# Package 'GWASpoly'

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```
Title Genome-wide Association Studies for Autopolyploids

Version 2.10

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Description Designed for genome-wide association studies in autopolyploids.

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Imports rrBLUP, methods, ggplot2, tidyr, stats, parallel, rlang, scam, Matrix

Suggests knitr, rmarkdown

VignetteBuilder knitr

Collate 'GWASpoly.data.R'
    'GWASpoly.K.R'
    'GWASpoly.R'
```

'GWASpoly.thresh.R' 'Keff.R' 'LD.plot.R' 'VCF2dosage.R' 'design.score.R' 'fit.QTL.R' 'get.QTL.R' 'get\_x.R' 'make.full.R' 'makeLOCO.R' 'manhattan.plot.R' 'qq.plot.R' 'qvalue.R' 'read.GWASpoly.R' 'score.calc.R' 'set.K.R' 'set.params.R' 'set.threshold.R'

'write.GWASpoly.R'

'GWASpoly.fitted.R'

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# **R** topics documented:

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# Description

Test markers as QTL under backward elimination

## Usage

```
fit.QTL(data, trait, qtl, fixed = NULL)
```

#### **Arguments**

data	variable inheriting from class GWASpoly.K
trait	name of trait
qtl	data frame to specify the multi-QTL model (see Details)
fixed	data frame to specify the fixed effects (see Details)

## **Details**

qt1 is a data frame with columns "Marker" and "Model", where each row corresponds to a QTL. fixed is a data frame with columns "Effect" and "Type": the first column is the name of the effect, which must match a column in the phenotype input file, and the second column is either "factor" or "numeric". The p-value and R2 for each marker are based on the likelihood ratio test under backward elimination, comparing the deviance to the chi-squared distribution.

### Value

data frame with partial r2 and p-values

get.QTL 3

get.QTL	Extract significant QTL

## Description

Output a table with significant markers

#### Usage

```
get.QTL(data, traits = NULL, models = NULL, bp.window = NULL)
```

## Arguments

data	Output from set.threshold
traits	Vector of trait names (by default, all traits)
models	Vector of model names (by default, all models)
bp.window	prune output to return only the most significant marker within this window size

#### **Details**

To return all significant markers (original behavior of the function), use bp.window=NULL. Assumes input map position in bp.

#### Value

Data frame with results. Score =  $-\log 10(p)$ . Effect = marker effect (not available for the general and diplo-general models because there are multiple effects).

GWASpoly Compute marker significance scores	GWASpoly	Compute marker significance scores	
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## Description

Compute marker significance scores

## Usage

```
GWASpoly(data, models, traits = NULL, params = NULL, n.core = 1, quiet = F)
```

## Arguments

(	data	Output from set.K
ı	models	Vector of model names
	traits	Vector trait names (by default, all traits)
	params	Optional list of params created by set.params
ı	n.core	Number of cores for parallel computing
	quiet	TRUE/FALSE whether to suppress output charting progress

#### **Details**

The following marker-effect models are available:

- "additive": Indicates the marker effect is proportional to the dosage of the alternate allele
- "X-dom": where X can be any integer between 1 and ploidy/2 and refers to the allele dosage needed for complete dominance (e.g., "1-dom" = simplex dominance, "2-dom" = duplex dominance). The software tries both dominance patterns for a given dosage model, e.g., whether the reference or alternate allele is dominant
- "diplo-general": All heterozygotes have the same effect
- "diplo-additive": All heterozygotes have the same effect, constrained to be halfway between the homozygous effects
- "general": There are no constraints on the effects of the different dosage levels

To specify additional model parameters, such as the inclusion of fixed effects (Q matrix) and the minimum minor allele frequency, use set.params

#### Value

Variable of class GWASpoly.fitted

GWASpoly.data-class S4 class with genotype and phenotype data

### **Description**

S4 class with genotype and phenotype data

## Slots

