 2).

$$r_{i}(T) = E(T)f_{i}(T)$$
where $E(T) = d \times 0.81e^{0.0631T_{\circ_{c}}}$
and $f_{i}(T) = \begin{cases} \exp(-|T_{K} - T_{K,i}^{opt}|^{3}/b_{i}), & T_{K} \leq T_{K,i}^{opt} \\ \exp(-5|T_{K} - T_{K,i}^{opt}|^{3}/b_{i}), & T > T_{K,i}^{opt} \end{cases}$

where $r_i(T)$ can be decomposed in two parts: the taxon-independent metabolism part E(T) and the taxon-specific niche part $f_i(T)$. The metabolism part describes the maximum achievable growth rate based on Bissinger et al. (2008), as an update of the formula by Eppley (1972) used in Scranton & Vasseur (2016). This maximum daily growth rate is weighted by the daylength d as no growth occurs at night. The niche part $f_i(T)$ describes the decrease in growth rate due to the difference between the temperature in the environment and the taxon-specific thermal optimum $T_{K,i}^{opt}$, and is controlled by the specific thermal decay b_i which depends of the niche width. Parameterisation is detailed in the SI.

In model II, oceanic and coastal dynamics are governed by eq. 2.

$$N_{t',i,c/o} = \frac{\exp(r_i(T))N_{t,i,c/o}}{1 + \sum_{j \in \mathbb{C}} \frac{a_C N_{t,j,c/o}}{H_{ij} + N_{t,i,c/o}} + \sum_{j \in \mathbb{F}} \frac{a_F N_{t,j,c/o}}{H_{ij} + N_{t,ic/o}} - lN_{t,i,c/o}} - lN_{t,i,c/o}}$$
(3)

where a_C and a_F are the maximum competition and facilitation strengths, respectively, with \mathbb{C} and \mathbb{F} the sets of competitors and facilitators of taxon i. The half-saturation coefficients H_{ij} characterize the effect of j on i. The switch from model I to model II is described in the SI.

After growth and mortality happen, exchanges take place between the three compartments during the second step of the model (eq. 4).

$$\begin{cases}
N_{t+1,i,c} = (1 - s_i - e)N_{t',i,c} + \gamma N_{t',i,b} + eN_{t',i,o} \\
N_{t+1,i,o} = (1 - s_i - e)N_{t',i,o} + eN_{t',i,c} \\
N_{t+1,i,b} = (1 - \gamma)N_{t',i,b} + s_iN_{t',i,c}
\end{cases} (4)$$

| Param | Name | Value (unit) | Status |
|-------------------|--|--|------------|
| $N_{t,i,c/o/b}$ | Abundance of taxon i at time t in the coast (c) or ocean (o) , or in the coastal benthos (b) | NA (Number of cells) | Dynamic |
| $T_{K/^{\circ}C}$ | Temperature | NA $(K/^{\circ}C)$ | Dynamic |
| $r_i(T)$ | Growth rate of taxon i | NA | Dynamic |
| b_i | Thermal decay | Field-based, taxon-specific (K^3) | Calibrated |
| T_i^{opt} | Optimal temperature for taxon i | Field-based, taxon-specific (K) | Calibrated |
| d | Daylength | 0.5 (%) | Fixed |
| $\alpha_{ij,c/o}$ | Interaction strength of taxon j on i in model I | Field-based, taxon-specific (Cells ⁻¹) | Calibrated |
| k_{c2o} | Ocean/Coast interaction strength ratio in model I | 1.5 | Fixed |
| a_C/a_F | Maximum competitive/facilitative interaction strength in model II | Field-based, taxon-specific (NA) | Calibrated |
| H_{ij} | Half-saturation for the interaction strength of taxon j on i in model II | Field-based, taxon-specific (Cells) | Calibrated |
| s_i | Sinking rate of taxon i | $\{0.1; 0.3; 0.5\}\beta(0.55, 1.25)$ | Fixed |
| e | Exchange rate between ocean and coast | 0.4; 0 in scenario | Scenario |
| l | Loss rate of vegetative phytoplanktonic cells | 0.04; 0.1; 0.2 | Fixed |
| m | Cyst mortality rate | $\approx 10^{-4}/10^{-5}$; 1 – ζ in scenario | Scenario |
| ζ | Cyst burial rate | $10^{-3}, \mathbf{10^{-2}}, 10^{-1}$ | Fixed |
| γ | Germination \times Resuspension rate | $10^{-3}, \mathbf{10^{-2}}, 10^{-1} \times 10^{-5}, 10^{-3}, \mathbf{10^{-1}}$ | Fixed |

Table 1: Definition of main state variables and parameters of the models. Calibrated parameters are either directly estimated on data for this study or parameters for which initial estimates exist, but are improved through calibration. Fixed values or distributions are estimated from the literature and references are given in the main text. When a range of values is given, the bold numbers indicate the reference values while the others are used to test the sensitivity of the model. Scenario parameters are the parameters which are used to build ecological scenarii.

