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### **Original Article**

# Coupled response of stomatal and mesophyll conductance to light enhances photosynthesis of shade leaves under sunflecks

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### **ABSTRACT**

Light gradients within tree canopies play a major role in the distribution of plant resources that define the photosynthetic capacity of sun and shade leaves. However, the biochemical and diffusional constraints on gas exchange in sun and shade leaves in response to light remain poorly quantified, but critical for predicting canopy carbon and water exchange. To investigate the CO<sub>2</sub> diffusion pathway of sun and shade leaves, leaf gas exchange was coupled with concurrent measurements of carbon isotope discrimination to measure net leaf photosynthesis  $(A_n)$ , stomatal conductance  $(g_s)$  and mesophyll conductance (gm) in Eucalyptus tereticornis trees grown in climate controlled whole-tree chambers. Compared to sun leaves, shade leaves had lower  $A_n$ ,  $g_m$ , leaf nitrogen and photosynthetic capacity  $(A_{\text{max}})$  but  $g_s$  was similar. When light intensity was temporarily increased for shade leaves to match that of sun leaves, both  $g_s$  and  $g_m$  increased, and  $A_n$  increased to values greater than sun leaves. We show that dynamic physiological responses of shade leaves to altered light environments have implications for up-scaling leaf level measurements and predicting whole canopy carbon gain. Despite exhibiting reduced photosynthetic capacity, the rapid up-regulation of  $g_{\rm m}$  with increased light enables shade leaves to respond quickly to sunflecks.

*Key-words*: *Eucalyptus tereticornis*; canopy light gradients; climate warming; leaf nitrogen; shade acclimation.

### INTRODUCTION

Light availability is one of the most important environmental drivers of leaf carbon (C) uptake in trees. Predicting C uptake of forests usually involves up-scaling leaf level measurements to assess whole canopy function. Because of the costs and limitations of efficient light harvesting within plant canopies, not all leaves are exposed to full sun (Niinemets 2010), making simple up-scaling based on solar irradiance problematic. Incident photosynthetic photon flux density (PPFD) declines exponentially with cumulative leaf area index (ratio of leaf area to ground area), creating a steep light gradient from the canopy top to bottom (Monsi & Saeki 2005). Consequently, structural and functional properties of leaves within canopies are

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modified to efficiently use light (Vogelman *et al.* 1996; Niinemets & Valladares 2004), as changing irradiance with canopy depth strongly affects rates of leaf net photosynthesis  $(A_n)$  (Evans 1995). To calculate whole canopy C gain, it is thus necessary to account for the non-linear response of  $A_n$  to light by distinguishing between shaded and sunlit leaves (De Pury & Farquhar 1997; Linderson *et al.* 2012).

The distribution of resources required for  $A_n$ , including leaf nitrogen (N) and supply of water, is also partially defined by canopy light gradients. As  $A_n$  has a saturating response with light and maximum rates depend, in part, on N-rich photosynthetic machinery, it has been argued that leaf N should be proportional to PPFD along the canopy light gradient to maximize canopy C gain at a given total canopy N (Field 1983; Field & Mooney 1986; Peltoniemi et al. 2012; Buckley et al. 2013). Changes in chlorophyll per unit N, chlorophyll a:b ratios, electron transport capacity per unit chlorophyll and ratios of electron transport capacity to Rubisco activity can also occur in response to changes in irradiance of the growth environment (Evans & Poorter 2001). Sun leaves frequently experience greater water limitations in the upper canopy, despite effective vascular systems that accommodate high radiation loads and transpiration (Sellin et al. 2008; Niinemets 2012). Optimal photosynthetic N investment in the upper canopy will be ineffective in enhancing  $A_n$  if water supply is insufficient (Niinemets 2012; Peltoniemi et al. 2012); thus, leaf-specific hydraulic conductance  $(K_{\rm I})$  should also be higher in the upper canopy to supply sunlit leaves with sufficient water (Hubbard et al. 2001; Burgess et al. 2006; Sellin & Kupper 2007; Sellin et al. 2008).

The photosynthetic rate in  $C_3$  plants is limited by the  $[CO_2]$  available for fixation by Rubisco within the chloroplast and this  $[CO_2]$  is a function of the drawdown of  $CO_2$  from the atmosphere to the site of carboxylation (Warren 2008). There are two major resistance pathways to  $CO_2$  diffusion: the  $CO_2$  diffusion from the atmosphere through stomata into intercellular air spaces (stomatal conductance,  $g_s$ ) and the pathway from the intercellular air spaces into the chloroplast of the mesophyll cells (mesophyll conductance,  $g_m$ ). Based on optimality theory, regulation of  $g_s$  within a tree canopy should act to efficiently utilize available supplies of light, N and water to maximize  $A_n$  (Peltoniemi *et al.* 2012). This is because stomata are hypothesized to exhibit an optimal behaviour to maximize C gain while simultaneously minimizing water loss through transpiration (Cowan & Farquhar 1977). Mesophyll conductance

can also impose limitations on  $A_n$  that can be as large as  $g_s$ (Warren 2008; Ubierna & Marshall 2011), reducing the efficiency of leaf N use in  $A_n$  (Niinemets 2007). Differences in photosynthetic capacity between sun and shade leaves may arise, in part, from differences in  $g_{\rm m}$  (Piel et al. 2002; Warren et al. 2007), vet the trade-offs that constrain this diffusion pathway are vet to be explicitly quantified. It is likely that leaf anatomical costs associated with minimizing the length and tortuosity of the  $g_{\rm m}$ diffusion pathway are necessary to maintain the benefits of a high  $g_{\rm m}$  (Hassiotou et al. 2009). Nonetheless, like  $g_{\rm s}$ ,  $g_{\rm m}$  may also vary dynamically in response to environmental drivers (Flexas et al. 2008). Thus, stomatal and mesophyll conductance should not be considered independent of each other (Griffiths & Helliker 2013), but a lack of empirical data currently hinders our ability to interpreting their coupled responses to  $A_n$  across sun and shade leaves.

