Tools for Polyploids Workshop Computational Support

SNP and Dosage Calling

Cristiane Taniguti
Gabriel Gesteira
Jeekin Lau
Maria Caraza-Harter



Getting Prepared for the Workshop



- Polyploids
- Molecular Markers
- Genome variations applications
 - Quantitative traits mapping
 - Genome Wide Association studies
 - Phenotypic predictions Genome Selection
 - Evolution and diversity studies
 - Gene expression studies



Genome variations

- Short sequences (SNPs, indels)
- Structural variants (number of copies, inversions, translocations)

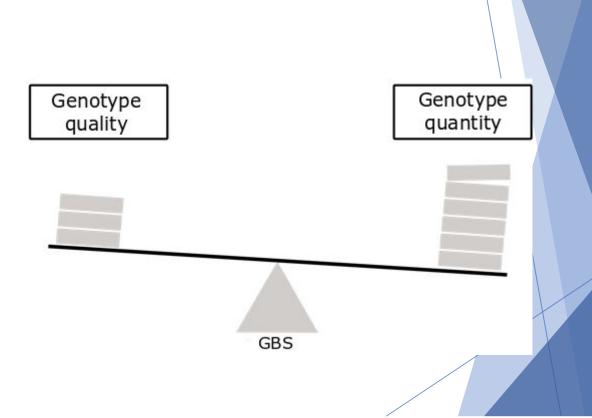
Molecular markers

- RFLP, RAPD, AFLP, and SSR
- Arrays (For Roses: \$\$\$\$\$)
- Sequencing (For Roses: \$)



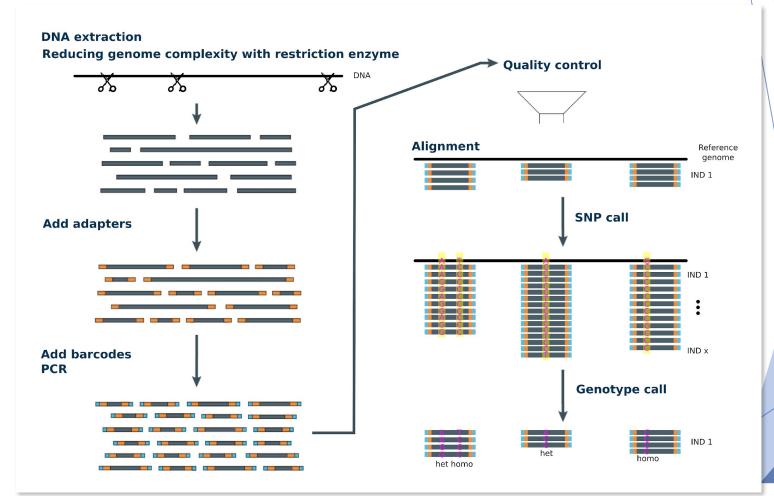
Sequencing Experiment Design

- Study goal
- Sequencer capacity
- Number of individuals per lane
- Number of sequenced loci





GBS Overview

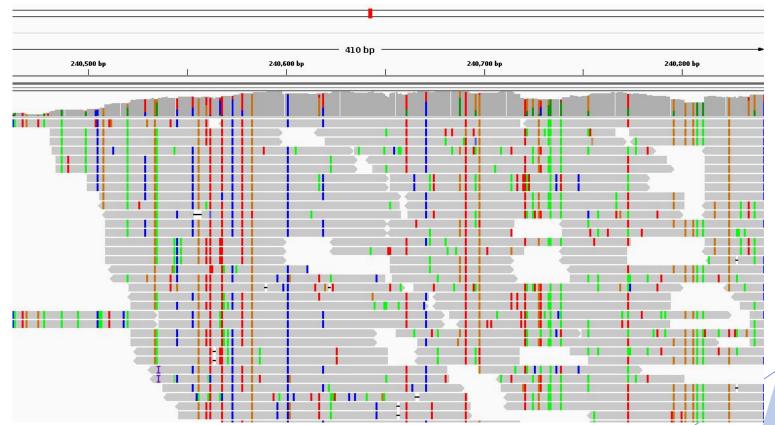




SNP Calling

Whole Genome Sequencing (WGS)