Each compartment (ocean, coast, seed bank) contains 10^3 cells at the beginning of the simulation, which is run for 1000 time steps. The temperature at each time step simulates the temperature observed in our example dataset, keeping the mean and variance (see below).

Parameterisation

Empirical dataset used for calibration

[[Write here]]

Parameter definition and initial values

Loss rate The loss rate of vegetative cells can be attributed to natural mortality, predation or parasitism. A maximum value of 0.2 is fixed for the model.

Sinking rate Phytoplanktonic particles have a higher density than water and cannot swim to prevent sinking (although see Reynolds (2006) for a discussion on the settling of phytoplanktonic cells compared to inorganic particles). Sinking is mostly affected by hydrodynamics, but at the species-level, size, shape and colony-formation capacity are key determinants of the particle floatation. In this model, the sinking rate of each taxon is drawn from a beta distribution with a mean value of 9%, and a maximum around 30%, that is $s \sim 0.3\beta(0.55, 1.25)$ (see Fig. SXX), adapted from observations by Passow (1991) and Wiedmann et al. (2016).

Exchange rate The exchange rate between the ocean and the coast depends on the shape and location of the estuary. In Marennes-Oléron and more specifically in Auger, the renewal time ranges between 1 and 2.5 days (Ascione Kenov *et al.*, 2015), which corresponds to a daily rate between 40 and 100 %.

Cyst mortality and burial Cyst loss is the result of cyst mortality m and burial by sedimentation ζ . Mortality values range between 10^{-5} and 10^{-4} (more details on the approximation of mortality rates from McQuoid et~al. (2002) are given in the SI). However, cyst burial by sedimentation might be a prevailing phenomenum in driving phytoplanktonic dynamics. Once cysts have been buried, they are not accessible for resuspension even if they could have germinated if put in an accessible location. Burial depends on the hydrodynamics of the site, but also on biotic processes (i.e., bioturbation) and anthropogenic disturbances such as fishing or leisure activities (e.g., jet skiing). This parameter is thus heavily dependent on the environmental context and varies here between 0.001 and 0.1.

Germination/resuspension Germination and resuspension are both needed for cyst to get back to the water column (γ = resuspension×germination). Following McQuoid *et al.* (2002) and Agrawal (2009), we assume a temperature threshold: germination is triggered by temperatures going above 15°C. As actual rates of germination are not easily deduced from the literature, a set of credible values are tested (1%, 0.1%, 0.01%). Similarly, resuspension values are seldom computed for phytoplanktonic cells, but models for other particles such as sediments can be used. In this paper, we explore values between 10^{-5} (stratified water column) to 0.1 (highly mixed environment).

Parameter calibration

In addition to phenology parameters whose estimation process is described in the SI, the 49 non-null interactions that form the community matrix of the model are calibrated on field-data. These interactions are computed from

previous models (Picoche & Barraquand, 2020, see SI for details on the equations) but need to be adjusted to take into account the differences in structure and time-step between studies.

The calibration procedure consisted in lauching 1000 simulations, each characterized by a specific interaction set. More precisely, for each simulation, each interaction coefficient (α_{ij} in model I, H_{ij} in model II) has the same probability of keeping its value, being increased or decreased by 10%, or being halved or doubled. The abundances of the coastal compartment are then extracted over the last 2 years of the simulation and compared to observations to compute the following summary statistics.

- average abundance: $f_1 = \sqrt{\frac{1}{S} \sum_{i}^{S} (\bar{N}_{i,obs} \bar{N}_{i,sim})^2}$ where S is the number of taxa.
- amplitude of the cycles: $f_2 = \sqrt{\frac{1}{S} \sum_{i=1}^{S} \left[\left(\max(N_{i,obs}) \min(N_{i,obs}) \right) \left(\max(N_{i,sim}) \min(N_{i,sim}) \right) \right]^2}$
- beginning of the bloom: the year is divided in 3 periods, i.e. summer, winter and the spring/autumn group (as taxa blooming in these periods can appear in either or both seasons). We give a score of 0 if the taxon blooms in the same period as its observed counterpart and 1 otherwise.