How shade leaves utilize sunflecks for short-term carbon gain depends on the combined response time of  $g_s$  and  $g_m$ and the underlying photosynthetic machinery acclimated to a low light environment (Pearcy 1990; Tausz et al. 2005). The utilization of sunflecks in shade leaves is initially limited by delayed responses of stomata opening, which may take minutes, limiting the assimilation rate that can be achieved (Pearcy 1990; Vico et al. 2011; Way & Pearcy 2012). Mesophyll conductance has been shown to respond to environmental factors (e.g. CO<sub>2</sub>, temperature or vapour pressure deficit) at timescales of minutes, possibly faster than g<sub>s</sub> (Flexas et al. 2008 and references therein), yet the short-term response to light availability is unclear. For example, gm was found to be independent of incident light intensity in wheat leaves (Tazoe et al. 2009) but was responsive to light in tobacco (Flexas et al. 2007) and Eucalyptus seedlings (Douthe et al. 2011, 2012). Anatomical parameters which regulate  $g_m$  with changing growth irradiance such as chloroplast surface area (Terashima et al. 2006) and mesophyll thickness (Boardman 1977; Terashima et al. 2001; Hanba et al. 2002) are also unlikely to adjust during transient fluctuations in light. The physiological behaviour of shade leaves to maximize C gain must be assessed as both a degree of acclimation to local irradiance and as a potential response to transitory light availability.

Climate warming may also affect the physiological behaviour of leaves within a canopy. Leaves are exposed to different heat, water and high light stresses as temperature and vapour pressure deficit (VPD) vary with canopy light availability (Baldocchi et al. 2002; Niinemets & Valladares 2004; Niinemets 2007). How these stresses affect the diffusion of  $CO_2$ , through either  $g_s$  or  $g_{\rm m}$ , will have implications for up-scaling  $A_{\rm n}$  for sun and shade leaves. How these parameters, in combination with photosynthetic capacity, are differentially affected by warming may impact constraints of N distribution and leaf photosynthetic capacity across light gradients. The impacts of warming on plant physiological processes are diverse, yet differentiating their impacts on leaf physiology within a canopy will be essential to evaluate whole tree responses to a changing climate.

In this study, we use Eucalyptus tereticornis trees, planted in naturally sunlit climate controlled whole-tree chambers with ambient and elevated temperature (ambient +3 °C) treatments, to test whether the distribution of N, water supply

capacity and leaf physiological traits result in higher photosynthetic capacity in sun leaves compared to shade leaves. We further aimed to quantify the constraints on  $A_n$  in sun and shade leaves arising from photosynthetic capacity and the CO<sub>2</sub> diffusion pathway via stomatal and mesophyll conductance. As leaves which were acclimated to shade conditions were expected to have lower photosynthetic capacity and correspondingly lower diffusive conductance (both  $g_s$  and  $g_m$ ) than sun leaves, we predicted that sunfleck simulations would increase  $A_n$  and  $g_s$  in shade leaves, but reach values lower than that attained by sun leaves in similar conditions. We further predicted that climate warming would decrease g<sub>s</sub> and leaf C gain in sun leaves more so than shade leaves during summer months, as increased evaporative demand from higher temperatures and irradiance lead to stomatal closure.

### **MATERIALS AND METHODS**

### Whole-tree chamber experimental design

Twelve E. tereticornis Sm. seedlings, chosen from a single local Cumberland plain cohort, were planted in March 2013 into 12 whole-tree chambers (WTC) at the Hawkesbury Forest Experiment site near Richmond, New South Wales, Australia. The top soils at this site, used in the chambers, are an alluvial formation of low-fertility sandy loam soils (380 and 108 mg kg<sup>-1</sup> total N and phosphorus respectively) with low organic matter (0.7%) and low water holding capacity. A root exclusion barrier extended from chamber walls to the hard layer (ca. 1 m) and roots were allowed to grow freely below the barrier. Each chamber was 9 m tall, which accommodated growth of trees for 15 months. A detailed description of the WTC operation and design is available in Barton et al. (2010) and methods for this experiment in Drake et al. (2016). Coppied E. saligna trees were present in the area between the whole-tree chambers, and these coppiced trees grew at rates similar to the trees in the chambers, thus providing ample shade. Six chambers were set to match outside ambient air temperatures (AT) while the remaining six experienced an elevated air temperature treatment of ambient +3 °C (ET, Figure S1). The trees grew quickly and developed large canopies, with height growth reaching the top of the WTCs at the end of the experiment. Trees were watered weekly with 70 L from March 2013 to September 2013. From October 2013 to the end of the experiment trees were watered every 15 days with the mean monthly rainfall amount for Richmond, NSW. In February 2014 half of the chambers (three each of AT and ET) were subjected to a drought treatment by withholding watering. Because of the difficulty of measuring  $g_{\rm m}$  in the droughted trees (the necessary CO2 drawdown was not achieved), we here report only data from well-watered trees, which reduces the sample size from n = 6 to n = 3 for the final 3 months of the experiment.

Leaf gas exchange measurements were initiated in October 2013 when trees had both ample height growth and canopy development for realistic canopy light gradients to be measured. At this point, trees under AT treatment had a mean stem diameter (measured at 65 cm from the stem base) of  $28.2 \pm 1.1$  mm (SE), height of  $348 \pm 15.1$  cm and a total leaf area of  $3.9 \pm 0.1 \,\mathrm{m}^2$ . For ET treatments, trees had a mean diameter of  $34.1 \pm 2.1 \,\mathrm{mm}$ , height of  $418.3 \pm 23.1 \,\mathrm{cm}$  and a leaf area of  $6.2 \pm 0.2 \,\mathrm{m}^2$ . Leaf area was calculated based on complete leaf counts and mean leaf size from a subsample from each tree (Drake *et al.* 2016).

### Leaf gas exchange, coupled with concurrent measurements of carbon isotope discrimination to estimate mesophyll conductance

Leaf gas exchange measurements were performed six times, beginning in October 2013 and monthly from December 2013 to April 2014. Measurements were taken on a representative sun and shade leaf for each tree during each measurement campaign. The newest fully expanded leaf from the branch apex was chosen for gas exchange measurements, and sun leaves were measured in the upper third of the canopy. Here, shade leaves are defined as inner-canopy leaves developing on secondary branches in a low light environment. Shade leaves were always measured in the lower canopy, but leaves were sampled on subsequent higher branches across measurement campaigns to minimize confounding effects of leaf age. As shade leaves most likely developed more slowly, this assured that older leaves in the bottom canopy were avoided.

Prior to gas exchange measurements, PPFD was recorded both as a point measurement at the individual leaf level and a spatially averaged measurement at the canopy position for each selected leaf. A hand-held quantum sensor (Li-Cor, Lincoln, NE, USA) was used to record leaf level PPFD to ensure that chosen leaves were positioned in the desired light environment, either sun or shade. A ceptometer (AccuPAR LP-80, Decagon Devices, Pullman, USA) was then used to measure a spatially averaged PPFD at the canopy height of each chosen leaf type. Each ceptometer reading integrated an array of 80 sensors over a total length of 84 cm. Five ceptometer readings were recorded at different locations within the canopy, but at the same height and close to each selected leaf. The mean of these readings was assumed to represent the local light environment of sun and shade leaves for each tree. All measurements of PPFD and gas exchange were performed on sunny days between 10:00 and 14:30 h.