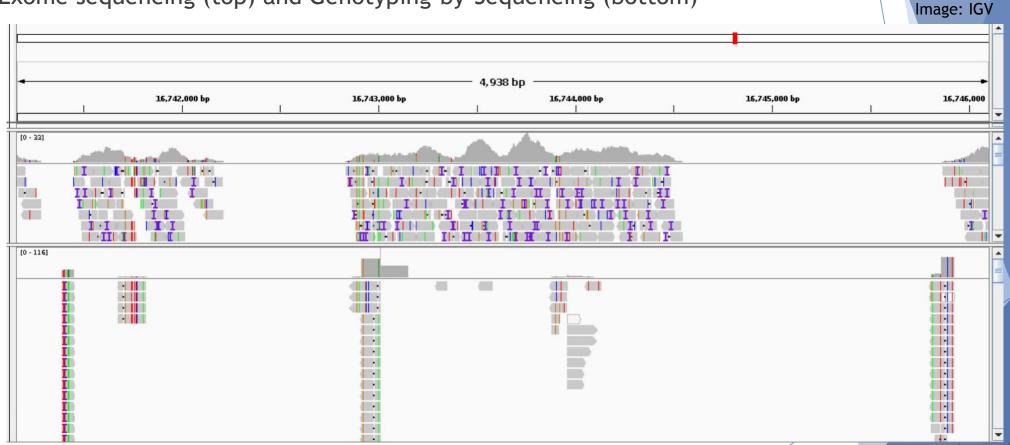






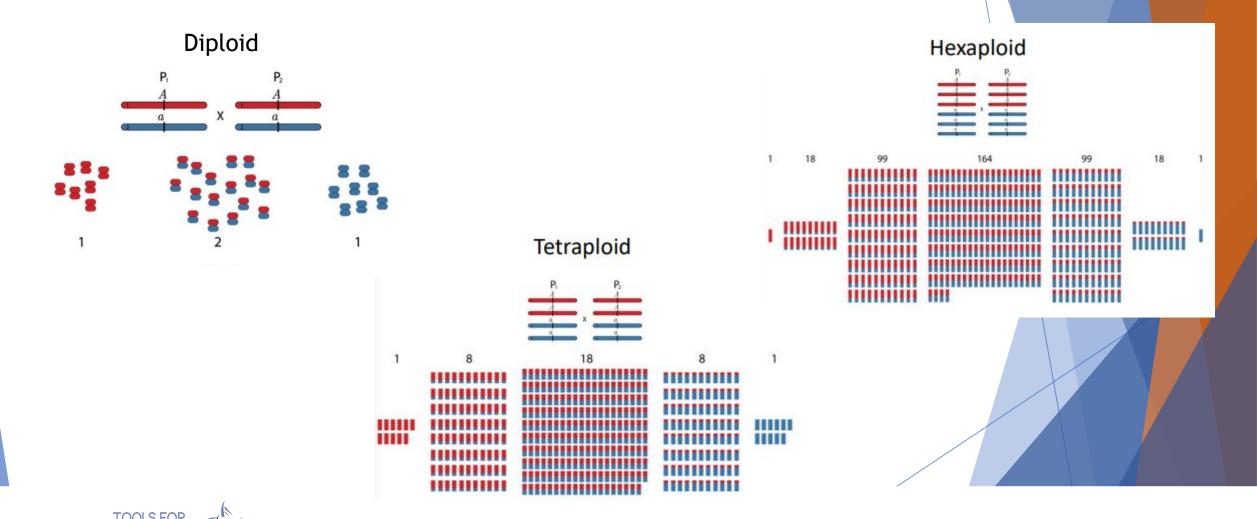
SNP Calling

Exome sequencing (top) and Genotyping-by-Sequencing (bottom)