```
map data frame with columns Marker, Chrom, Position, Ref, Alt pheno data frame of phenotypes geno matrix (individuals x markers) of allele dosages (0,1,2,...ploidy) fixed data frame of fixed effects ploidy ploidy
```

GWASpoly.fitted-class S4 class with results from genome-wide scan

### Description

S4 class with results from genome-wide scan

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#### **Slots**

map data frame with columns Marker, Chrom, Position, Ref, Alt pheno data frame of phenotypes geno matrix with allele dosages fixed data frame of fixed effects ploidy ploidy

K list of covariance matrices scores -log10(p) results effects estimated marker effects params parameters used for the analysis

GWASpoly.K-class

S4 class with genotypes, phenotypes, and polygenic covariance

#### **Description**

S4 class with genotypes, phenotypes, and polygenic covariance

#### **Slots**

map data frame with columns Marker, Chrom, Position, Ref, Alt pheno data frame of phenotypes geno matrix with allele dosages fixed data frame of fixed effects ploidy ploidy

K list of covariance matrices (one for each chromosome)

GWASpoly.thresh-class S4 class with results from genome-wide scan and detection threshold

## **Description**

S4 class with results from genome-wide scan and detection threshold

#### **Slots**

map data frame with columns Marker, Chrom, Position, Ref, Alt pheno data frame of phenotypes geno matrix with allele dosages fixed data frame of fixed effects ploidy ploidy

K list of covariance matrices scores -log10(p) results effects estimated marker effects params parameters used for the analysis threshold thresholds for significance

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LD.plot

Plot LD vs distance

#### **Description**

Plot LD vs distance

#### Usage

```
LD.plot(data, max.pair = 10000, dof = 8)
```

#### **Arguments**

data variable inheriting from class GWASpoly max.pair maximum number of r2 pairs for the spline

dof degrees of freedom for the spline

#### **Details**

A monotone decreasing, convex spline is fit using R package scam.

#### Value

ggplot2 object

manhattan.plot

Create Manhattan plot

#### **Description**

Create Manhattan plot

# Usage

```
manhattan.plot(data, traits = NULL, models = NULL, chrom = NULL)
```

#### **Arguments**

data Variable of class GWASpoly.fitted

traits Vector of trait names (by default, all traits plotted)
models Vector of model names (by default, all models plotted)

chrom optional, to plot only one chromosome

#### **Details**

Results for the ref and alt versions of the dominance model are combined. If data is the output from set.threshold, then the threshold is displayed as a horizontal dashed line when models contains a single model. Because the threshold varies between models, it is not drawn when multiple models are included. Although the ref and alt versions of each dominance model are slightly different (as seen with qq.plot), they are treated as a single model for the Manhattan plot, and the average threshold is shown.

qq.plot

#### Value

ggplot2 object

qq.plot

Quantile-Quantile (QQ) Plot

#### **Description**

Inspect p-value inflation using a QQ plot

## Usage

```
qq.plot(data, trait, models = NULL)
```

## **Arguments**

data Variable of class GWASpoly.fitted

trait Trait name

models Vector of model names (by default, all models plotted)

#### Value

ggplot2 object

read.GWASpoly

Read in marker and phenotype data

#### **Description**

Read in marker and phenotype data

# Usage

```
read.GWASpoly(ploidy, pheno.file, geno.file, format, n.traits, delim = ",")
```

#### **Arguments**

ploidy Ploidy (e.g., 2 for diploid, 4 for tetraploid)

pheno.file Name of the phenotype file geno.file Name of the genotype file

format Format for the marker data. See details.

n.traits Number of traits

delim Character to indicate the delimiter in the data files (e.g., "," for csv, "\t" for

tab-delimited)

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#### **Details**

The first column of the phenotype file contains the genotype identifier, columns 2 through (n.traits + 1) contain trait values, and subsequent columns contain the levels (for factors) or numeric values (for covariates) of any fixed effects. The first three columns of the genotype file are (1) marker name, (2) chromosome, and (3) position. Optionally, columns 4 and 5 can be REF and ALT, respectively. Subsequent columns contain the marker data for each individual in the population. Marker data can be coded in one of three formats:

- "numeric": markers are coded based on the dosage of the alternate allele, taking on values between 0 and ploidy
- "AB": e.g., AAAB, ABBB for tetraploids
- "ACGT": e.g., AAAT, GGCC for tetraploids

Only bi-allelic markers are allowed. As of version 2.02 of the package, fractional values of dosage are allowed for the "numeric" format, with missing values imputed by the population mean for each marker. The fractional values are only used for the additive genetic model; for the other models, dosages are rounded to the nearest whole number. If the input allele dosages are whole numbers, then missing values are imputed with the population mode (most frequent value) for each marker.

#### Value

Variable of class GWASpoly.data

set.K

Set covariance matrix for polygenic effect

## Description

Set covariance matrix for polygenic effect

#### Usage