Concurrent gas exchange and C isotope discrimination measurements were conducted based on methods described in Tazoe et al. (2011) and Evans & von Caemmerer (2013). Leaf level gas exchange was measured with a standard (2×3 cm) leaf cuvette and red/blue light source using a portable gas exchange system (LI-6400XT, Li-Cor, Lincoln, NE, USA). This system was coupled with a tunable diode laser (TDL; TGA100, Campbell Scientific, Inc., Logan, UT, USA) for concurrent measurements of <sup>13</sup>C isotope discrimination. The CO<sub>2</sub> in the leaf cuvette was set at ambient atmospheric [CO<sub>2</sub>]  $(400 \,\mu\text{l l}^{-1})$  with a flow rate of  $200 \,\mu\text{mol s}^{-1}$ . Two identical gas exchange systems were run simultaneously, one in each of a randomly chosen WTC for each temperature treatment. Leaf temperatures were controlled at the current AT or ET WTC air temperature, and measurements were made at ambient air humidity. Measurements were thus made across a range of air temperatures and vapour pressure deficits (kPa) across the six campaigns. PPFD in the cuvette was set to match the individual light environment of each leaf type (explained above). Sustained periods of high irradiance (sunflecks) were simulated for shade leaves by increasing the leaf cuvette PPFD (Li-Cor red/blue light source) to match the light environment of the sun leaf in the same tree. The maximum sunfleck response of shade leaves was then recorded once CO<sub>2</sub> and water vapour fluxes re-stabilized in the leaf cuvette (ca. 25 min).

Once CO<sub>2</sub> and water vapour flux values were stable for each leaf measurement, the sample and reference gas lines were diverted to the TDL via T-junctions inserted into the reference gas tube and match valve outlet of the LI-6400XT. The gas streams were dried by passing through napion gas dryers in the respective gas lines, and then <sup>12</sup>CO<sub>2</sub> and <sup>13</sup>CO<sub>2</sub> concentrations were measured for each gas stream by the TDL. Reference, sample and two calibration gases were run on alternating 80 s loops (20 s each), one for each AT and ET leaf at a matched canopy position, for a total of 12 min. This allowed for four or five measurements per leaf and data were averaged over the last 10s of reference line and sample line gas streams for calculations. The two calibration gases were drawn from compressed air tanks (330 and 740 ppm CO<sub>2</sub>) in order to correct for gain drift of the TDL on each measurement cycle. All gas exchange variables were auto-logged every 15 s for each gas exchange system over the 12 min interval.

Mesophyll conductance was calculated from carbon isotope discrimination with equations and fractionation factors as presented in Evans & von Caemmerer (2013), including the ternary corrections proposed by Farquhar & Cernusak (2012), such that:

$$\Delta_o = \Delta_i - \Delta_{gm} - \Delta_e - \Delta_f \tag{1}$$

where  $\Delta_0$  is the observed discrimination and  $\Delta_i$ ,  $\Delta_{gm}$ ,  $\Delta_e$  and  $\Delta_f$ are the contributions to fractionation if  $C_i = C_c$ ,  $g_m$ , respiration and photorespiration, respectively.  $C_i$  and  $C_c$  are the intercellular and chloroplast CO<sub>2</sub> partial pressures. For this study, the  $CO_2$  compensation point  $(\Gamma_*)$  and respiration during the day (R<sub>d</sub>) parameters originally derived for tobacco plants (von Caemmerer et al. 1994) were replaced with parameters derived for Eucalyptus globulus from Crous et al. (2012) when calculating  $g_{\rm m}$ . Full descriptions of the carbon isotope discrimination equations, with ternary corrections, are presented in Supporting Information Methods S1. The variation in  $\Delta_0$  between sun and shade leaves and the simulated sunfleck were compared as a function of  $C_i/C_a$ . Once  $g_m$  was calculated,  $C_c$ and the drawdown of CO<sub>2</sub> from the intercellular air spaces to the site of carboxylation were estimated from the relationship between  $g_m$  and leaf photosynthesis rate  $(A_n)$  by:

$$g_m = \frac{A_n}{C_i - C_c}. (2)$$

### Photosynthetic capacity

Photosynthetic capacity  $(A_{\text{max}})$  of sun and shade leaves was measured at saturating light and  $[CO_2]$  for one sun and shade

leaf for each WTC. Measurements were completed in February 2014, when all trees were well-watered in each temperature treatment (n = 6).  $A_{\text{max}}$  measurements were made at a PPFD of  $1800 \,\mu\text{mol m}^{-1}\,\text{s}^{-1}$  and  $[\text{CO}_2]$  of  $1800 \,\mu\text{l l}^{-1}$ , with leaf temperature held constant at 25 °C.

### Leaf nitrogen and hydraulic conductance

Following gas exchange measurements, each leaf was collected, measured for leaf water potential (explained below), scanned for leaf area, oven-dried and weighed. These leaves were then milled and analysed for leaf N content and  $\delta^{13}$ C. Leaf samples were analysed on a Delta V Advantage coupled to a Flash HT and Conflo IV (Thermo Fisher Scientific, Bremen, Germany) in dual-reactor setup. Samples were flash combusted at 1000 °C and converted to CO2 and N2 and then subjected to stable isotope ratio mass spectrometry. Leaf N is reported on an area basis ( $N_a$ , g m<sup>-2</sup>) and isotopic signatures of dry matter are reported relative to standard Vienna Pee Dee Belemnite.

Prior to the gas exchange measurements, predawn  $(\Psi_{pd})$  leaf water potentials (MPa) were measured for a separate set of sun and shade leaves using a PMS 1505D pressure chamber (PMS Instruments, Albany, OR, USA). The leaf closest to the leaf used for gas exchange was sampled for measurement of  $\Psi_{pd}$ before sunrise on the same day as gas exchange measurements. Following the gas exchange measurements, leaves used for gas exchange were immediately sampled for midday  $(\Psi_L)$  leaf water potentials. All leaves were detached and immediately stored inside foil covered bags before water potential measurements were performed. Leaf water potentials and transpiration  $(E_{\rm L}, {\rm mmol\,m^{-2}\,s^{-1}})$  from gas exchange in the leaf cuvette were then used to calculate leaf-specific hydraulic conductance ( $K_{\rm I}$ , mmol m<sup>-2</sup> s<sup>-1</sup> MPa<sup>-1</sup>) from the equation (Whitehead 1998; Flexas et al. 2013):

$$K_L = \frac{E_L}{\Psi_{pd} - \Psi_L}. (3)$$

As light intensity was increased for shade leaves before they were detached for measurement of  $\Psi_L$ , the  $E_L$  measured at high light was used to calculate  $K_{\rm L}$  for all shade leaves.