These summary statistics are summed up to compute an objective function that needs to be minimized. Simulations with taxon extinction are discarded. The parameter set corresponding to the lowest value of the objective function is kept throughout the rest of the simulation.

Sensitivity analysis Certain parameters, which were evaluated from the literature, may be site- or model-specific, or vary over one order of magnitude, e.g. rates of sinking s, resuspension/germination γ , cyst mortality m and burial ζ , as well as the loss rate l. The dependence between variation in values and outputs of present models needs to be investigated before drawing conclusions in specific ecological scenarii. The set of tested values for each parameter is given in Table 1. Variations in average abundances and amplitudes at the community- and taxon levels for the last 2 years of simulations are the major diagnostics.

Scenarii

Maintenance of biodiversity by exchanges between compartments can be gauged with two main scenarios:

- 1. modifying the storage by the cyst bank (by setting cyst mortality/burial to 1)
- 2. changing the exchanges between the coast and the ocean (e=0)

Final richness and average abundance/biomass productivity (approximated by the final total abundances, to create a link with ecosystem functions) can be diagnostics.

We can make 2 a-priori ecological hypotheses. First, removing the cyst bank would increase the sensitivity to competition, i.e. decrease the maximum sustainable interspecific competition strength, and decrease the resilience of the community confronted to changes in the environment. (Fig. 1). Under high competition, variability in final richness may increase because exclusion is more likely, especially without a cyst bank. This is also true for higher temperatures.

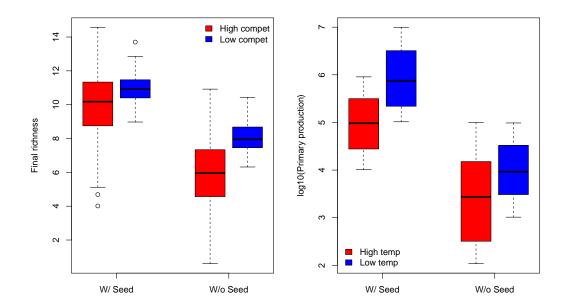


Figure 1: Expected changes in the model outputs in the ocean. Left panel, final richness with and without cyst bank, with high (red) and low (blue) interspecific competition. Right panel, final primary productivity with and without cyst bank, with a high (red) and low (blue) mean temperature.

Second, reducing the exchange with the ocean might lead to depletion of the ocean biomass and richness. In the ocean, individuals disappear through sinking and cannot reappear thanks to coastal production fueled by a cyst bank. Fig. 2 gives a more precise view of the expectation for possible interactions of cyst mortality and exchange between the coast and the ocean.

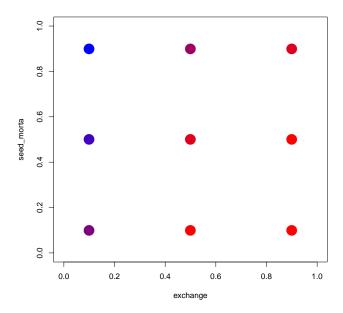


Figure 2: Expected changes in community richness in the ocean for different values of cyst mortality/burial (=access to the seed compartment) and exchange with the ocean. Point color indicates final richness in the ocean, from low (blue) to high (red).

Results

Phytoplankton dynamics

There are still issues in the calibration of the model: abundances can be lower than expected and the variation in abundances due to seasonality is, for now, underestimated (Fig. 3).

Figure 3: Examples of phytoplankton dynamics (*Chaetoceros* on the left, *Skeletonema* on the right). The solid black line is the observed average abundance of the species.

Parameter effect

Find a way to represent the variation in total abundances in the models (Fig. 2) (mostly bar plots?)

Scenarios

Discussion

[[See outline]]

[[Need to compare our values to Jewson et al. (1981) who took into account most of the phenomena we describe here and the cyst bank as well as the water column]

References

Agrawal, S.C. (2009). Factors affecting spore germination in algae - review. Folia Microbiologica, 54, 273–302.