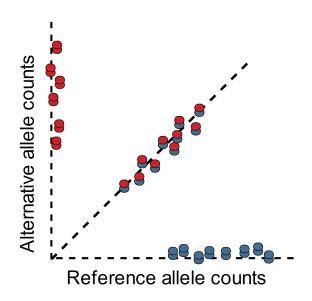
Dosage calling



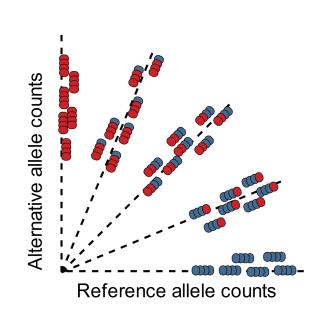
Dosage Calling

The theory

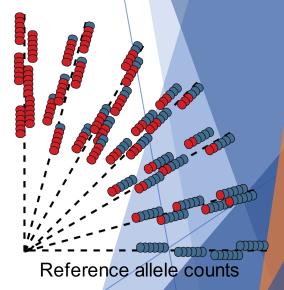
Diploid



Tetraploid



Hexaploid

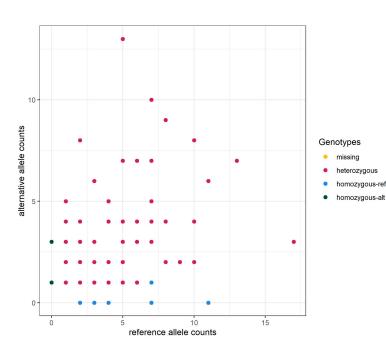




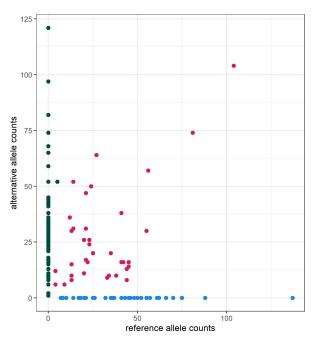
Dosage Calling

The reality

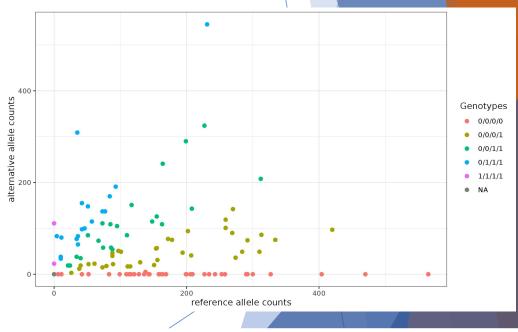
Diploid (mean depth 6) N = 200 Aa x Aa



Diploid (mean depth 96) N = 138 Aa x Aa



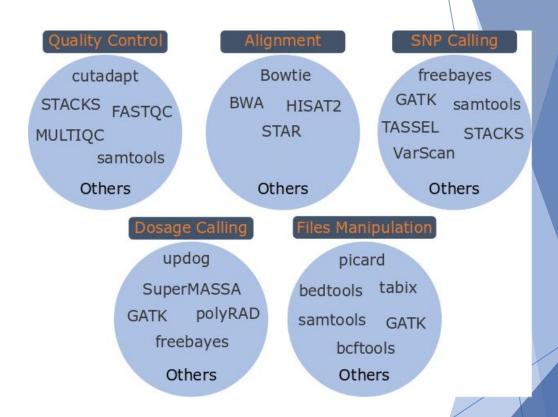
Tetraploid (mean depth 83) N = 114 AAaa x AAaa





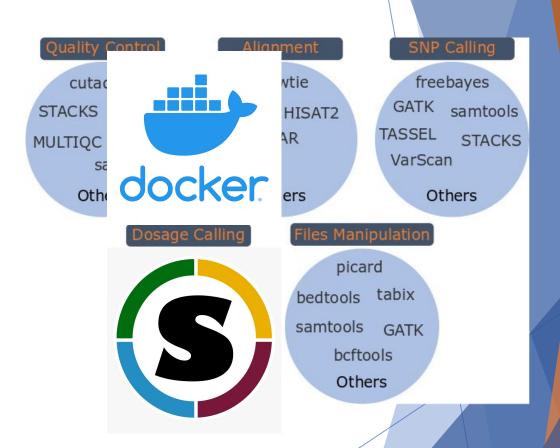
Large files

- Many software
- Many programming languages
- Different Operational Systems
- Updates



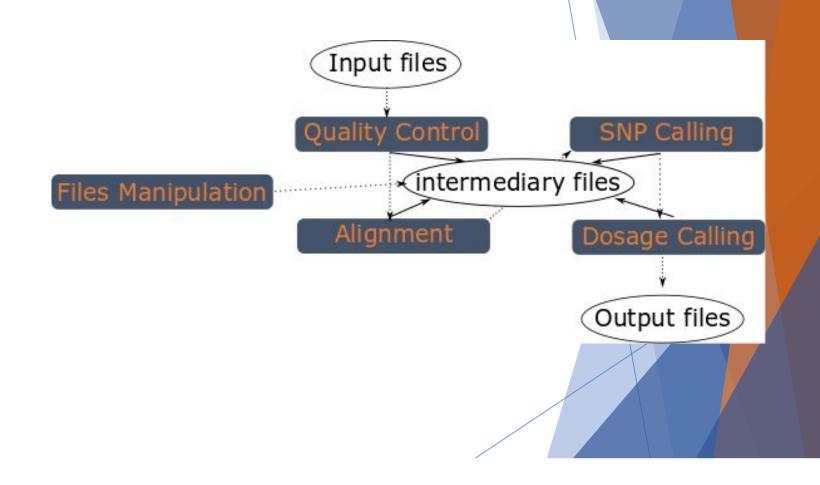


- Large files
 - High Performance Computing (HPC)
 - Management systems (SLURM, SGE)
 - Cloud (Google, Amazon)
- Many software
- Many programming languages
- Different Operational Systems
- Updates
 - Containers
 - Docker
 - Singularity (usually available in HPC)
 - ► <u>BioContainers</u>