Leaf level instantaneous transpiration efficiency (ITE) was calculated as  $A_n$  divided by  $E_L$ . The  $g_1$  parameter, describing plant water-use strategy, was estimated from ITE versus VPD response curves by fitting a rearranged optimal g<sub>s</sub> model for ITE (Medlyn et al. 2011) using non-linear regression (see Duursma et al. 2013). Small values of  $g_1$  indicate that transpiration is comparatively costly in C terms and reflecting conservative water use (increased water-use efficiency), whereas large values imply a lower C cost and decreased water-use efficiency (Lin et al. 2015).

#### Data analysis

Differences in responses of dependent variables to either the warming treatment or leaf type were analysed with linear mixed-effects models with WTC as a random effect, and leaf

type (sun, shade or shade in the sun) and warming treatment, and the interaction as fixed effects. The interaction was never significant, so we only report results of the tests of the main effects. Explained variance  $(R^2)$  of mixed models was computed as in Nakagawa & Schielzeth (2013), in which the marginal  $R^2$ represents variance explained by fixed factors and the conditional  $R^2$  by both fixed and random factors. Confidence intervals (95%) of linear mixed-effects models were generated using bootstrapping methods with 999 simulations, using the 'bootMer' function in the 'lme4' package (Bates et al. 2015). For non-linear relationships, confidence intervals were estimated by fitting a generalized additive model to the data with the 'mgcv' package (Wood 2006), using WTC as a random effect. All tests of statistical significance were conducted at an  $\alpha$ of 0.05. For multiple post-hoc comparisons, we used Tukey pairwise comparison tests with the 'multcomp' package (Hothorn et al. 2008). In all results, we report the mean  $\pm$  one standard error for the mean (SE). All analyses were performed with R 3.3.0 (R Development Core Team 2011).

### **RESULTS**

### Leaf resource distribution

Across six measurement campaigns over the 7 month period, PPFD was reduced on average by >75% in the shade (Fig. 1). Leaf  $N_a$  was approximately 20% higher in sun leaves compared

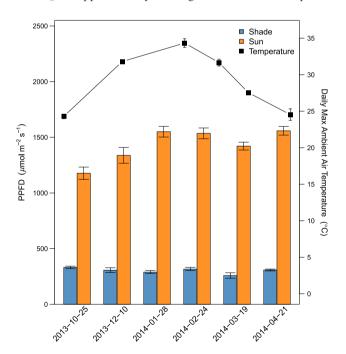


Figure 1. Environmental conditions during the six measurement campaigns. Bars represent the local light environment for sun and shade leaves during six gas exchange campaigns from October 2013 to April 2014 (as photosynthetic photon flux density, PPFD). Means  $\pm 1$ standard error represent PPFD measured with a ceptometer at the height of each selected leaf in the canopy. Points connected by a line represent the mean (±1 standard error) daily maximum air temperature during each campaign period. Each date represents the starting date for each measurement campaign; each campaign was completed in 3 days.

**Table 1.** Leaf morphological and physiological traits of sun and shade leaves under ambient and elevated temperature treatments for *Eucalyptus tereticornis*. Leaf mass per area (LMA, g m<sup>2</sup>), leaf nitrogen per unit area ( $N_a$ , gN m<sup>-2</sup>),  $\delta^{13}$ C (‰), pre-dawn leaf water potential ( $\Psi_{pd}$ , MPa), midday leaf water potential ( $\Psi_L$ , MPa) and leaf-specific hydraulic conductance ( $K_L$ , mmol m<sup>-2</sup> s<sup>-1</sup> MPa<sup>-1</sup>) are given as treatment means (±1 standard error) across six measurement campaigns. Values of  $A_{max}$  ( $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>) were measured once on all trees for each leaf type at 25 °C and at both saturating light and [CO<sub>2</sub>]. Different letters represent significant differences between leaf type and temperature treatments (based on a Tukey test). The P value represents the overall effect between each unique combination of leaf type and temperature treatment for each trait.

Leaf	Temperature	LMA	$A_{ m max}$	$N_{ m a}$	$K_1$	$\Psi_{\mathrm{pd}}$	$\Psi_{ m L}$	$\delta^{13}$ C
Sun	AT	114.1 (4.5) a	28.4 (2.2) b	2.63 (0.08) b	1.69 (0.18) b	-0.32 (0.04) a	-1.64 (0.11) a	-28.1 (0.18) b
	ET	109.9 (4.8) a	27.5 (1.2) b	2.60 (0.09) b	1.74 (0.15) b	-0.32(0.03) a	-1.74 (0.10) a	−28.3 (0.17) b
Shade	AT	118.3 (4.4) a	21.8 (0.9) a	2.13 (0.07) a	3.00 (0.21) a	-0.27 (0.02) a	-1.56 (0.09) a	-29.9 (0.17) a
	ET	113.1 (4.3) a	19.2 (0.9) a	1.88 (0.06) a	3.09 (0.21) a	-0.29 (0.02) a	-1.64 (0.11) a	-30.4 (0.22) a
P value		0.781	0.001	0.001	0.001	0.3486	0.6385	0.001

to shade leaves (Table 1). Leaf mass per area (LMA) was not different between leaf types (Table 1). Leaf-specific hydraulic conductance ( $K_L$ ) was significantly higher in shade leaves (+44%) compared to sun leaves (Table 1). This was because of differences in transpiration rates ( $E_L$ , Table 2) between leaf types at high light, as neither leaf water potentials ( $\Psi_{pd}$  and  $\Psi_L$ , Table 1) differed between leaf types. No effect of the warming treatment was detected on PPFD,  $\Psi_{pd}$ ,  $\Psi_L$ ,  $K_L$ ,  $E_L$ ,  $N_a$  or LMA either within or across leaf types (P > 0.05).

## Photosynthetic capacity and leaf photosynthesis rates

Mean photosynthetic capacity ( $A_{\rm max}$ ) of sun leaves was significantly higher (+27%) compared to shade leaves, when measured at 25 °C (Table 1). Within leaf types, no effect of the warming treatment was detected on  $A_{\rm max}$ .  $A_{\rm max}$  was positively related to mean leaf  $N_{\rm a}$  across leaf types and temperature treatments (P=0.01, Fig. 2).

Mean  $A_{\rm n}$  was significantly higher in sun compared to shade leaves (+23%), when measured at their local light environment and temperatures (Table 2). Following an increase in light intensity to match high-light conditions ('sunfleck simulations'),  $A_{\rm n}$  of shade leaves increased to values significantly greater than sun leaves at high light (P < 0.001, Table 2). No effect of the warming treatment was detected on  $A_{\rm n}$  of sun leaves measured at high light or shade leaves at either low or high light. Rates of photosynthesis within leaf types and warming treatments were similar through time and across the range of leaf temperatures measured (Fig. S3a).