- Ascione Kenov, I., Muttin, F., Campbell, R., Fernandes, R., Campuzano, F., Machado, F., Franz, G. & Neves, R. (2015). Water fluxes and renewal rates at Pertuis d'Antioche/Marennes-Oléron Bay, France. Estuarine, Coastal and Shelf Science, 167, 32–44.
- Bissinger, J., Montagnes, D., Harples, J. & Atkinson, D. (2008). Predicting marine phytoplankton maximum growth rates from temperature: Improving on the Eppley curve using quantile regression. *Limnology and Oceanography*, 53, 487–493.
- Cáceres, C.E. (1997). Temporal variation, dormancy, and coexistence: A field test of the storage effect. Proceedings of the National Academy of Sciences, 94, 9171–9175.
- Chesson, P. (1986). Environmental variation and the coexistence of species. In: *Community ecology* (eds. Diamond, J. & Case, T.). Harper & Row, New-York, chap. 14, pp. 240–256.
- Comita, L., Queenborough, S., Murphy, S., Eck, J., Xu, K., Krishnadas, M., Beckman, N. & Zhu, Y. (2014).
 Testing predictions of the Janzen-Connell hypothesis: a meta-analysis of experimental evidence for distance- and density-dependent seed and seedling survival. *Journal of Ecology*, 102, 845–856.
- Ellegaard, M. & Ribeiro, S. (2018). The long-term persistence of phytoplankton resting stages in aquatic seed banks'. *Biological Reviews*, 93, 166–183.

- Ellner, S., Snyder, R. & Adler, P. (2016). How to quantify the temporal storage effect using simulations instead of math. *Ecology Letters*, 19, 1333–1342.
- Eppley, R. (1972). Temperature and phytoplankton growth in the sea. Fishery Bulletin, 70, 1063–1085.
- Hinners, J., Hense, I. & Kremp, A. (2019). Modelling phytoplankton adaptation to global warming based on resurrection experiments. *Ecological Modelling*, 400, 27–33.
- Jewson, D.H., Rippey, B.H. & Gilmore, W.K. (1981). Loss rates from sedimentation, parasitism, and grazing during the growth, nutrient limitation, and dormancy of a diatom crop. *Limnology and Oceanography*, 26, 1045–1056.
- Marcus, N. & Boero, F. (1998). Minireview: The importance of benthic-pelagic coupling and the forgotten role of life cycles in coastal aquatic systems. *Limnology and Oceanography*, 43, 763–768.
- McQuoid, M.R., Godhe, A. & Nordberg, K. (2002). Viability of phytoplankton resting stages in the sediments of a coastal Swedish fjord. *European Journal Phycology*, 37, 191–201.
- Nguyen, V., Buckley, Y.M., Salguero-Gómez, R. & Wardle, G.M. (2019). Consequences of neglecting cryptic life stages from demographic models. *Ecological Modelling*, 408, 108723.
- Passow, U. (1991). Species-specific sedimentation and sinking velocities of diatoms. Marine Biology, 108, 449–455.
- Patrick, R. (1948). Factors effecting the distribution of diatoms. Botanical Review, 14, 473–524.
- Picoche, C. & Barraquand, F. (2019). How self-regulation, the storage effect, and their interaction contribute to coexistence in stochastic and seasonal environments. *Theoretical Ecology*.
- Picoche, C. & Barraquand, F. (2020). Strong self-regulation and widespread facilitative interactions between genera of phytoplankton. preprint, bioRxiv.
- Reynolds, C.S. (2006). The ecology of phytoplankton. Cambridge University Press.
- Sanyal, A., Larsson, J., van Wirdum, F., Andrén, T., Moros, M., Lönn, M. & Andrén, E. (2018). Not dead yet: Diatom resting spores can survive in nature for several millennia. preprint, bioRxiv.
- Scranton, K. & Vasseur, D.A. (2016). Coexistence and emergent neutrality generate synchrony among competitors in fluctuating environments. *Theoretical Ecology*, 9, 353–363.
- Shoemaker, L.G. & Melbourne, B.A. (2016). Linking metacommunity paradigms to spatial coexistence mechanisms. *Ecology*, 97, 2436–2446.
- Smayda, T.J. (2002). Turbulence, watermass stratification and harmful algal blooms: an alternative view and frontal zones as "pelagic seed banks". *Harmful Algae*, 1, 95–112.
- Wiedmann, I., Reigstad, M., Marquardt, M., Vader, A. & Gabrielsen, T. (2016). Seasonality of vertical flux and sinking particle characteristics in an ice-free high arctic fjord-Different from subarctic fjords? *Journal of Marine* Systems, 154, 192–205.
- Wisnoski, N.I., Leibold, M.A. & Lennon, J.T. (2019). Dormancy in metacommunities. *The American Naturalist*, 194, 135–151.