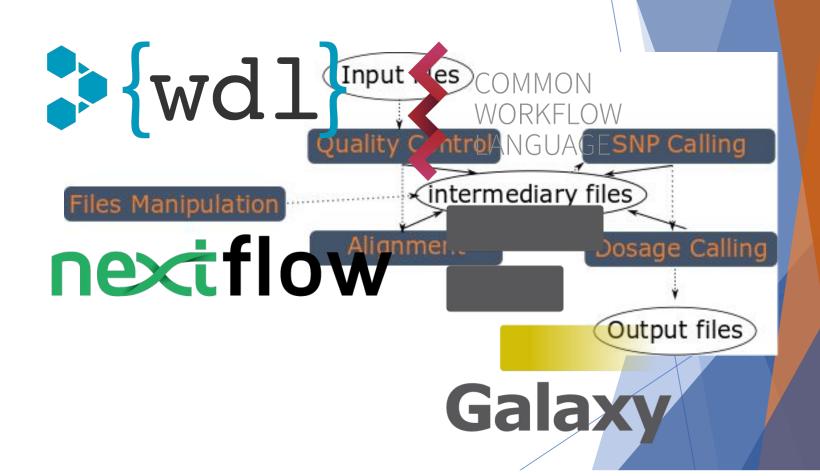


- Many steps
- Many file formats





- Many steps
- Many file formats
 - Workflows systems
 - Galaxy
 - Nextflow
 - Snakemake
 - CWL
 - WDL
 - Workflows repositories
 - Dockerstore
 - WorkflowHub
 - Run workflows on Cloud
 - Galaxy
 - DNAnexus
 - Terra
 - AnVIL
 - SevenBridges

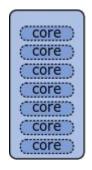




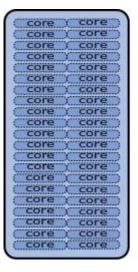
- Resources optimization
 - Time
 - Cores
 - Nodes
 - RAM memory

Personal Computer:

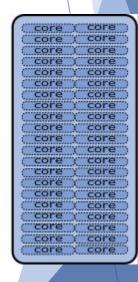
4GB RAM; 8 cores; 1 node



High Performance Computing (Texas A&M): 384GB; 48 cores per node; 900 nodes

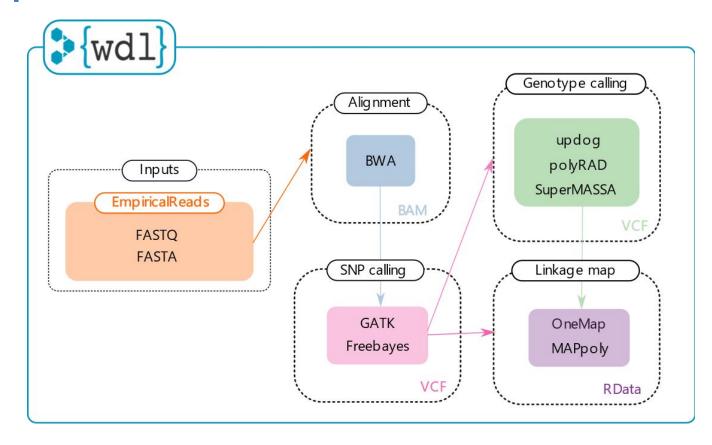


	_
/ COFO	core
Core /	
core (core
STREET, STREET	core)
(core)	core
(core)	core)
(core)	core ;
(core)	core ;
core"	core
core	core
core	core
Core Y	core
core Y	core
core	core
core	COPO
core	core
	core
core i	core;
core ;	core