### Stomatal conductance and leaf water-use efficiency

On average,  $g_s$  was 18% higher in shade compared to sun leaves under their local light environment (Table 2). Photosynthesis was positively correlated with  $g_s$  in all leaves measured under high light conditions; however,  $g_s$  and  $A_n$  were not correlated in shade leaves under low light (Fig. 3a). In high light,  $g_s$  of shade leaves increased compared to low light and was significantly greater than sun leaves, pooled across all measurement dates (Fig. 4a). No effect of the warming treatment was detected on  $g_s$  within or across leaf types. Stomatal conductances

within leaf types and treatments were similar through time and across the range of leaf temperatures measured (Fig. S4b).

Measured under ambient light and temperature, leaf instantaneous transpiration efficiency (ITE) was significantly greater in sun leaves than in shade leaves at low light (+21%, P = 0.001). Following an increase in PPFD to high-light conditions, ITE of shade leaves did not differ from shade leaves at low light and was therefore still significantly lower than sun leaves (P < 0.001). ITE in sun leaves was reduced in the warming treatment compared to ambient, but no warming effect was detected in shade leaves measured at low or high light (Table 2). The mean estimated  $g_1$  for sun leaves was  $1.51 \pm 0.11$ . Mean  $g_1$  was  $2.59 \pm 0.12$  for shade leaves measured at low light and  $2.74 \pm 0.04$  when measured at high light. For all leaf types and light treatments there was a strong response of ITE to VPD and individual data points broadly corresponded to the fitted response curves from the optimal ITE model with a specified  $g_1$  value for each leaf type and treatment (Fig. 5a). Within leaf types and light treatments, the response of VPD to leaf temperature was similar across all measurement campaigns (Fig. S4a).

Bulk-leaf  $\delta^{13}$ C was significantly lower in shade leaves compared to sun leaves by ca. 2‰ (Table 1). No effects of the warming treatment on leaf  $\delta^{13}$ C were detected. Leaf  $\delta^{13}$ C and  $N_a$  were positively correlated for all leaves (P < 0.001, Fig. 5b), with less negative  $\delta^{13}$ C and higher N investment in sun leaves than shade leaves.

### Leaf carbon isotope discrimination and mesophyll conductance

The observed carbon isotope discrimination ( $\Delta_o$ ) measured during photosynthesis was positively correlated with  $C_i/C_a$  for both leaf types (P < 0.001), with larger  $\Delta_o$  detected for sun leaves and shade leaves at high light than shade leaves at low light (Fig. S2). Carbon isotope discrimination associated with  $g_{\rm m}$  accounted for the majority of  $\Delta_o$  (69.7 ± 0.3%) and varied little across measurement temperatures, leaf types, or warming treatments. The remainder of the variation in  $\Delta_o$  consists of the contributions of  $g_{\rm s}$ , respiration and photorespiration to discrimination.

Mean  $g_{\rm m}$  was higher in sun compared to shade leaves (+25%) under their local light environment (P < 0.001).

Leaf gas exchange parameters for sun and shade leaves under ambient and elevated temperature treatments for Eucalyptus tereticornis. Leaf net photosynthesis rate (An., µmol m<sup>-2</sup> s<sup>-1</sup>), leaf  $^2$ s $^{-1}$ bar $^{-1}$ ), intercellular CO $_2$  concentration (C $_1$ ,  $\mu$ bar), chloroplastic CO $_2$  concentration (C $_2$ ,  $\mu$ bar) and leaf-to-(±1 standard error) for each treatment across six gas exchange campaigns. Different letters represent significant differences between leaf type, light environment and temperature treatments (based on a Tukey test). The P value represents the overall effect between each unique combination of leaf type, light environment and temperature  $[s^{-1}), g_{\rm m} \, ({\rm mol \, m}^{-1})$  $^{2}$  s<sup>-1</sup>), stomatal conductance (g<sub>s</sub>, mol m<sup>-1</sup> air vapour pressure deficit (VPD, kPa) are given as the mean ( transpiration rate  $(E_{\rm L}, {\rm mmol \ m}^{-})$ Table 2.

treatment	treatment tor each parameter.	ameter.								
Leaf	Light	Light Temperature	$A_{ m n}$	Ss	8m	ITE	$E_{ m L}$	VPD	$C_i$	$C_{c}$
Sun	High	AT	13.5 (0.3) b	0.122 (0.005) a	0.195 (0.006) ac	8.26 (0.48) b	1.78 (0.07) a	1.60 (0.04) ab	179.8 (3.2) a	106.4 (2.8) a
		ET	13.1 (0.3) b	0.123 (0.005) a	0.182 (0.008) ac	6.57 (0.39) ab	2.21 (0.09) a	1.90(0.05) b	187.9 (2.9) a	108.4 (2.6) a
Shade	Low	AT	10.4 (0.1) a		0.162 (0.006) ab	6.24 (0.50) a	1.93 (0.07) a	1.40 (0.04) a	255.4 (3.8) b	184.3 (4.2) cd
		ET	10.0 (0.1) a		0.161 (0.007) a	5.43 (0.51) a	2.23 (0.09) a	1.60 (0.05) a	253.8 (4.1) b	185.2 (3.8) d
	High	AT	18.1 (0.3) c	0.255 (0.007) b	0.212 (0.004) c	5.85 (0.33) a	3.42 (0.12) b	1.40 (0.04) a	237.4 (2.2) b	151.3 (2.0) b
	ı	ET	16.7(0.2) c	0.246 (0.009) b	0.208 (0.004) bc	5.02 (0.35) a	3.81 (0.15) b	1.70 (0.04) ab	238.1 (3.2) b	156.4 (2.9) bc
P value			0.001	0.001	0.001	0.001	0.001	0.005	0.001	0.001

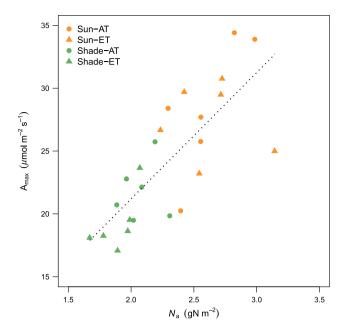
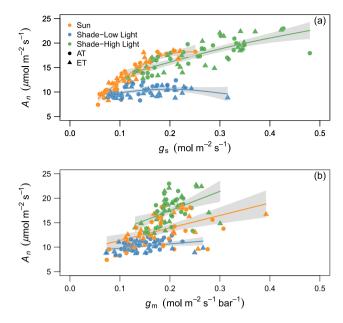


Figure 2. Photosynthetic capacity  $(A_{\text{max}})$  was positively correlated with mean leaf N per area  $(N_a)$  for sun and shade leaves grown under elevated (ET) and ambient (AT) temperature treatments. Leaf  $N_a$  is the mean of each leaf type, in well-watered treatments, sampled across six gas exchange campaigns in each whole-tree chamber.  $A_{\rm max}$  was measured once on all trees for each leaf type, before the initiation of the drought treatment, at 25 °C and at both saturating light and [CO<sub>2</sub>]. The dashed line represents the significant linear model fit for all leaves, with a marginal and conditional  $R^2$  of 0.59 and 0.62.