```
set.K(data, K = NULL, n.core = 1, LOCO = NULL)
```

#### **Arguments**

data Output from read.GWASpoly

K Optional: user-supplied matrix

n.core Number of cores for parallel computing

LOCO TRUE/FALSE, whether to use leave-one-chromosome-out

#### **Details**

When LOCO = TRUE, K is computed for each chromosome as \$K=MM'\$, where M is the centered genotype matrix (lines x markers), and scaled to have unit diagonal (the overall scaling is not important for GWAS). When LOCO = FALSE, a single K matrix is computed for all markers (this was the original behavior of the function). Alternatively, the user can supply their own positive semidefinite K, with row.names that match the genotype identifiers (this option cannot be used with LOCO).

#### Value

Variable of class GWASpoly.K

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et parameters
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## **Description**

Set parameters

# Usage

```
set.params(
  fixed = NULL,
  fixed.type = NULL,
  n.PC = 0,
  MAF = NULL,
  geno.freq = NULL,
  P3D = TRUE
)
```

# Arguments

fixed	Vector of names of fixed effects
fixed.type	Vector of effect types ("numeric" or "factor"), corresponding to the effects listed in "fixed"
n.PC	Number of principal components to include as covariates
MAF	Minimum minor allele frequency
geno.freq	Maximum genotype frequency (after applying dominance relations)
P3D	TRUE/FALSE whether to use the P3D approximation (variance components not re-estimated for every marker)

## **Details**

The list returned by the function should be passed to GWASpoly function.

## Value

A list with the following components

fixed	Names of fixed effects
fixed.type	Types of fixed effects
n.PC	Number of principal components to include as covariates
min.MAF	Minimum minor allele frequency
max.geno.freq	Maximum genotype frequency (after applying dominance relations)
P3D	TRUE/FALSE whether to use the P3D approximation

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set.threshold Set the significance threshold

## **Description**

Set the significance threshold

#### Usage

```
set.threshold(
  data,
  method = "M.eff",
  level = 0.05,
  n.permute = 1000,
  n.core = 1
)
```

## **Arguments**

data Variable of class GWASpoly.fitted

method One of the following: "M.eff", "Bonferroni", "FDR", "permute"

level Genome-wide false positive or false discovery rate (depending on method).

n.permuteNumber of permutations for method "permute"n.coreNumber of cores to use for multicore processing

#### **Details**

The default method, "M.eff", is a Bonferroni-type correction but using an effective number of markers that accounts for LD between markers (Moskvina and Schmidt, 2008). The FDR method is based on version 1.30.0 of the qvalue package.

# Value

Variable of class GWASpoly. thresh

## References

Moskvina V, Schmidt KM (2008) On multiple-testing correction in genome-wide association studies. Genetic Epidemiology 32:567-573. doi:10.1002/gepi.20331

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VCF2dosage

Prepare GWASpoly dosage input file from VCF file

#### **Description**

Prepare GWASpoly dosage input file from VCF file

#### Usage

```
VCF2dosage(
   VCF.file,
   dosage.file,
   geno.code,
   ploidy,
   samples = NULL,
   min.DP = 1,
   max.missing,
   min.minor = 5
)
```

## Arguments

VCF.file	VCF filename to read in
dosage.file	name of CSV file to output with allele dosage
geno.code	genotype code in the FORMAT field: either "GT" or "DS" (see Details)
ploidy	ploidy
samples	optional vector of sample names, to export subset of the population
min.DP	minimum per sample depth (DP) to export genotype. Default is 1, for no filter-
	ing.
max.missing	threshold for missing data per marker, as a proportion.
min.minor	minimum number of samples with the minor allele. Default is 5.

## **Details**

The "GT" option for geno.code is the posterior maximum genotype (e.g., 0/0/1/1), while "DS" represents the posterior mean dosage of the alternate allele. (At present, only bi-allelic variants are supported.) The VCF file must conform to version 4.1 of the VCF specification or later.

write.GWASpoly

Write results to file

#### **Description**

Write results to file

#### Usage

```
write.GWASpoly(data, trait, filename, what = "scores", delim = ",")
```

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# Arguments

data Variable of class GWASpoly.fitted

trait Trait name filename Filename

what Either "scores" or "effects"

delim Delimiter to use in the output file (default is comma)

# **Details**

 $Score = -log10(p). \ Effect = marker \ effect \ (not \ available \ for \ the \ general \ and \ diplo-general \ models).$ 

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