Reads2Map



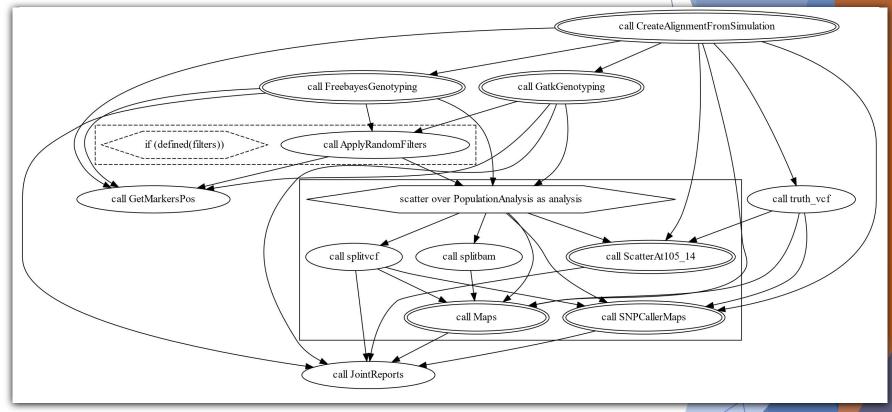
Available in Github, Dockerstore and WorkflowHub



Implementation

Tasks Sub-workflows Parallelized loop Conditionals

- Workflows
 - Sub-workflows
 - Tasks



\$ java -jar /path/to/womtool.jar graph tasks/SimulatedSingleFamily.wdl > SimulatedSingleFamily.dot \$ dot -Tsvg SimulatedSingleFamily.dot -o SimulatedSingleFamily.svg



Implementation

- Cloud environments
 - ► <u>terra.bio</u>
- High Performance Computing (HPC)
 - Cromwell
 - MiniWDL
 - ► <u>dxWDL</u>

\$ java -jar /path/to/cromwell.jar run -i inputs/EmpiricalSNPCalling.inputs.json EmpiricalSNPCalling.wdl



Tutorials

- polyRAD tutorial
- updog tutorial
- ► <u>fitPoly tutorial</u>
- ► (TASSEL) Variant and Genotype Calling in Highly Duplicated Genomes (Lindsay Clark)
- Step-by-step of SNP and dosage calling using containers and WDL workflows



References

- Catchen, J., Hohenlohe, P. A., Bassham, S., Amores, A.; Cresko, W. A. (2013). Stacks: an analysis tool set for population genomics. Molecular Ecology, 22(11), 3124-3140. https://doi.org/10.1111/mec.12354
- Glaubitz, J. C., Casstevens, T. M., Lu, F., Harriman, J., Elshire, R. J., Sun, Q.; Buckler, E. S. (2014). TASSEL-GBS: a high capacity genotyping by sequencing analysis pipeline. PLoS ONE, 9(2), 1-11. https://doi.org/10.1371/journal.pone.0090346
- Garrison, E.; Marth, G. (2012). Haplotype-based variant detection from short-read sequencing. ArXiv E-Prints, 9. https://doi.org/1207.3907
- McKenna, A., Hanna, M., Banks, E., Sivachenko, A., Cibulskis, K., Kernytsky, A., Garimella, K., Altshuler, D., Gabriel, S., Daly, M.; DePristo, M. A. (2010). The Genome Analysis Toolkit: A MapReduce framework for analyzing next-generation DNA sequencing data. Genome Research, 20(9), 1297-1303. https://doi.org/10.1101/gr.107524.110



References

- Gerard, D., Ferrão, L. F. V., Garcia, A. A. F., & Stephens, M. (2018). Genotyping Polyploids from Messy Sequencing Data. Genetics, 210(3), 789-807. doi: 10.1534/genetics.118.301468.
- Wadl, P. A., Olukolu, B. A., Branham, S. E., Jarret, R. L., Yencho, G. C.; Jackson, D. M. (2018). Genetic Diversity and Population Structure of the USDA Sweetpotato (Ipomoea batatas) Germplasm Collections Using GBSpoly. Frontiers in Plant Science, 9, 1166. https://doi.org/10.3389/fpls.2018.01166
- Serang, O., Mollinari, M.; Garcia, A. A. F. (2012). Efficient exact maximum a posteriori computation for bayesian SNP genotyping in polyploids. PLoS ONE, 7(2), 1-13. https://doi.org/10.1371/journal.pone.0030906
- Clark, L. v., Lipka, A. E.; Sacks, E. J. (2019). polyRAD: Genotype Calling with Uncertainty from Sequencing Data in Polyploids and Diploids. G3: Genes|Genomes|Genetics, 9(March), g3.200913.2018. https://doi.org/10.1534/g3.118.200913



Project Members



























Other Collaborators











Woolf Roses

L.L.C.



















BAILEY

Royal Van Zanten











CIAT