Following the increase in PPFD from low to high light on the same leaf,  $g_{\rm m}$  values of shade leaves increased on average 37% after approximately 25 min (Fig. 6). These measured values of g<sub>m</sub> for shade leaves at high light were also indistinguishable than those of sun leaves (Table 2). Proportional increases in  $g_m$  were matched by proportional increases in  $A_n$ from low to high light in shade leaves (Fig. 4b,c). Photosynthesis scaled positively with increases in  $g_{\rm m}$  for all leaves, with similar intercepts but different slopes between leaf type and light treatment (P = 0.019). The large increases in  $g_{\rm m}$  in shade leaves under high light likely resulted in the highest rates of  $A_n$ (Fig. 3b). No differences in  $g_{\rm m}$  were detected with the warming treatment within leaf types. Mesophyll conductance did not vary across measurements campaigns within leaf types and light treatments (Fig. S3b), but a weak negative relationship with increasing leaf temperature was detected with sun and shade leaves under their local light environment (P = 0.002for sun leaves, P = 0.044 for shade leaves).

### Variation in intercellular and chloroplastic CO<sub>2</sub> concentrations

Higher  $g_s$  in shade leaves under low and high light led to significant increases in  $C_i$  compared to sun leaves (Fig. 7a). The chloroplast CO<sub>2</sub> partial pressure was comparable between shade leaves when measured at both low and high light conditions (Fig. 7c). In sun leaves  $C_c$  was significantly lower than shade

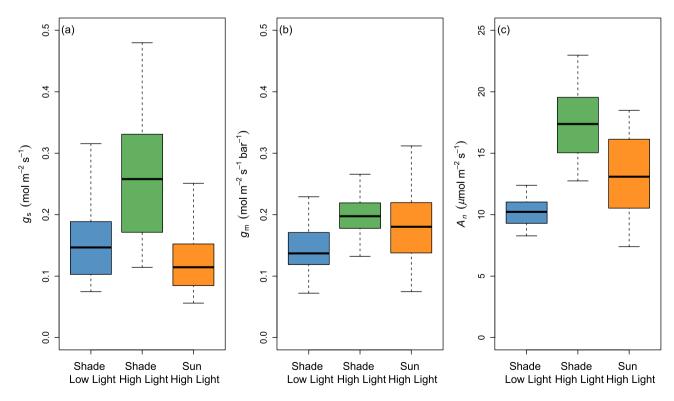


**Figure 3.** Relationships between leaf net photosynthesis rate  $(A_n)$ , stomatal conductance  $(g_s, panel (a))$  and mesophyll conductance  $(g_m, panel (b))$  for sun leaves and shade leaves in low or high light. Sun leaves were measured at high light, shade leaves were measured at both low and high light. Data are combined across elevated (ET, triangles) and ambient (ET, circles) temperature treatments, and six measurement campaigns. Lines represent either smoothed regressions from a generalized additive model fit (a) or linear model fits (b), fitted across all data but accounting for random effects (see Methods). Grey areas are 95% confidence intervals for the mean.

leaves, consistent with a lower  $C_i$ . The drawdown of  $\mathrm{CO}_2$  from intercellular spaces to the chloroplast,  $C_i - C_c$ , measures the coordination between  $g_{\mathrm{m}}$  and  $A_{\mathrm{n}}$  (von Caemmerer & Evans 2014). This drawdown was similar between sun and shade leaves measured at their local light and temperature environment and increased marginally in shade leaves at high light (Fig. 7c). This result demonstrates the approximately proportional relationship between  $g_{\mathrm{m}}$  and  $A_{\mathrm{n}}$  across all leaves. The  $\mathrm{CO}_2$  drawdown from  $C_{\mathrm{a}}$  to  $C_{\mathrm{i}}$  and  $C_{\mathrm{i}}$  to  $C_{\mathrm{c}}$  were both relatively stable across the range of temperatures measured and gas exchange campaigns (Fig. S4c and S3c, respectively).

### DISCUSSION

Here we show that  $A_n$  in leaves within canopies of E. tereticornis is limited by the local light environment; however, shade leaves increased rates of leaf C gain exceeding sun leaves when light availability increased. Although shade leaves in lower light environments exhibited relatively high  $g_s$  and  $C_i$ , the increases in  $g_m$  under periods of high light availability enabled the up-regulation of  $A_n$ . Although it is well-known that shade leaves experience transient periods of sun and shade (Pearcy 1990), a lack of empirical data within tree canopies currently impedes our ability to predict whole canopy C gain. These findings offer new insights into how aspects of leaf gas exchange physiology may be optimized differently in sun and shade leaves and reveal how the total leaf  $CO_2$  conductance pathway should be accounted for when testing optimizations of canopy C uptake and water



**Figure 4.** Box plots of stomatal conductance  $(g_s)$  (a), mesophyll conductance  $(g_m)$  (b) and leaf net photosynthesis rate  $(A_n)$  (c) of sun leaves and shade leaves at low and high light pooled across six measurement dates and both temperature treatments. For statistical comparisons, see Table 2.

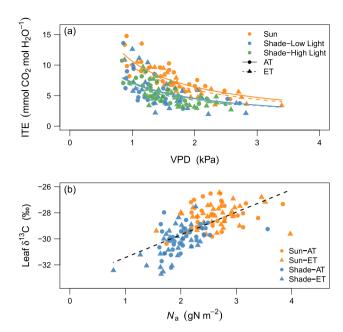
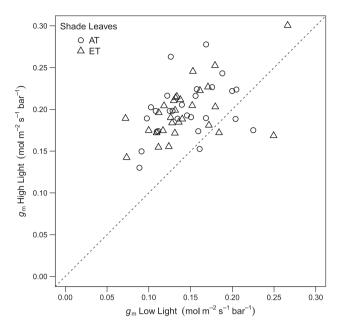


Figure 5. Two measures of water-use efficiency related to vapour pressure deficit and leaf nitrogen per area  $(N_a)$ . (a) Instantaneous transpiration efficiency (ITE) declines with VPD, and is lower for shade leaves (either when measured in high or low light). VPD is the leaf to air pressure difference inside the gas exchange cuvette and lines represent predictions from the optimal ITE model with a  $g_1$  value for each leaf type and temperature treatment (AT, ambient temperature; ET, elevated temperature). (b) The relationship between leaf  $\delta^{13}$ C (higher values indicate higher water-use efficiency) and leaf  $N_a$  for sun leaves at high light and shade leaves at low light. The dashed line represents the significant linear model fit across all leaves with a marginal and conditional  $R^2$  of 0.41 and 0.45, respectively.

use. Additionally, with measurements recorded across a large natural range of air temperatures only minimal effects of a +3 °C warming treatment were detected on leaf physiology, consistent with temperature acclimation demonstrated in this experiment (Aspinwall et al. 2016).

### Resource distribution and photosynthetic capacity

The allocation of  $N_a$  along canopy gradients constrains  $A_n$  and is thus a key trait in determining the relative contribution of individual leaves to canopy C gain. Decreasing light availability should decrease the investment of N into photosynthetic enzymes within a canopy (Mooney & Gulmon 1979). As a result, acclimation of photosynthetic capacity to irradiance is typically reflected in the key photosynthetic biochemical parameters  $V_{\rm cmax}$  and  $J_{\rm max}$  (Farquhar et al. 1980). Our data agree with these conventional conclusions as the distribution of  $N_a$ ,  $A_{max}$ and  $A_n$  were all reduced in shade leaves. Leaf mass per area (LMA), however, was not different between sun and shade leaves, consistent with small differences reported in a clonal Eucalyptus plantation (Nouvellon et al. 2010), but in contrast to data for many other species (Niinemets et al. 2015). Similar LMA in sun and shade leaves could be the result of initial leaf formation and expansion under comparable light conditions



**Figure 6.** Mesophyll conductance  $(g_m)$  measured on shade leaves in low light, and in high light (on the same leaves). After measurement in the shade, the photosynthetic photon flux density (PPFD) was increased to match the light environment of full sun leaves in the same tree. All six measurement dates are included, and symbols reflect the temperature treatment (AT, ambient temperature; ET, elevated temperature). Measurements of gm were recorded after CO2 and water vapour fluxes were stable in the leaf cuvette, which took approximately 25 min after light intensity was increased. The dashed line is the 1:1 relationship.

and subsequent shade arising during canopy development in this evergreen species. Unmeasured differences in total nonstructural carbohydrate contents between leaf types may also have masked differences in leaf thickness or density. For example, sun leaves of Abies amabilis foliage that became shaded exhibited physiology similar to shaded leaves, while retaining the morphology of sun leaves (Brooks et al. 1996).

Photosynthesis is also limited by the supply of water to the upper canopy. Ultimately, the ability of tree hydraulic architecture to supply water to foliage across increasing pathlengths affects productivity and survival (Sellin et al. 2008). Using a two-leaf model, Peltoniemi et al. (2012) theorized that optimal N distribution will be proportional to light distribution only if  $K_{\rm L}$  is also optimally distributed. We found higher  $K_{\rm L}$  in shade leaves compared to sun leaves, contrary to the prediction by Peltoniemi et al. (2012), but consistent with higher  $g_s$  attained by shade leaves in high light conditions. This finding is consistent with the idea that  $g_s$  is regulated, at least in part, by  $K_{\rm L}$  (Hubbard *et al.* 2001).

### Physiological behaviour of sun and shade leaves

Unexpected higher  $g_s$  in shade leaves compared to sun leaves led to decreased ITE in shade leaves throughout the experiment. Consistently lower leaf  $\delta^{13}$ C in shade leaves also suggests that observed higher  $C_i$  and  $C_c$  in shade leaves was prevalent long term (Figure S2) (Marshall et al. 2007), despite the

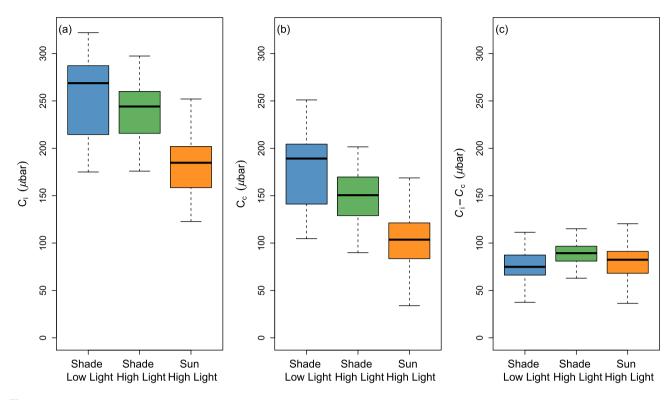


Figure 7. Boxplots of (a) intercellular  $CO_2$  concentration ( $C_i$ ), (b)  $CO_2$  concentration in the chloroplasts ( $C_c$ ) and (c)  $CO_2$  drawdown from substomatal cavities to sites of carboxylation of sun leaves and shade leaves at both low and high light ( $C_i - C_c$ ). All data across six measurement dates and both temperature treatments are included. For statistical comparisons, see Table 2.

observation that  $g_m$  accounted for nearly 70% of the variation in  $\Delta_0$ . Higher  $g_s$  in shade leaves relative to the differences in  $A_n$ , appears to result in comparatively inefficient water use as more water is used per unit photosynthesis. As whole canopy C gain integrates the efficiency of all leaves, this raises the question of why shade leaves exhibit a lower water-use efficiency compared to sun leaves.

In sun leaves,  $A_n$  and  $g_s$  were strongly correlated, exhibiting behaviour broadly consistent with optimal stomatal theory (Fig. 5). However, lower rates of  $A_n$  in shade leaves were not coupled with decreases in  $g_s$ , leading to the observed decreases in ITE. This is significant as optimal stomatal regulation to balance C gain with water loss has been reported across a wide range of ecosystems and plant functional types; however, empirical data is often collected only on sun leaves (e.g. Prentice et al. 2014; Lin et al. 2015). For the entire canopy to exhibit optimal regulation of stomatal conductance, that is, to maximize photosynthesis minus transpiration cost (Cowan & Farquhar 1977), sun and shade leaves should have equal ITE – provided environmental drivers are similar (Buckley et al. 2014). In our study, higher  $g_s$  in shade leaves may in part be explained by higher  $K_{I,s}$ which effectively reduces the cost of water transport, allowing more profligate water use in shade leaves. Consistent with our results, Buckley et al. (2014) found that shade leaves in a grapevine canopy exhibited higher  $g_s$  than expected based on their environment and comparison to sun leaves. Further work is needed to confirm the generality of this observation.

It is also possible that higher  $g_s$  in shade leaves is a strategy to take advantage transient periods of high light quickly in shade

leaves, avoiding the delay associated with stomatal opening (Tausz et al. 2005; Vico et al. 2011). Our results support this hypothesis, as shade leaves increased  $A_n$  to values equal or even outperforming sun leaves when subjected to a brief period of identical high-light intensity. Transpiration-induced cooling in shade leaves, by keeping stomata open, has also been suggested as an effective strategy to reduce sunfleck-induced rapid increase in leaf temperature (Schymanski et al. 2013), which has been shown to inhibit C gain (Leakey et al. 2003). However, this response likely occurs at very high air temperatures and may not explain the observed g<sub>s</sub> in shade leaves across the large natural seasonal range of temperatures included in our study. How prevalent each of these strategies are within tree canopies is still unknown, as empirical studies assessing photosynthetic responses to sunflecks generally focus on seedlings (Küppers & Schneider 1993; Pepin & Livingston 1997; Leakey et al. 2002) and understory plants, often in deep shade (Chazdon & Pearcy 1991; Allen & Pearcy 2000; Brantley & Young 2009), although leaves within a forest canopy exhibited a similar uncoupling of  $A_n$  and  $g_s$  across a light gradient (Tjoelker et al. 1995). Thus, our findings highlight a critical need for empirical measurements of shade leaves under dynamic light environments in order to accurately scale C gain from leaf to canopy (see De Pury & Farquhar 1997).

We found that  $A_n$  and  $g_m$  scaled positively across leaf types (Fig. 3b) and, unexpectedly, increased in shade leaves when light intensity was increased to values similar in sun leaves. Comparable LMA within the canopy likely allowed for this up-regulation of  $g_m$ , which played a critical role in the

photosynthetic response of shade leaves to sunflecks. Components of LMA, such as palisade thickness and cell surface exposed to intercellular air space, set an upper limit for  $g_m$ , but considerable variation in  $g_m$  can exist for a given LMA (Flexas et al. 2008; Milla-Moreno et al. 2016). Possible mechanisms for the rapid ( $\sim$ 25 min) up-regulation of  $g_{\rm m}$  to light intensity include regulation of aquaporins, which can facilitate increases in the CO<sub>2</sub> permeability of the cell membranes (Hanba et al. 2004; Heinen et al. 2009; Li et al. 2014) or movement of chloroplasts to facilitate CO<sub>2</sub> diffusion (Tholen et al. 2008). It has also been proposed that  $g_{\rm m}$  and  $K_{\rm L}$  should be coordinated, as optimizing hydraulic supply allows both  $g_s$  and  $g_m$  to enhance  $A_n$ (Flexas et al. 2013). Here, higher  $K_L$  in shade leaves under high light lends support to the potential coordination  $K_L$  and  $g_m$  and also to the association of higher  $A_n$  with higher  $K_L$  when light becomes saturating, as reported by Franks (2006).

If shade leaves 'lie in wait' for sunflecks, then perhaps we should consider an alternate leaf economic strategy to maximize C gain, beyond conventional trade-offs associated with canopy resource distribution. This is because the role of  $g_s$  in regulating photosynthetic induction impacts the capacity of a leaf to utilize sunflecks (Way & Pearcy 2012). If the valuation of sunflecks as a C resource is large enough, then costs of sub-optimal stomatal behaviour could be offset over the leaf lifespan or across the entire canopy when considering both sun and shade leaf types (Vico et al. 2011). For example, the potential C gain in leaves where sunflecks constitute a large proportion of total daily PPFD may be large enough to accommodate the cost of increased water use in the shade. However, accounting for the heterogeneous nature of light within a canopy remains a current challenge for empirical and modelling studies. Thus, models which predict leaf photosynthesis from N distribution within a canopy will be incomplete unless inclusion of canopy light extinction and the integration of sunflecks on shade leaves are included (De Pury & Farquhar 1997).

### **CONCLUSIONS**

Here we show that dynamic physiological responses of shade leaves to altered light environments have important implications for up-scaling leaf level measurements to the canopy. Although resource allocation constrains leaf photosynthetic capacity, it is the physiological behaviour of individual leaves that determines C gain. These findings suggest that current theories of leaf optimal behaviour should be extended to include dynamic light environments, which will have implications for process-based models that predict canopy C gain from rates of leaf photosynthesis. Furthermore, the dynamic nature of  $g_{\rm m}$  cannot be simply parameterized in vegetation models, and further experimental evidence is needed so that it can be represented properly. Additional empirical data, across multiple tree species, are needed to determine both the mechanisms and the capacity of  $g_{\rm m}$  to respond to environmental drivers.

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### SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article at the publisher's web-site:

- Figure S1. Daily maximum and minimum temperature (a), daily maximum VPD (b) and total daily PPFD (c) for each chamber across the experiment duration.
- Figure S2. Relationship between the observed discrimination of  $^{13}\text{CO}_2$  measured during photosynthesis ( $\Delta_o$ ) and measured  $C_i/C_a$  for sun leaves measured at high light and shade leaves measured at both low and high light. The solid line represents the theoretical line for C3 plants from Evans et al. (1986).
- **Figure S3.** Response of  $A_n$  (a),  $g_m$  (b) and  $C_i C_c$  to leaf temperature for sun leaves and shade leaves at low and high light. Shaded symbols represent each monthly measurement campaign. Solid lines, coloured by leaf and light type, are fitted line for the relationship with each parameter and leaf temperature across all measurement campaigns. All parameters with no relationship are fitted with zero slope and the overall mean value for each treatment combination. Weak negative relationships with  $g_{\rm m}$  and increasing leaf temperature were detected with sun and shade leaves under their local light environment  $(R^2 = 0.16 \text{ and } 0.08, \text{ respectively}).$
- **Figure S4.** Response of VPD (a),  $g_s$  (b) and  $C_a C_i$  to leaf temperature for sun leaves and shade leaves at low and high light. Shaded symbols represent each monthly measurement campaign. Solid lines, coloured by leaf and light type, are fitted line for the relationship with each parameter and leaf temperature across all measurement campaigns. All parameters with no relationship are fitted with zero slope and the overall mean value for each treatment combination. Leaf VPD inside the gas exchange cuvette was positively correlated with increasing leaf temperature for sun leaves and shade leaves at low and high light ( $R^2 = 0.73$ , 0.58 and 0.72, respectively).

**Methods S1.** Description of the calculation of  $g_{\rm m}$  from carbon isotope discrimination during C3 photosynthesis